

Electronic Supplementary Information

Tuning the Interlocking in Partially Saturated Copper(I) Rotaxane Complexes for O₂-to-H₂O Electrocatalytic Reduction

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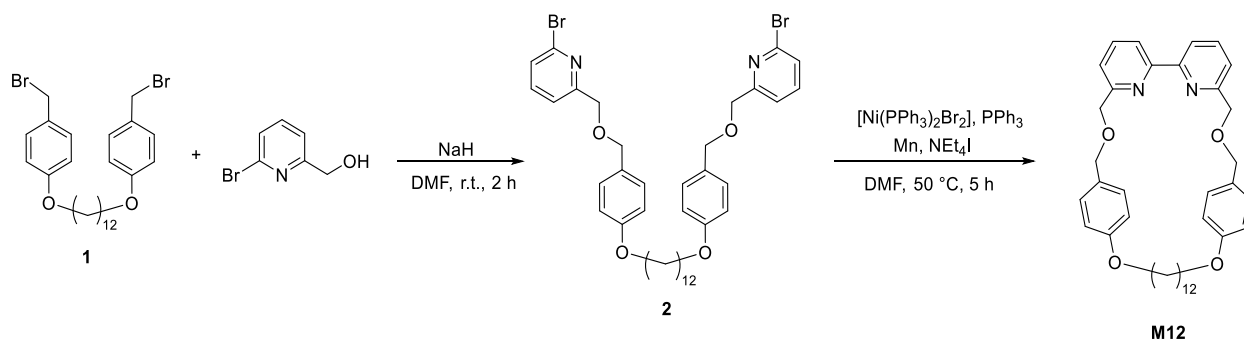
List of Contents

1. Syntheses and Characterisations
2. Electrochemical Measurements
3. Supplementary References

1. Molecular Syntheses and Characterisations.

1.1 Syntheses.

General. All reagents (purity $\geq 98\%$ unless otherwise noted) were purchased from commercial suppliers (J&K, Sigma-Aldrich, TCI, Aladdin, Energy, Macklin, and Dkmchem) and used without further purification unless otherwise noted. All the solvents were of analytical grade (ACI Labscan and DUKSAN Pure Chemicals). Deuterated solvents CDCl_3 and CD_3CN were purchased from Cambridge Isotope Laboratories (CIL) (≥ 99.8 atom% D). Compounds **1**, **3**, **5**, **M4**, **M6**, **M8**, and **M10** were synthesized according to literature procedures.¹⁻³ Thin layer chromatography (TLC) was performed on silica gel 60 F254 (Merck, Germany, Aluminium sheet) and column chromatography was carried out using silica gel F60 (Silicycle, Canada). HR-ESI-MS were obtained from a Bruker Impact II Ultra-High Resolution QTOF mass spectrometer. NMR spectra were recorded on Bruker DPX spectrometers with working frequencies at 400 MHz, 500 MHz, or 600 MHz for ^1H , and 101 MHz, 126 MHz, or 151 MHz for ^{13}C , respectively. Chemical shifts were reported in ppm and referenced to solvent residues (for ^1H : CDCl_3 : $\delta = 7.26$ ppm, CD_3CN : $\delta = 1.94$ ppm; for ^{13}C : CDCl_3 : $\delta = 77.16$ ppm, CD_3CN : $\delta = 1.32$ ppm and 118.26 ppm).

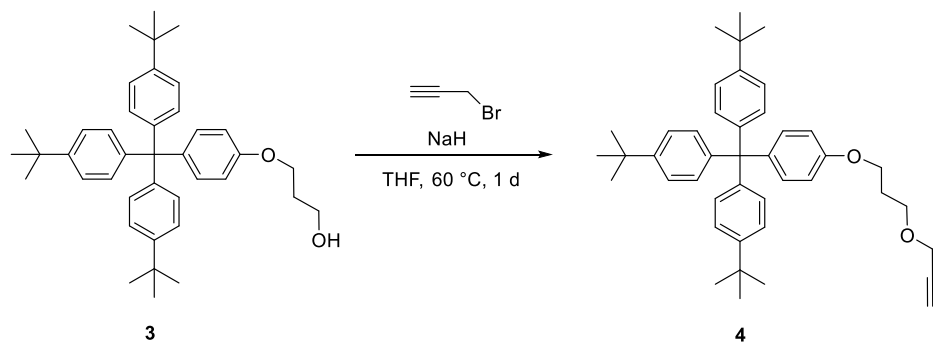


Scheme S1. Synthesis of **M12**.

Synthesis of 2. To a solution of 2-bromo-6-pyridinemethanol (432 mg, 0.8 mmol) in 10 mL of dimethylformamide (DMF) at 0 °C was added 60 wt% NaH (64 mg, 1.6 mmol) in three portions with continuous stirring. After stirring at 0 °C for 30 minutes, compound **1** (358 mg, 0.7 mmol) was added and the mixture was stirred at 0 °C for 1 hour. The reaction mixture was warmed up to room temperature, and water was added (10 mL) dropwise. The solvents were removed using a rotary evaporator, and the residue was redissolved in CH_2Cl_2 (50 mL), washed with water (2×10 mL) and brine (10 mL), dried over Na_2SO_4 and filtered. Solvents were removed from the filtrate using a rotary evaporator to afford a yellow oil, which was purified using a silica column (eluent:

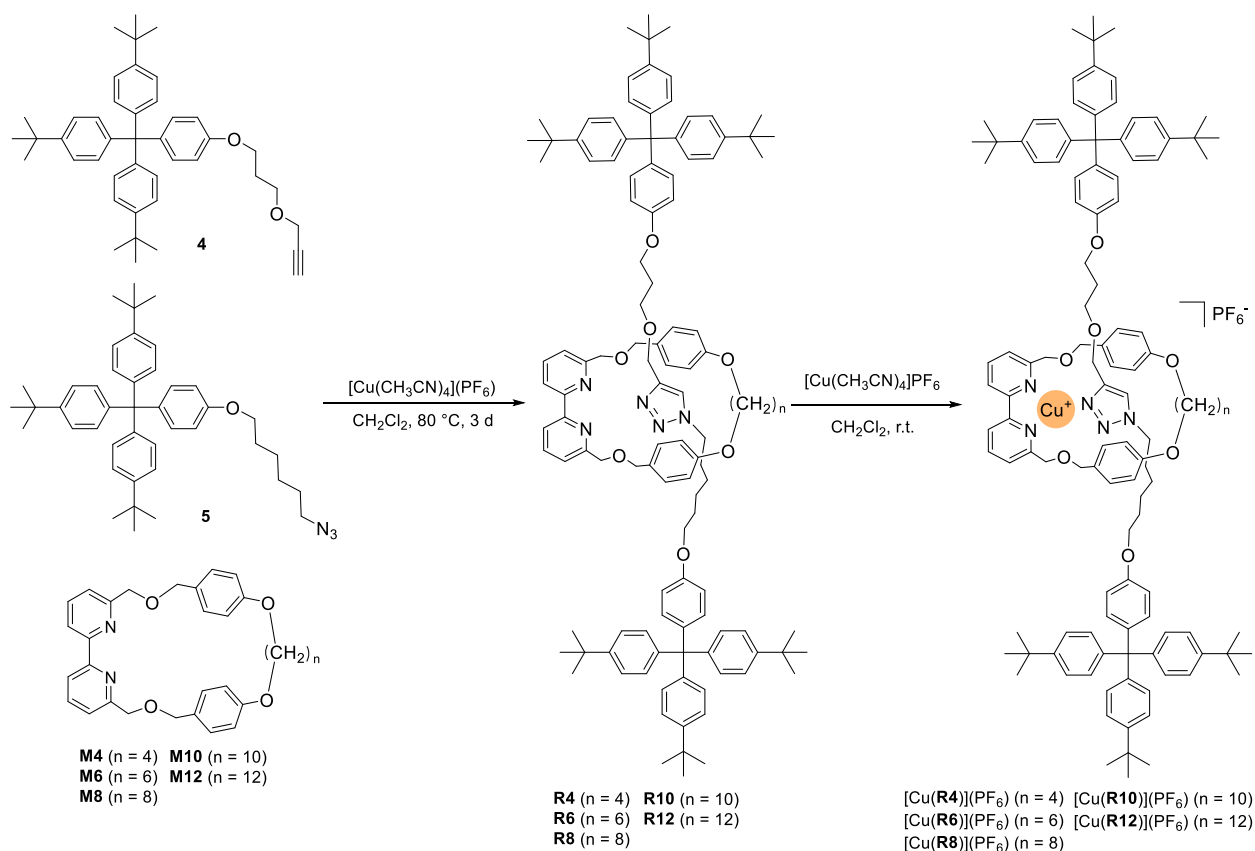
CH₂Cl₂/hexane, v/v = 1:1) to give **2** as a white solid. Yield = 0.49 g, 65%. ¹H NMR (500 MHz, CDCl₃, 298 K) δ 7.55 (t, *J* = 8.0 Hz, 2H), 7.46 (d, *J* = 7.5 Hz, 2H), 7.37 (d, *J* = 8.0 Hz, 2H), 7.28 (d, *J* = 8.5 Hz, 4H), 6.88 (d, *J* = 8.5 Hz, 4H), 4.62 (s, 4H), 4.56 (s, 4H), 3.95 (t, *J* = 7.0 Hz, 4H), 1.77 (p, *J* = 7.0 Hz, 4H), 1.45 (p, *J* = 7.8 Hz, 4H), 1.36–1.23 (m, 12H). ¹³C {¹H} NMR (126 MHz, CDCl₃, 298 K) δ 159.0, 141.4, 139.1, 129.7, 126.7, 120.1, 114.6, 73.0, 72.1, 68.2, 29.7, 29.5, 29.4, 26.2. HRMS (ESI⁺): *m/z* calcd. for C₃₈H₄₇N₂O₄Br₂ [M+H]⁺: 755.1881, found: 755.1879.

Synthesis of M12. Under an argon atmosphere, a mixture of [Ni(PPh₃)₂Br₂] (1.49 g, 2.0 mmol), PPh₃ (1.05 g, 4.0 mmol), manganese powder (1.10 g, 20.0 mmol) and NEt₄I (0.51 g, 2.0 mmol) in DMF (20 mL) was sonicated for 10 minutes, and the mixture was heated at 50 °C for 1 hour. To this mixture was slowly added a solution of compound **2** (1.51 g, 2.0 mmol) in DMF (20 mL) over 4 hours, and the reaction mixture was heated at 50 °C for another 1 hour. The reaction mixture was cooled to room temperature, CH₂Cl₂ (50 mL) and a saturated solution of Na₂EDTA in aqueous ammonia (17.5 wt%, 50 mL) were added, the mixture was then filtered through a Celite pad. The filtrate was washed with water (2 × 50 mL) and brine (50 mL), dried over Na₂SO₄, and the solvents were removed using a rotary evaporator. The crude product was purified by a silica column with a gradient elution with an increasing amount of CH₃CN from 0% to 10% in CH₂Cl₂/hexane (v/v 1:1). Macrocycle **M12** was obtained as a white solid. Yield = 0.27 g, 54%. ¹H NMR (500 MHz, CD₃CN, 298 K) δ 8.14 (d, *J* = 7.5 Hz, 2H), 7.81 (t, *J* = 8.0 Hz, 2H), 7.40 (d, *J* = 7.5 Hz, 2H), 7.22 (d, *J* = 8.5 Hz, 4H), 6.79 (d, *J* = 8.5 Hz, 4H), 4.63 (s, 4H), 4.58 (s, 4H), 3.93 (t, *J* = 6.5 Hz, 4H), 1.65 (p, *J* = 7.0 Hz, 4H), 1.37 (p, *J* = 8.0 Hz, 4H), 1.27–1.19 (m, 12H). ¹³C {¹H} NMR (126 MHz, CD₃CN, 298 K) δ 159.6, 159.3, 156.3, 138.4, 131.4, 130.8, 123.1, 120.4, 115.3, 73.4, 73.0, 68.4, 30.1, 29.9, 29.5, 29.4, 26.3. HRMS (ESI⁺): *m/z* calcd. for C₃₈H₄₇N₂O₄ [M+H]⁺: 595.3530, found: 595.3520.



Scheme S2. Synthesis of **4**.

Synthesis of 4. To a solution of compound **3** (456 mg, 0.8 mmol) in THF (10 mL) at 0 °C was added 60 wt% NaH (64 mg, 1.6 mmol) in three portions with continuous stirring. After stirring at 0 °C for 30 minutes, propargyl bromide (0.1 mL, 0.96 mmol) was added, the solution was stirred at 0 °C for 1 hour. The reaction mixture was warmed to room temperature, and subsequently quenched by adding water (10 mL) dropwise. Solvents were removed using a rotary evaporator and the residue was redissolved in CH₂Cl₂ (50 mL), washed with water (2 × 10 mL) and brine (10 mL). The organic layer was dried over Na₂SO₄, filtered, solvents of the filtrate was removed using a rotary evaporator to afford a yellow oil residue, which was purified using a silica column (eluent: CH₂Cl₂/hexane, v/v = 1:5) to afford **4** as a pale yellow solid. Yield = 0.35 g, 73%. ¹H NMR (500 MHz, CDCl₃, 298 K) δ 7.23 (d, *J* = 8.5 Hz, 6H), 7.09–7.07 (m, 8H), 6.77 (d, *J* = 8.5 Hz, 2H), 4.15 (d, *J* = 2.5 Hz, 2H), 4.04 (t, *J* = 6.5 Hz, 2H), 3.70 (t, *J* = 6.0 Hz, 2H), 2.39 (t, *J* = 2.5 Hz, 1H), 2.06 (p, *J* = 6.0 Hz, 2H), 1.30 (s, 27H). ¹³C {¹H} NMR (126 MHz, CDCl₃, 298 K) δ 156.8, 148.4, 144.3, 139.7, 132.4, 130.8, 124.2, 113.2, 80.0, 74.4, 66.9, 64.7, 63.2, 58.4, 34.4, 31.5, 29.8. HRMS (ESI+): *m/z* calcd. for C₄₃H₅₃O₂ [M+H]⁺: 601.4040, found: 601.4043.



Scheme S3. Active template synthesis of the rotaxane ligands and Cu^I complexes.

General procedures for the synthesis of [2]rotaxane ligands and the Cu^I complexes. Under an argon atmosphere, a mixture of the macrocycle (25 μmol) and [Cu(MeCN)₄](PF₆) (9.3 mg, 25 μmol) in CH₂Cl₂ (1 mL) was stirred at room temperature for 30 minutes to give an orange colour solution, and then a mixture of **3** (45.1 mg, 75 μmol) and **5** (47.2 mg, 75 μmol) in CH₂Cl₂ (1.5 mL) was added and heated at 80 °C for 3 days. The reaction mixture was cooled to room temperature, diluted with CH₂Cl₂ (50 mL), and washed with a saturated solution of Na₂EDTA in aqueous ammonia (17.5 wt%, 50 mL). The organic layer was collected and the aqueous phase was extracted with CH₂Cl₂ (3 × 10 mL). The organic phases were combined and dried over Na₂SO₄, filtered, and solvents from the filtrate were removed using a rotary evaporator. The crude product was purified using a silica column (gradient elution: from v/v = 1:1 CH₂Cl₂/hexane to v/v/v = 2:2:1 CH₂Cl₂/hexane/MeCN) to give the [2]rotaxanes.

To obtain the Cu^I complexes, an equimolar mixture of the rotaxane ligand (10 μmol) and [Cu(MeCN)₄](PF₆) (10 μmol) in CH₂Cl₂ (10 mL) was stirred at room temperature for 1 hour under

an argon atmosphere. The reaction mixture was then filtered through a Celite pad, and solvents in the filtrate were removed using a rotary evaporator to give the Cu^I rotaxane complexes (as hexafluorophosphate salts) as a red solid.

R4: White solid, yield = 31 mg, 74%. ¹H NMR (600 MHz, *v/v* = 1:2 CDCl₃/CD₃CN, 298 K) δ 7.74 (s, 1H), 7.62 (t, *J* = 7.8 Hz, 2H), 7.46 (d, *J* = 7.8, 2H), 7.35 (d, *J* = 7.8 Hz, 2H), 7.32–7.25 (m, 12H), 7.16 (d, *J* = 8.4 Hz, 6H), 7.11 (d, *J* = 9.0 Hz, 6H), 7.04 (d, *J* = 9.0 Hz, 2H), 6.99 (d, *J* = 9.0 Hz, 2H), 6.96 (d, *J* = 8.4 Hz, 4H), 6.63–6.60 (m, 8H), 4.47 (d, *J* = 12.0 Hz, 2H), 4.41 (d, *J* = 11.4 Hz, 2H), 4.39 (s, 2H), 4.24 (d, *J* = 12.6 Hz, 2H), 4.21 (d, *J* = 12.6 Hz, 2H), 4.05–4.00 (m, 2H), 3.97–3.92 (m, 2H), 3.81 (t, *J* = 6.6 Hz, 2H), 3.47 (t, *J* = 6.0 Hz, 2H), 3.40 (t, *J* = 6.6 Hz, 2H), 3.37 (t, *J* = 7.8 Hz, 2H), 1.85–1.79 (m, 6H), 1.27 (s, 27H), 1.26 (s, 27H), 1.02 (p, *J* = 7.2 Hz, 2H), 0.75 (p, *J* = 7.8 Hz, 2H), 0.59 (p, *J* = 7.8 Hz, 2H), 0.44 (p, *J* = 7.8 Hz, 2H). ¹³C {¹H} NMR (151 MHz, *v/v* = 1:2 CDCl₃/CD₃CN, 298 K) δ 159.9, 159.7, 157.8, 157.7, 157.5, 149.4, 145.6, 145.5, 145.0, 140.2, 139.9, 138.2, 132.4, 132.4, 131.3, 131.2, 131.2, 130.7, 130.6, 125.4, 125.4, 124.4, 121.9, 121.9, 115.8, 114.3, 114.2, 73.1, 71.7, 68.1, 67.5, 67.4, 65.7, 65.0, 64.0, 63.9, 50.1, 34.9, 31.5, 30.3, 30.2, 29.8, 29.0, 27.7, 26.2, 25.5, 25.4. IR (cm⁻¹): 2955, 2920, 2859, 1607, 1578, 1506, 1457, 1363, 1248, 1182, 1092, 1018, 821, 704, 626, 579. HRMS (ESI⁺): *m/z* calcd. for C₁₁₆H₁₃₈N₅O₇ [M+H]⁺: 1714.0624, found: 1714.0601.

[Cu(**R4**)](PF₆): Orange solid, yield = 18 mg, 93%. ¹H NMR (600 MHz, CD₃CN, 298 K) δ 7.82 (br, 4H), 7.59 (br, 2H), 7.48 (s, 1H), 7.31 (d, *J* = 6.6 Hz, 6H), 7.28 (d, *J* = 8.4 Hz, 6H), 7.19 (d, *J* = 8.4 Hz, 6H), 7.12 (d, *J* = 9.0 Hz, 6H), 7.07 (d, *J* = 9.0 Hz, 2H), 7.03 (d, *J* = 8.4 Hz, 2H), 6.93 (d, *J* = 7.8 Hz, 4H), 6.77–6.74 (m, 6H), 6.31 (d, *J* = 8.4 Hz, 2H), 4.66 (br, 2H), 4.38 (t, *J* = 7.2 Hz, 2H), 4.25–4.13 (m, 6H), 4.07–4.04 (m, 4H), 3.95 (t, *J* = 6.6 Hz, 2H), 3.25 (br, 2H), 3.02 (br, 2H), 2.77 (br, 2H), 2.03–1.95 (m, 6H), 1.78 (p, *J* = 6.6 Hz, 2H), 1.55 (p, *J* = 7.2 Hz, 2H), 1.40 (p, *J* = 3.0 Hz, 2H), 1.26 (d, *J* = 1.8 Hz, 54H), 1.14–1.11 (m, 2H). ¹³C {¹H} NMR (151 MHz, CD₃CN, 298 K) δ 159.8, 157.8, 157.0, 149.4, 149.4, 145.6, 145.5, 140.6, 140.5, 140.3, 132.6, 132.4, 131.1, 130.7, 125.5, 125.4, 116.0, 114.2, 113.8, 72.8, 71.3, 68.4, 67.8, 64.5, 64.0, 63.9, 34.9, 34.9, 31.5, 30.4, 30.3, 29.7, 27.7, 26.8, 26.0, 25.7. IR (cm⁻¹): 2958, 2928, 2864, 1606, 1578, 1505, 1467, 1393, 1363, 1300, 1245, 1174, 1080, 1016, 839, 819, 703, 624, 583, 556. HRMS (ESI⁺): *m/z* calcd. for C₁₁₆H₁₃₇N₅O₇Cu [M-PF₆]⁺: 1775.9842, found: 1775.9812.

R6: White solid, yield = 31 mg, 71%. ^1H NMR (600 MHz, $\nu/\nu = 1:2$ $\text{CDCl}_3/\text{CD}_3\text{CN}$, 298 K) δ 7.64 (d, $J = 7.8$ Hz, 2H), 7.59 (t, $J = 7.8$ Hz, 2H), 7.55 (s, 1H), 7.37 (d, $J = 7.8$ Hz, 2H), 7.27–7.21 (m, 12H), 7.13 (d, $J = 9.0$ Hz, 6H), 7.09 (d, $J = 8.4$ Hz, 6H), 7.01 (d, $J = 8.4$ Hz, 2H), 6.97–6.93 (m, 6H), 6.55 (d, $J = 9.0$ Hz, 4H), 6.51–6.45 (m, 4H), 4.47 (s, 4H), 4.37 (s, 6H), 3.79 (t, $J = 6.6$ Hz, 4H), 3.74 (t, $J = 6.6$ Hz, 2H), 3.59 (t, $J = 7.8$ Hz, 2H), 3.43 (t, $J = 6.0$ Hz, 2H), 3.38 (t, $J = 6.6$ Hz, 2H), 1.80 (p, $J = 6.6$ Hz, 2H), 1.58 (p, $J = 6.6$ Hz, 4H), 1.31–1.28 (m, 4H), 1.26 (d, $J = 3.0$ Hz, 54H), 1.09 (p, $J = 6.6$ Hz, 2H), 1.04 (p, $J = 7.8$, 2H), 0.71 (p, $J = 7.2$ Hz, 2H), 0.57 (p, $J = 7.8$ Hz, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, $\nu/\nu = 1:2$ $\text{CDCl}_3/\text{CD}_3\text{CN}$, 298 K) δ 159.8, 157.8, 157.7, 149.4, 145.6, 145.4, 140.2, 140.1, 138.3, 132.7, 131.5, 130.8, 125.4, 124.1, 121.8, 121.4, 115.6, 114.3, 114.2, 72.9, 71.9, 68.5, 68.2, 67.8, 65.7, 65.2, 64.1, 64.0, 50.5, 35.1, 32.0, 30.5, 29.8, 29.4, 28.0, 26.6, 26.2, 25.8. IR (cm^{-1}): 2951, 2920, 2854, 1644, 1607 1579, 1507, 1459, 1361, 1244, 1181, 1090, 1016, 821, 786, 703, 624, 579. HRMS (ESI+): m/z calcd. for $\text{C}_{118}\text{H}_{142}\text{N}_5\text{O}_7$ $[\text{M}+\text{H}]^+$: 1742.0937, found: 1742.0916.

[Cu(R6)](PF₆): Orange solid, yield = 18 mg, 92%. ^1H NMR (600 MHz, CD_3CN , 298 K) δ 8.04 (d, $J = 8.4$ Hz, 2H), 7.96 (t, $J = 7.8$ Hz, 2H), 7.68 (d, $J = 7.8$ Hz, 2H), 7.29–7.27 (m, 12H), 7.15–7.13 (m, 12H), 7.10 (d, $J = 9.0$ Hz, 2H), 7.07 (d, $J = 9.0$ Hz, 2H), 7.05 (s, 1H), 6.88 (d, $J = 8.4$ Hz, 4H), 6.77 (d, $J = 9.0$ Hz, 2H), 6.64–6.61 (m, 6H), 4.52 (d, $J = 12.6$ Hz, 2H), 4.37 (d, $J = 12.6$ Hz, 2H), 4.34–4.28 (m, 4H), 4.11 (t, $J = 7.2$ Hz, 2H), 3.95–3.90 (m, 6H), 3.72 (t, $J = 6.0$ Hz, 2H), 3.61 (t, $J = 6.0$ Hz, 2H), 3.19 (t, $J = 6.0$ Hz, 2H), 1.77–1.74 (m, 6H), 1.72–1.69 (m, 2H), 1.63 (t, $J = 6.0$ Hz, 2H), 1.57–1.52 (m, 4H), 1.45 (p, $J = 7.8$ Hz, 2H) 1.26 (d, $J = 2.6$ Hz, 54H), 1.24–1.22 (m, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, CD_3CN , 298 K) δ 159.4, 158.9, 157.8, 157.5, 149.4, 149.4 145.5, 145.2, 140.5, 140.4, 140.2, 132.6, 132.5, 131.1, 130.1, 130.0, 125.4, 124.2, 123.4, 121.4, 115.2, 114.3, 114.2, 72.3, 70.7, 68.4, 68.2, 68.0, 65.0, 64.2, 63.9, 51.5, 34.9, 31.5, 30.4, 30.3, 29.9, 29.8, 29.6, 26.7, 26.3, 26.0. IR (cm^{-1}): 2956, 2927, 2864, 1609, 1576, 1505, 1467, 1394, 1361, 1244, 1173, 1105, 1083, 1016, 839, 821, 791, 705, 624, 583, 556, 528. HRMS (ESI+): m/z calcd. for $\text{C}_{118}\text{H}_{141}\text{N}_5\text{O}_7\text{Cu}$ $[\text{M}-\text{PF}_6]^+$: 1805.0159, found: 1805.0133.

R8: White solid, yield = 35 mg, 79%. ^1H NMR (600 MHz, $\nu/\nu = 1:2$ $\text{CDCl}_3/\text{CD}_3\text{CN}$, 298 K) δ 7.85 (d, $J = 7.8$ Hz, 2H), 7.56 (t, $J = 7.8$ Hz, 2H), 7.42 (s, 1H), 7.35 (d, $J = 7.2$ Hz, 2H), 7.26–7.23 (m, 12H), 7.13–7.07 (m, 12H), 6.98–6.95 (m, 6H), 6.91 (d, $J = 9.0$ Hz, 2H), 6.54 (d, $J = 8.4$ Hz, 4H), 6.41 (d, $J = 9.0$ Hz, 2H), 6.38 (d, $J = 9.0$ Hz, 2H), 4.50 (s, 4H), 4.47 (s, 4H), 4.40 (s, 2H),

3.82 (t, $J = 7.8$ Hz, 2H), 3.74–3.70 (m, 6H), 3.46 (t, $J = 6.6$ Hz, 2H), 3.38 (t, $J = 6.0$ Hz, 2H), 1.80 (p, $J = 6.0$ Hz, 2H), 1.52 (p, $J = 6.6$ Hz, 4H), 1.33–1.30 (m, 2H), 1.27 (d, $J = 3.6$ Hz, 54H), 1.21–1.16 (m, 6H), 1.09 (p, $J = 3.6$ Hz, 4H), 0.88 (m, $J = 7.8$ Hz, 2H), 0.79 (q, $J = 8.4$ Hz, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, $v/v = 1:2$ $\text{CDCl}_3/\text{CD}_3\text{CN}$, 298 K) δ 159.6, 157.8, 157.7, 149.3, 145.7, 145.6, 140.2, 140.1, 138.3, 132.7, 131.5, 130.9, 130.7, 125.4, 123.9, 122.3, 120.8, 115.4, 114.2, 72.9, 72.5, 68.4, 68.2, 67.9, 65.6, 65.2, 64.1, 64.0, 50.7, 35.2, 32.0, 30.8, 30.5, 30.1, 29.8, 29.6, 26.8, 26.0. IR (cm^{-1}): 2955, 2930, 2862, 1607, 1576, 1503, 1460, 1440, 1393, 1360, 1298, 1242, 1181, 1105, 1087, 1017, 817, 786, 703, 622, 579, 528. HRMS (ESI⁺): m/z calcd. for $\text{C}_{120}\text{H}_{146}\text{N}_5\text{O}_7$ $[\text{M}+\text{H}]^+$: 1770.1250, found: 1770.1212.

$[\text{Cu}(\mathbf{R8})](\text{PF}_6)$: Orange solid, yield = 19 mg, 95%. ^1H NMR (600 MHz, CD_3CN , 298 K) δ 8.11 (d, $J = 7.8$ Hz, 2H), 8.00 (t, $J = 7.8$ Hz, 2H), 7.71 (d, $J = 7.8$ Hz, 2H), 7.28–7.25 (m, 12H), 7.14–7.08 (m, 17H), 6.90 (d, $J = 8.4$ Hz, 4H), 6.76 (d, $J = 9.0$ Hz, 2H), 6.71 (d, $J = 9.0$ Hz, 2H), 6.59 (d, $J = 8.4$ Hz, 4H), 4.51–4.37 (m, 8H), 3.97 (t, $J = 7.2$ Hz, 2H), 3.95–3.81 (m, 10H), 3.22 (t, $J = 6.0$ Hz, 2H), 1.75–1.71 (m, 2H), 1.69–1.63 (m, 6H), 1.58 (p, $J = 7.8$ Hz, 2H), 1.50 (q, $J = 7.8, 6.2$ Hz, 4H), 1.40–1.37 (m, 6H), 1.25 (d, $J = 3.0$ Hz, 54H), 1.18–1.14 (m, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, CD_3CN , 298 K) δ 159.5, 158.8, 157.8, 157.6, 151.8, 149.4, 145.8, 145.5, 140.5, 140.4, 140.0, 132.6, 131.2, 130.2, 129.8, 125.4, 124.1, 123.9, 121.4, 115.0, 114.3, 72.2, 70.8, 68.3, 68.3, 67.8, 65.3, 64.0, 63.9, 51.4, 34.9, 31.5, 30.3, 30.1, 29.7, 29.7, 29.5, 26.7, 26.7, 26.0. IR (cm^{-1}): 2955, 2902, 2864, 1607, 1576, 1503, 1465, 1393, 1363, 1244, 1174, 1105, 1082, 1016, 839, 819, 703, 622, 583, 555, 525. HRMS (ESI⁺): m/z calcd. for $\text{C}_{120}\text{H}_{145}\text{N}_5\text{O}_7\text{Cu}$ $[\text{M}-\text{PF}_6]^+$: 1833.0472, found: 1833.0428.

R10: White solid, yield = 38 mg, 85%. ^1H NMR (600 MHz, $v/v = 1:2$ $\text{CDCl}_3/\text{CD}_3\text{CN}$, 298 K) δ 7.98 (d, $J = 7.8$ Hz, 2H), 7.55 (t, $J = 7.8$ Hz, 2H), 7.39 (s, 1H), 7.30 (d, $J = 7.2$ Hz, 2H), 7.26–7.23 (m, 12H), 7.11–7.07 (m, 12H), 6.98–6.93 (m, 8H), 6.52 (d, $J = 8.4$ Hz, 4H), 6.46 (d, $J = 9.0$ Hz, 2H), 6.39 (d, $J = 8.4$ Hz, 2H), 4.51 (d, $J = 4.8$ Hz, 8H), 4.41 (s, 2H), 3.94 (t, $J = 7.8$ Hz, 2H), 3.74–3.71 (m, 6H), 3.47 (t, $J = 6.0$ Hz, 2H), 3.41 (t, $J = 6.0$ Hz, 2H), 1.81 (p, $J = 6.0$ Hz, 2H), 1.54 (p, $J = 6.6$ Hz, 4H), 1.45 (p, $J = 7.8$ Hz, 2H), 1.27 (d, $J = 4.2$ Hz, 54H), 1.25–1.20 (m, 6H), 1.13–1.07 (m, 8H), 0.99 (p, $J = 7.8$ Hz, 2H), 0.94–0.90 (m, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, $v/v = 1:2$ $\text{CDCl}_3/\text{CD}_3\text{CN}$, 298 K) δ 159.6, 159.4, 157.8, 157.7, 156.4, 149.4, 145.7, 140.3, 140.2, 138.4, 132.7, 131.5, 130.8, 125.5, 123.9, 122.8, 120.7, 115.3, 114.2, 73.1, 73.1, 68.6, 68.3, 68.0, 65.7,

65.2, 64.1, 50.8, 35.2, 32.1, 30.9, 30.5, 30.5, 29.9, 29.8, 29.7, 26.9, 26.8, 26.2. IR (cm⁻¹): 2953, 2930, 2863, 1609, 1578, 1506, 1461, 1439, 1393, 1363, 1298, 1244, 1182, 1108, 1077, 1018, 821, 788, 706, 626, 581, 530. HRMS (ESI⁺): *m/z* calcd. for C₁₂₂H₁₅₀N₅O₇ [M+H]⁺: 1798.1563, found: 1798.1538.

[Cu(**R10**)](PF₆): Orange solid, yield = 19 mg, 93%. ¹H NMR (600 MHz, CD₃CN, 298 K) δ 8.12 (d, *J* = 7.8 Hz, 2H), 8.01 (t, *J* = 7.8 Hz, 2H), 7.71 (d, *J* = 7.8 Hz, 2H), 7.37 (s, 1H), 7.30–7.24 (m, 12H), 7.16–7.07 (m, 16H), 6.91 (d, *J* = 8.4 Hz, 4H), 6.75 (d, *J* = 9.0 Hz, 2H), 6.68 (d, *J* = 9.0 Hz, 2H), 6.62 (d, *J* = 9.0 Hz, 4H), 4.51–4.42 (m, 8H), 3.97–3.91 (m, 4H), 3.89–3.81 (m, 6H), 3.79 (t, *J* = 6.0 Hz, 2H), 3.19 (t, *J* = 6.0 Hz, 2H), 1.68 (p, *J* = 7.2, 6.6 Hz, 6H), 1.61–1.51 (m, 4H), 1.45 (p, *J* = 7.2 Hz, 4H), 1.36–1.31 (m, 10H), 1.26 (d, *J* = 1.8 Hz, 54H), 1.09 (q, *J* = 7.8 Hz, 2H). ¹³C {¹H} NMR (151 MHz, CD₃CN, 298 K) δ 159.6, 158.8, 157.8, 157.6, 151.8, 149.4, 145.7, 145.5, 145.5, 140.5, 140.4, 140.0, 132.6, 132.6, 131.2, 130.2, 130.0, 125.4, 124.6, 124.2, 121.5, 114.9, 114.3, 72.4, 71.3, 68.5, 68.3, 67.8, 65.4, 63.9, 63.8, 51.3, 34.9, 31.5, 30.3, 30.3, 30.0, 29.7, 29.6, 29.5, 29.4, 26.6, 26.6, 25.9. IR (cm⁻¹): 2956, 2920, 2862, 1647, 1606, 1579, 1497, 1466, 1391, 1362, 1300, 1245, 1175, 1103, 1083, 1019, 840, 822, 703, 625, 579, 554, 528. HRMS (ESI⁺): *m/z* calcd. for C₁₂₂H₁₄₉N₅O₇Cu [M-PF₆]⁺: 1860.0781, found: 1860.0753.

R12: White solid, yield = 37 mg, 81%. ¹H NMR (600 MHz, *v/v* = 1:2 CDCl₃/CD₃CN, 298 K) δ 8.06 (d, *J* = 7.8 Hz, 2H), 7.60 (t, *J* = 7.8 Hz, 2H), 7.39 (s, 1H), 7.33 (d, *J* = 7.8 Hz, 2H), 7.25–7.22 (m, 12H), 7.12–7.08 (m, 12H), 7.03 (d, *J* = 8.4 Hz, 4H), 6.99–6.96 (m, 4H), 6.58 (d, *J* = 8.4 Hz, 4H), 6.50 (d, *J* = 9.0 Hz, 2H), 6.44 (d, *J* = 8.4 Hz, 2H), 4.53 (s, 4H), 4.50 (s, 4H), 4.39 (s, 2H), 3.97 (t, *J* = 7.8 Hz, 2H), 3.78–3.73 (m, 6H), 3.49–3.46 (m, 4H), 1.81 (p, *J* = 6.0 Hz, 2H), 1.57 (p, *J* = 6.6 Hz, 4H), 1.49 (p, *J* = 7.8 Hz, 2H), 1.33–1.31 (m, 2H), 1.27–1.26 (m, 58H), 1.16–1.14 (m, 12H), 1.07–1.05 (m, 2H), 0.99–0.96 (m, 2H). ¹³C NMR (151 MHz, *v/v* = 1:2 CDCl₃/CD₃CN, 298 K) δ 159.6, 159.3, 157.8, 157.7, 149.4, 145.6, 145.6, 140.4, 140.2, 132.7, 132.7, 131.5, 131.5, 131.0, 130.9, 130.8, 125.4, 123.8, 123.2, 120.8, 115.4, 114.2, 114.2, 73.0, 72.8, 68.5, 68.3, 67.9, 65.6, 65.1, 64.1, 64.1, 50.8, 35.2, 32.0, 30.9, 30.5, 30.3, 29.8, 29.7, 28.0, 26.9, 26.7, 26.2. IR (cm⁻¹): 2957, 2928, 2859, 1609, 1578, 1506, 1471, 1438, 1391, 1361, 1297, 1244, 1174, 1108, 1077, 1018, 821, 788, 704, 624, 579. HRMS (ESI⁺): *m/z* calcd. for C₁₂₄H₁₅₄N₅O₇ [M+H]⁺: 1826.1876, found: 1826.1859.

[Cu(**R12**)](PF₆): Orange solid, yield = 19 mg, 92%. ¹H NMR (600 MHz, CD₃CN, 298 K) δ 8.12 (d, *J* = 8.0 Hz, 2H), 7.99 (t, *J* = 7.9 Hz, 2H), 7.70 (d, *J* = 7.7 Hz, 2H), 7.52 (s, 1H), 7.27 (dd, *J* = 8.7, 1.9 Hz, 12H), 7.15–7.08 (m, 16H), 6.93 (d, *J* = 8.6 Hz, 4H), 6.72 (d, *J* = 8.9 Hz, 2H), 6.64 (dd, *J* = 8.8, 2.0 Hz, 6H), 4.56–4.47 (m, 4H), 4.43 (s, 4H), 3.97 (d, *J* = 6.7 Hz, 4H), 3.86–3.80 (m, 6H), 3.76 (t, *J* = 6.3 Hz, 2H), 3.19 (t, *J* = 6.1 Hz, 2H), 1.67 (q, *J* = 6.5 Hz, 6H), 1.56 (td, *J* = 7.7, 3.0 Hz, 4H), 1.45–1.40 (m, 4H), 1.31 (d, *J* = 7.4 Hz, 14H), 1.26 (d, *J* = 1.1 Hz, 54H), 1.12–1.08 (m, 2H). ¹³C{¹H} NMR (151 MHz, CD₃CN, 298 K) δ 159.6, 158.8, 149.4, 145.5, 145.5, 139.9, 132.6, 132.6, 131.2, 130.2, 125.4, 115.0, 114.2, 114.2, 72.6, 71.7, 68.6, 68.3, 67.8, 65.4, 63.9, 51.3, 34.9, 31.5, 30.3, 30.0, 29.8, 29.7, 29.6, 29.5, 29.4, 26.6, 26.5, 25.9. IR (cm⁻¹): 2953, 2924, 2859, 1609, 1578, 1506, 1465, 1393, 1361, 1300, 1246, 1176, 1103, 1081, 1018, 839, 823, 704, 624, 579, 556, 525. HRMS (ESI⁺): *m/z* calcd. for C₁₂₄H₁₅₃N₅O₇Cu [M-PF₆]⁺: 1889.1100, found: 1889.1059.

1.2 NMR Characterisations.

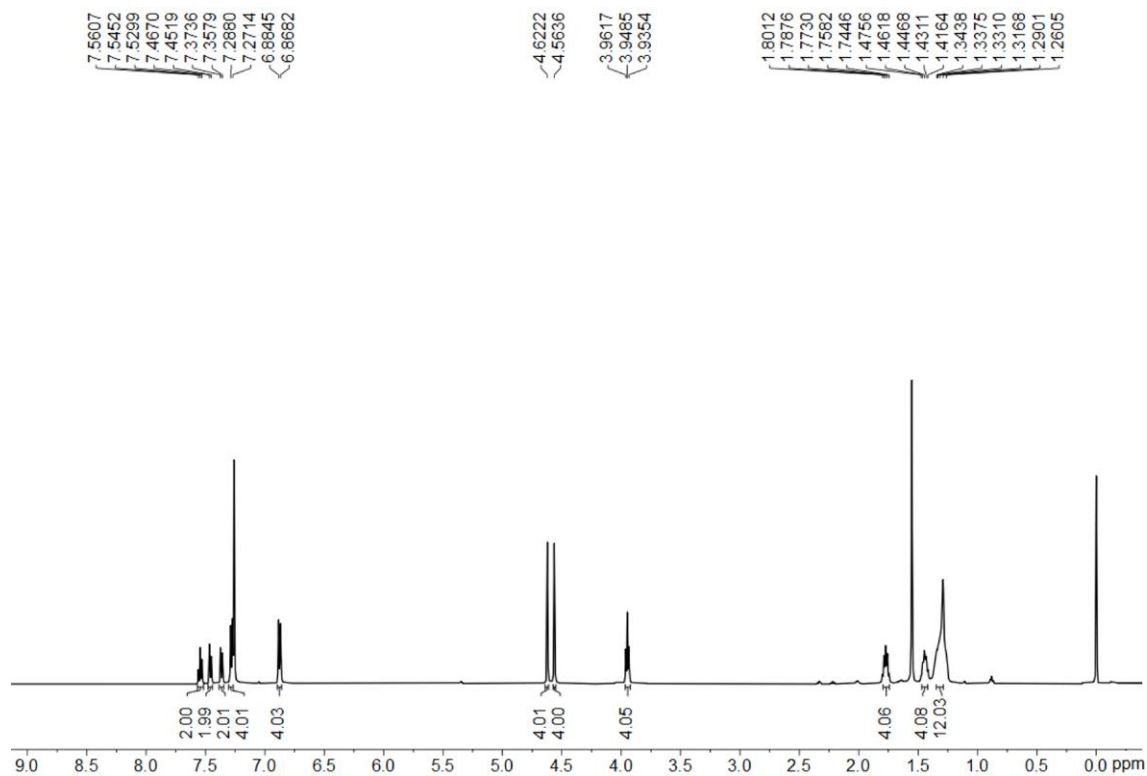


Fig. S1. ^1H NMR (500 MHz, CD_3CN , 298 K) spectrum of **2**.

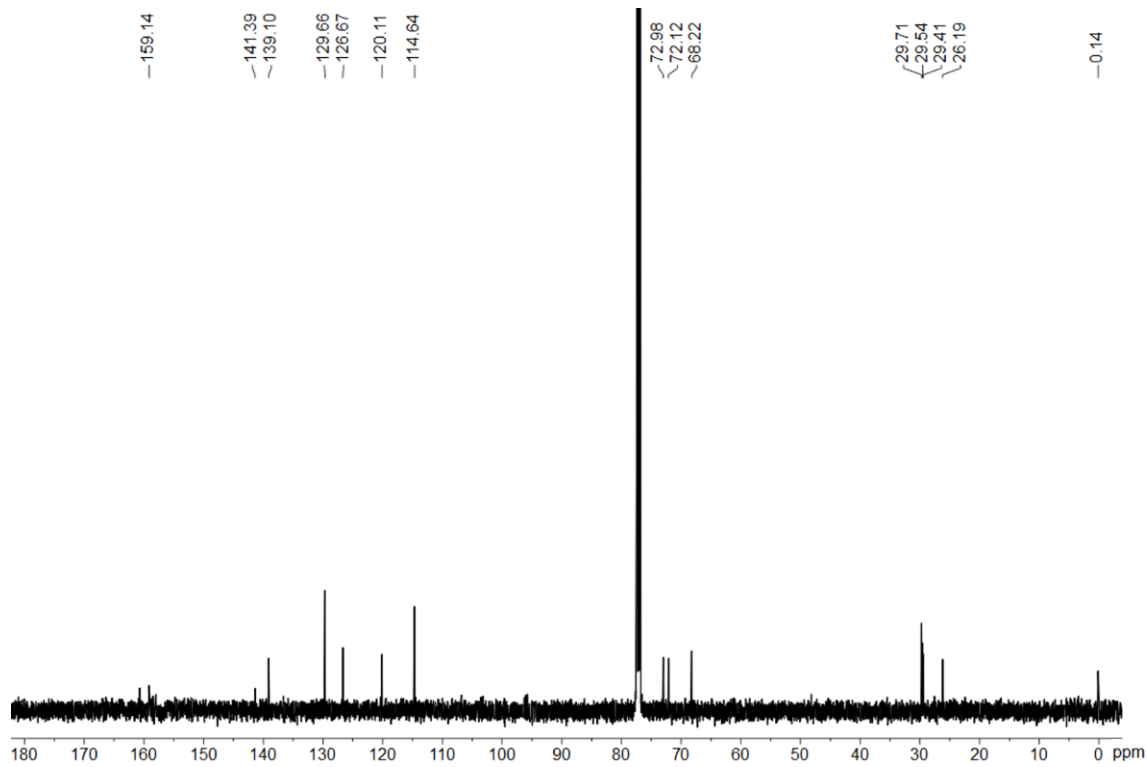


Fig. S2. $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CD_3CN , 298 K) spectrum of **2**.

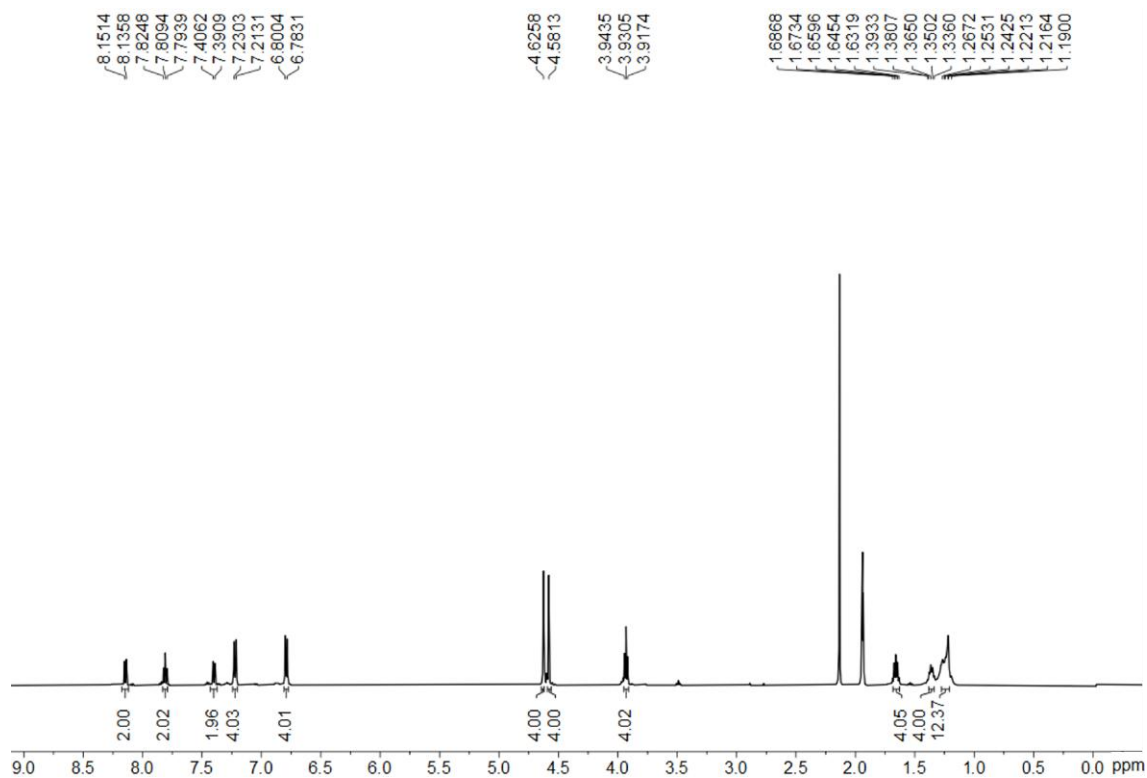


Fig. S3. ^1H NMR (500 MHz, CD_3CN , 298 K) spectrum of **M12**.

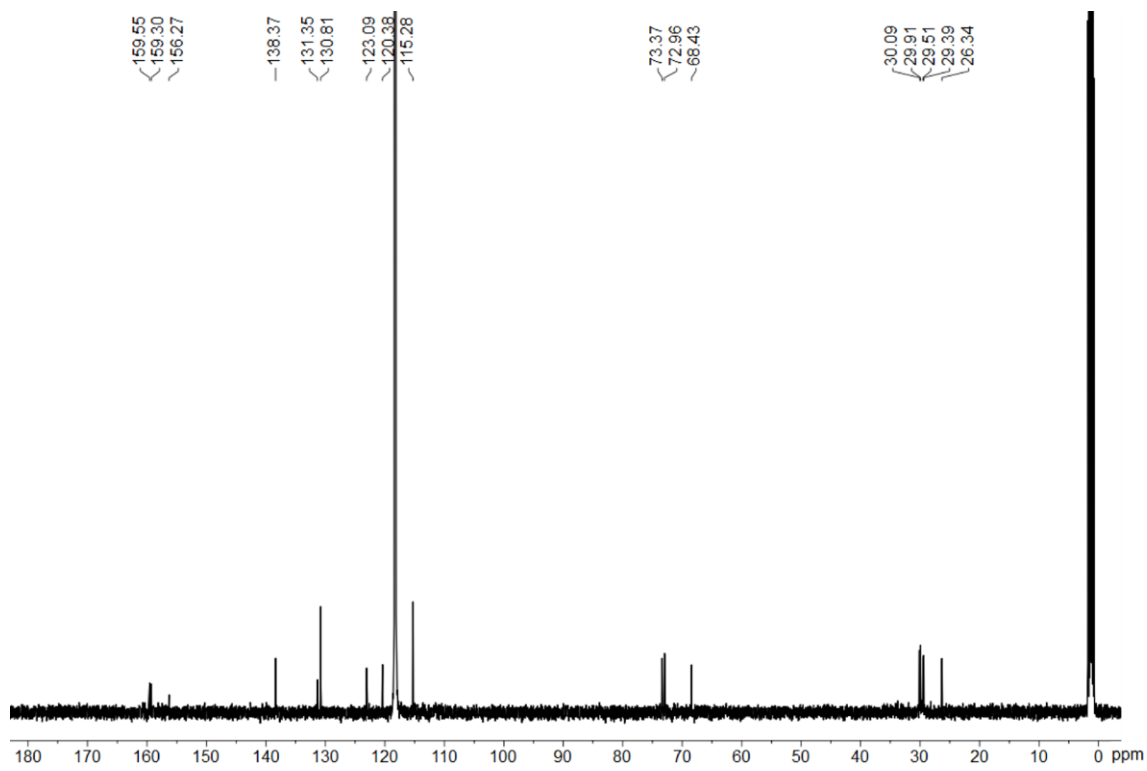


Fig. S4. $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CD_3CN , 298 K) spectrum of **M12**.

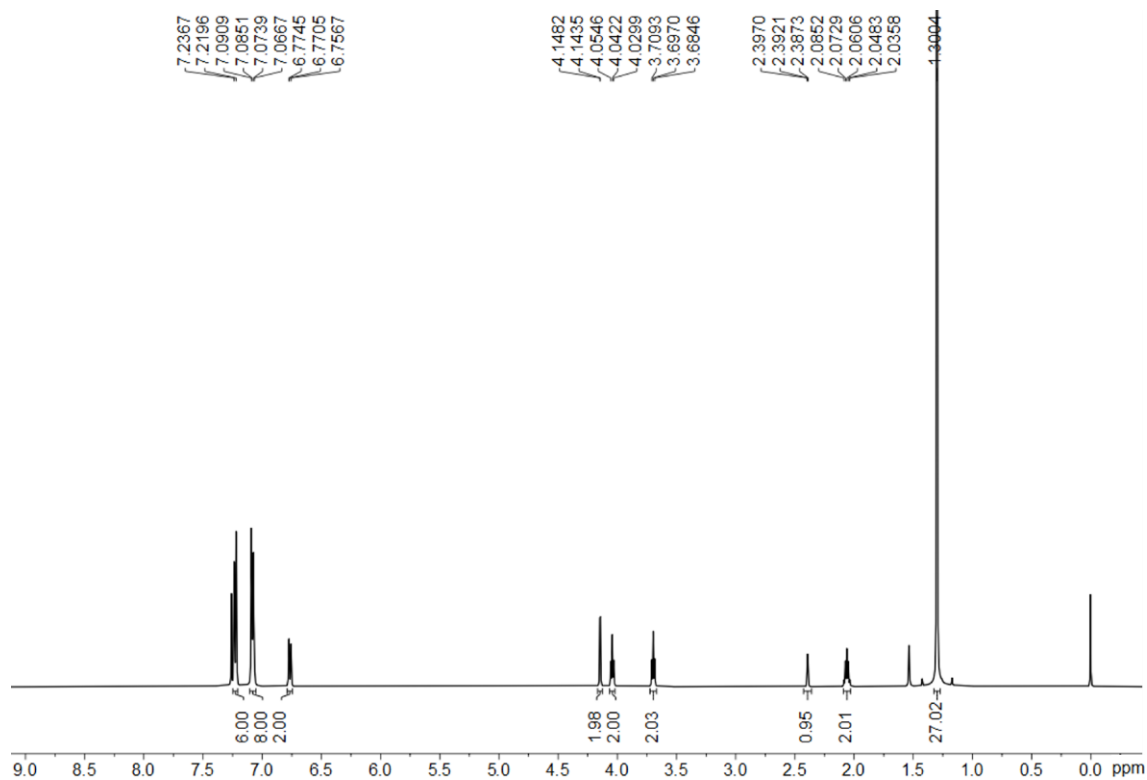


Fig. S5. ^1H NMR (500 MHz, CDCl_3 , 298 K) spectrum of **4**.

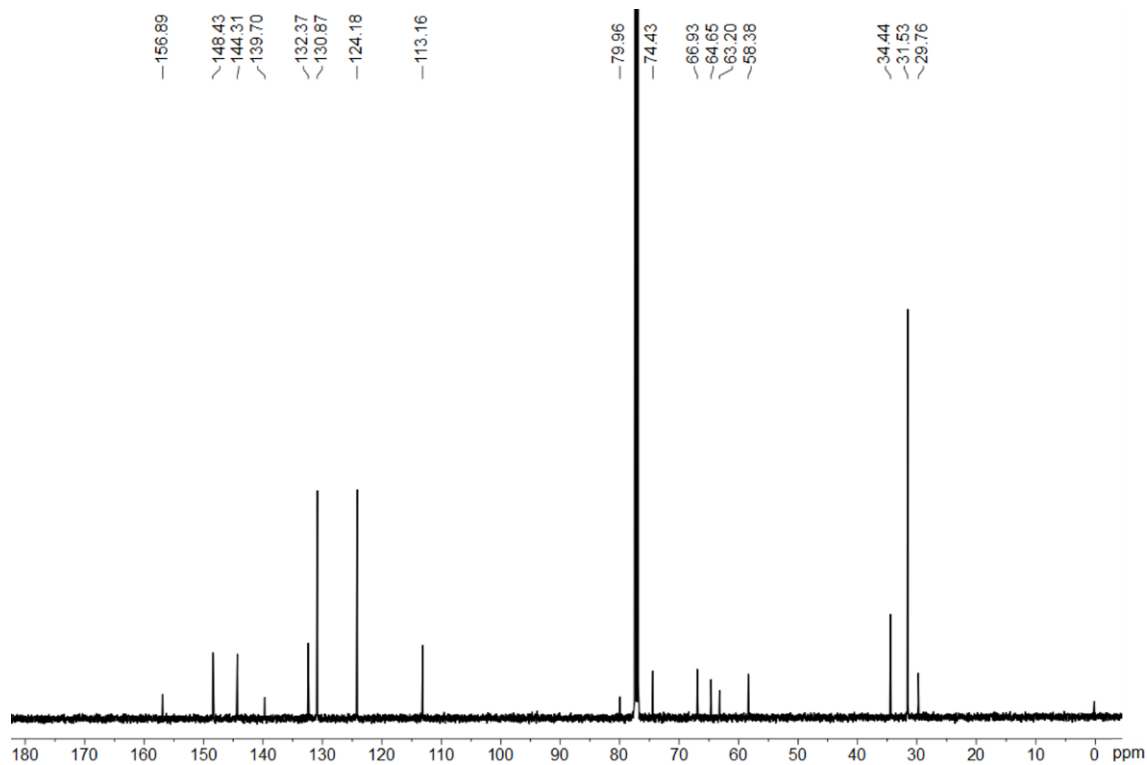


Fig. S6. $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3 , 298 K) spectrum of **4**.

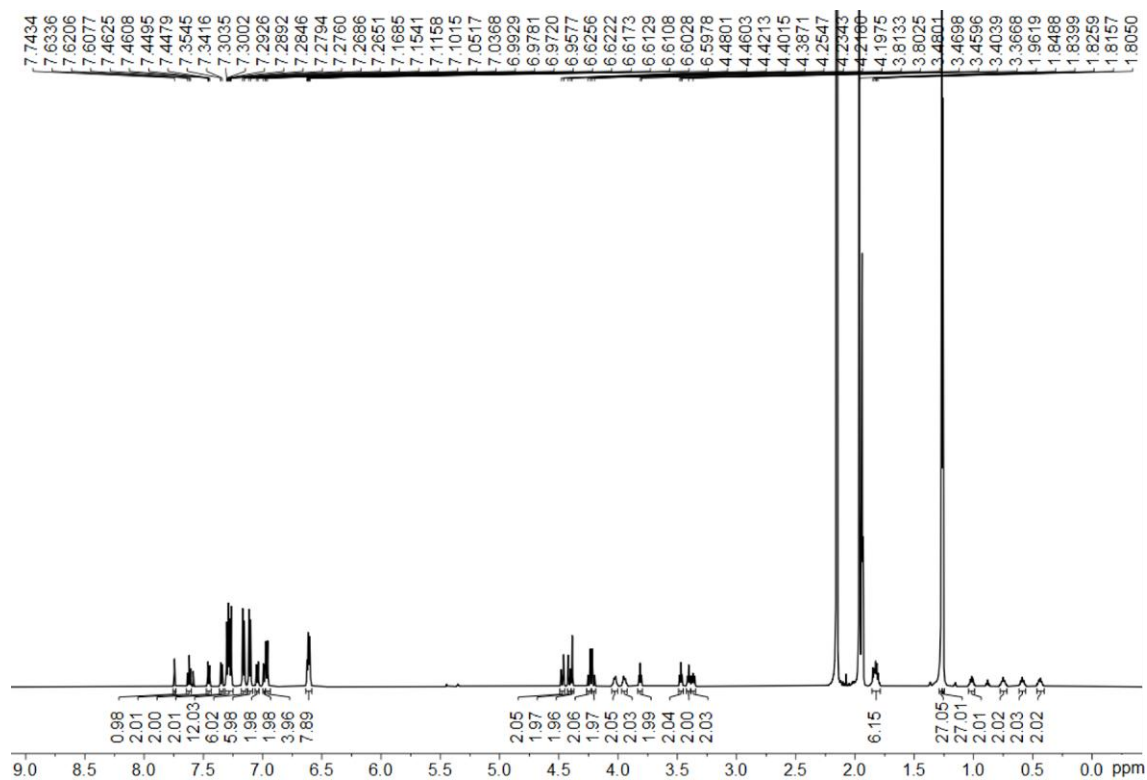


Fig. S7. ^1H NMR (600 MHz, $v/v = 1:2$ $\text{CDCl}_3/\text{CD}_3\text{CN}$, 298 K) spectrum of **R4**.

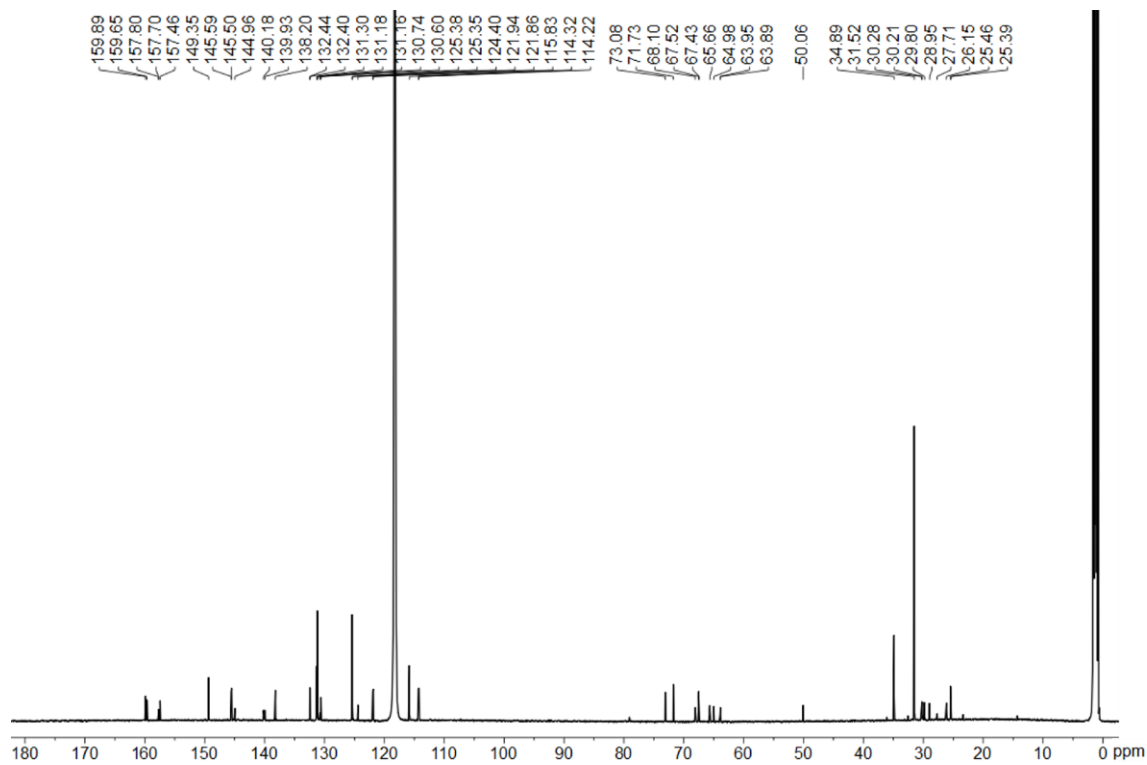


Fig. S8. $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, $v/v = 1:2$ $\text{CDCl}_3/\text{CD}_3\text{CN}$, 298 K) spectrum of **R4**.

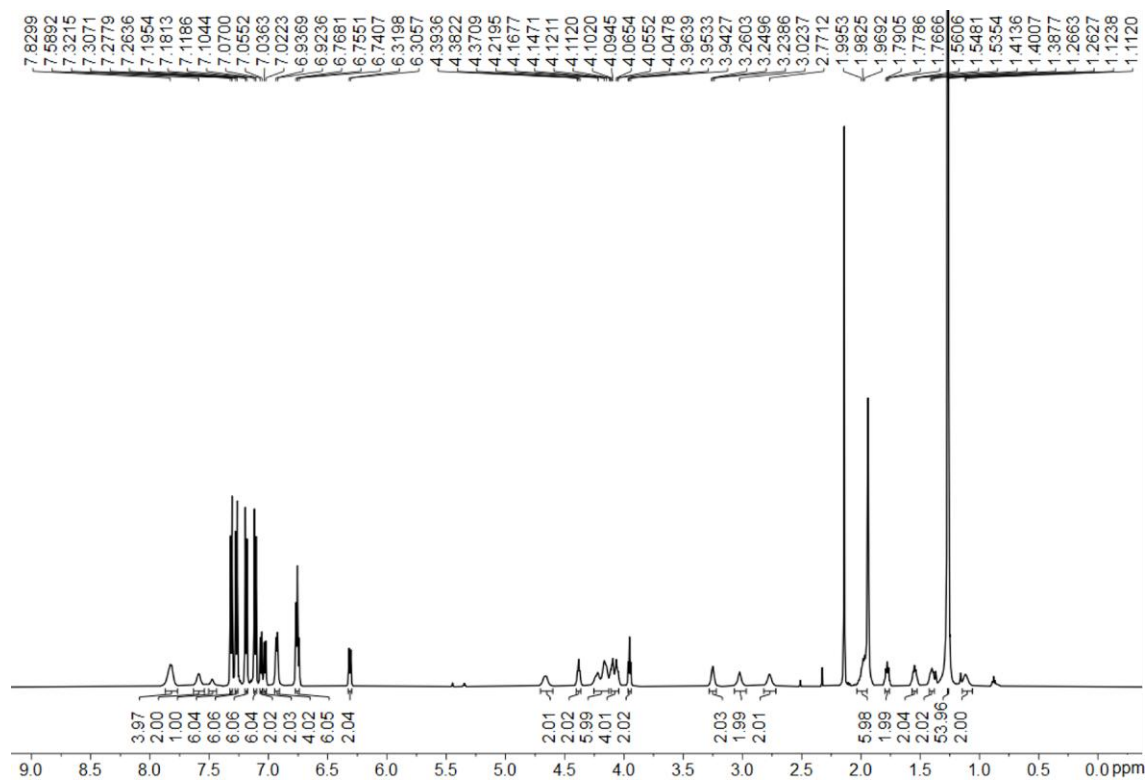


Fig. S9. ^1H NMR (600 MHz, CD_3CN , 298 K) spectrum of $[\text{Cu}(\mathbf{R4})](\text{PF}_6)$.

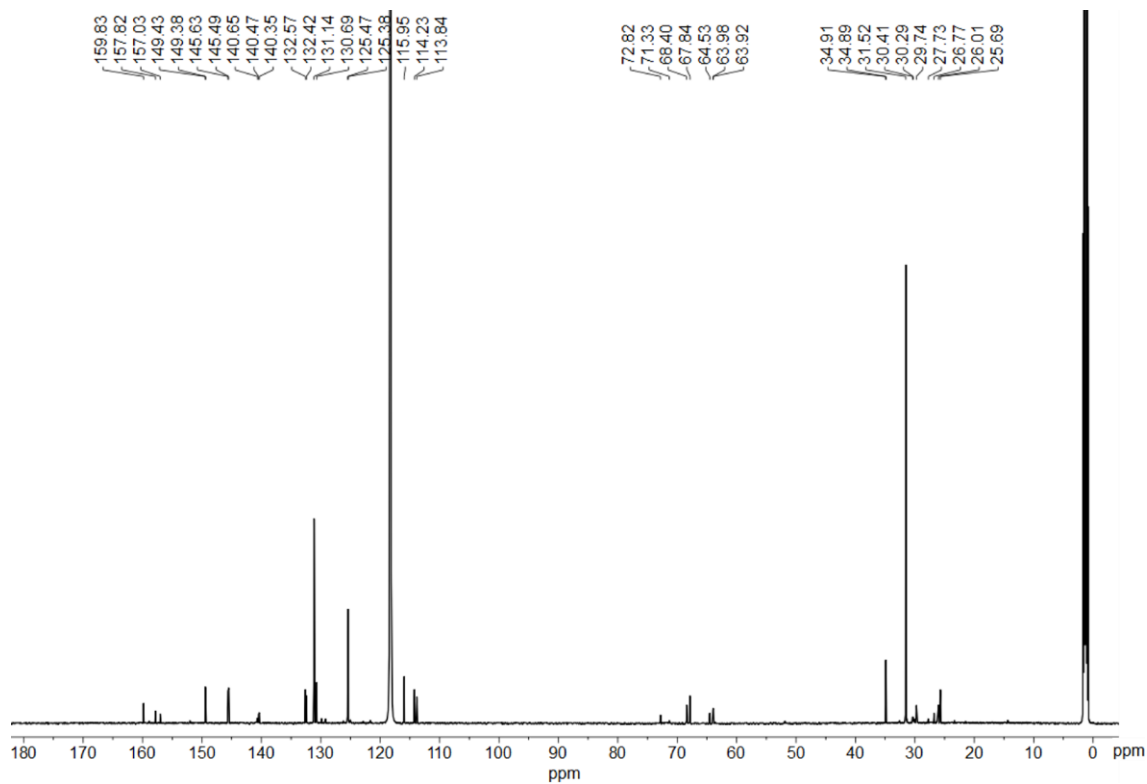


Fig. S10. $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, CD_3CN , 298 K) spectrum of $[\text{Cu}(\mathbf{R4})](\text{PF}_6)$.

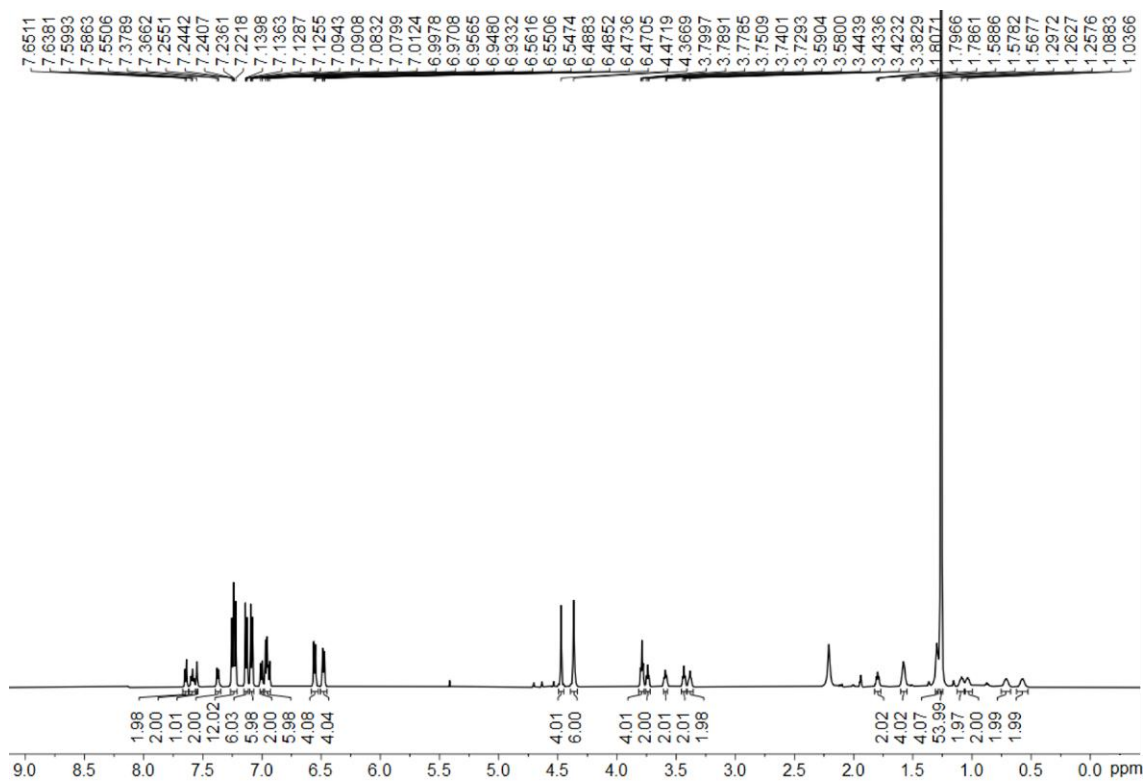


Fig. S11. ^1H NMR (600 MHz, $v/v = 1:2$ $\text{CDCl}_3/\text{CD}_3\text{CN}$, 298 K) spectrum of **R6**.

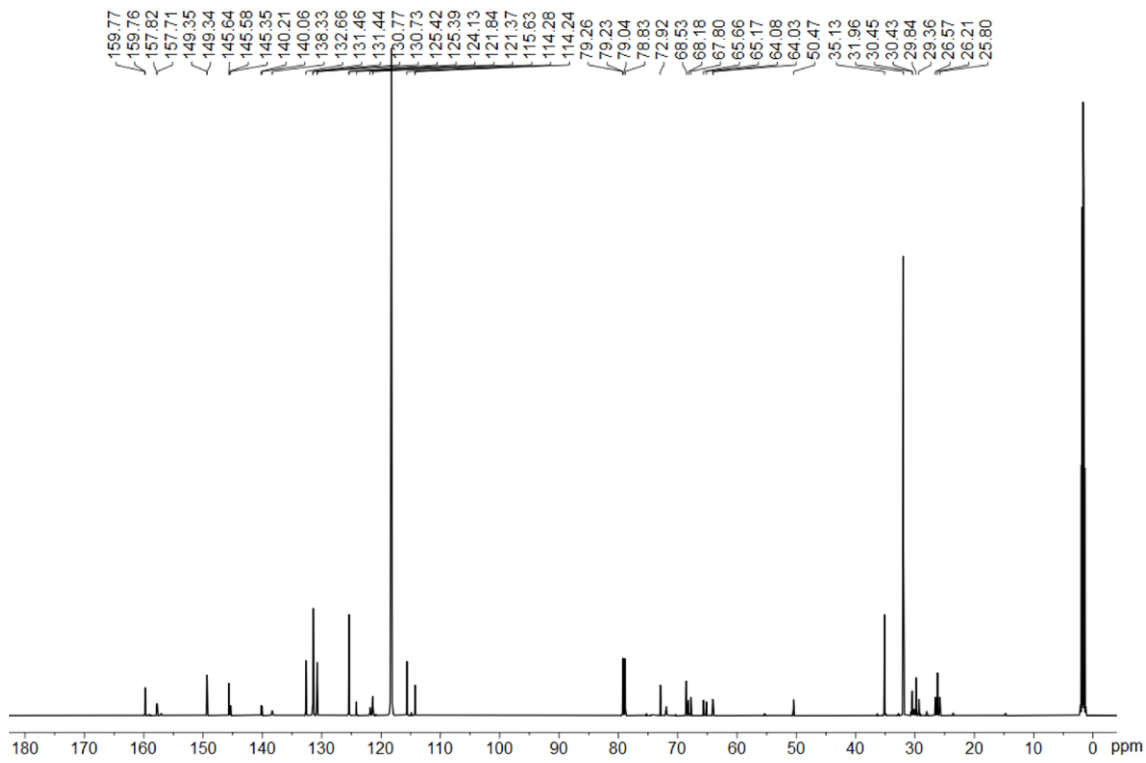


Fig. S12. $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, $v/v = 1:2$ $\text{CDCl}_3/\text{CD}_3\text{CN}$, 298 K) spectrum of **R6**.

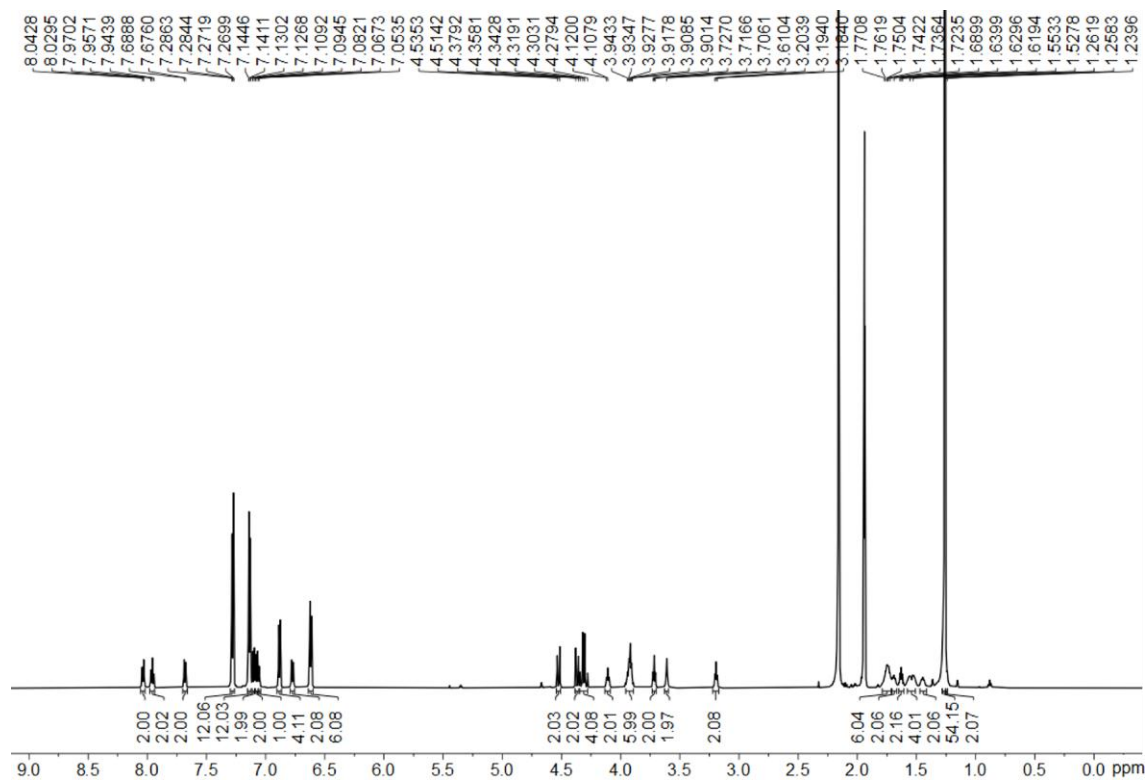


Fig. S13. ^1H NMR (600 MHz, CD_3CN , 298 K) spectrum of $[\text{Cu}(\text{R6})](\text{PF}_6)$.

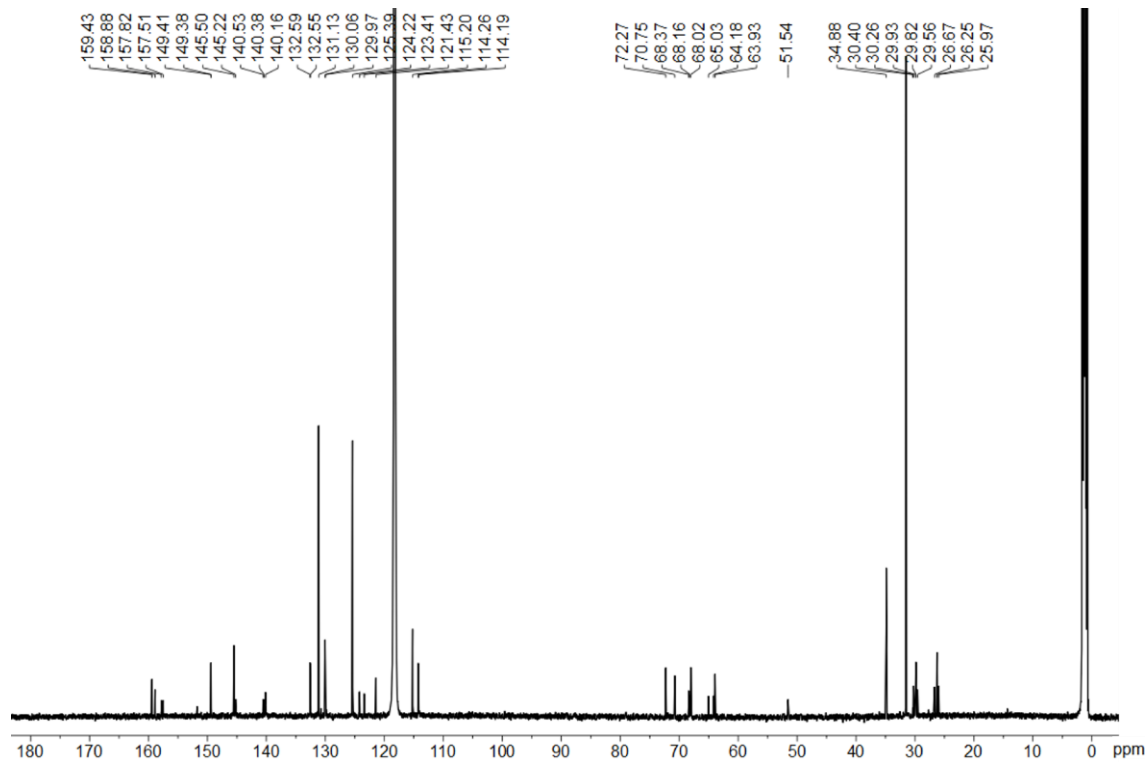


Fig. S14. $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, CD_3CN , 298 K) spectrum of $[\text{Cu}(\text{R6})](\text{PF}_6)$.

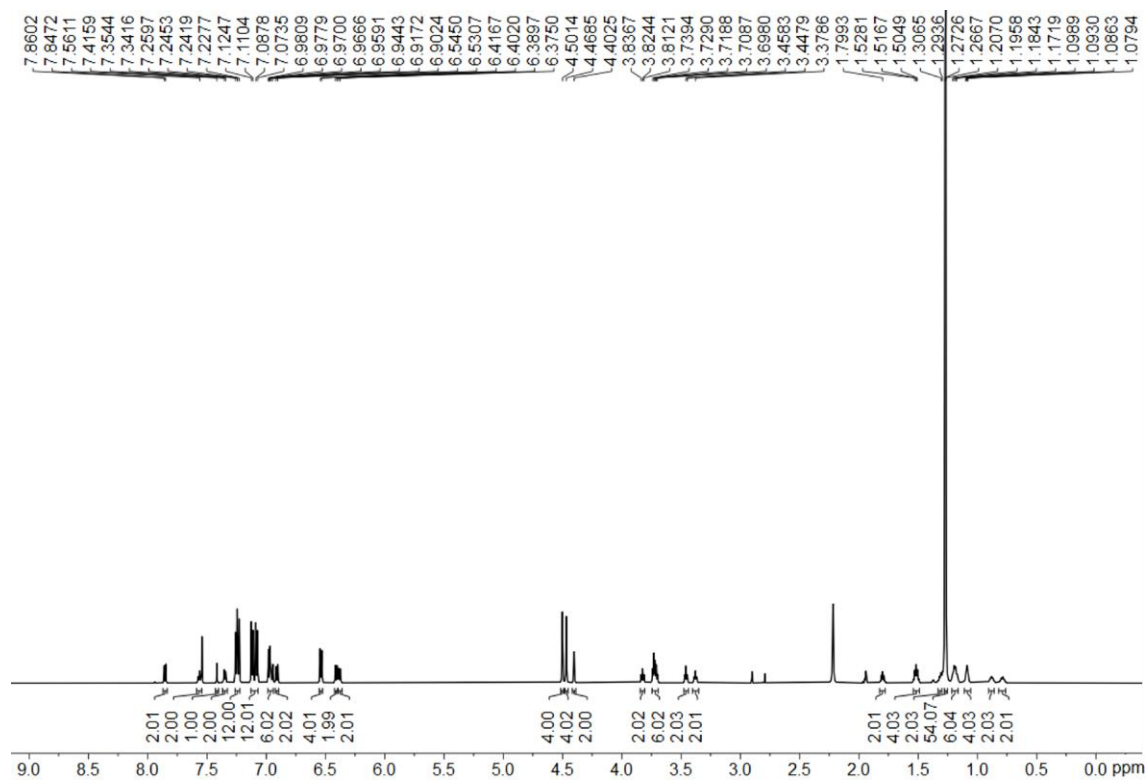


Fig. S15. ^1H NMR (600 MHz, $v/v = 1:2$ $\text{CDCl}_3/\text{CD}_3\text{CN}$, 298 K) spectrum of **R8**.

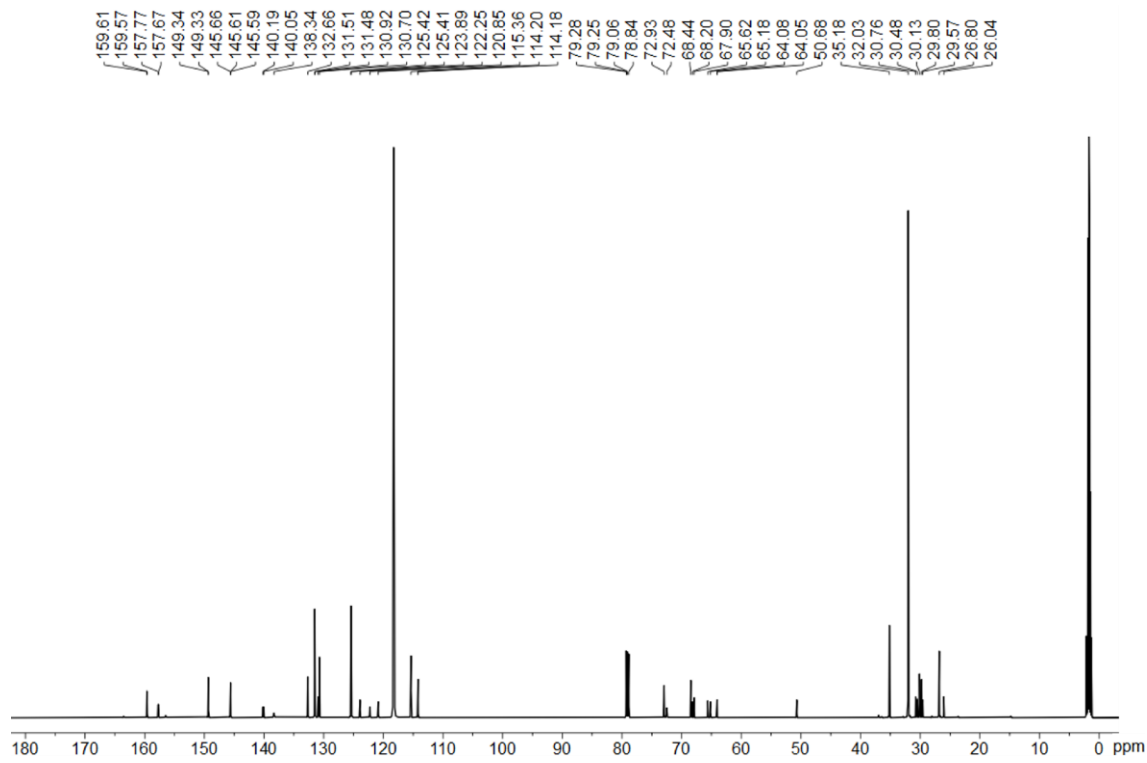


Fig. S16. $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, $v/v = 1:2$ $\text{CDCl}_3/\text{CD}_3\text{CN}$, 298 K) spectrum of **R8**.

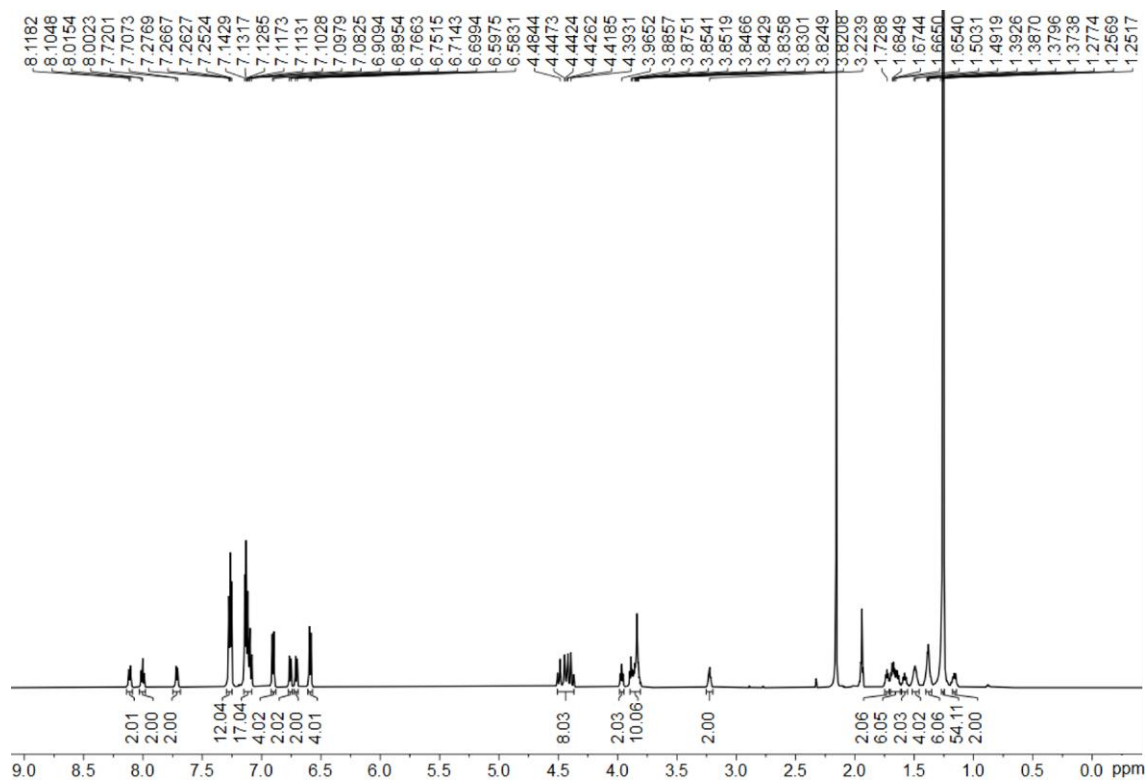


Fig. S17. ^1H NMR (600 MHz, CD_3CN , 298 K) spectrum of $[\text{Cu}(\mathbf{R8})](\text{PF}_6)$.

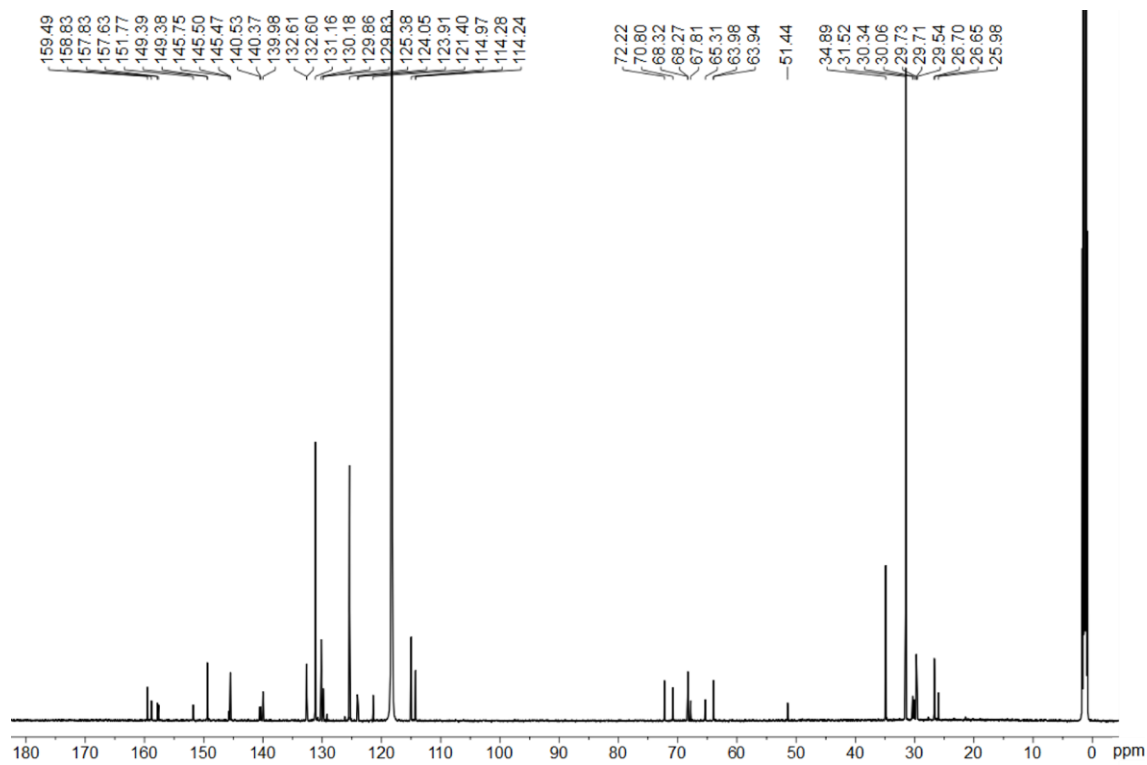


Fig. S18. $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, CD_3CN , 298 K) spectrum of $[\text{Cu}(\mathbf{R8})](\text{PF}_6)$.

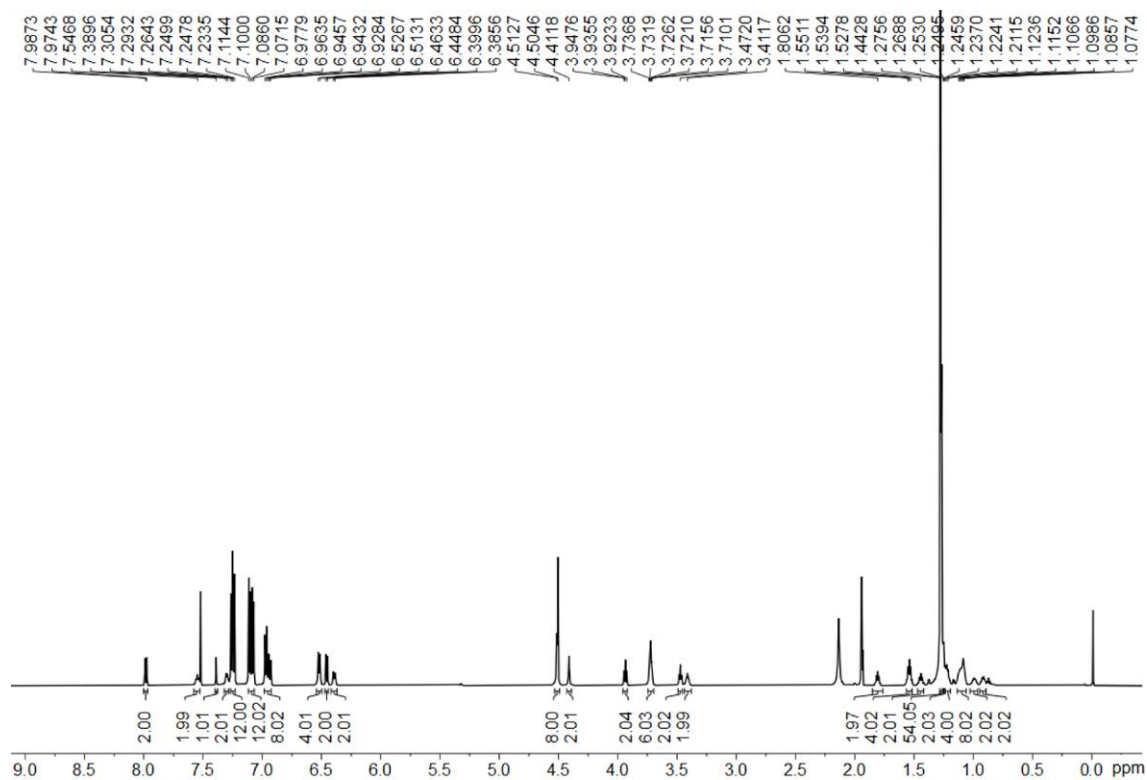


Fig. S19. ^1H NMR (600 MHz, $v/v = 1:2$ $\text{CDCl}_3/\text{CD}_3\text{CN}$, 298 K) spectrum of **R10**.

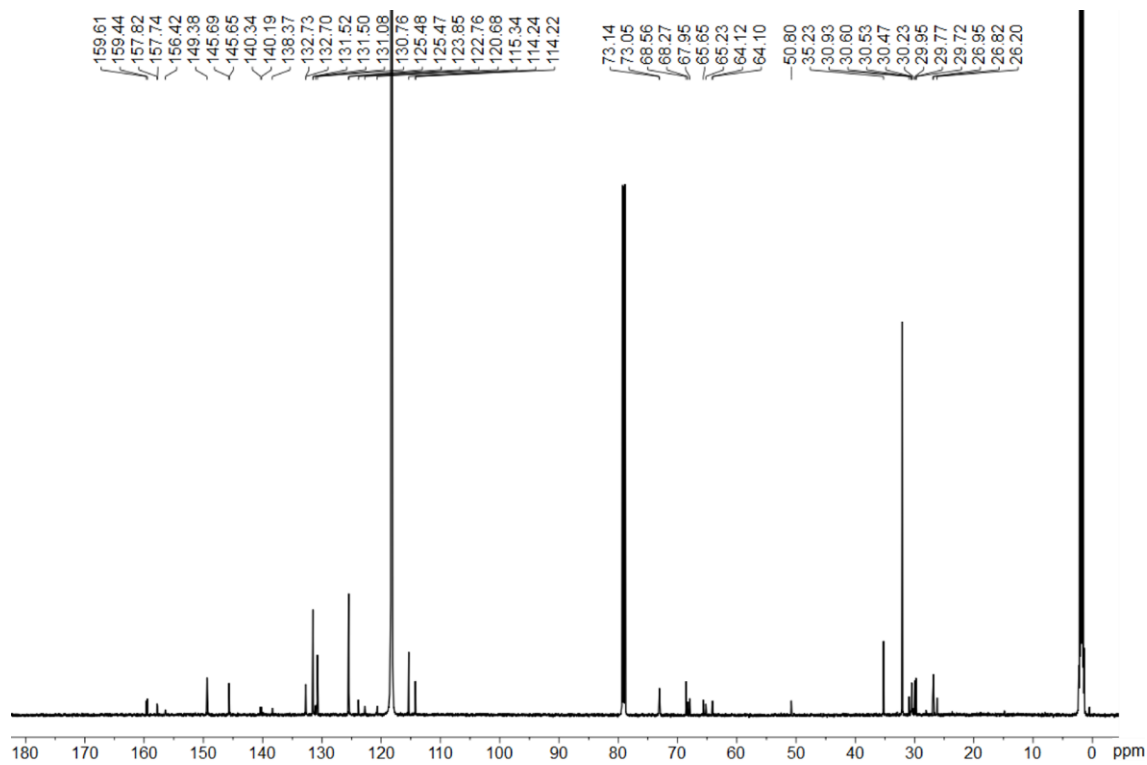


Fig. S20. $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, $v/v = 1:2$ $\text{CDCl}_3/\text{CD}_3\text{CN}$, 298 K) spectrum of **R10**.

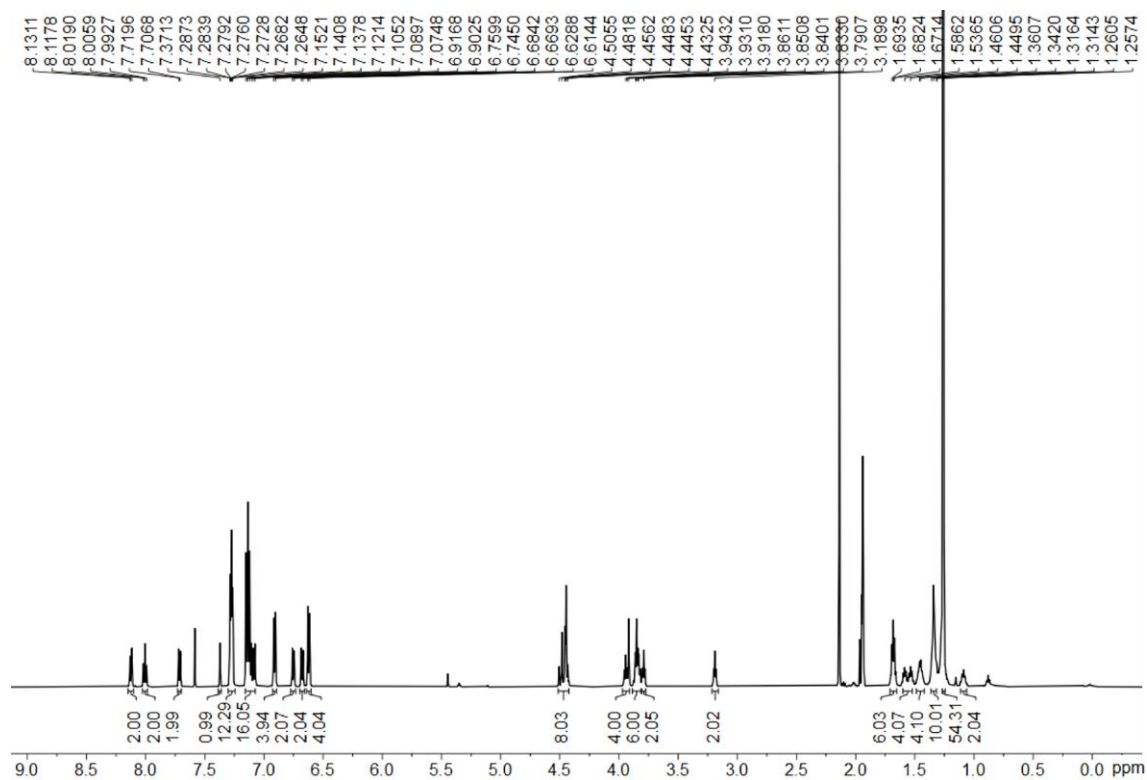


Fig. S21. ^1H NMR (600 MHz, CD_3CN , 298 K) spectrum of $[\text{Cu}(\mathbf{R10})](\text{PF}_6)$.

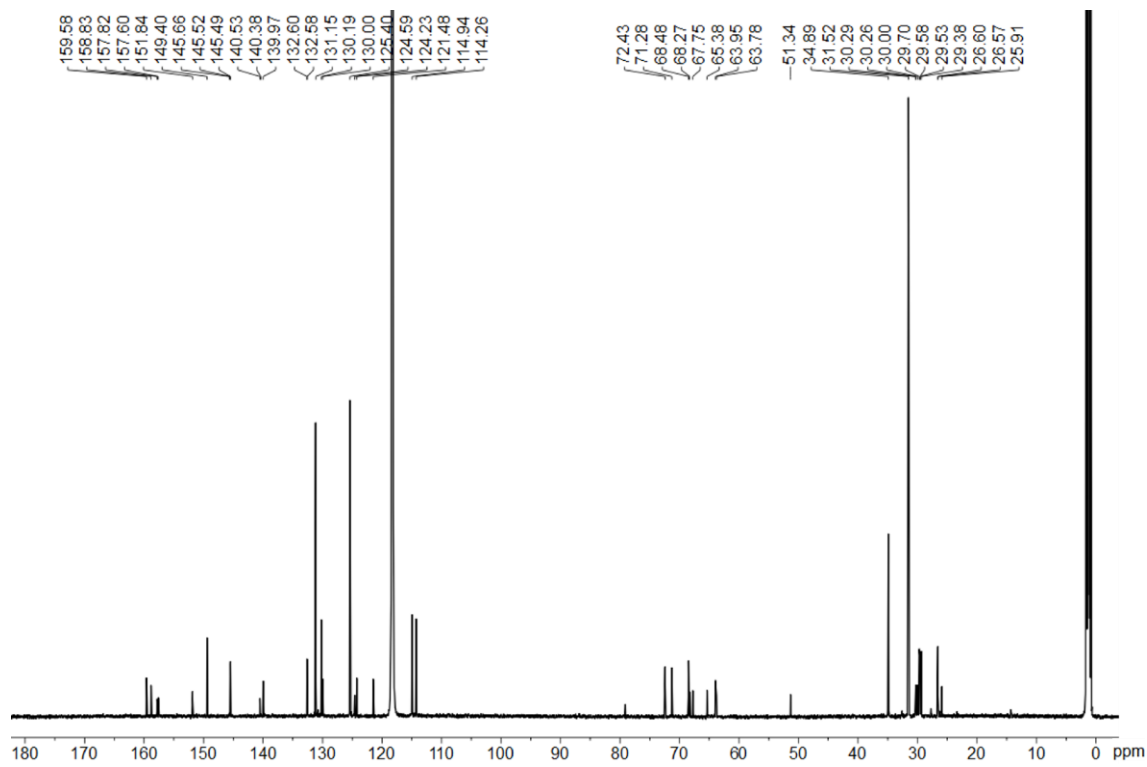


Fig. S22. $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, CD_3CN , 298 K) spectrum of $[\text{Cu}(\mathbf{R10})](\text{PF}_6)$.

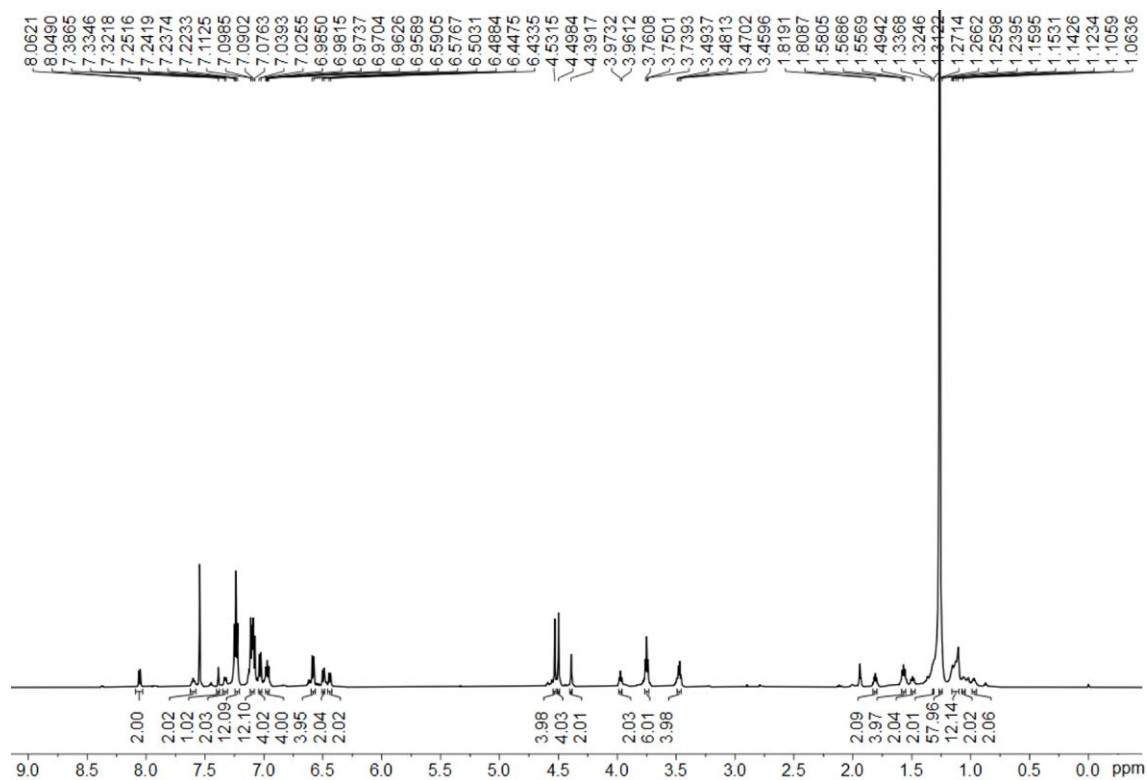


Fig. S23. ¹H NMR (600 MHz, *v/v* = 1:2 CDCl₃/CD₃CN, 298 K) spectrum of **R12**.

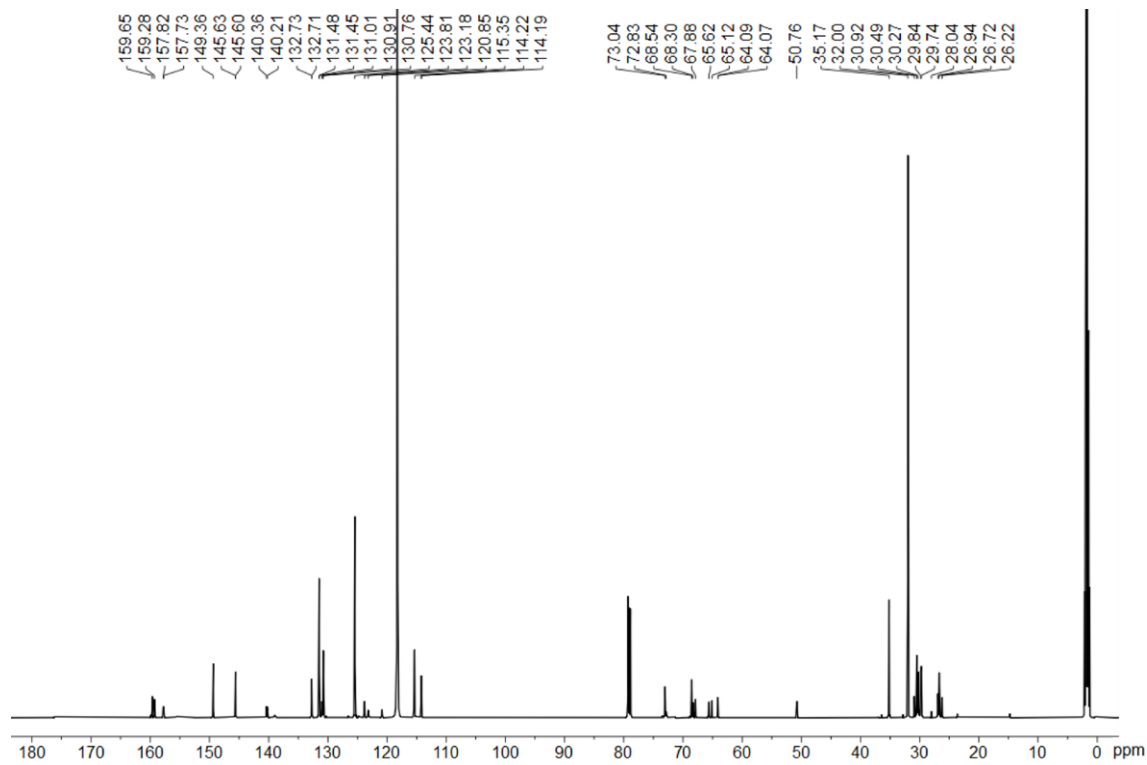


Fig. S24. ¹³C {¹H} NMR (151 MHz, *v/v* = 1:2 CDCl₃/CD₃CN, 298 K) spectrum of **R12**.

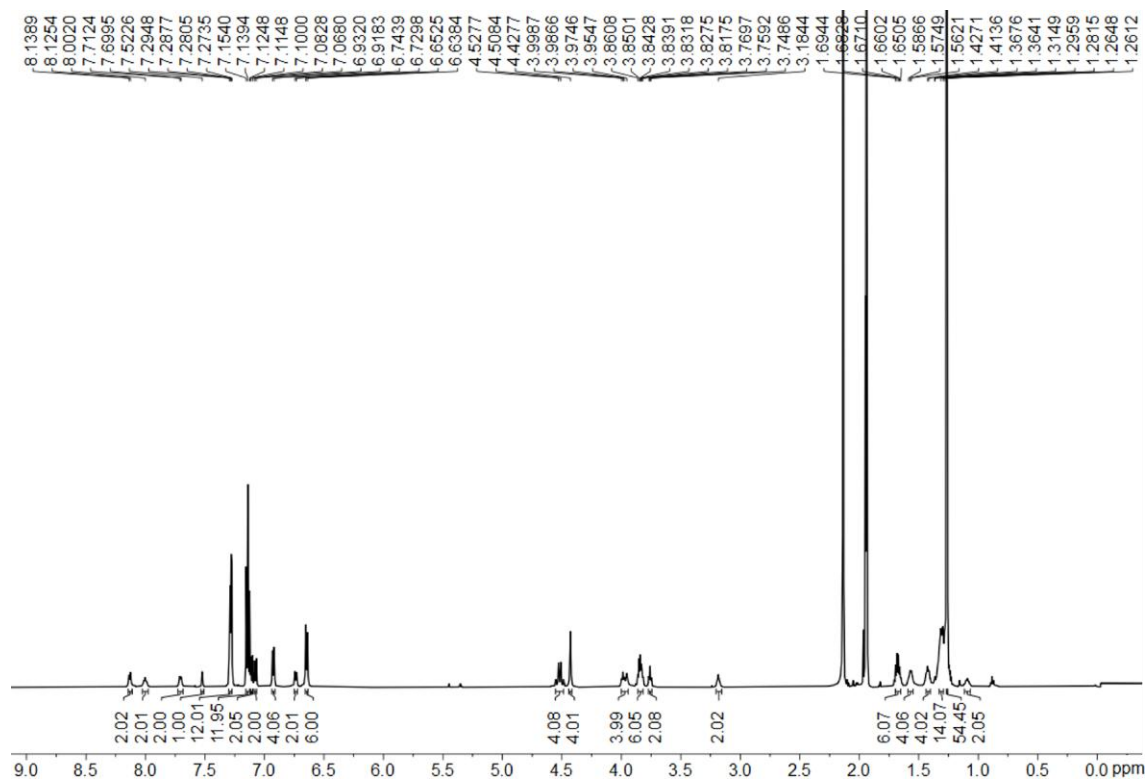


Fig. S25. ^1H NMR (600 MHz, CD_3CN , 298 K) spectrum of $[\text{Cu}(\text{R12})](\text{PF}_6)$.

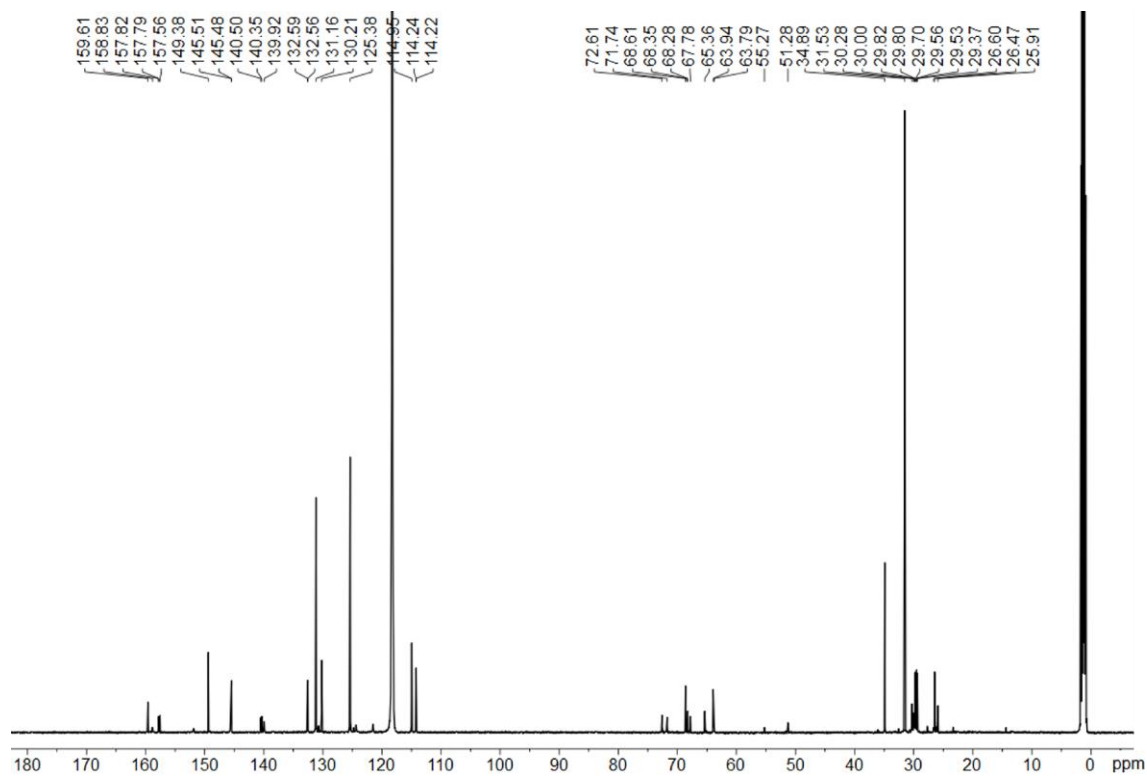
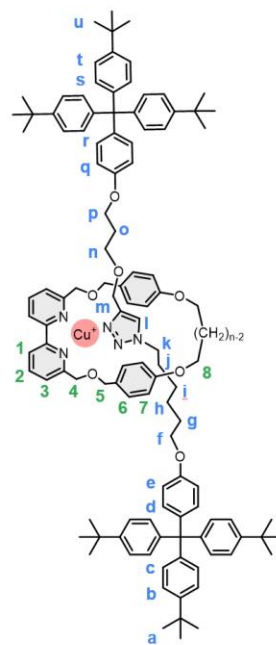


Fig. S26. $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz, CD_3CN , 298 K) spectrum of $[\text{Cu}(\text{R12})](\text{PF}_6)$.

Table S1. Partial ^1H NMR shift analysis of the PF_6^- salts of $[\text{Cu}(\mathbf{R}_n)]^+$, ($n = 4, 6, 8, 10, 12$).

	^1H NMR (ppm)						
	δH_6	δH_7	δH_8	δH_m	δH_p	δH_k	δH_n
$[\text{Cu}(\mathbf{R}_4)]^+$	6.93	6.77–6.74	4.12–4.04	3.02	3.25	4.38	2.77
$[\text{Cu}(\mathbf{R}_6)]^+$	6.88	6.64–6.61	3.95–3.90	3.61	3.95	4.11	3.19
$[\text{Cu}(\mathbf{R}_8)]^+$	6.90	6.59	3.95–3.81	3.83	3.88	3.97	3.22
$[\text{Cu}(\mathbf{R}_{10})]^+$	6.91	6.62	3.89–3.81	3.85	3.79	3.94	3.19
$[\text{Cu}(\mathbf{R}_{12})]^+$	6.93	6.64	3.86–3.80	3.96	3.76	3.98	3.19



1.3 Mass Spectrometry.

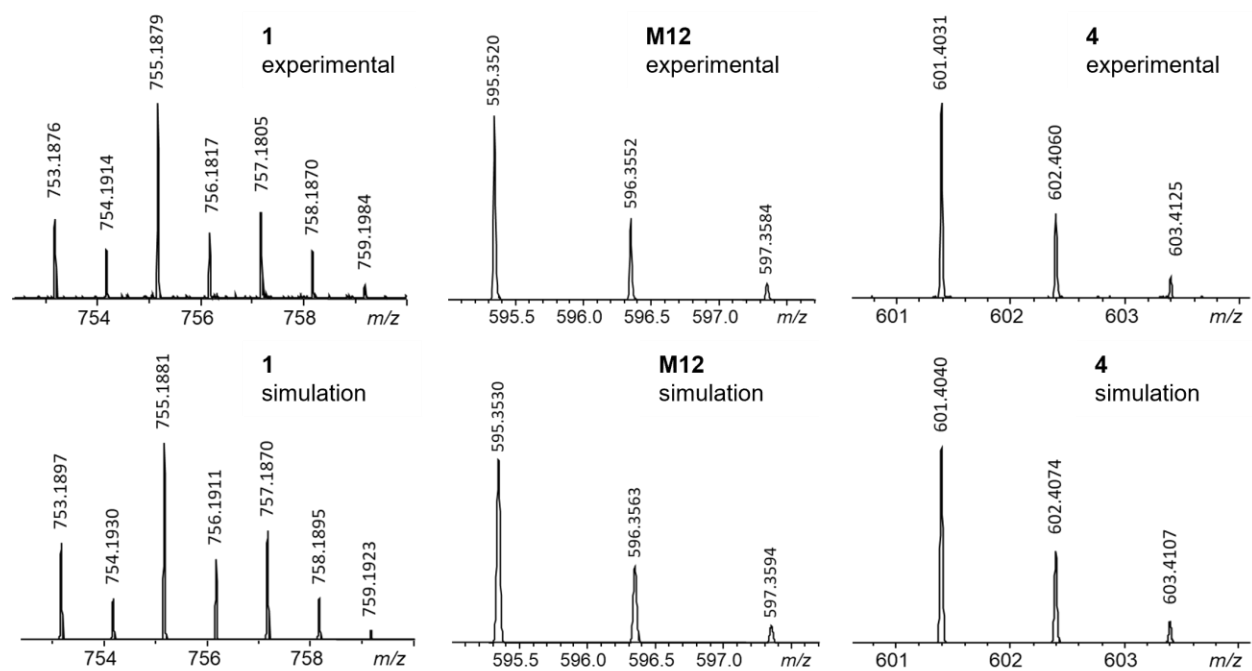


Fig. S27. HR-ESI-MS spectra of rotaxane precursors **1** (left), **M12** (middle) and **4** (right) reported in this work.

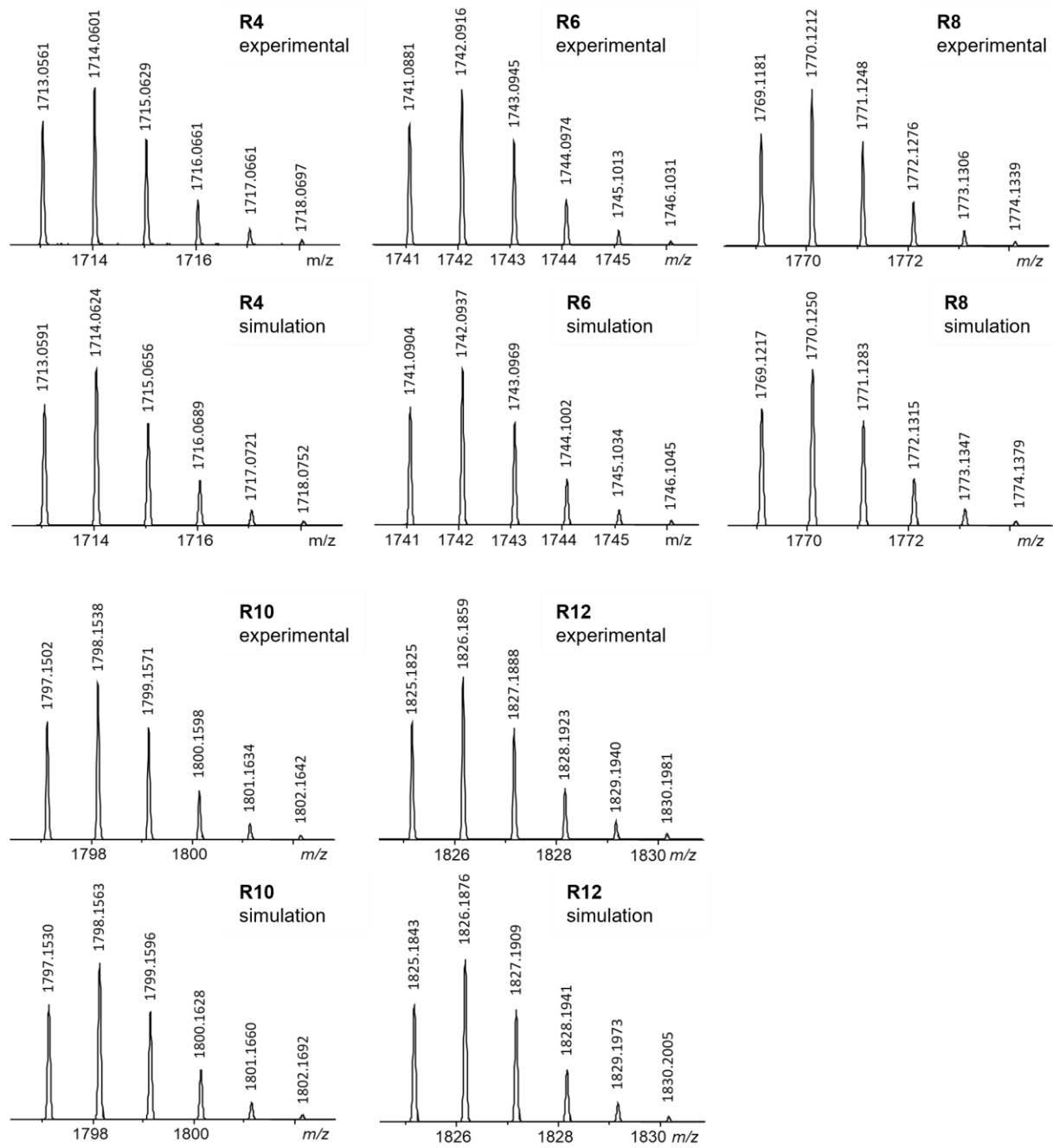


Fig. S28. HR-ESI-MS spectra of the [2]rotaxanes **R4**, **R6**, **R8**, **R10** and **R12**.

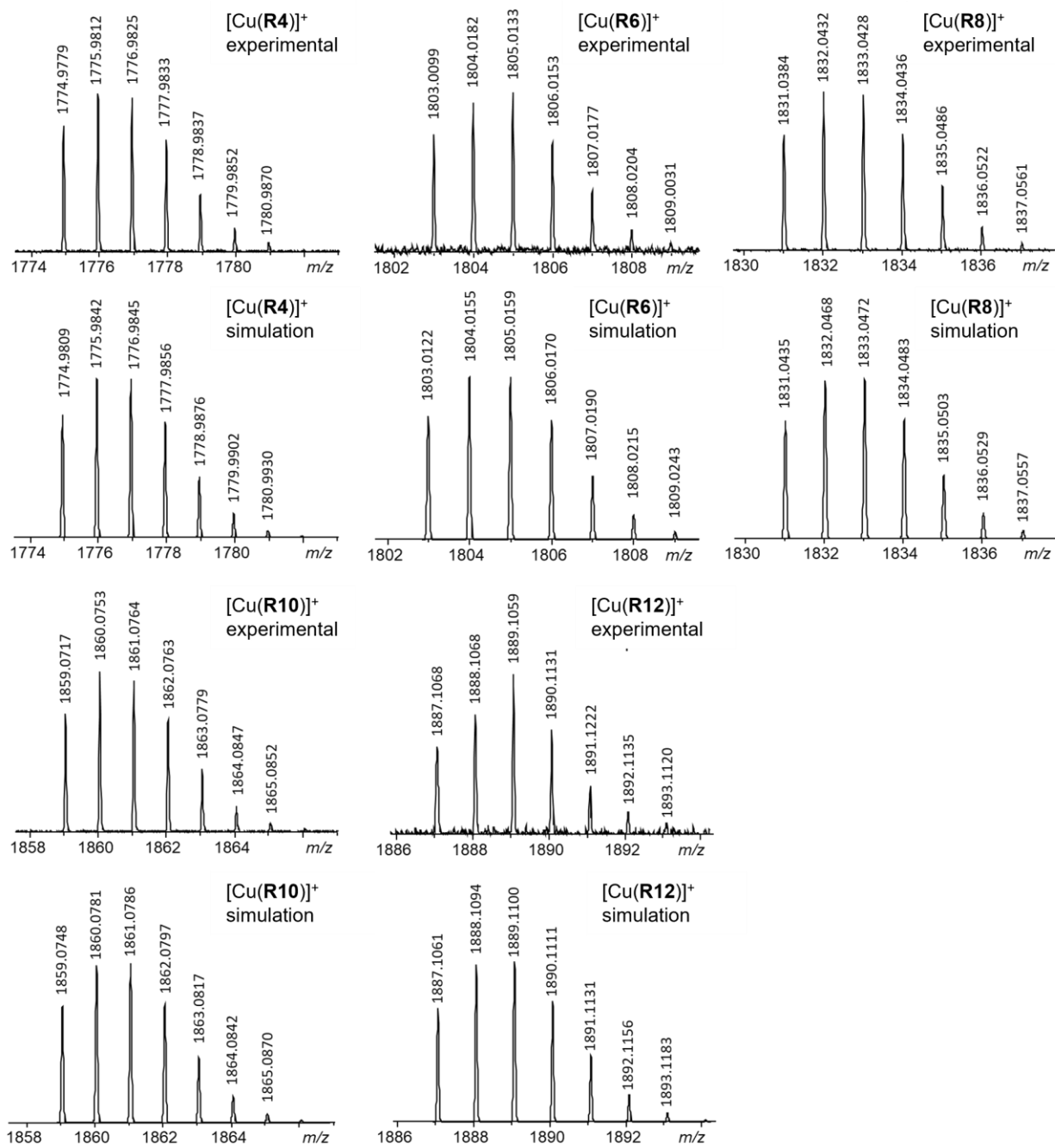


Fig. S29. HR-ESI-MS spectra of the Cu^I [2]rotaxane complexes [Cu(R4)]⁺, [Cu(R6)]⁺, [Cu(R8)]⁺, [Cu(R10)]⁺, and [Cu(R12)]⁺.

1.4 X-ray Crystallography.

Bruker D8 VENTURE X-ray diffractometer was used for crystal screening, unit cell determination, and data collection for the X-ray crystal structure of [Cu(R6)](PF₆). Data was collected using a Mo K α X-ray radiation ($\lambda = 0.71073 \text{ \AA}$) at 100 K. The ShelXT program was used to solve the structure, and *Olex2* was used for the graphical interface.⁴ The models were refined with ShelXL 2014/7 (Sheldrick, 2015) using full matrix Least Squares minimisation on F^2 . SQUEEZE routine in PLATON was employed in the structural refinements. The X-ray crystallographic coordinates have been deposited at CCDC, under the deposition number 2457336, 2504020. These data can be obtained free of charge from CCDC via <https://www.ccdc.cam.ac.uk/structures/>.

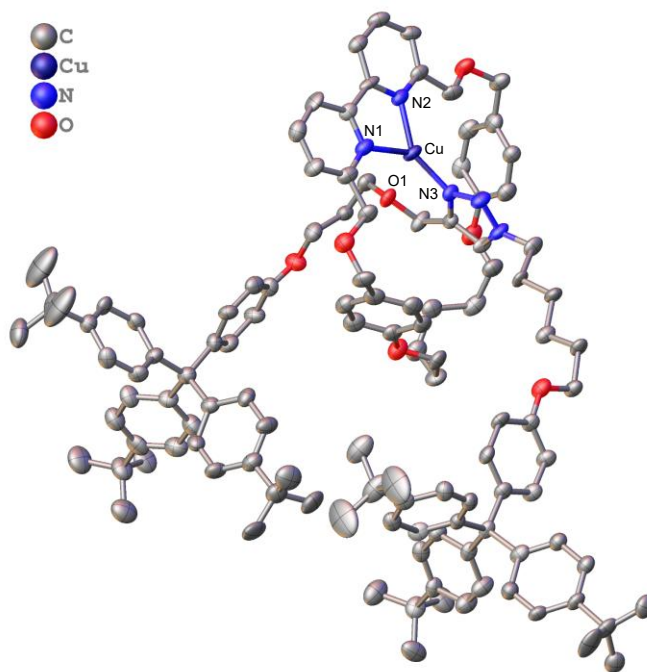


Fig. S30. Crystal structure of [Cu(R6)](PF₆) (CCDC: 2457336). All H atoms have been omitted for clarity. Thermal ellipsoids are scaled to a 50% probability level.

Table S2. Selected crystal data and structure refinement data of [Cu(**R6**)](PF₆).

Moiety formula	C ₁₁₈ H ₁₄₁ CuF ₆ N ₅ O ₇ P
Formula weight	1947.98
Crystal system	Monoclinic
Space group	<i>P2₁/n</i>
Color of crystal	Orange
Temperature/K	100
<i>a</i> /Å	34.636(7)
<i>b</i> /Å	16.348(3)
<i>c</i> /Å	20.073(4)
α /°	90
β /°	92.404(5)
γ /°	90
Volume/Å ³	11356(4)
<i>Z</i>	4
$\rho_{\text{calc}}/\text{gcm}^3$	1.213
μ/mm^{-1}	0.275
F(000)	4416.0
Crystal size/mm ³	0.04 × 0.02 × 0.02
Radiation/Å	MoK α ($\lambda = 0.71073$)
$\theta_{\text{min}}, \theta_{\text{max}}/\text{°}$	2.756, 48.998
Index ranges	-40 ≤ <i>h</i> ≤ 40, 0 ≤ <i>k</i> ≤ 19, 0 ≤ <i>l</i> ≤ 23
Data/restraints/parameters	18427/11/1322
Final R indexes [<i>I</i> ≥ 2 σ (<i>I</i>)]	<i>R</i> ₁ = 0.1500, <i>wR</i> ₂ = 0.3039
Final R indexes [all data]	<i>R</i> ₁ = 0.1847, <i>wR</i> ₂ = 0.3263
Largest diff. peak/hole / e Å ⁻³	0.97/-1.03
Restrained goodness of fit	1.223

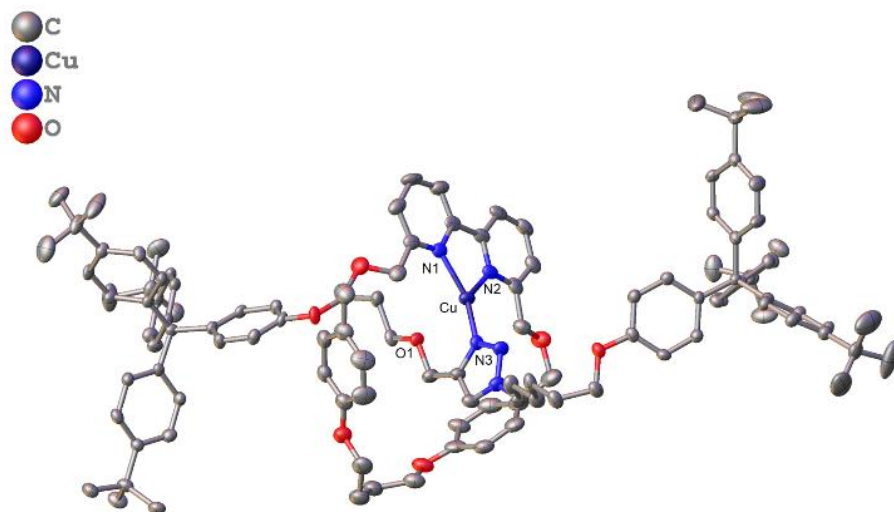


Fig. S31. Crystal structure of [Cu(**R4**)](PF₆) (CCDC: 2504020). All H atoms have been omitted for clarity. Thermal ellipsoids are scaled to a 50% probability level.

Table S3. Selected crystal data and structure refinement data of [Cu(**R4**)](PF₆).

Moiety formula	C ₁₁₆ H ₁₃₇ CuF ₆ N ₅ O ₇ P
Formula weight	1919.95
Crystal system	Triclinic
Space group	P-1
Color of crystal	Orange
Temperature/K	100
<i>a</i> /Å	14.1927(10)
<i>b</i> /Å	16.1321(10)
<i>c</i> /Å	27.927(2)
<i>α</i> /°	90.618(5)
<i>β</i> /°	104.435(5)
<i>γ</i> /°	108.908(4)
Volume/Å ³	5828.9(7)
<i>Z</i>	2
ρ_{calc} /gcm ³	1.203

μ/mm^{-1}	0.945
F(000)	2256.0
Crystal size/ mm^3	$0.1 \times 0.05 \times 0.03$
Radiation/ \AA	CuK α ($\lambda = 1.54178$)
$\theta_{\text{min}}, \theta_{\text{max}}/^\circ$	3.284, 118.684
Index ranges	$-15 \leq h \leq 15, -17 \leq k \leq 17, -31 \leq l \leq 31$
Data/restraints/parameters	16816/105/1365
Final R indexes [$I \geq 2\sigma(I)$]	$R_1 = 0.0767, wR_2 = 0.1886$
Final R indexes [all data]	$R_I = 0.1262, wR_2 = 0.2189$
Largest diff. peak/hole / $e \text{\AA}^{-3}$	1.18/-0.64
Restrained goodness of fit	1.024

2. Electrochemical Measurements.

2.1 General Methods.

Chemicals for synthesis were obtained from commercial sources and used as received unless otherwise stated. Electrochemical studies at pH 4 and pH 7 were performed in Britton–Robinson buffer (BR) containing H_3BO_3 (0.04 M, 99.999%, Fisher Scientific), CH_3COOH (0.04 M, 100%, VWR), H_3PO_4 (0.04 M, 85 wt % in H_2O , J&K Scientific), NaClO_4 (0.1 M, 99.9%, Sigma-Aldrich) and Milli-Q ultrapure water ($>18.2 \text{ M}\Omega \text{ cm}$). Solutions were adjusted to pH 4 and pH 7 respectively using NaOH (10 M, analytical grade, Merck Millipore), and subjected to 30-min sparging with either N_2 or O_2 (99.995% high-purity grade, Linde HKO) before electrochemical measurements.

2.2 Catalyst Ink Preparations.

Multiwalled carbon nanotubes were pretreated by immersing in 6 M HCl (Duksan, 37% GR) for 24 hours. The acid-treated carbon nanotubes were subsequently filtered, rinsed with Milli-Q ultrapure water, and dried overnight in a vacuum oven at $80 \text{ }^\circ\text{C}$. The Cu catalyst (2.0 mg) was dissolved in CH_3CN (RCI Labscan, AR) and mixed with the acid-treated carbon nanotubes (8.0 mg) to form a catalyst/carbon mixture. The mixture was sonicated for 5 min and subsequently vacuum-dried at $37 \text{ }^\circ\text{C}$ overnight. The catalyst/carbon mixture (4.0 mg) was finely ground and then suspended in absolute CH_3OH (1.0 mL, Scharlab, Abs) and sonicated for 15 min. Nafion

perfluorinated resin (10 μL , 5 wt% in lower aliphatic alcohols with 15–20% water, Sigma-Aldrich) was added to the well-dispersed catalyst ink. The catalyst ink was sonicated for an additional 10 min and was drop-casted (15.0 μL) onto a polished glassy carbon disk electrode ($A = 0.196 \text{ cm}^2$). The electrode was then dried under a stream of N_2 .

2.3 Electrochemical Activity Measurements.

The electrocatalytic oxygen reduction activity was evaluated in a three-electrode electrochemical cell. A polished glassy carbon (GC) electrode served as the working electrode, a graphite rod as the counter electrode, and an aqueous “no-leak” Ag/AgCl (3 M KCl, EDAQ Inc) as the reference electrode. At both pH 4 and pH 7, ORR onset potential is defined as the potential at which 10 $\mu\text{A}/\text{cm}^2$ current density is reached. The $\text{Cu}^{\text{II/I}}$ redox properties of Cu^{I} rotaxanes were characterised in 0.1 M tetrabutylammonium hexafluorophosphate (NBu_4PF_6) in CH_3CN , using GC as the working electrode and a graphite rod as the counter electrode, with an Ag wire immersed in 10 mM AgNO_3 (in CH_3CN) as a pseudo-reference. The $\text{Cu}^{\text{II/I}}$ redox couples were referenced against 1 mM ferrocene (Fc/Fc^+) in NBu_4PF_6 in CH_3CN . Cyclic voltammetry (CV) and linear sweep voltammetry (LSV) experiments were conducted using a CH Instruments 660E potentiostat at a scan rate of 10 mV/s for ORR activity measurement and 100 mV/s for $\text{Cu}^{\text{II/I}}$ and Fc/Fc^+ redox measurements. All experiments were performed in triplicates, and error bars represent the standard deviations of the triplicate trials. The GC electrode was polished using a 3 to 0.5 μm alumina suspension polishing kit (Allied Tech) and cleaned by electrochemical cycling from -0.4 to 1.7 V vs. Ag/AgCl in 0.1 M HClO_4 . Electrochemical potentials were referenced to the reversible hydrogen electrode (RHE) according to established protocols.^{5,6}

2.4 Product Quantification.

The electrocatalytic oxygen reduction selectivity was assessed through a colorimetric assay for H_2O_2 quantification, following a previously reported protocol.^{7,8} Briefly, the concentration of H_2O_2 was determined using cerium(IV) sulfate ($\text{Ce}(\text{SO}_4)_2$) titration at pH 4 and Amplex Red assay at pH 7. A solution of $\text{Ce}(\text{SO}_4)_2$ (1.0 mM) was prepared by dissolving $\text{Ce}(\text{SO}_4)_2 \cdot 4\text{H}_2\text{O}$ (4.0 mg) in 0.5 M H_2SO_4 (10.0 mL). A standard curve for H_2O_2 was generated by adding the prepared cerium titrant (0.1 mL) to H_2O_2 solutions (0.9 mL) with known concentrations (0.5, 1.0, 2.0, 5.0, 10.0 μM). A working solution of Amplex Red was prepared by mixing of 10 mM Amplex Red (40 μL) in DMSO with of 0.05 M pH 7 sodium phosphate buffer (3.88 mL) containing of 10 U/mL

horseradish peroxidase (HRP) (80 μL). A standard curve for H_2O_2 was generated by adding the prepared Amplex Red (0.5 mL) to H_2O_2 solutions (0.5 mL) with known concentrations (1.0, 2.5, 5.0, 10.0 μM). The absorbance at 316 nm ($\text{Ce}(\text{SO}_4)_2$ titration) and 571 nm (Amplex Red assay) were recorded respectively to determine the H_2O_2 concentration based on the calibration curve.

2.5 Calculations of H_2O Selectivity.

The number of electrons transferred per O_2 molecule (n) during oxygen reduction reaction by the Cu^{I} rotaxane complexes was determined following a reported method,^{9,10} where x represents the mole fraction of O_2 consumed for H_2O_2 generation, relative to total O_2 consumed for H_2O_2 and H_2O generation.

$$n = 4(1 - x) + 2x$$

$$x = \frac{\text{mole of } \text{O}_2 \text{ reduced (H}_2\text{O}_2\text{)}}{\text{total mole of } \text{O}_2 \text{ reduced (H}_2\text{O}_2 + \text{H}_2\text{O})}$$

$$\% \text{H}_2\text{O} = \frac{n - 2}{2} \times 100 \%$$

2.6 Metal Quantifications.

The quantity of Cu in catalyst-loaded was determined using inductively coupled plasma mass spectrometry (ICP-MS) (Agilent 7700x). The catalyst ink (10 μL) was added in aqua regia (9.99 mL, 1:3 v/v HNO_3 : HCl) and left standing for 24 h. The resulting catalyst/acid mixture was filtered using a PES syringe filter (Labfil, 0.22 μm). The filtrate (490 μL) and an internal standard (10000 ppb Rh, 10 μL) were added to 1% HNO_3 (9.5 mL).

2.7 Experimental Results.

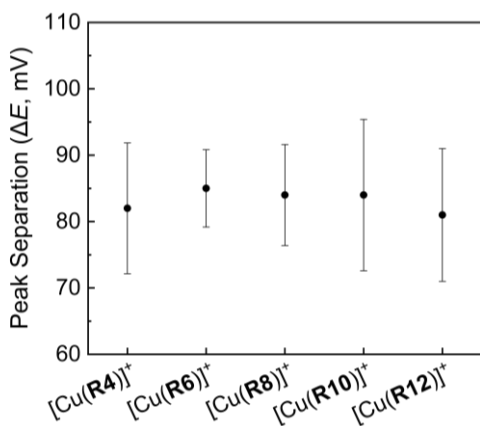


Fig. S32. Cu^{II/I} redox characterisation of Cu^I rotaxanes. Peak separation (ΔE) of the Cu^{II/I} redox couple in 0.1 M NBu₄PF₆ in CH₃CN. Error bars represent the standard deviations of the triplicate trials.

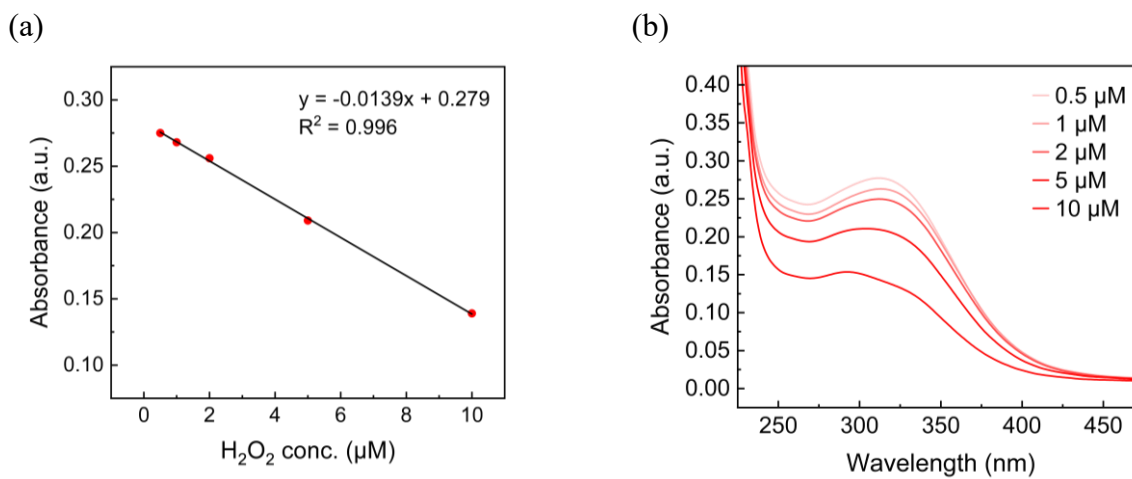


Fig. S33. H₂O₂ colorimetric assay using Ce(SO₄)₂ titration method. (a) Standard curve of H₂O₂ with known concentrations. (b) UV-vis absorption spectra of the H₂O₂ standards.

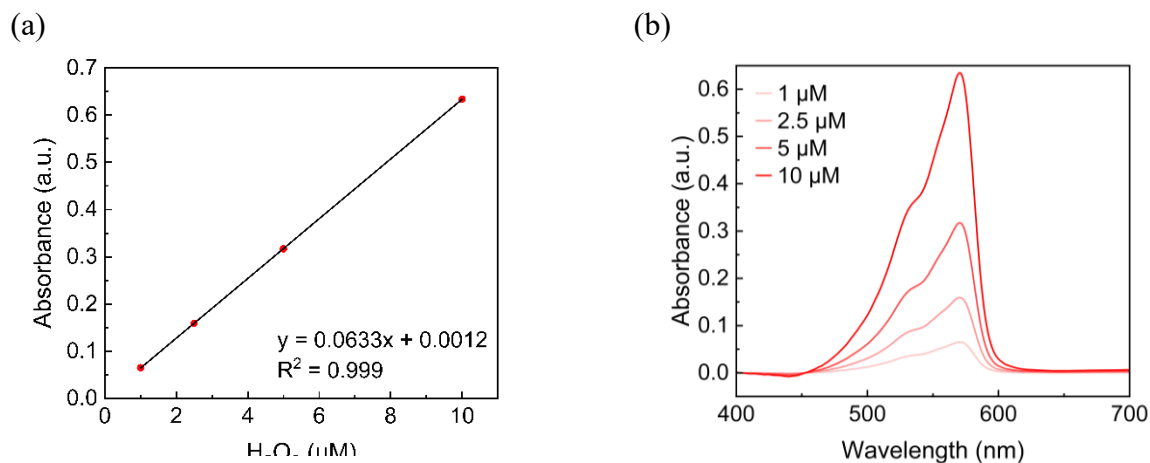


Fig. S34. H₂O₂ colorimetric assay using Amplex Red assay. (a) Standard curve of H₂O₂ with known concentrations. (b) UV-vis absorption spectra of the H₂O₂ standards.

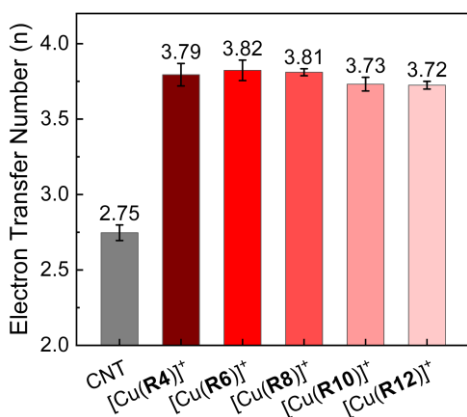


Fig. S35. Electron transfer number (n) of ORR catalysed by carbon nanotubes (CNT), [Cu(R4)]⁺, [Cu(R6)]⁺, [Cu(R8)]⁺, [Cu(R10)]⁺, and [Cu(R12)]⁺ supported on glassy carbon electrode (GC) in pH 4 BR buffer. Error bars represent the standard deviations of the triplicate trials.

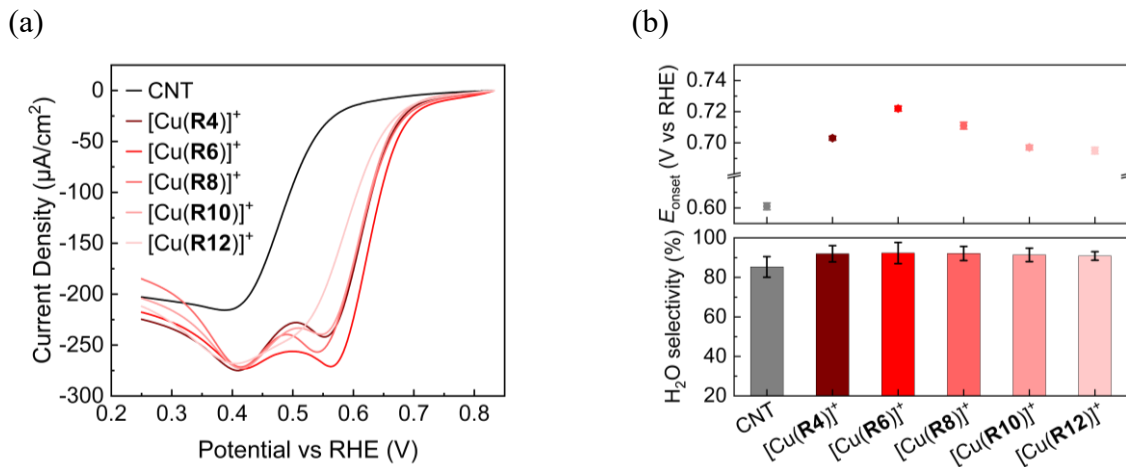


Fig. S36. (a) Linear sweep voltammograms of carbon nanotubes (CNT), [Cu(R4)]⁺, [Cu(R6)]⁺, [Cu(R8)]⁺, [Cu(R10)]⁺, and [Cu(R12)]⁺ supported on glassy carbon electrode (GC) in O₂-saturated pH 7 BR buffer, and (b) their respective ORR onset potential and product selectivity. The ORR onset potential is defined as the potential at which 10 µA/cm² is reached. Error bars represent the standard deviations of the triplicate trials.

At both pH 4 and pH 7, a second ORR reduction peak was observed in the LSV curves for all Cu^I rotaxanes, occurring at *c.a.* 0.25 V (vs. RHE) and *c.a.* 0.40 V (vs. RHE), respectively (Fig. 4, S36). This peak is attributed to the background ORR activity of the CNT support.

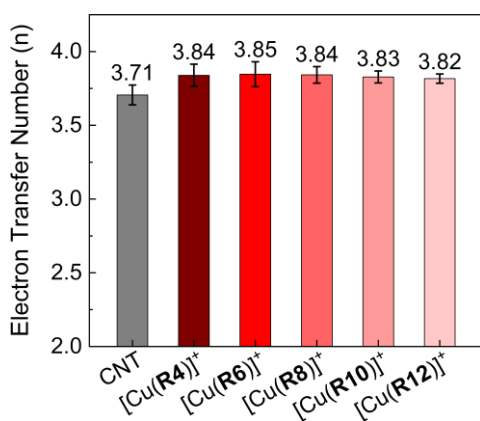


Fig. S37. Electron transfer number (n) of ORR catalysed by carbon nanotubes (CNT), [Cu(R4)]⁺, [Cu(R6)]⁺, [Cu(R8)]⁺, [Cu(R10)]⁺, and [Cu(R12)]⁺ supported on glassy carbon electrode (GC) in pH 7 BR buffer. Error bars represent the standard deviations of the triplicate trials.

Table S4. ORR onset potentials and electron transfer number of reported molecular Cu catalysts.

Molecular Cu complexes	Onset potential (V vs RHE)		Electron transfer number (n)		References
	pH 4	pH 7	pH 4	pH 7	
[Cu(R6)] ⁺	0.58	0.72	3.8	3.9	This work
[Cu(2-(5-(pyren-1-yl) pentyl)-9-methyl-1,10-phenanthroline) ₂] ²⁺	0.86 (pH 5)	-	3.9 (pH 5)	-	11
[Cu(2,9-Me ₂ -phen) ₂] ⁺	0.72 (pH 5.2)	-	2.7 (pH 5.2)	-	12
[Cu(2,9-Me ₂ -4,7-Ph ₂ -phen) ₂] ⁺	0.69 (pH 5.2)	-	2.9 (pH 5.2)	-	12
[Cu(3,8-Br ₂ -phen) ₂] ⁺	0.61 (pH 5.2)	-	3.8 (pH 5.2)	-	12
[Cu(LH)] ²⁺	0.60 (pH 5)	-	3.3 (pH 5)	-	13
[Cu(5-Cl-phen) ₂] ⁺	0.60 (pH 5.2)	-	3.1 (pH 5.2)	-	12
Au Cu(DAT)	<i>ca.</i> 0.6 (pH 5.2)	-	-	-	14
[Cu(phen) ₂] ⁺	0.58 (pH 5.2)	-	3.1 (pH 5.2)	-	12
[Cu(2,9-Et ₂ -phen)] ²⁺	0.59 (pH 4.8)	-	-	-	15,16
[Cu(TPA)] ²⁺	0.57	0.69	3.6	-	5
[Cu ₂ (BPMP-pyrene)] ²⁺	0.57	-	2.7	-	17
[Cu(PMAP)] ²⁺	0.57	-	2.5	-	5
[Cu(4,7-Me ₂ -phen) ₂] ⁺	0.56 (pH 5.2)	-	3.7 (pH 5.2)	-	12
[Cu(3,4,7,8-Me ₄ -phen) ₂] ⁺	0.55 (pH 5.2)	-	3.8 (pH 5.2)	-	12
[Cu(2,9-nBu ₂ -phen)] ²⁺	0.55 (pH 4.8)	-	-	-	15,16
Cu(phen ^C)	0.53 (pH 4.8)	-	3.6 (pH 4.8)	-	18

[Cu(TPT)] ²⁺	0.51 (pH 5.3)	-	1.9	-	19
[Cu(5-Cl-phen)] ²⁺	0.49 (pH 5.2)	-	<i>ca.</i> 3.5	-	20
[Cu(TPP)] ²⁺	0.44	-	-	-	21
[Cu(2-Cl-phen) ₂] ⁺	0.42 (pH 5.2)	-	2.8 (pH 5.2)	-	12
[Cu(4,7-dppds)] ²⁺	0.41 (pH 5.3)	-	-	-	22
PG Cu(DAT)	<i>ca.</i> 0.4 (pH 5.2)	--	1.2 (pH 5.2)	-	14
[Cu(phen)]/Au	0.40	-	-	-	23
[Cu(3-ethynyl-phen)] ⁺	0.39 (pH 4.8)	-	3.6	-	18
[Cu(phen)] ²⁺	0.30 (pH 4.8)	-	-	-	24
MWCNT@BTAH-Cu ²⁺	-	0.74	-	<i>ca.</i> 3.8	25
Cu(ImBenz-NO ₂)Cl ₂	-	0.69	-	3.5–3.8	26
[Cu(C10)] ⁺	-	0.62	-	<i>ca.</i> 3.8	6
[Cu(3,5-dimethyl-4-amino-1,2,4-triazole)] ²⁺	-	0.58	-	-	16
[Cu(trpn)(Im)] ²⁺	-	0.58	-	-	27
[Cu(baEtO)(Im)] ²⁺	-	0.57	-	-	28
Cu ²⁺ :L ₂	-	0.55	-	3.2–3.8	29
[Cu(dpaq)] ⁺	-	0.50	-	3.8	30
CuPoly-CB	-	0.50	-	2.7	31
[Cu(bmpa)] ²⁺	-	0.49	-	<i>ca.</i> 2.5–2.7	32
Cu-tpmen	-	0.47	-	<i>ca.</i> 2.3-3.2	33
Cu-tpbn-Me ₂ bpy	-	0.46	-	2.9–3.4	34

[Cu(terpy)] ²⁺	-	0.45	-	ca. 2.5–2.7	32
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LH = (2-tert-butyl-6-{{[(6-methyl-pyridin-2-ylmethyl)-pyridin-2-ylmethyl-amino]-methyl}-4-({4-[(pyren-1-ylmethyl)-amino]-butylamino}-methyl)-phenol), Au| = modified gold electrode; DAT = 3,5-diamino-1,2,4-triazole, TPA = tris(2-pyridylmethyl)amine, BPMP = 2,6-bis[(bis(2-pyridylmethyl)amino)methyl]-phenol, PMAP = bis[2-(2-pyridyl)ethyl]-(2-pyridyl)methylamine, phen^c = 3-ethynyl-1,10-phenanthroline was covalently attached to an azide-modified glassy carbon electrode by cycloaddition, TPT = 2,4,6-tris(2-pyridyl)-1,3,5-triazine, TPP = 5,10,15,20-tetraphenyl-21H,23H-porphyrin, dppds = 4,7-diphenyl-1,10-phenanthroline disulfonate, PG| = modified pyrolytic graphite electrode, BTAH = 1,2,3-benzotriazole, ImBenz-NO₂ = 2-(1-(2,6-diisopropylphenyl)-1H-imidazol-2-yl)-5-nitro-1H-benzo[d]imidazole], C10 = [2]catenane derivatives, trpn = tris(3-aminopropyl)amine, baEtO = 2-[bis(2-aminoethyl)amino]ethanol, L₂ = ligand obtained from polymerizations of poly(ethylene glycol) diglycidyl ether (PEGDGE) and 2,2'-bipyridine-5,5'-dicarboxylic acid, dpaq = 2-[bis(pyridine-2-ylmethyl)]amino-N-quinolin-8-yl-acetamidate, Poly, = pyrene-terminated poly(2-hydroxy-3-dipicolylamino) propyl methacrylate (Py-PGMADPA); CB = carbon black, bmpa = bis(2-pyridylmethyl)amine, tpbn = tetrapyridylalkylamine N,N,N',N'-tetrakis(2-pyridyl-methyl)-1,4-butanediamine, terpy = 2,2':6',2''-terpyridine, tpmen = N,N,N'-tris(2-pyridylmethyl)-N'-methylethylenediamine.

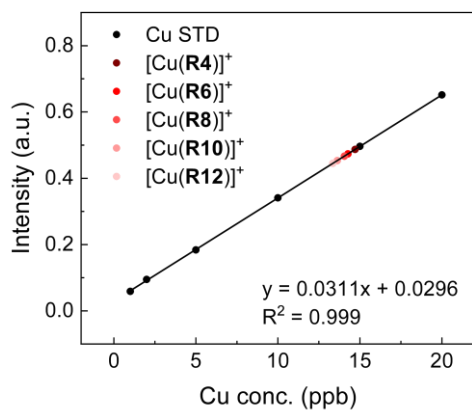


Fig. S38. ICP-MS results of Cu^I rotaxane complexes.

Table S5. Concentration of Cu metal in rotaxane complexes.

Samples	Cu/Rh ratio	Cu conc. ($\mu\text{mol}/\text{cm}^2$)
[Cu(R4)] ⁺	0.487	0.0236
[Cu(R6)] ⁺	0.473	0.0229
[Cu(R8)] ⁺	0.467	0.0226
[Cu(R10)] ⁺	0.453	0.0219
[Cu(R12)] ⁺	0.445	0.0214

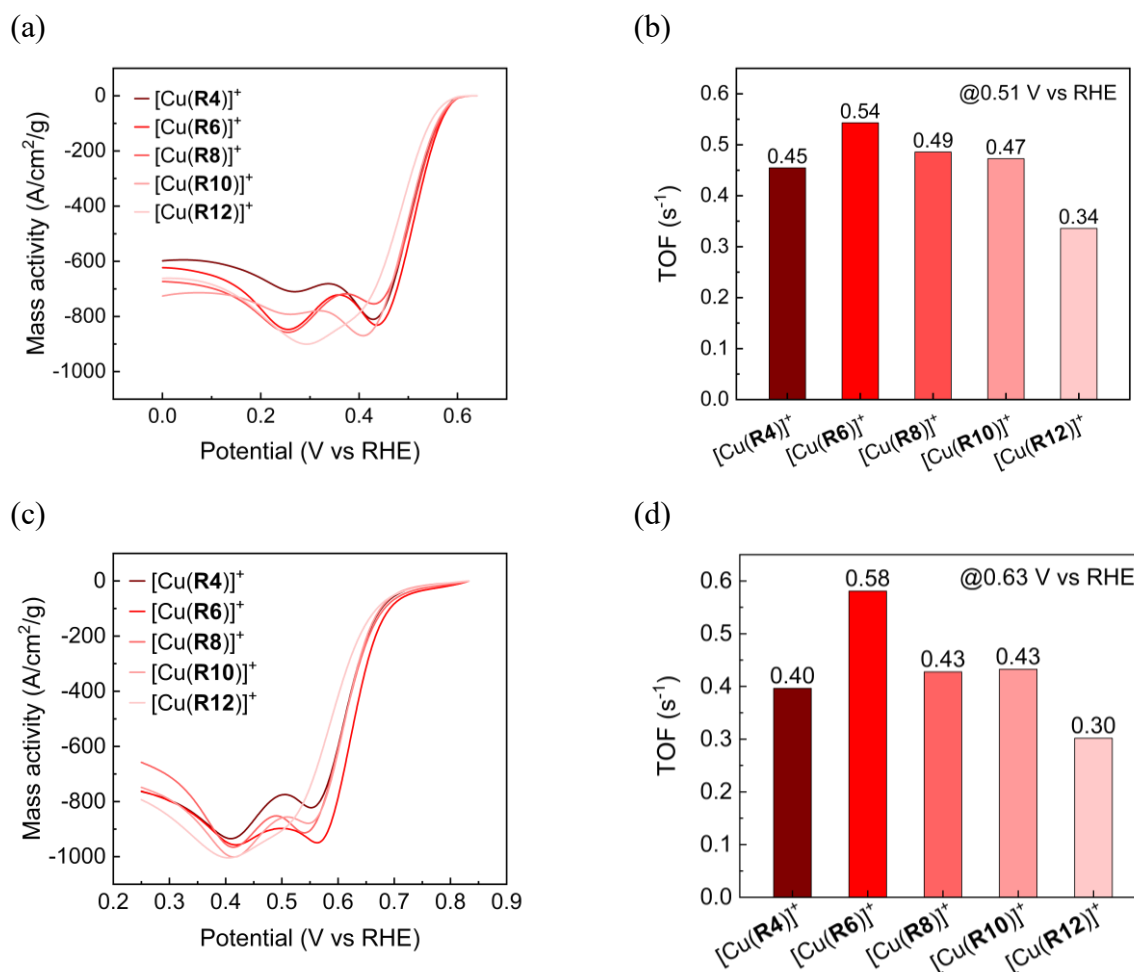


Fig. S39. Mass activity and turnover frequency (TOF) of [Cu(R4)]⁺, [Cu(R6)]⁺, [Cu(R8)]⁺, [Cu(R10)]⁺, and [Cu(R12)]⁺ with their ORR current densities normalized by Cu quantity in (a, b) pH 4 BR buffer and (c, d) pH 7 BR buffer.

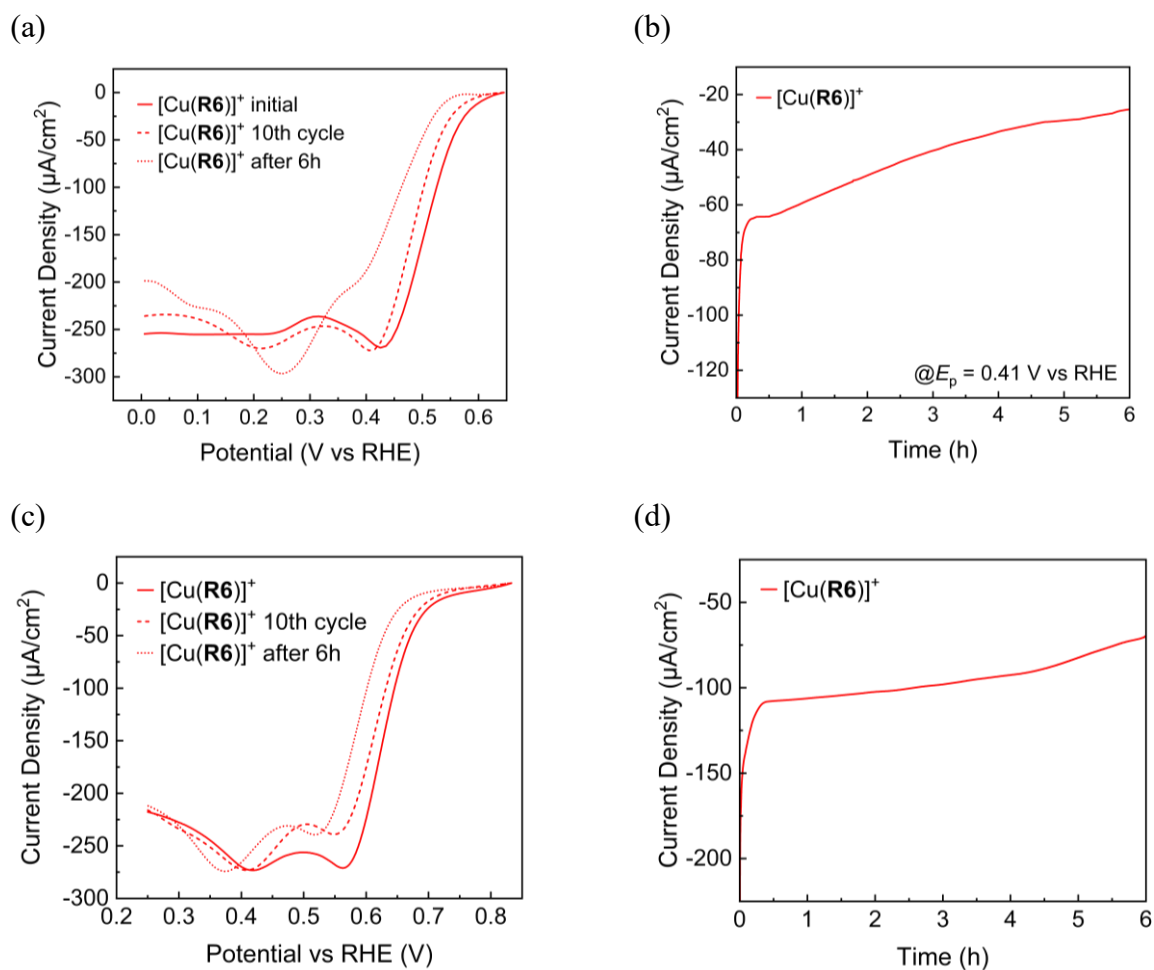
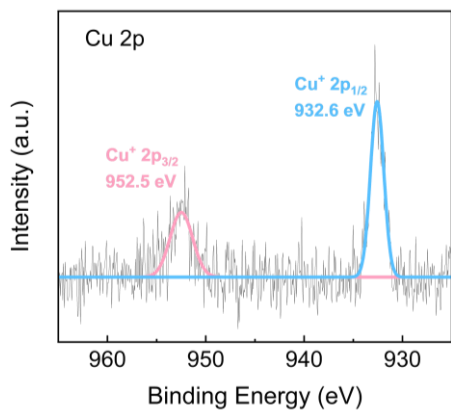
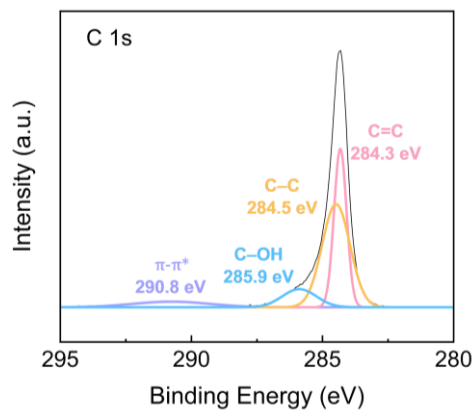


Fig. S40. Linear sweep voltammograms (LSV) of $[\text{Cu}(\text{R6})]^+$ at the initial stage (solid red), at 10th consecutive LSV scan (dashed red), and after 6 hours of chronoamperometry test (dotted red). Chronoamperometry curve of $[\text{Cu}(\text{R6})]^+$ recorded over 6 hours at E_p . Data in (a) and (b) were collected in pH 4 BR buffer, while data in (c) and (d) were recorded in pH 7 BR buffer.

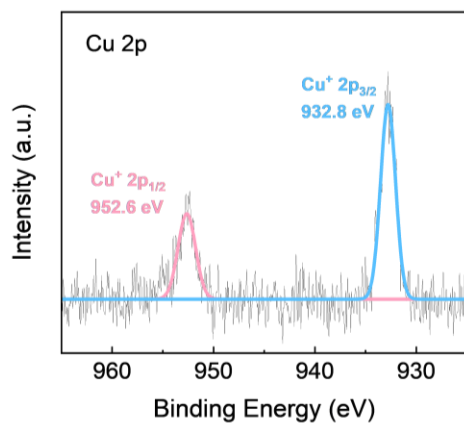
(a)



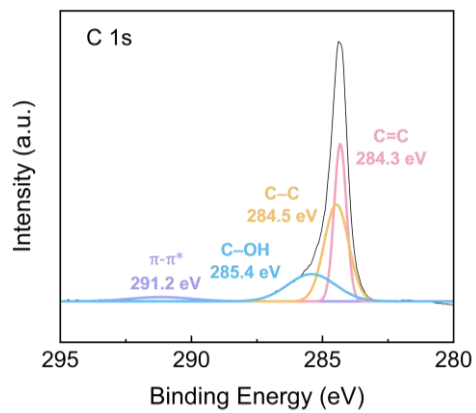
(b)



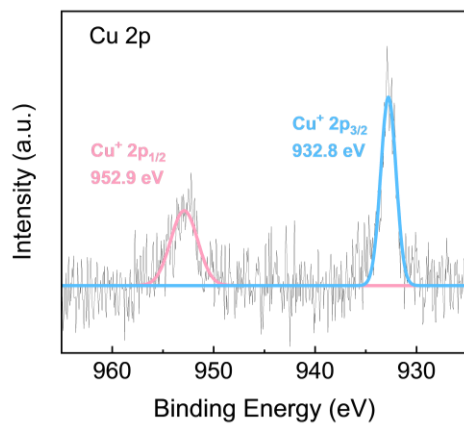
(c)



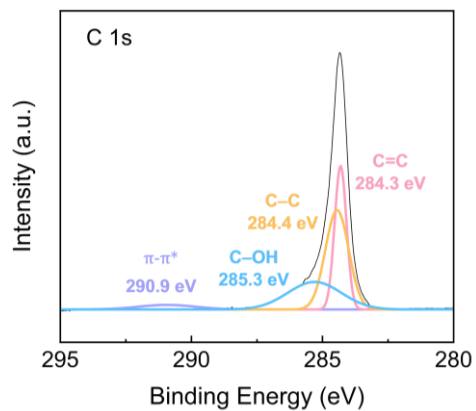
(d)



(e)



(f)



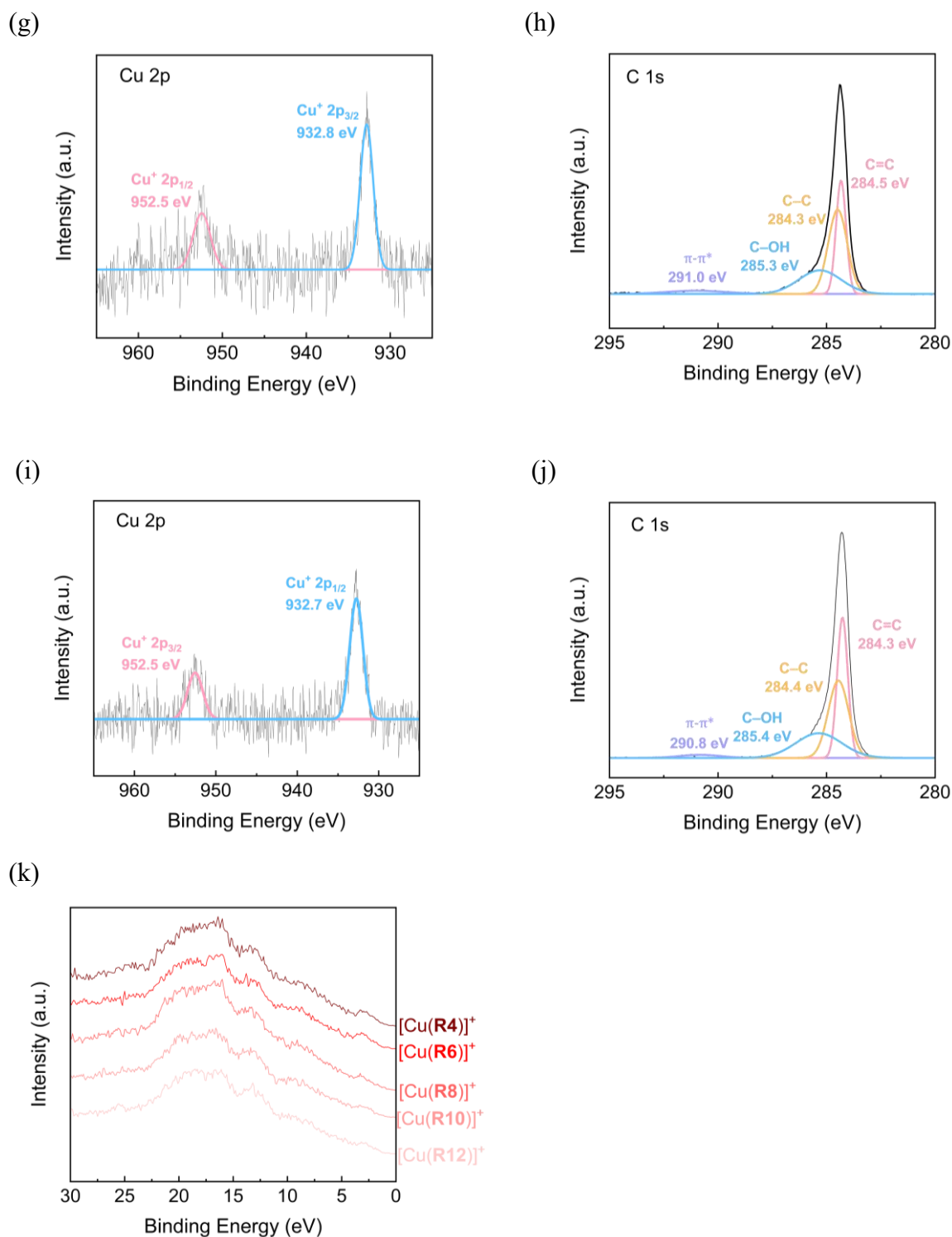


Fig. S41. High-resolution X-ray photoelectron (XPS) spectra of Cu 2p, C 1s, and (k) valence band for (a,b) [Cu(R4)]⁺, (c,d) [Cu(R6)]⁺, (e,f) [Cu(R8)]⁺, (g,h) [Cu(R10)]⁺, and (i,j) [Cu(R12)]⁺ before ORR.

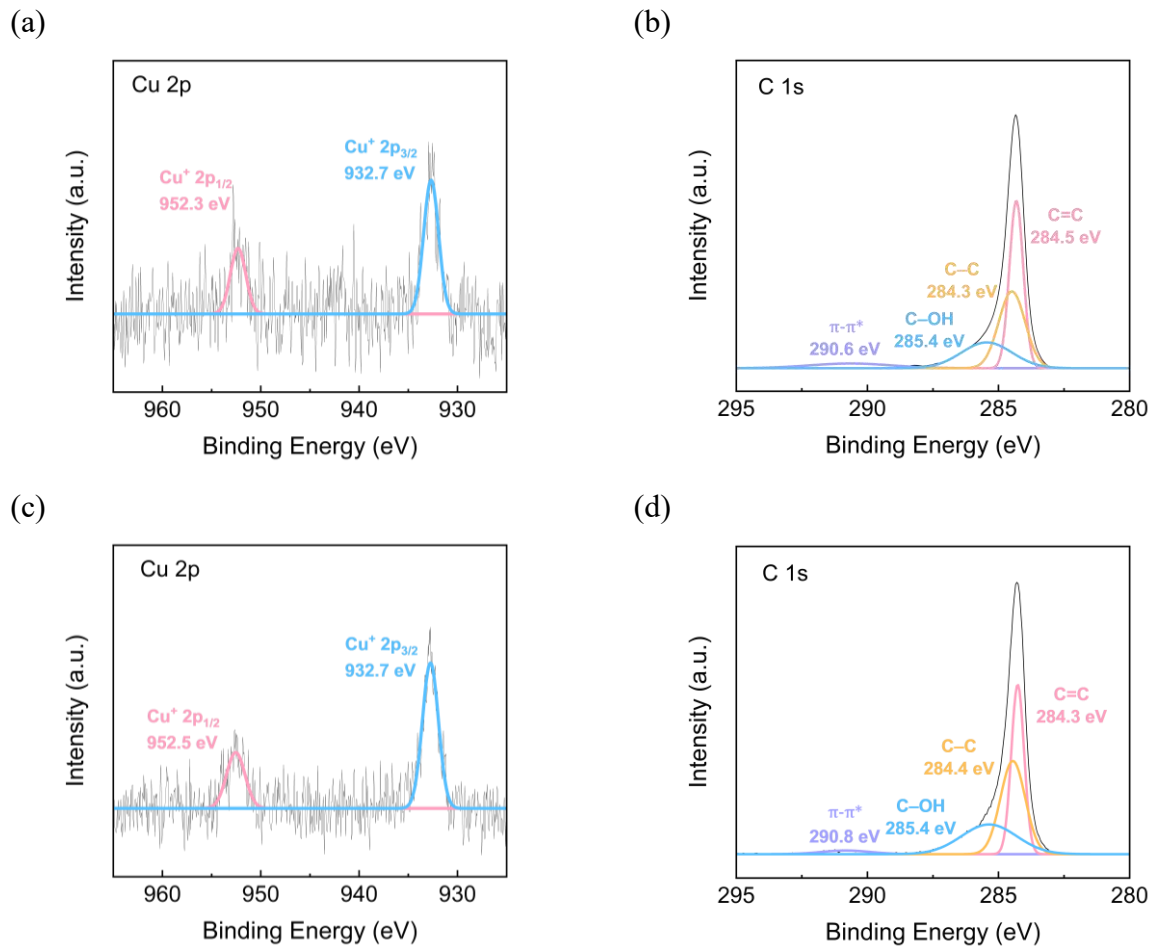
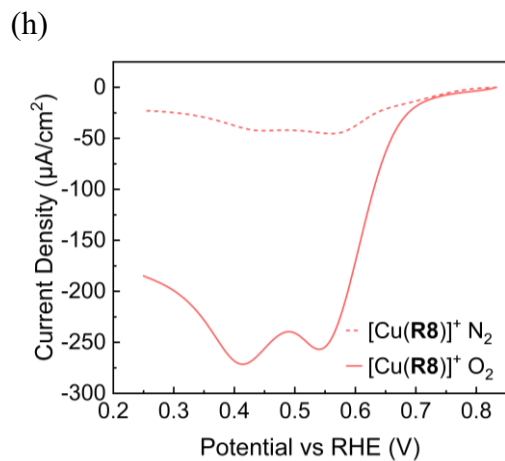
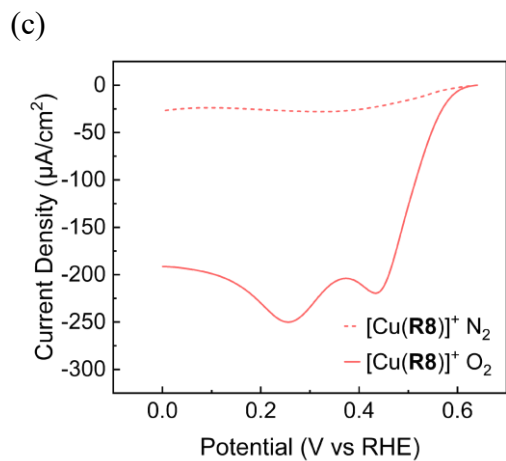
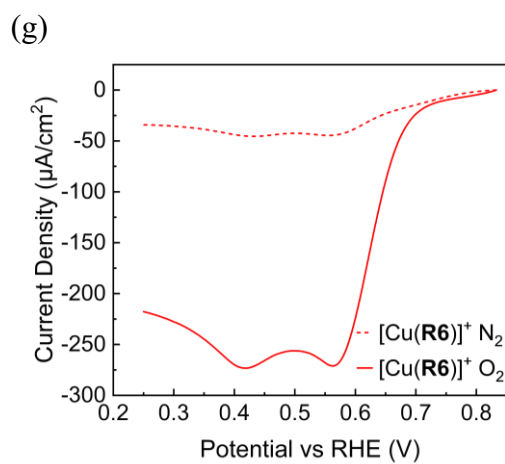
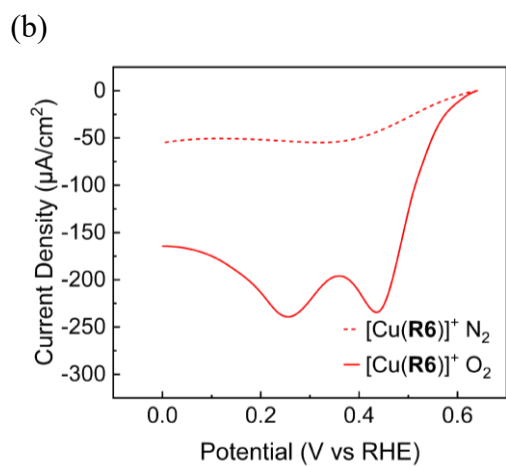
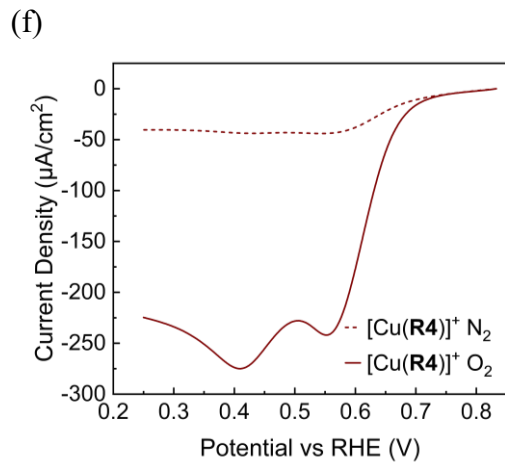
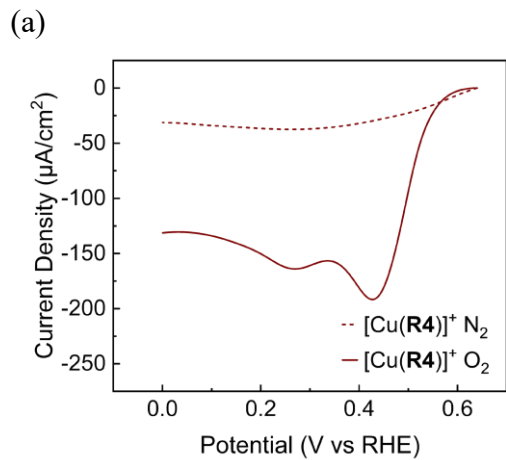


Fig. S42. High-resolution XPS spectra of Cu 2p and C 1s for $[\text{Cu}(\mathbf{R6})]^+$ after 10 consecutive ORR LSV scans (a,b) at pH 4 and (c, d) at pH 7.



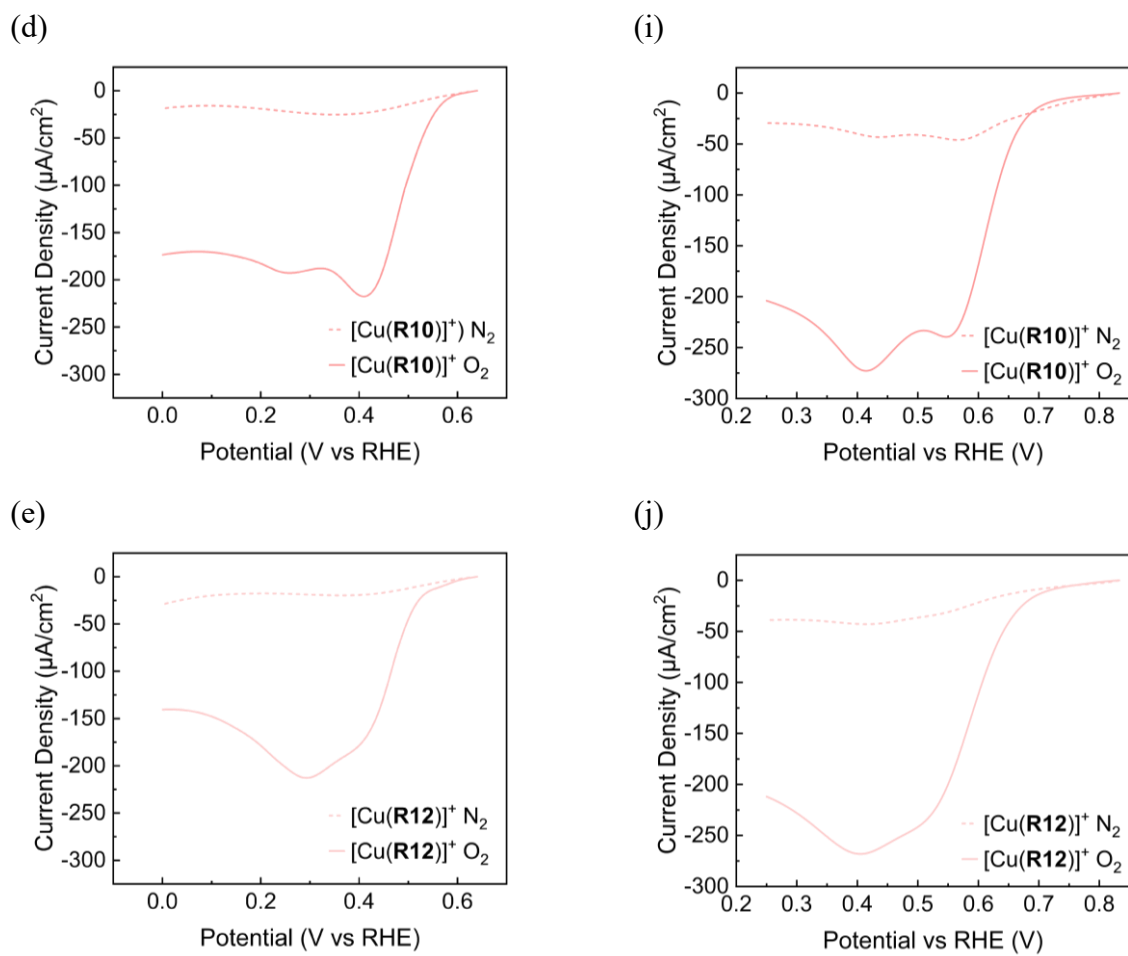


Fig. S43. Cyclic voltammograms of all Cu^I rotaxane complexes under N₂-saturated (dotted) and O₂-saturated (solid) electrolytes in BR buffers at (a-e) pH 4 and (f-j) pH 7.

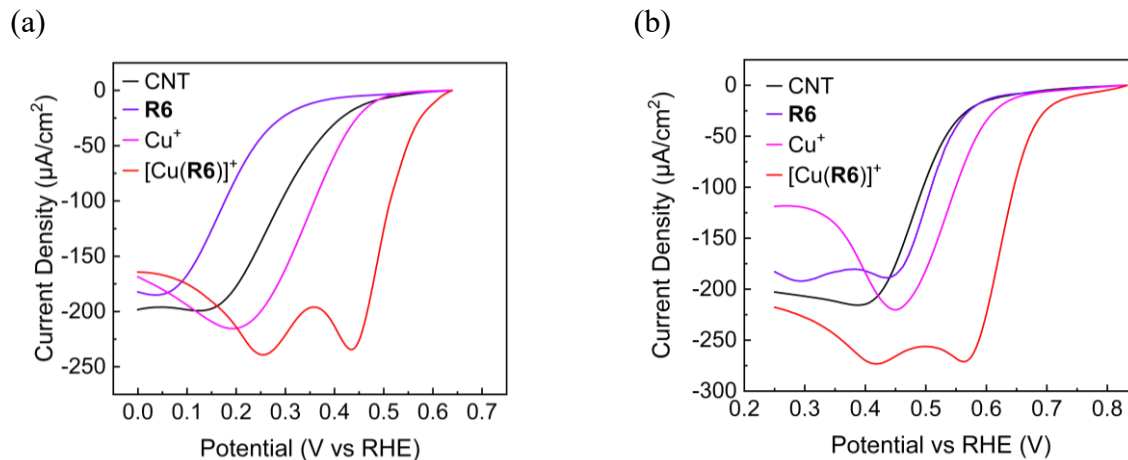


Fig. S44. Linear sweep voltammograms of ORR catalysed by heterogenous **R6** and $[\text{Cu}(\text{R6})]^+$ supported on carbon nanotubes (CNT), homogenous Cu^+ ions (2 mM $[\text{Cu}(\text{MeCN})_4](\text{PF}_6)$), and CNT only in BR buffer at (a) pH 4 and (b) pH 7.

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