

## Supporting Information

### Tandem and mild synthesis of symmetric secondary alcohol toward ionizable lipid analogue to DLin-MC3-DMA

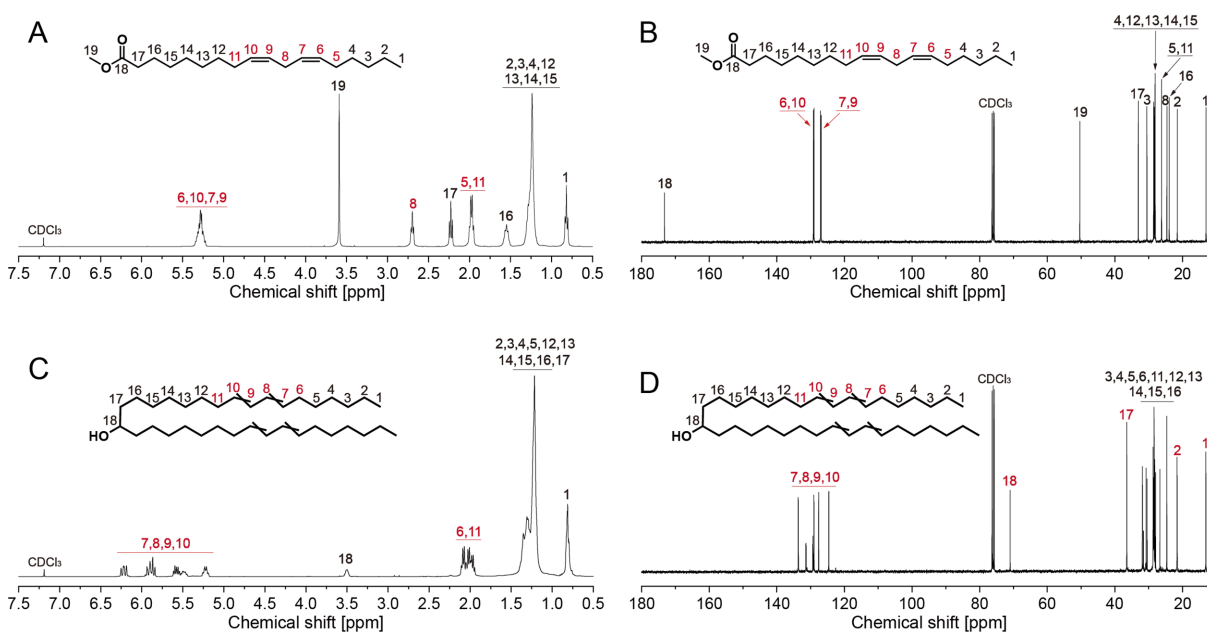
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**Figure S1.** NMR spectra of methyl linoleate and secondary alcohol (A)  $^1\text{H}$  and (B)  $^{13}\text{C}$  NMR spectra of methyl linoleate. (C)  $^1\text{H}$  and (D)  $^{13}\text{C}$  NMR spectra of the secondary alcohol obtained from three-step reaction. First, Claisen condensation reaction mediated by the classical strong base sodium hydride using methyl linoleate as feedstock; subsequent hydrolysis and reduction achieved secondary alcohol.

**Table S1.** Screening of Lewis acid/base pairs for the self-condensation of methyl linoleate<sup>a</sup>.

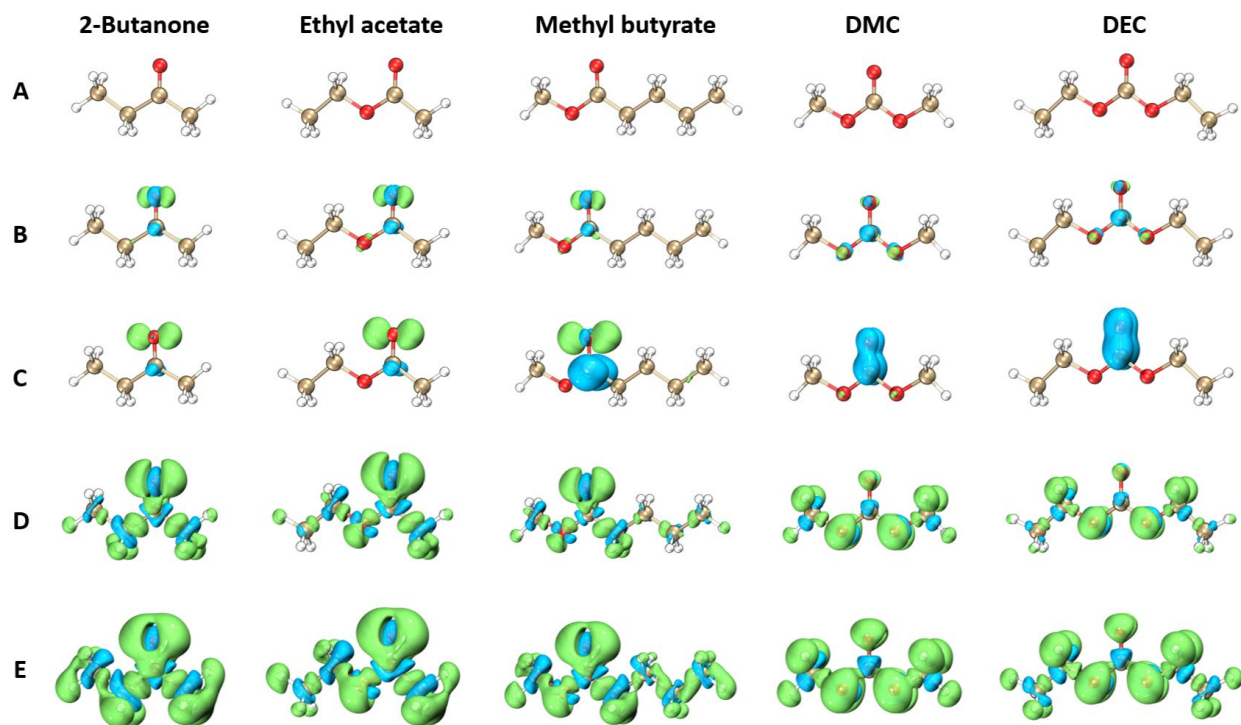
Entry	Lewis acid	Base	Solvent	yield <sup>b</sup> (%)
1	TiCl <sub>4</sub>	TEA	Toluene	57
2	SnCl <sub>4</sub>	TEA	Toluene	0
3	TiCl <sub>4</sub>	TEA	DCM	61
4	SnCl <sub>4</sub>	TEA	DCM	0
5	TiCl <sub>4</sub>	Diisopropylethylamine	DCM	54
6	TiCl <sub>4</sub>	Imidazole	DCM	Trace
7	TiCl <sub>4</sub>	Tributylamine	DCM	55
8	TiCl <sub>4</sub>	TEA	THF	0
9	TiCl <sub>4</sub>	TEA	Ether	19
10	TiCl <sub>4</sub>	TEA	1,2-Dichloroethane	47

<sup>a</sup> The Lewis acid (10 mmol) in solvent (30 ml) was added dropwise over a period of 60 min to a solution of methyl linoleate (10 mmol) and base (20 mmol) at 20 °C, followed by 1 h of reaction. <sup>b</sup> Isolated yield of β-ketoester.

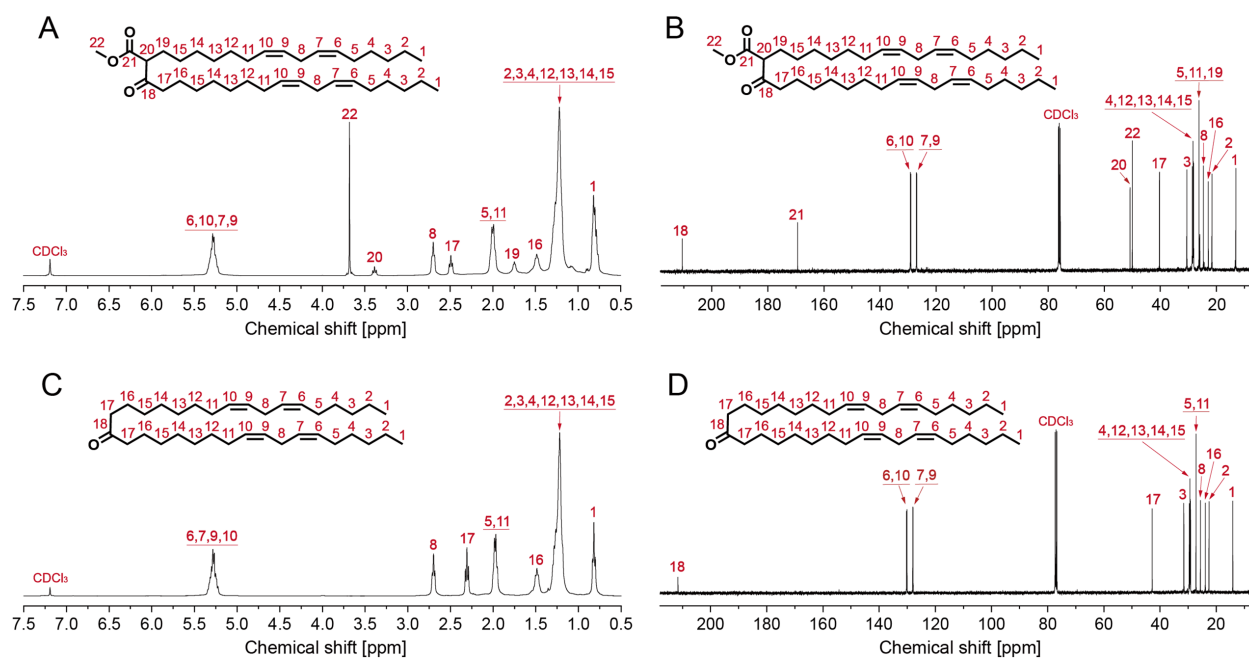
**Table S2.** Screening of conditions for the solvent-free self-Claisen condensation of methyl linoleate.<sup>a</sup>

Entry	TiCl <sub>4</sub> (eq.)	TEA (eq.)	Time <sup>b</sup> (min)	Temperature (°C)	Conversion <sup>c</sup> (%)	Yield <sup>d</sup> (%)
1	2	4	60	20	90	34
2	1.5	3	60	20	88	37
3	1	1.5	60	20	65	45
4	1	1.5	80	20	70	53
5	1	1.5	100	20	69	52
6	1	1.5	100	-20	55	43

<sup>a</sup> General procedure: TiCl<sub>4</sub> was added dropwise to a mixture of TEA and methyl linoleate. The reaction was stirred for another 1 h after complete addition, where methyl linoleate was used as 1 equivalent (eq). <sup>b</sup> The time taken for the dropwise addition of TiCl<sub>4</sub> into the mixture of TEA and methyl linoleate. <sup>c</sup> Conversion of methyl linoleate; determined by recovery. <sup>d</sup> Isolated yield of  $\beta$ -ketoester.



**Figure S2.** Isosurface diagram of Fukui function for solvents. (A) Molecular structure diagram of the different solvent. (B) Dual descriptor (isosurface = -0.03), (C) dual descriptor potential (isosurface = -0.08), (D) Fukui function  $f^{\cdot}$  (isosurface = 0.05) and (E) Local softness  $s^{-}$  (isosurface = 0.05) of the different solvent. All calculations were produced at the B3LYP/6-31G++ (d,p) level. The blue area represents the electrophilic region susceptible to enolate anion attack.



**Figure S3.** NMR spectra of the tandem reaction intermediates. (A)  $^1\text{H}$  and (B)  $^{13}\text{C}$  NMR spectra of the  $\beta$ -ketoester intermediate obtained from the Claisen condensation; (C)  $^1\text{H}$  and (D)  $^{13}\text{C}$  NMR spectra of the ketone intermediate obtained after hydrolysis.

**Table S3.** Mass balance for calculating the E-factor of the tandem process present in this work.<sup>a</sup>

Material	Role	Amount (g)
Methyl linoleate	Feedstock	40
TEA	Reagent	23.3
DEC	Solvent	64
TiCl <sub>4</sub>	Condensation Reagent	33.5
Ethanol	Solvent	550
NaOH	Hydrolysis reagent	32
NaBH <sub>4</sub>	Reduction reagent	15
NH <sub>4</sub> Cl	Quenching reagent	27
Petroleum ether	Extraction solvent	160
Silica gel	Chromatography	130
Ethyl acetate and hexane	Chromatography	1800
Symmetric secondary alcohol	Product	29.2

E-factor for the tandem synthesis of symmetric secondary alcohol was performed according to the following equation:

$$E\text{-factor} = \frac{m_{\text{total waste}} - m_{\text{product}}}{m_{\text{product}}} = 98$$

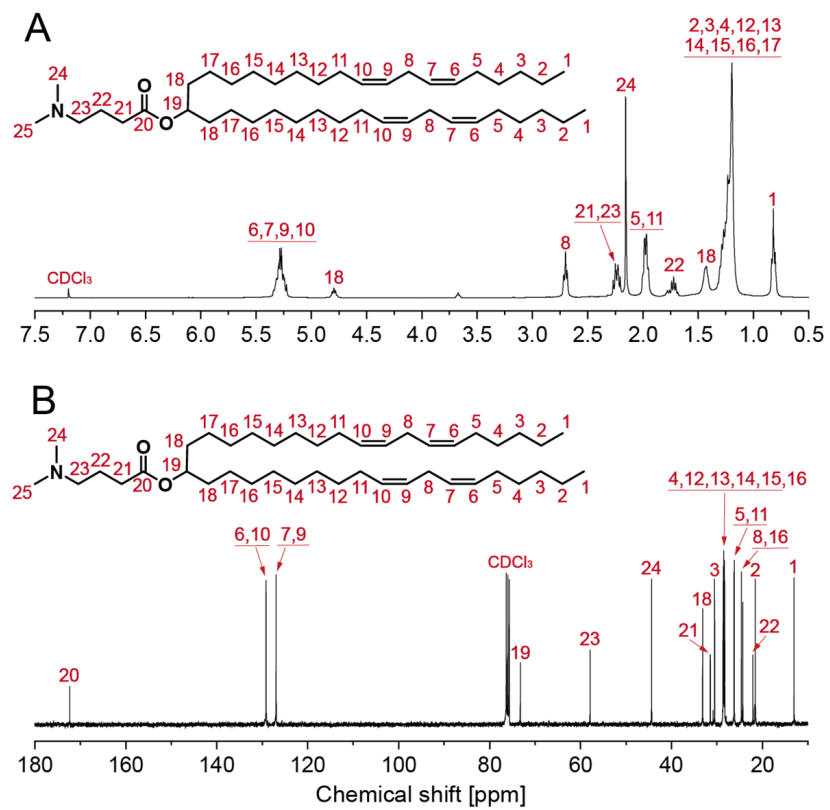
**Table S4.** Mass balance for calculating the E-factor of the Grignard-based route.<sup>a</sup>

Step	Material	Role	Amount (g)
1	Linoleyl alcohol	Feedstock	40
1	DCM	Solvent	720
1	TEA	Reagent	19.7
1	Mesyl chloride	Reagent	18.9
1	NaHCO <sub>3</sub>	Washing	27.1
1	NaCl	Washing	52.6
1	Silica gel	Chromatography	150
1	Hexane/Et <sub>2</sub> O	Chromatography	3000
2	mesylate	Intermediate product	43.1
2	Et <sub>2</sub> O	Solvent	2275
2	MgBr.Et <sub>2</sub> O	Reagent	105
2	K <sub>2</sub> CO <sub>3</sub>	Washing	3.4
2	NaCl	Washing	120
2	Silica gel	Chromatography	150
2	Hexane/Et <sub>2</sub> O	Chromatography	3000
3	Bromide	Intermediate product	43.1
3	Magnesium	Reagent	3.9
3	Et <sub>2</sub> O	Solvent	697
3	Ethyl formate	Reagent	4.4
3	Acetone	Quenching	13
3	H <sub>2</sub> SO <sub>4</sub>	Acidification	54
4	Sodium	Reagent	1.6
4	Methanol	Reagent	257.6
4	Hydrochloric acid	Acidification	7.5
4	NaCl	Washing	57
4	Silica gel	Chromatography	150
4	Hexane/Et <sub>2</sub> O	Chromatography	3000
4	DLin-MeOH	Product	26.7

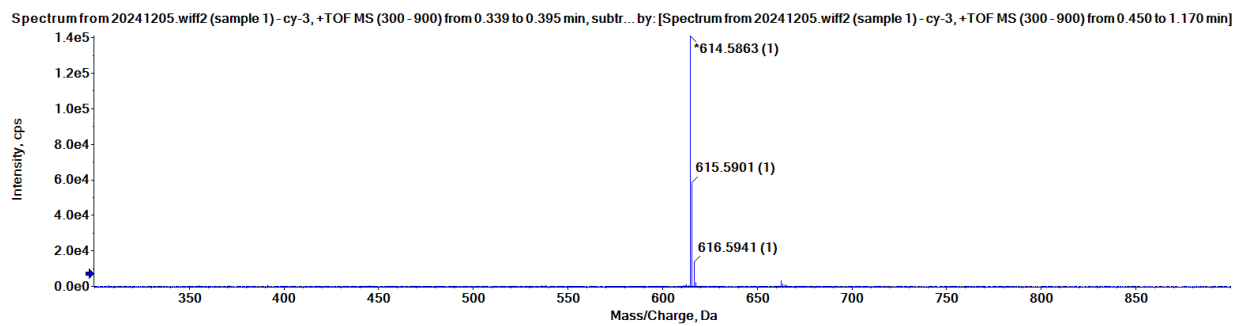
<sup>a</sup> Determined by using the minimum practical amounts for these procedures, as these data were not available in the original literature.

E-factor for the Grignard-based route to synthesize DLin-MeOH was performed according to the following equation:

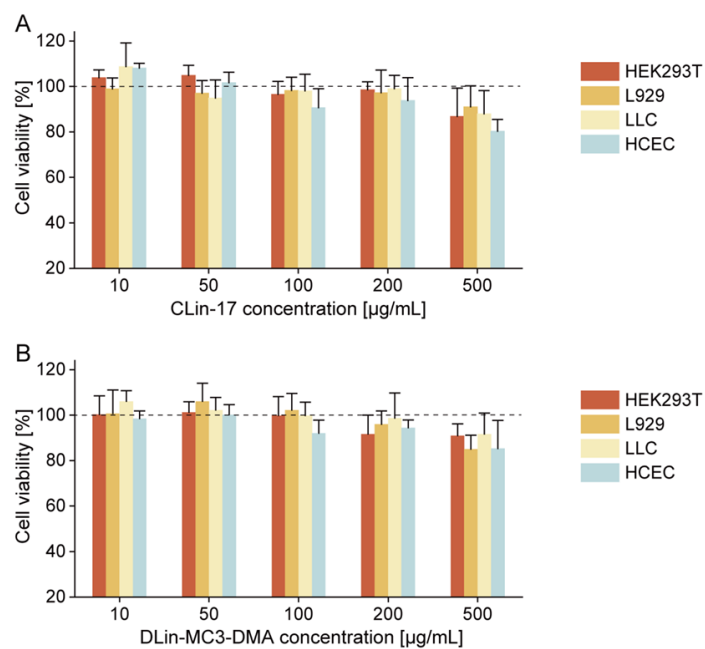
$$E\text{-factor} = \frac{m_{\text{total waste}} - m_{\text{product}}}{m_{\text{product}}} = 524$$



**Figure S4.** (A)  $^1\text{H}$  and (B)  $^{13}\text{C}$  NMR spectra of DLin-MC3-DMA.



**Figure S5.** ESI mass spectrum of CLin-17.



**Figure S6.** *In vitro* cytotoxicity of (A) CLin-17 and (B) DLin-MC3-DMA measured by CCK8 assay. Data were presented as mean  $\pm$  S.D. of biological replicates (n = 5).