

Supplementary Information

An enzyme cascade for green synthesis of chrysanthemol and chrysanthemic acid: key precursors for pyrethrin and pyrethroids

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General information

Ethyl laurate, prenol, and (1*R*,3*R*)-chrysanthemic acid were purchased from Shanghai Haohong Scientific Co., Ltd. (Shanghai, China). Isoprenol, NAD⁺, NADH, methyl tert-butyl ether (MTBE), and (±)-chrysanthemic acid were purchased from Shanghai Macklin Biochemical Technology Co., Ltd. (Shanghai, China). Ethyl chrysanthemate was purchased from Shaoyuan Technology Co., Ltd. (Shanghai, China). Sodium pyruvate was purchased from Bide Pharmatech Co., Ltd. (Shanghai, China). phosphoenolpyruvate (PEP) and ATP were purchased from Shanghai Dibo Biotech Co., Ltd. (Shanghai, China). pyruvate kinase (PK) was purchased from J&K Chemical Co., Ltd. (Shanghai, China). Other chemicals and reagents are purchased from commercial suppliers and used without further purification.

The genes of *SpLicA*, *SyPhoN*, *SmDAGK*, *MtIPK*, *TcADH2*, *TcADH2low*, *TcCDS* were synthesized by SynBio Technologies (Suzhou, China) with codon optimization for *E. coli*. Other genes are cloned from the genome of microbes or the cDNA of plants (*Arabidopsis thaliana*, *Tanacetum cinerariifolium*).

Construction of Plasmids Used in This Study

Detailed cloning steps are described in the Molecular Biology Methods section within the Method section of the article. The details of primers

Linearized pETDuet, pACYCDuet, and pRSFDuet vectors were cloned using primers Duet-XhoI-F and Duet-1stATG-R. Linearized pET28a was cloned using primers pet28aChis-F and pet28a-R (for C-terminal His-tag), or pET28a-F and pET28aNhis-R (for N-terminal His-tag).

The *EcThiM* gene in pRSFDuet-*EcThiM*-CHis was cloned from the *Escherichia coli* genomic library using primers RS-C-Thim-F and RS-C-Thim-R.

The *BsThiM* gene in pRSFDuet-*BsThiM*-CHis was cloned from the *Bacillus subtilis* genomic library using primers RS-C-BsThim-F and RS-C-BsThim-R.

The *SpLicA* gene in pRSFDuet-*SpLicA*-CHis was cloned from synthesized *SpLicA* gene using primers RS-C-LicA-F and RS-C-LicA-R.

The *ScCK* gene in pRSFDuet-*ScCK*-CHis was cloned from the *Saccharomyces cerevisiae* genomic library using primers RS-C-ScCK-F and RS-C-ScCK-R.

The *SfPhoN* gene in pRSFDuet-*SfPhoN*-CHis was cloned from synthesized *SfPhoN* gene using primers RS-C-PhoN-F and RS-C-PhoN-R.

The *BceThiM* gene in pET28a-*BceThiM*-CHis was cloned from the *Bacillus cereus* genomic library using primers ET-C-Bce-F and ET-C-Bce-R.

The *BliThiM* gene in pET28a-*BliThiM*-CHis was cloned from the *Bacillus licheniformis* genomic library using primers ET-C-Bli-F and ET-C-Bli-R.

The *CbuThiM* gene in pET28a-*CbuThiM*-CHis was cloned from the *Clostridium butyricum* genomic library using primers ET-C-Cbu-F and ET-C-Cbu-R.

The *EhuThiM* gene in pET28a-*EhuThiM*-CHis was cloned from the *Enterobacter huaxiensis* genomic library using primers ET-C-Ehu-F and ET-C-Ehu-R.

The *GstThiM* gene in pET28a-*GstThiM*-CHis was cloned from the *Geobacillus stearothermophilus* genomic library using primers ET-C-Gst-F and ET-C-Gst-R.

The *SmaThiM* gene in pET28a-*SmaThiM*-CHis was cloned from the *Serratia marcescens* genomic library using primers ET-C-Sma-F and ET-C-Sma-R.

The *StyThiM* gene in pET28a-*StyThiM*-CHis was cloned from the *Salmonella typhimurium* genomic library using primers ET-C-Sty-F and ET-C-Sty-R.

The *MtIPK* gene in pETDuet-IPK was cloned from the synthesized *MtIPK* gene using primers PET-Ipk-F and KLD-IPK-R.

The *SmaThiM* gene in pETDuet-*SmaThiM*-IPK was cloned from pET28a-*SmaThiM*-CHis using primers RS-Smathim-F and RBS-Smathim-R. The IPK gene was cloned from pETDuet-IPK using primers IDI-IPK-RBS-F and Duet-1stATG-R.

The *BsThiM* gene in pETDuet-*BsThiM*-IPK was cloned from pRSFDuet-*BsThiM*-CHis using primers RS-Bsthim-F and RBS-Bsthim-R. The IPK gene was cloned from pETDuet-IPK using primers IDI-IPK-RBS-F and Duet-1stATG-R.

The *StyThiM* gene in pETDuet-*StyThiM*-IPK was cloned from pET28a-*StyThiM*-CHis using primers RS-Stythim-F and RBS-Stythim-R. The IPK gene was cloned from pETDuet-IPK using primers IDI-IPK-RBS-F and Duet-1stATG-R.

The *SmDAGK* gene in pETDuet-*SmDAGK*-IPK was cloned from synthesized *SmDAGK* gene using primers Duet-DAGKop-F and Duet-DAGKop-IPK-R. The IPK gene was cloned from pETDuet-*SmaThiM*-IPK using primers IK-IPK-F and Duet-1stATG-R.

The *BceThiM* gene in pETDuet-*BceThiM*-IPK was cloned from pET28a-*BceThiM*-CHis using primers Duet-Bce-F and Duet-Bce-IPK-R. The IPK gene was cloned from pETDuet-*SmaThiM*-IPK using primers IK-IPK-F and Duet-1stATG-R.

The *BliThiM* gene in pETDuet-*BliThiM*-IPK was cloned from pET28a-*BliThiM*-CHis using primers Duet-Bli-F and Duet-Bli-IPK-R. The IPK gene was cloned from pETDuet-*SmaThiM*-IPK using primers IK-IPK-F and Duet-1stATG-R.

The *CbuThiM* gene in pETDuet-*CbuThiM*-IPK was cloned from pET28a-*CbuThiM*-CHis using primers Duet-Cbu-F and Duet-Cbu-IPK-R. The IPK gene was cloned from pETDuet-*SmaThiM*-IPK using primers IK-IPK-F and Duet-1stATG-R.

The *EhuThiM* gene in pETDuet-*EhuThiM*-IPK was cloned from pET28a-*EhuThiM*-CHis using primers Duet-Ehu-F and Duet-Ehu-IPK-R. The IPK gene was cloned from pETDuet-*SmaThiM*-IPK using primers IK-IPK-F and Duet-1stATG-R.

The *GstThiM* gene in pETDuet-*GstThiM*-IPK was cloned from pET28a-*GstThiM*-CHis using primers Duet-Gst-F and Duet-Gst-IPK-R. The IPK gene was cloned from pETDuet-*SmaThiM*-IPK using primers IK-IPK-F and Duet-1stATG-R.

The *EcThiM* gene in pETDuet-*EcThiM*-IPK was cloned from pRSFDuet-*EcThiM*-CHis using primers Duet-Ec-F and Duet-Ec-IPK-R. The IPK gene was cloned from pETDuet-*SmaThiM*-IPK using primers IK-IPK-F and Duet-1stATG-R.

The *SfPhoN* gene in pETDuet-*SfPhoN*-IPK was cloned from pRSFDuet-*SfPhoN*-CHis using primers Duet-PhoN-F and Duet-PhoN-IPK-R. The IPK gene was cloned from pETDuet-*Sma*ThiM-IPK using primers IK-IPK-F and Duet-1stATG-R.

The *SpLicA* gene in pETDuet-*SpLicA*-IPK was cloned from pRSFDuet-*SpLicA*-CHis using primers Duet-LicA-F and Duet-LicA-IPK-R. The IPK gene was cloned from pETDuet-*Sma*ThiM-IPK using primers IK-IPK-F and Duet-1stATG-R.

The *ScCK* gene in pETDuet-*ScCK*-IPK was cloned from pRSFDuet-*ScCK*-CHis using primers Duet-ScCK-F and Duet-ScCK-IPK-R. The IPK gene was cloned from pETDuet-*Sma*ThiM-IPK using primers IK-IPK-F and Duet-1stATG-R.

The *TcALDH1* gene in pRSFDuet-*TcALDH1* was cloned from the *Tanacetum cinerariifolium* cDNA library using primers RS-ALDH3-F and RS-ALDH3-R.

The *TcADH2* gene in pRSFDuet-*TcADH2*-*TcALDH1* was cloned from synthesized *TcADH2* gene using primers RS-ADH-F and RS-ADH-ALDH-R; The *TcALDH1* gene was cloned from pRSFDuet-*TcALDH1* using primers RSALDH-F and Duet-1stATG-R.

The *TcADH2* gene in pET28a-NHis-*TcADH2* was cloned from synthesized *TcADH2* gene using primers ET-His-ADH-F and ET-ADH-R.

The *TcADH2low* gene in pRSFDuet-*TcADH2low*-*TcALDH1* was cloned from synthesized *TcADH2low* gene using primers RS-ADH3-F and RS-ADH3-R. The *TcALDH1* gene was cloned from pRSFDuet-*TcADH2*-*TcALDH1* using primers RSALDH4-F and Duet-1stATG-R.

The *TcNudix* gene in pRSFDuet-*TcNudix* was cloned from the *Tanacetum cinerariifolium* cDNA library using primers RS-Nudix-F and RS-Nudix-R.

The *TcCDS* gene in pRSFDuet-*TcCDS* was cloned from synthesized *TcCDS* gene using primers RS-CDS-F2 and RS-CDS-R2.

The *TcNudix* gene in pRSFDuet-*TcNudix*-*TcCDS* was cloned from pRSFDuet-*TcNudix* using primers RS-Nudix2-F and RS-Nudix2-R; The *TcCDS* gene was cloned from pRSFDuet-*TcCDS* using primers RSCDS-F and Duet-1stATG-R.

The *TcCDS* gene in pRSFDuet-*TcCDS*-*TcNudix* was cloned from pRSFDuet-*TcCDS* using primers Duet-1stATG-F and CDS-Nudix-R; The *TcNudix* gene was cloned from pRSFDuet-*TcNudix* using primers CDS-Nudix-F and Duet-1stATG-R.

The *TcNudix*-*TcCDS* gene in pACYCDuet-*TcNudix*-*TcCDS* and pCDFDuet-*TcNudix*-*TcCDS* was cloned from pRSFDuet-*TcNudix*-*TcCDS* using primers Duet-1stATG-F and Duet-XhoI-R.

The *TcCDS*-*TcNudix* gene in pACYCDuet-*TcCDS*-*TcNudix* and pCDFDuet-*TcCDS*-*TcNudix* was cloned from pRSFDuet-*TcCDS*-*TcNudix* using primers Duet-1stATG-F and Duet-XhoI-R.

The *TcADH2low-TcALDH1* gene in pRSFDuet-Sumo*TcADH2low-TcALDH1* was cloned from pRSFDuet-*TcADH2low-TcALDH1* using primers RSAAM-ADHm-F and RSAAM-R. The Sumo tag gene was cloned from commercially available plasmid using primers RSAAM-SUMO-F and RSAAM-SUMO-R.

The *TcADH2low-TcALDH1* gene in pRSFDuet-Gst*TcADH2low-TcALDH1* was cloned from pRSFDuet-*TcADH2low-TcALDH1* using primers RSAAM-ADHm-F and RSAAM-R. The Gst tag gene was cloned from commercially available plasmid using primers RSAAM-Gst-F and RSAAM-Gst-R.

The *TcADH2low-TcALDH1* gene in pRSFDuet-Trx*TcADH2low-TcALDH1* was cloned from pRSFDuet-*TcADH2low-TcALDH1* using primers RSAAM-ADHm-F and RSAAM-R. The Trx tag gene was cloned from the *Escherichia coli* genomic library using primers RSAAM-Trx-F and RSAAM-Trx-R.

The *TcADH2-TcALDH1* gene in pRSFDuet-Sumo*TcADH2-TcALDH1* was cloned from pRSFDuet-*TcADH2-TcALDH1* using primers RSAAS-ADHs-F and RSAAM-R. The Sumo tag gene was cloned from commercially available plasmid using primers RSAAM-SUMO-F and RSAAS-SUMO-R.

The *TcADH2-TcALDH1* gene in pRSFDuet-Gst*TcADH2-TcALDH1* was cloned from pRSFDuet-*TcADH2-TcALDH1* using primers RSAAS-ADHs-F and RSAAM-R. The Gst tag gene was cloned from commercially available plasmid using primers RSAAM-Gst-F and RSAAS-Gst-R.

The *TcADH2-TcALDH1* gene in pRSFDuet-Trx*TcADH2-TcALDH1* was cloned from pRSFDuet-*TcADH2-TcALDH1* using primers RSAAS-ADHs-F and RSAAM-R. The Trx tag gene was cloned from the *Escherichia coli* genomic library using primers RSAAM-Trx-F and RSAAS-Trx-R.

The trigger factor gene in pCDFDuet-TriggerFactor was cloned from the *Escherichia coli* genomic library using primers Duet-TFactor-F and Duet-TFactor-R.

The GroES-GroEL gene in pCDFDuet-GroES-GroEL was cloned from the *Escherichia coli* genomic library using primers Duet-GroESL-F and Duet-GroESL-R.

The DnaK-DnaJ gene in pCDFDuet-DnaK-DnaJ was cloned from the *Escherichia coli* genomic library using primers Duet-DnaKJ-F and Duet-DnaKJ-R.

The ClpB gene in pCDFDuet-ClpB was cloned from the *Escherichia coli* genomic library using primers Duet-clpB-F and Duet-clpB-R.

pACYCDuet-*TcNudix*-(GSG)₃-*TcCDS* was generated from pACYCDuet-*TcNudix-TcCDS* by inverse PCR using primers (GSG)₃-CDS-F and Nudix-R.

pACYCDuet-*TcNudix*-(PT)₄P-*TcCDS* was generated from pACYCDuet-*TcNudix-TcCDS* by inverse PCR using primers (PT)₄P-CDS-F and *Nudix*-R.

pACYCDuet-*TcCDS*-(GSG)₃-*TcNudix* was generated from pACYCDuet-*TcCDS-TcNudix* by inverse PCR using primers (GSG)₃-CDS-F and CDS-R.

pACYCDuet-*TcCDS*-(PT)₄P-*TcNudix* was generated from pACYCDuet-*TcCDS-TcNudix* by inverse PCR using primers (PT)₄P-*Nudix*-F and CDS-R.

The *TcCDS* gene in pET28a-NHis-*TcCDS* was cloned from pACYCDuet-*TcNudix-TcCDS* using primers 28a-NHis-CDS-F and 28a-CDS-R.

The *TcNudix* gene in pET28a-*TcNudix*-CHis was cloned from pACYCDuet-*TcNudix-TcCDS* using primers 28a-*Nudix*-F and 28a-*Nudix*-CHis-R.

The *MtIPK* gene in pET28a-NHis-IPK was cloned from pETDuet-IPK using primers 28a-NHis-IPK-F and 28a-IPK-R.

The *TcCDS* gene in pET28a-NHis-*TcCDS* was cloned from pACYCDuet-*TcNudix-TcCDS* using primers 28a-NHis-CDS-F and 28a-CDS-R.

The *TcCDS*-(PT)₄P-*TcNudix* gene in pRSFDuet-Sumo*TcCDS*-(PT)₄P-*TcNudix* was cloned from pACYCDuet-*TcCDS*-(PT)₄P-*TcNudix* using primers TAG-CDS-F and Duet-*Nudix*-R. The Sumo tag gene was cloned from pRSFDuet-Sumo*TcADH2-TcALDH1* using primers SUMO-CDS-R and Duet-XhoI-F.

The *TcCDS*-(PT)₄P-*TcNudix* gene in pRSFDuet-Gst*TcCDS*-(PT)₄P-*TcNudix* was cloned from pACYCDuet-*TcCDS*-(PT)₄P-*TcNudix* using primers TAG-CDS-F and Duet-*Nudix*-R. The Gst tag gene was cloned from pRSFDuet-Gst*TcADH2-TcALDH1* using primers GST-CDS-R and Duet-XhoI-F.

The Sumo*TcCDS*-(PT)₄P-*TcNudix* gene in pACYCDuet-Sumo*TcCDS*-(PT)₄P-*TcNudix* was cloned from pRSFDuet-Sumo*TcCDS*-(PT)₄P-*TcNudix* using primers Duet-1stATG-F and Duet-XhoI-R.

The Gst*TcCDS*-(PT)₄P-*TcNudix* gene in pACYCDuet-Gst*TcCDS*-(PT)₄P-*TcNudix* was cloned from pRSFDuet-Gst*TcCDS*-(PT)₄P-*TcNudix* using primers Duet-1stATG-F and Duet-XhoI-R.

The *TcNudix* gene in pACYCDuet-*TcNudix*-Sumo*TcCDS* was cloned from pACYCDuet-*TcNudix-TcCDS* using primers Duet-XhoI-F2 and *Nudix*-RBS-R. The Sumo*TcCDS* gene was cloned from pRSFDuet-Sumo*TcCDS*-(PT)₄P-*TcNudix* using primers RBS-SUMO-F and Duet-CDS-R.

The *AtFKI* gene in pETDuet-*AtFKI*-IPK was cloned from the *Arabidopsis thaliana* cDNA library using primers Duet-ATFKI-F and ATFKI-IPK-R. The IPK gene was cloned from pETDuet-*SmaThiM*-IPK using primers IK-IPK-F and Duet-1stATG-R.

Gene sequence

> Sequences encoding *SmDAGK*¹

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ATGCCGATGGATCTGCGCGACAACAAACAGTCCCAGAAAAAATGGAAAAACCGC
ACCCTGACCAGTAGTCTGGAATTTGCACTGACCGGTATCTTCACCGCGTTCAAAGA
AGAGCGCAACATGAAAAAACACGCGGTTAGCGCTCTGCTGGCGGTGATTGCGGGT
CTGGTTTTTAAAGTCTCCGTCATTGAGTGGCTGTTTCTGCTGCTGAGCATCTTCCTG
GTCATCACCTTCGAGATCGTCAACAGCGCGATTGAAAACGTCGTCGATCTGGCAA
GCGATTACTTTCAGCATGCTGGCGAAAAACGCCAAAGATATGGCAGCAGGTGC
GGTTCTGGTTATTAGCGGTTTTGACGACTGACCGGTCTGATCATCTTCCTGCTGAA
AATCTGGTTCCTGCTGTTCCACTAA
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> Sequences encoding *BsThiM*

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ATGGATGCACAATCAGCAGCAAAATGTCTTACGGCTGTCCGCCGGCATAGCCCCT
GGTGCATAGCATAACCAACAATGTCGTAACGAATTCACAGCAAACGGCCTGCTCG
CGCTCGGCGCATCGCCCGTTATGGCGTACGCAAAAGAAGAGGTCGCCGATATGGC
GAAAATTGCGGGTCACTCGTTTTAAATATCGGAACACTGAGCAAGGAGTCAGTC
GAAGCGATGATCATCGCGGGAAAATCAGCTAATGAACATGGCGTTCCCGTCATTCT
TGATCCTGTCCGTGCCGGAGCAACACCGTTCCGCACTGAATCGGCACGTGACATC
ATTCGTGAGGTGCGCCTTGCTGCAATCAGAGGAAATGCGGCGGAAATTGCCCATAC
CGTCGGCGTGACCGATTGGCTGATCAAAGGTGTTGATGCGGGTGAAGGTGGAGGC
GACATCATCCGGCTGGCTCAGCAGGCGGCACAAAAGCTAAACACGGTCATTGCGA
TAACTGGTGAAGTTGATGTCATAGCCGACACGTACATGTATACACCCTTCATAAC
GGCCACAAGCTGCTGACAAAAGTGACAGGCGCCGGTTGCCTGCTGACTTCCGTCG
TCGGTGCGTTTTGCGCTGTGGAAGAAAATCCATTGTTTGCTGCTATTGCGGCCATTT
CTTCGTATGGGGTTCGCCGCTCAGCTTGCCGCACAGCAGACGGCTGACAAAGGCC
TGGAAGCTTTCAGATTGAATTGCTGAACAAGCTTTCAACTGTTACTGAACAAGAC
GTCCAAGAATGGGCGACTATAGAAAGGGTGA CTGTCTCATGA.
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> Sequences encoding *EcThiM*²

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ATGCAAGTCGACCTGCTGGGTTCAAGCGCAATCTGCGCACGCGTTACACCTTTTTCA
CCAACATTCCCCTCTTGTGCACTGCATGACCAATGATGTGGTGCAAACCTTTACCG
CCAATACCTTGCTGGCGCTCGGTGCATCGCCAGCGATGGTTATCGAAACCGAAGAG
GCCAGTCAGTTTGCGGCTATCGCCAGTGCCCTTGTTGATTAACGTTGGCACACTGAC
GCAGCCACGCGCTCAGGCGATGCGTGCTGCCGTTGAGCAAGCAAAAAGCTCTCA
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AACACCCTGGACGCTTGATCCAGTAGCGGTGGGTGCGCTCGATTATCGCCGCCATT
TTTGTCAATGAACTTTTATCTTTTAAACCGGCAGCGATACGTGGTAATGCTTCGGAAA
TCATGGCATTAGCTGGCATTGCTAATGGCGGACGGGGAGTGGATAACCACTGACGCC
GCAGCTAACGCGATACCCGCTGCACAAACACTGGCACGGGAAACTGGCGCAATCG
TCGTGGTCACTGGCGAGATGGATTATGTTACCGATGGACATCGTATCATTGGTATTC
ACGGTGGTGATCCGTTAATGACCAAAGTGGTAGGAACTGGCTGTGCATTATCGGGCG
GTTGTGCTGCCTGCTGTGCGTTACCAGGCGATACGCTGGAAAATGTCGCATCTGC
CTGTCACTGGATGAAACAAGCCGGAGAACGCGCAGTCGCCAGAAGCGAGGGGCC
AGGCAGTTTTGTTCCACATTCCTTGATGCGCTCTGGCAATTGACGCAGGAGGTGC
AGGCATAA

> Sequences encoding *SfPhoN*³

ATGTCTATTCCACCAGGTAACGACGTAACACTACAAAGCCAGATTTATATTATTTGACT
AACGACAACGCTATTGACTCATTAGCCCTTCTTCCGCCGCCACCTCAGATCGGTTTC
CATTGCCTTCCTTAACGACCAGGCCATGTATGAAAAGGGTCGCTTGTTGCGGAACA
CTGAGCGGGGTAAGCTCGCTGCTGAGGACGCTAACCTGTCAAGCGGCGGGGTGG
CGAACGTCTTTAGCGCCGCTTTTCGGCAGCCCAATCACTGCCAAGGATTCCCCAGA
GCTTCACAAGTTACTGACTAATATGATTGAAGATGCAGGCGATTTGGCGACCCGCT
CGGCAAAAGAGTACTACATGCGTATCCGCCCATTCGCTTTTTTACGGTGTGAGTACC
TGTAACACTAAGGAGCAAGACACATTGAGCCGTAATGGCTCTTATCCATCAGGCCA
TACTAGTATTGGTTGGGCGACAGCCTTGGTGTGTCCGAAATCAATCCGGCACGTC
AAGACACAATTCTCAAGCGTGGCTACGAATTGGGGGACTCTCGCGTAATCTGTGG
GTATCACTGGCAGAGCGATGTAGATGCCGCCCGTATTGTTGGTTCAGCTATCGTGG
CGACATTACATTCTAATCCAGTTTTCCAGGCACAATTACAAAAGCAAAAGACGAA
TTTGCTAATAACCAGAAGAAGTAA

> Sequences encoding *ScCK*⁴

ATGGTACAAGAATCACGTCCAGGGAGTGTAAGAAGTTACTCGGTCGGTTACCAAG
CAAGGTCCAGATCGAGTTCTCAAAGAAGACATTCGTTAACACGCCAACGTTCCCTC
GCAAAGACTGATTAGAACCATCAGTATCGAGTCTGATGTGTCTAATATTACTGACG
ATGACGATTTGAGAGCTGTCAATGAGGGAGTAGCGGGTGTGCAACTGGACGTCTC
TGAAACCGCAAATAAGGGACCAAGAAGAGCATCAGCAACTGATGTCACAGATAGT
TTGGGTTCGACTTCGTCCGAATATATTGAGATTCCTTTGTAAAGGAAACATTGGAT
GCAAGTTTACCTTCGATTATCTGAAGCAGGACATATTAATCTCATTACAGAGTTTG
AAGATATCCAAATGGTATAACAACAAGAAAATCCAACCGGTAGCACAAGATATGAA
CTTAGTCAAGATCTCTGGTGCATGACAAACGCAATTTTCAAAGTTGAATACCCTA

AGTTACCATCGTTGCTATTGAGAATATACGGACCGAATATTGATAATATCATTGACAG
GGAATATGAATTGCAGATTTTGGCTAGGCTTTCATTGAAAAATATAGGTCCTTCCCT
TTACGGCTGTTTTGTAAACGGTAGATTTGAGCAGTTTCTGGAGAATTCTAAGACTT
TAACAAAAGACGACATTAGAACTGGAAGAACTCTCAAAGGATTGCAAGGAGAA
TGAAGGAGTTACATGTAGGTGTTCCCTCTCTTGAGTTCAGAAAGGAAGAACGGGTC
GGCTTGTGGCAAAAGATTAACCAGTGGTTGCGCACGATTGAAAAAGTCGACCAA
TGGGTGGGGGATCCTAAAAACATTGAAAACTCTTTATTATGTGAGAATTGGTCCAA
GTTTATGGATATTGTTCGATAGATATCACAAGTGGCTTATTTCTCAAGAACAGGGTAT
AGAGCAAGTCAACAAAAATCTTATATTCTGCCATAATGATGCCCAATACGGCAATTT
ACTTTTCACTGCTCCTGTGATGAACACACCGAGCCTATACTGCACCTTCGTCTA
CATCATTGACTTCCCAATCAAGTTCCTTATTTCCCTTCGAGCTCCAATGTCATTGTAG
ATGATATAATCAACCCGCCAAAGCAGGAGCAAAGCCAAGATTCCAAATTGGTCGTC
ATTGATTTTGAATATGCAGGTGCCAATCCCGCCGCATATGATTTAGCGAATCATCTTT
CCGAGTGGATGTATGATTACAACAATGCTAAGGCCCCACATCAGTGCCACGCTGAT
AGATATCCCGATAAAGAACAGGTTTTGAATTTCTTATACTCTTATGTTTCGCATCTAA
GGGTGGTGCTAAGGAACCCATAGATGAAGAGGTTCAAAGACTCTATAAGTCAAT
CATTCAATGGAGACCCACTGTACAACCTATTTTGGTCGCTCTGGGCCATCCTACAAA
GTGGTAAATTAGAGAAAAAAGAAGCCTCCACTGCCATCACTAGAGAAGAAATTGG
ACCCAATGGAAAAAATATATCATCAAGACTGAACCCGAATCCCCTGAAGAAGAC
TTTGTGAAAATGACGACGAGCCTGAAGCTGGCGTCAGCATTGACACGTTTCGATTA
TATGGCTTATGGTCGTGACAAGATTGCGGTCTTTTGGGGCGACCTTATTGGCTTAGG
CATAATCACCGAAGAAGAATGCAAAAATTTAGCTCTTTCAAGTTCCTCGATACTA
GTTATTTGTAA

> Sequences encoding *SpLicA*

ATGGAAGATCATCAAAGAGAAGATTAGCAGCCTGCTGAGCCAAGAAGAAGAA
GTTCTGAGCGTTGAACAGTTAGGTGGTATGACCAATCAGAATTATCTGGCCAAAAC
CACCAACAAACAGTACATCGTTAAATTTCTTTGGCAAAGGCACCGAGAACTGATT
AATCGTCAGGATGAAAAGTACAACCTGGAAGTCTGAAAGATTTAGGTCTGGATG
TGAAAACTACCTGTTTCGATATTGAAGCCGGTATCAAAGTGAACGAATATATTGAA
AGCGCGATTACCCTGGATAGCACCAGCATTAAAACCAAATTCGATAAAATCGCACC
GATCCTGCAAACCATTACATACCAGCGCAAAGAAGTGCCTGGTGAATTTGCACCGT
TTGAAGAGATCAAAAAATACGAAAGCCTGATCGAAGAACAATCCCGTATGCAAA
TTATGAAAGCGTTCGTAATGCAGTGTTGAGCCTGGAAAAACGTCTGGCCGATCTGG
GTGTTGATCGTAAAAGCTGTCATATTGATCTGGTGCCGAAAACCTTATTGAAAGT
CCGCAGGGTCTGCTGTATCTGATTGATTGGGAATATAGCAGCATGAATGATCCGATG
TGGGATTTAGCAGCACTGTTTCTGGAAAGCGAATTTACCAGTCAAGAAGAGGAAA

CCTTTCTGAGCCATTATGAGAGCGATCAGACACCGGTTAGCCATGAAAAAATTGCC
ATCTACAAAATCCTGCAAGACACCATTTGGAGCCTGTGGACGTTTATAAAGAAGA
ACAGGGCGAAGATTTTGGTGATTATGGTGTTAATCGTTATCAGCGTGCAGTTAAAG
GTCTGGCAAGCTATGGTGGTAGTGATGAAAAATAA

> Sequences encoding *Bce*ThiM

ATGAATATGAAAGAAATTAGTAAAGTAGTGGATTTGGTGAGAGAATCTAATCCGCT
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> Sequences encoding *Cbu*ThiM

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> Sequences encoding *MiPK*

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> Highly codon-optimized sequences encoding *TcADH2*

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> Moderately codon-optimized sequences encoding *TcADH2* (*TcADH2*_{low})

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Supporting Tables

Table S1 Kinetic parameters of *TcADH2* catalytic oxidation of **1** and **3**

Substrate	k_{cat} (s ⁻¹)	K_m (μM)	K_i (μM)	k_{cat}/K_m (μM ⁻¹ ·s ⁻¹)
1	6.21×10^{-1}	7.56×10^2	5.86×10^3	8.21×10^{-4}
3	2.73×10^{-1}	1.07×10^2	1.55×10^3	2.56×10^{-3}

Table S2 Kinetic parameters of *EcThiM* and *SmaThiM* catalytic phosphorylation of **1**

Enzyme	k_{cat} (s ⁻¹)	K_m (mM)	k_{cat}/K_m (μM ⁻¹ ·s ⁻¹)
<i>EcThiM</i>	5.70×10^{-2}	10.61	5.37×10^{-6}
<i>SmaThiM</i>	1.21×10^{-1}	13.07	9.26×10^{-6}

Supporting Figures

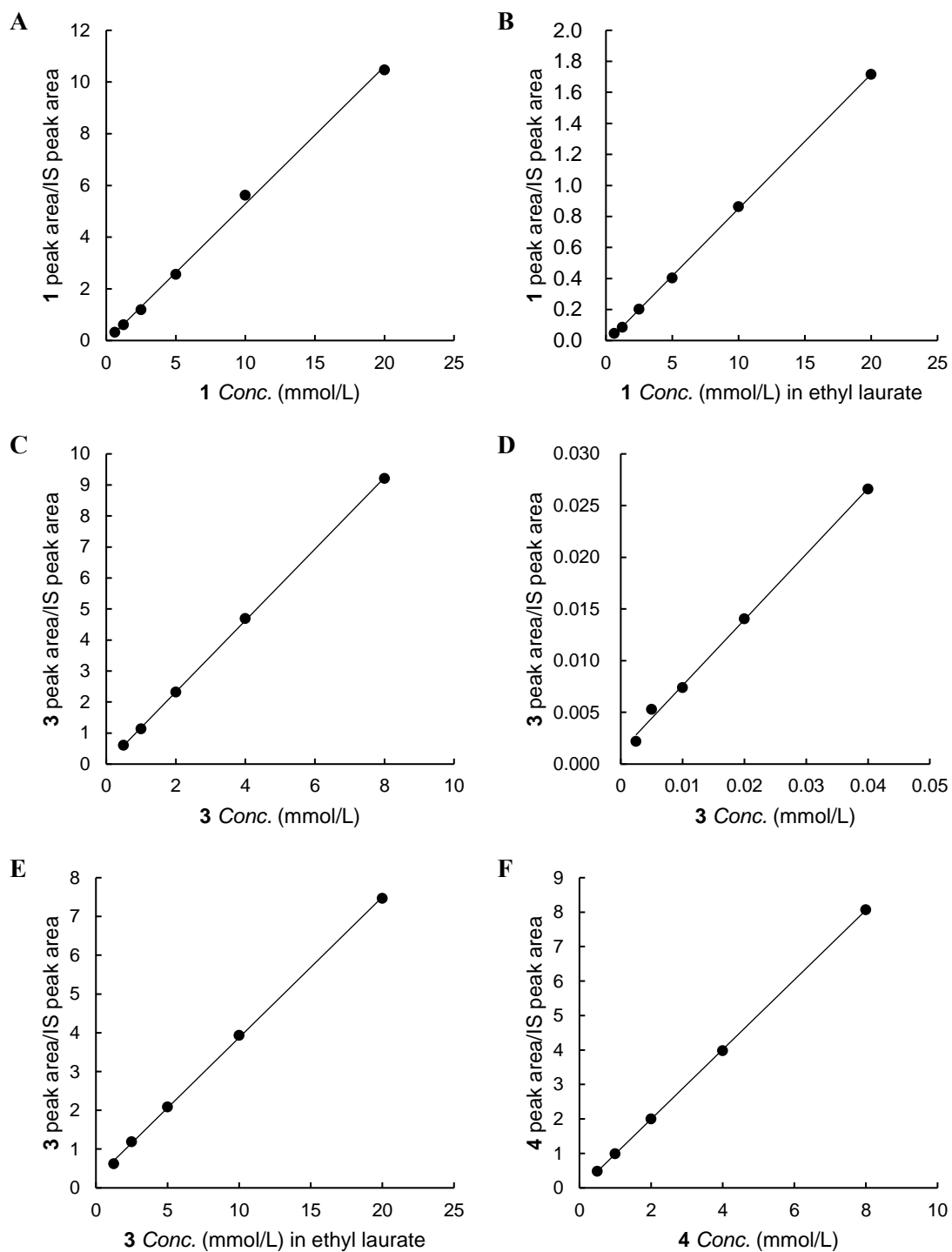
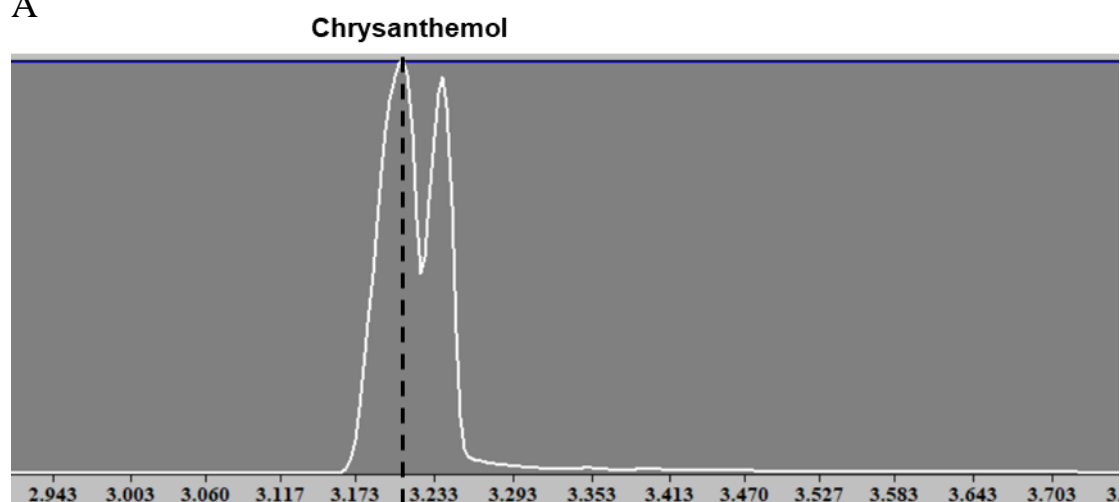


Fig. S1 A_p/A_{IS-p} Conc. standard curves of 1, 3, 4. (A) standard curves of 1; (B) standard curves of 1 in ethyl laurate; (C) standard curves of 3; (D) standard curves of 3 in the low-concentration range; (E) standard curves of 3 in ethyl laurate; (F) standard curves of 4.

A



B

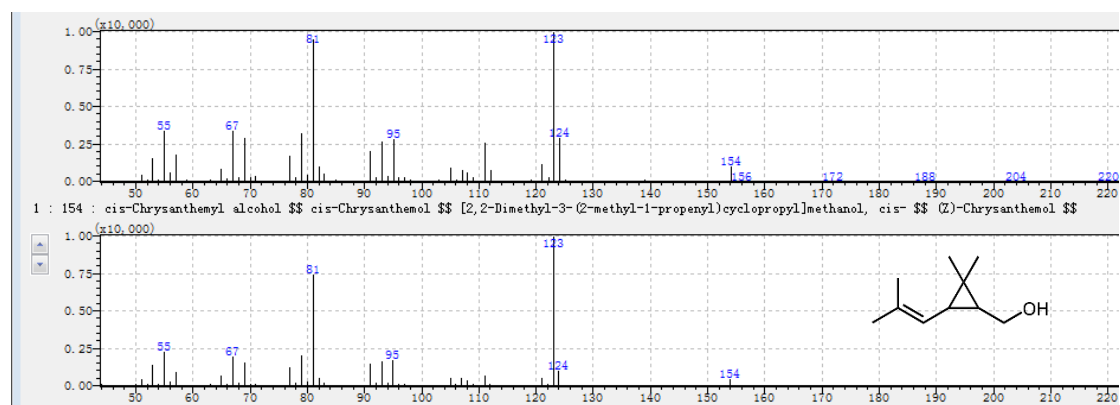


Fig. S2 GC-MS analysis of chrysanthemol **3** standard. (A) GC analysis of chrysanthemol standard; (B) Mass spectra are shown for the experimental sample (upper panel) and the standard chrysanthemol from the NIST database (lower panel).

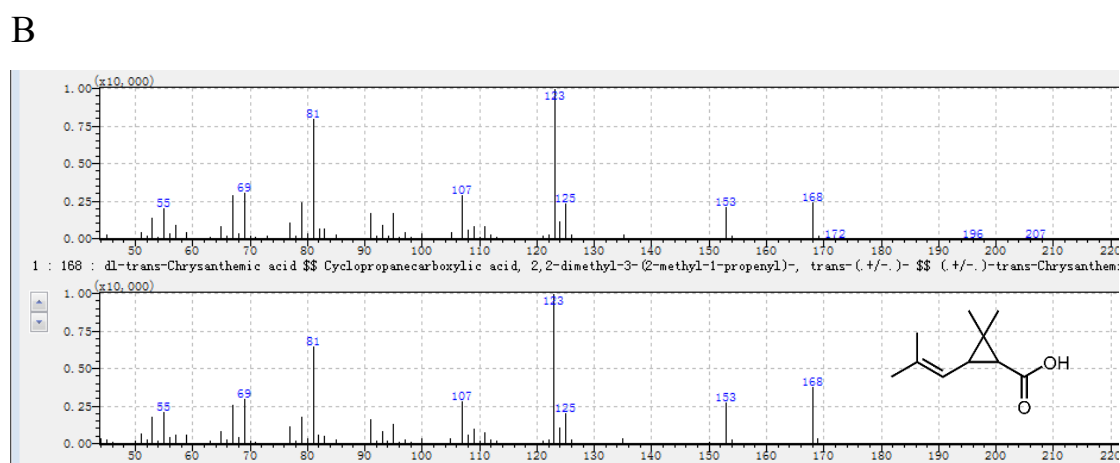
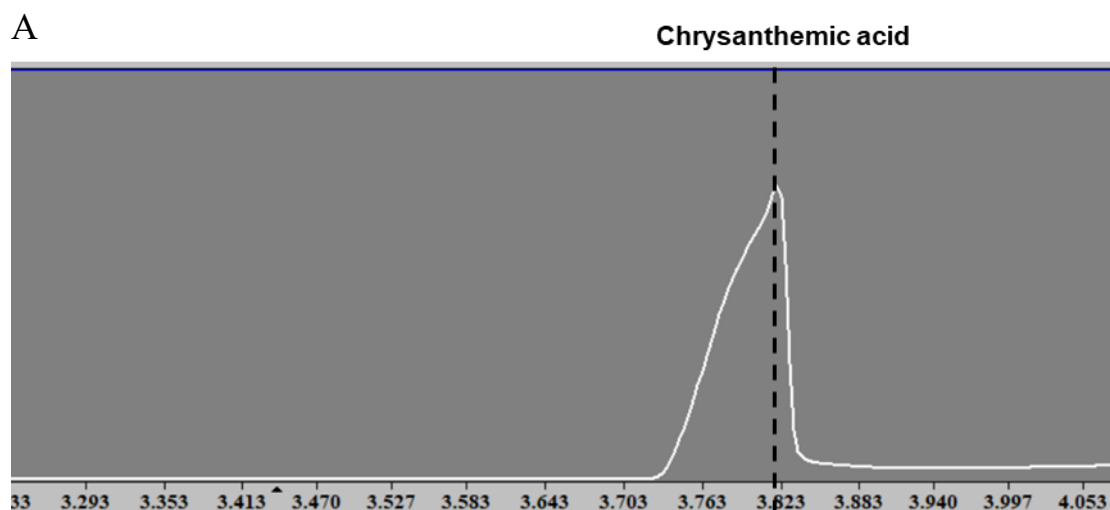


Fig. S3 GC-MS analysis of chrysanthemic acid **4** standard. (A) GC analysis of chrysanthemic acid standard; (B) Mass spectra are shown for the experimental sample (upper panel) and the standard chrysanthemic acid from the NIST database (lower panel).

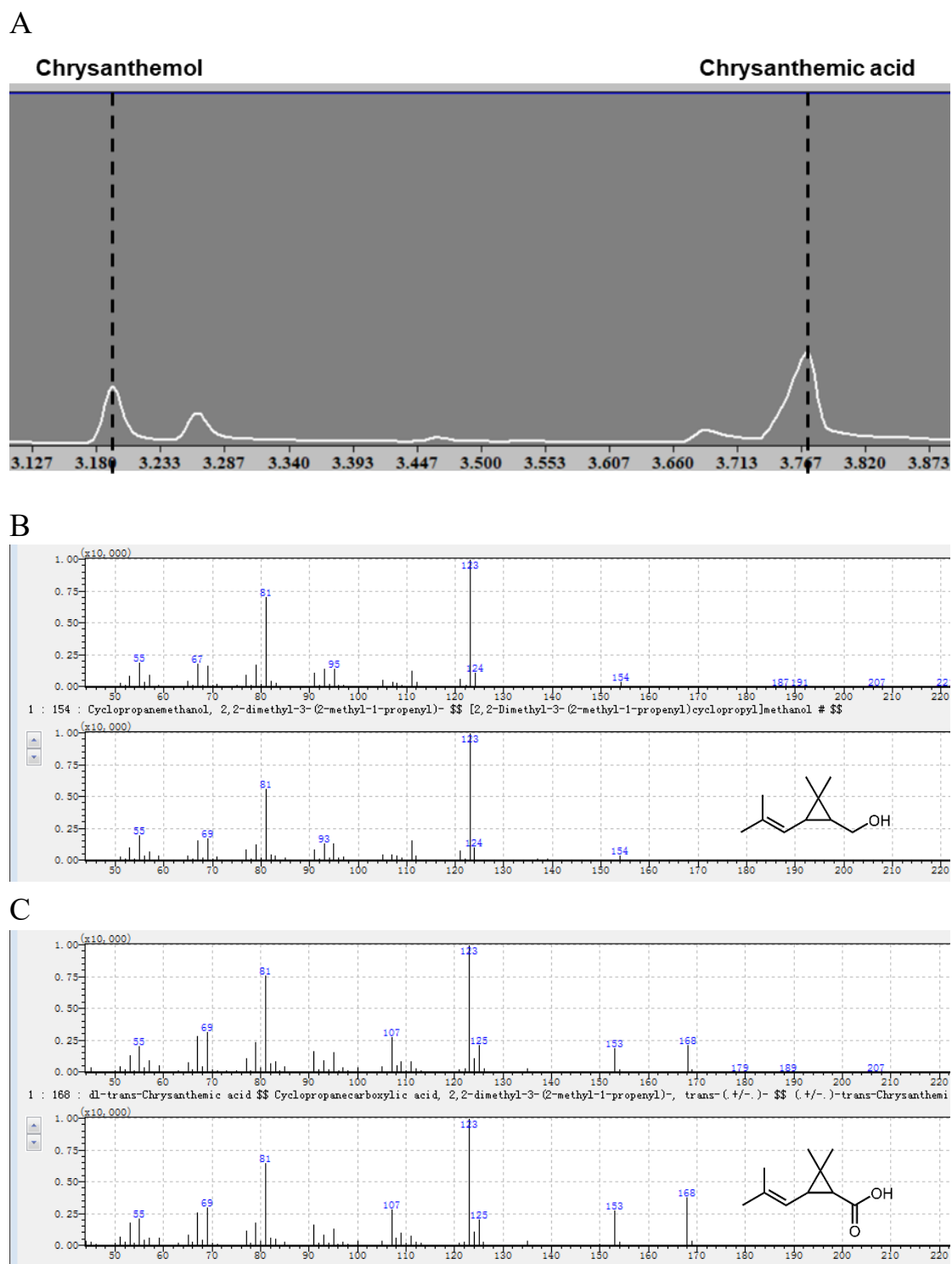


Fig. S4 GC-MS analysis of product **3** and **4**. (A) GC analysis of product sample; (B) Mass spectra are shown for the chrysanthemol peak (upper panel) and the standard chrysanthemol from the NIST database (lower panel); (C) Mass spectra are shown for the chrysanthenic acid peak (upper panel) and the standard chrysanthenic acid from the NIST database (lower panel).

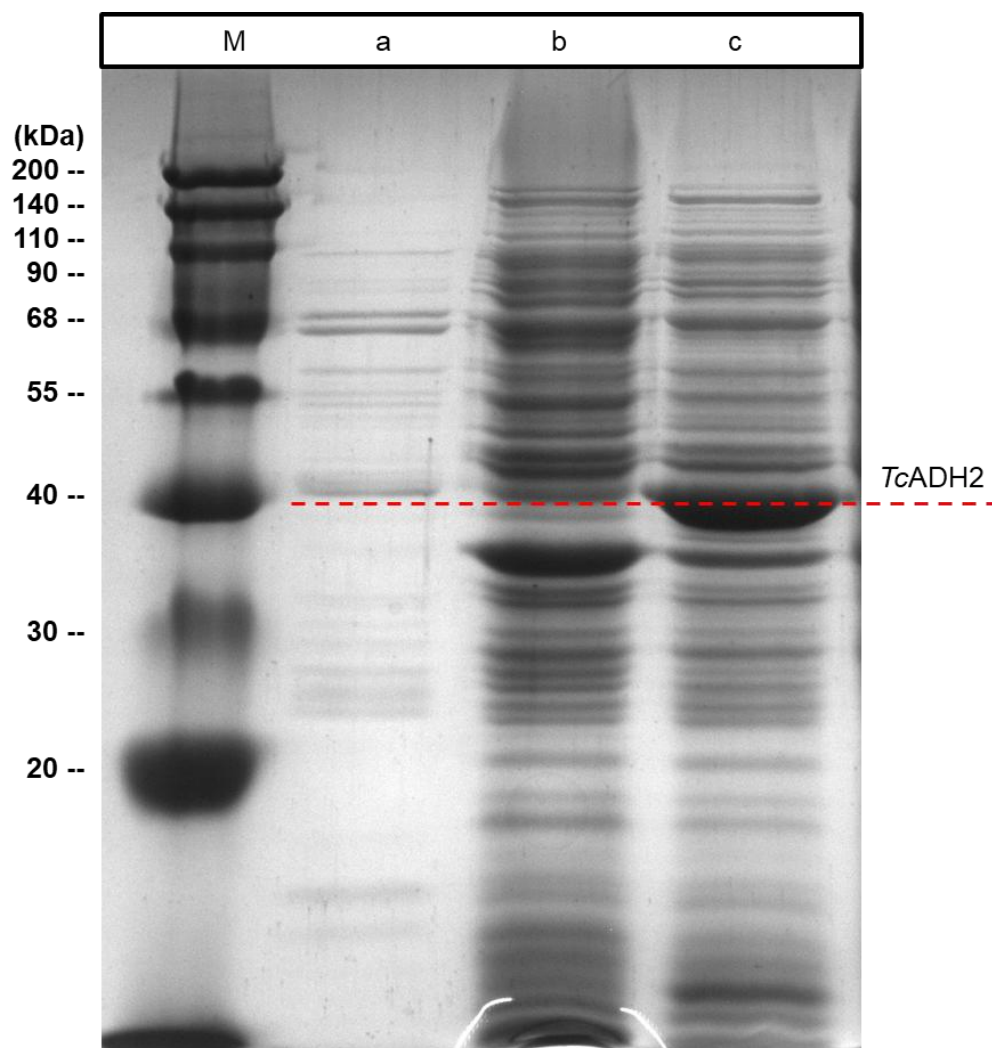


Fig. S6 SDS-PAGE analysis of *TcADH2* (41.4 kDa) purification process. M: Marker; a: Purified *TcADH2* solution; b: cell-free extract (supernatant) of *E. coli* expressing *TcADH2*; c: whole-cell protein of *E. coli* expressing *TcADH2*.

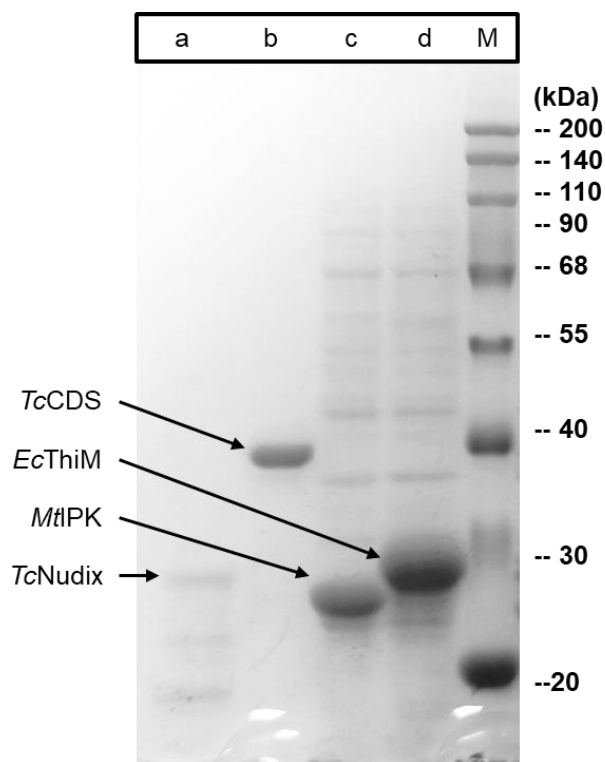


Fig. S7 SDS-PAGE analysis of the purified enzymes involved in the cascade for synthesis of chrysanthemol. a: *TcNudix*; b: *TcCDS*; c: *MtIPK*; d: *EcThiM*; M: Marker.

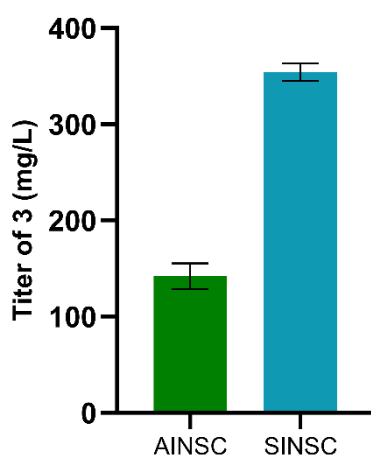


Fig. S8 Titer of **3** using different IKs. The AINSC strain, expressing *AtFKI* as the IK, was transformed with pETDuet-*AtFKI*-IPK and pACYCDuet-*TcNudix*-Sumo*TcCDS*. In contrast, the SINSC strain, expressing *SmaThiM* as the IK, was transformed with the same set of expression vector.

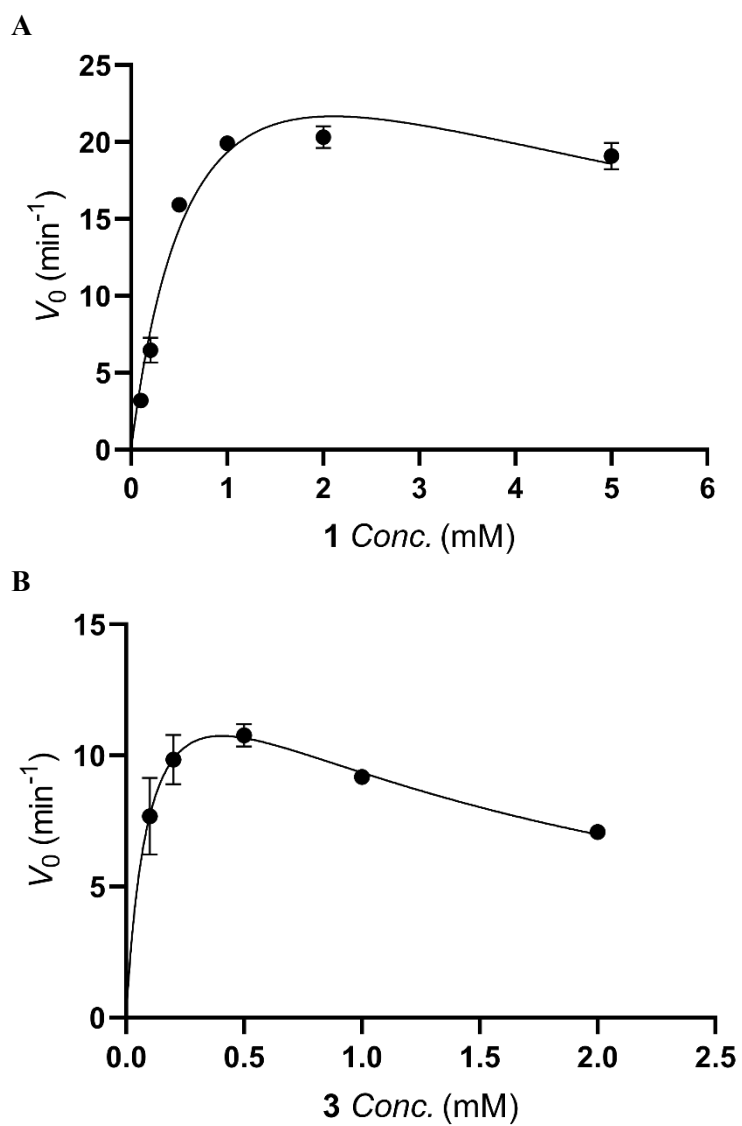


Fig. S9 Haldane curves of *TcADH2*-catalyzed oxidation of **1** and **3**. (A) Haldane curves of *TcADH2*-catalyzed oxidation of **1**; (B) Haldane curves of *TcADH2*-catalyzed oxidation of **3**.

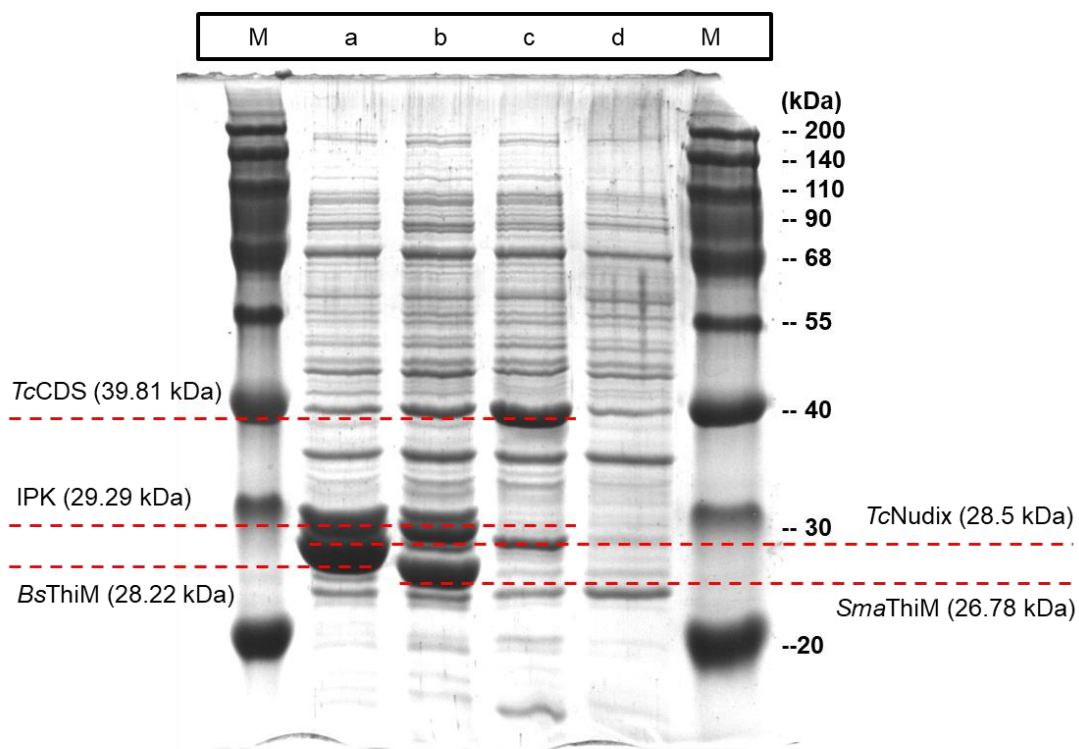


Fig. S10 SDS-PAGE analysis of **3**-producing strains, which co-expresses pETDuet-*SmaThiM*/*BsThiM*-IPK and pACYCDuet-*TcNudix*-*TcCDS*. a: **3**-producing strains (expressing *BsThiM*); b: **3**-producing strains (expressing *SmaThiM*); c: the control group that was transferred by pACYCDuet-*TcNudix*-*TcCDS* only; d: the blank control group that was transferred by pETDuet-1 and pACYCDuet-1 without any target gene; M: Marker. According to the electrophoresis results, the 4 pathway enzymes IK (including *SmaThiM* (26.78 kDa) and *BsThiM* (28.22 kDa)), IPK (29.29 kDa), *TcNudix* (25.46 kDa) and *TcCDS* (39.81 kDa) in the **3**-producing strains were successfully expressed.

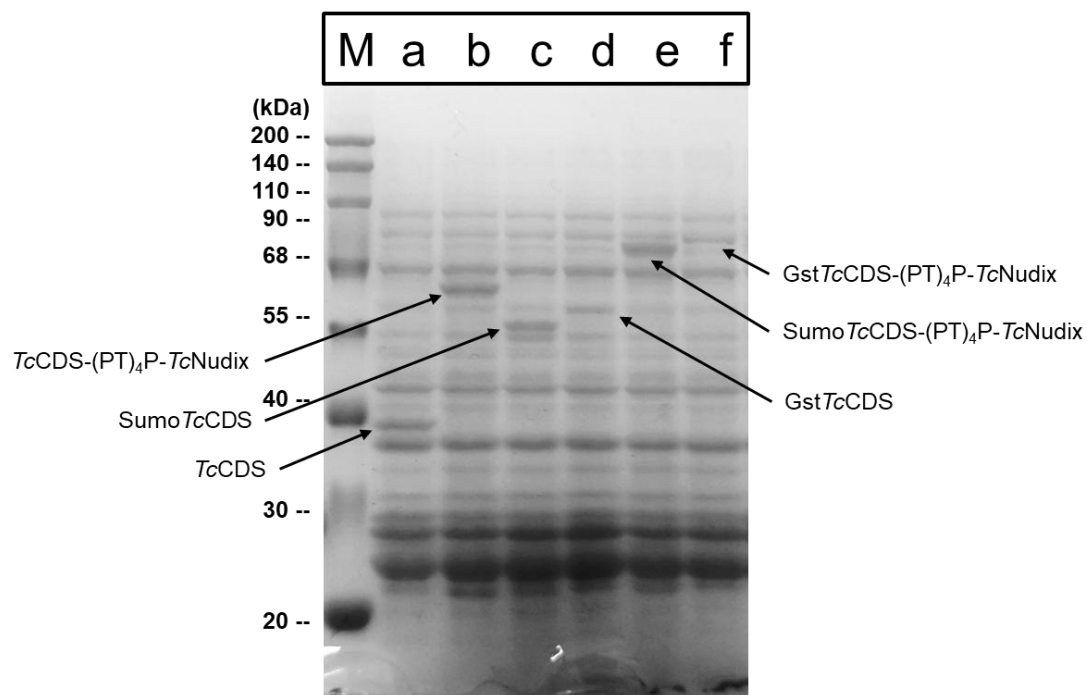


Fig. S11 SDS-PAGE analysis of **3**-producing strains that fusing different solubility tags to the *TcCDS* N-terminus (in the AC-N-C vector) or to the fusion protein N-terminus (in the AC-C-(PT)₄P-N vector): M: Marker; a: pACYCDuet-*TcNudix*-*TcCDS*; b: pACYCDuet-*TcCDS*-(PT)₄P-*TcNudix*; c: pACYCDuet-*TcNudix*-Sumo*TcCDS*; d: pACYCDuet-*TcNudix*-Gst*TcCDS*; e: pACYCDuet-Sumo*TcCDS*-(PT)₄P-*TcNudix*; f: pACYCDuet-Gst*TcCDS*-(PT)₄P-*TcNudix*.

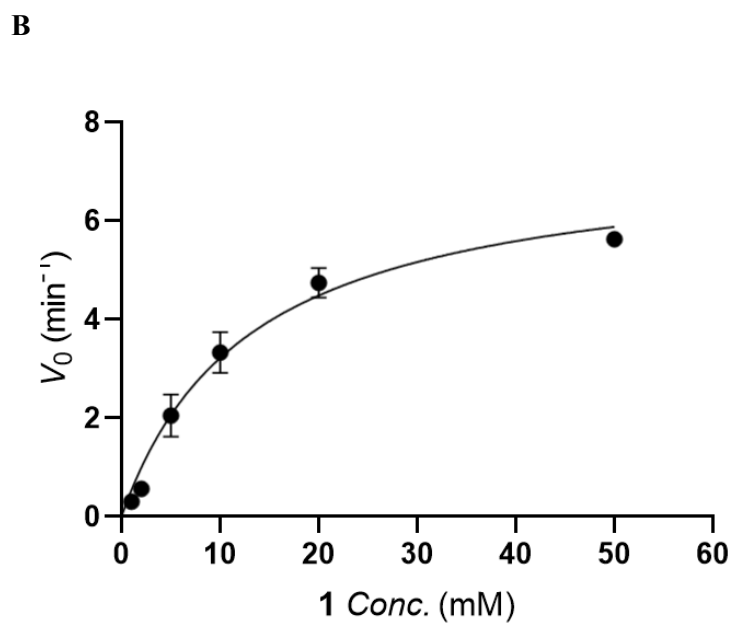
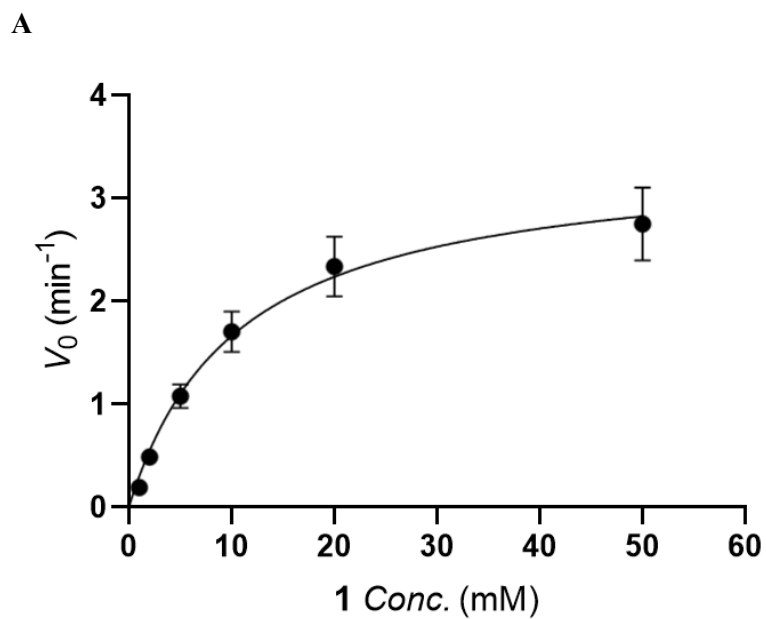


Fig. S12 Michaelis–Menten curves of phosphorylation of 1 by *EcThiM* (A) and *SmaThiM* (B).

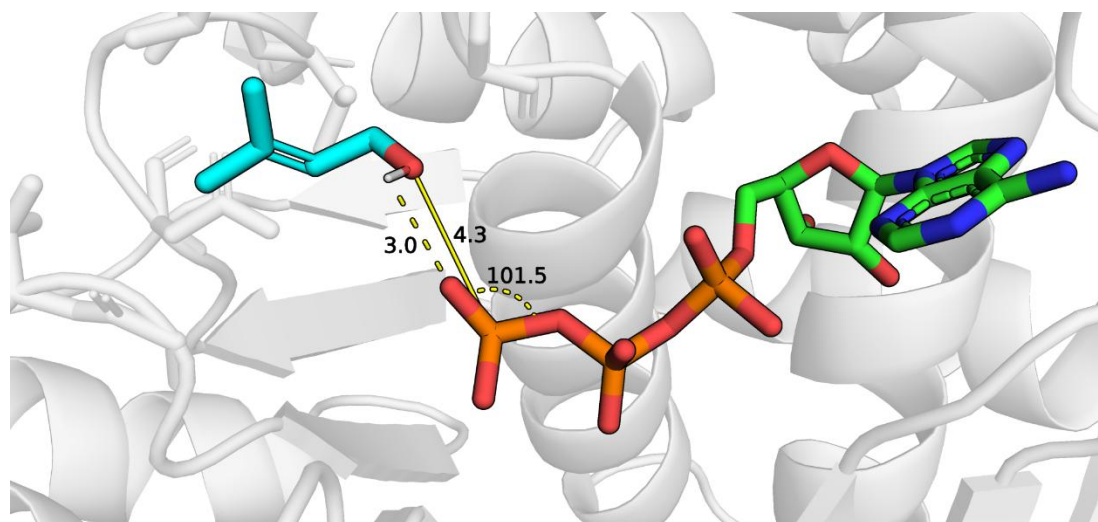
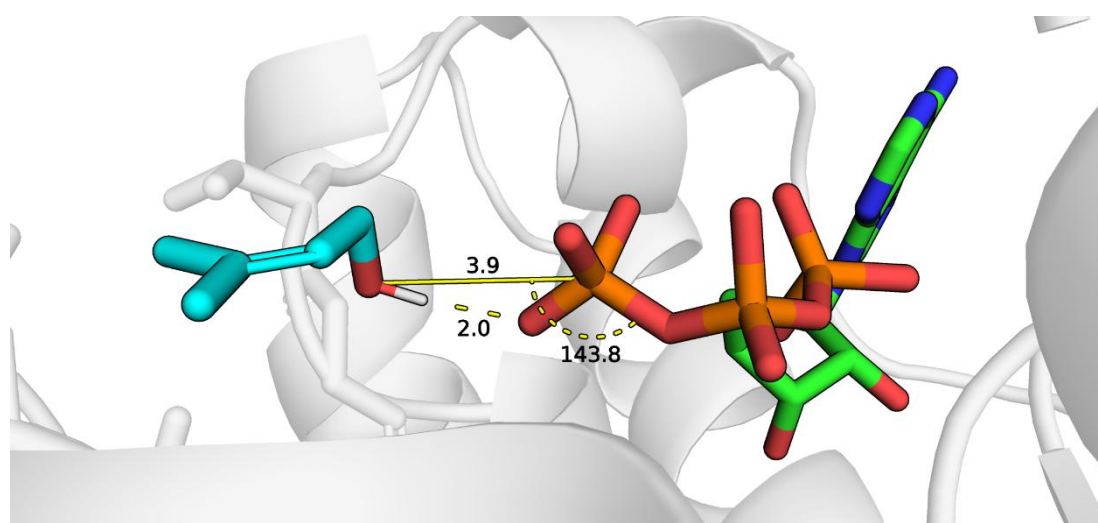
A**B**

Fig. S13 Molecular docking results of *EcThiM* (A) and *SmaThiM* (B). The angle formed by the prenyl hydroxyl oxygen, the ATP γ -phosphorus atom, and the leaving group oxygen is 143.8° in *SmaThiM*, compared to 101.5° in *EcThiM*. The former more closely approaches the optimal geometry for an in-line attack, thereby conferring a catalytic advantage.^{5, 6} Furthermore, hydroxyl hydrogen atom of **1** is positioned closer to the γ -phosphate oxygen atom (which may serve as a proton acceptor) in *SmaThiM* (2.0 Å) than in *EcThiM* (3.0 Å). Additionally, the hydroxyl oxygen atom is nearer to the γ -phosphorus atom of ATP in *SmaThiM* (3.9 Å) than in *EcThiM* (4.3 Å). Under the substrate-assisted catalysis mechanism,⁷ these proximity effects allow prenyl in *SmaThiM* to be more readily deprotonated to form an oxyanion, which can then more easily attack the γ -phosphate group of ATP to drive the reaction forward.

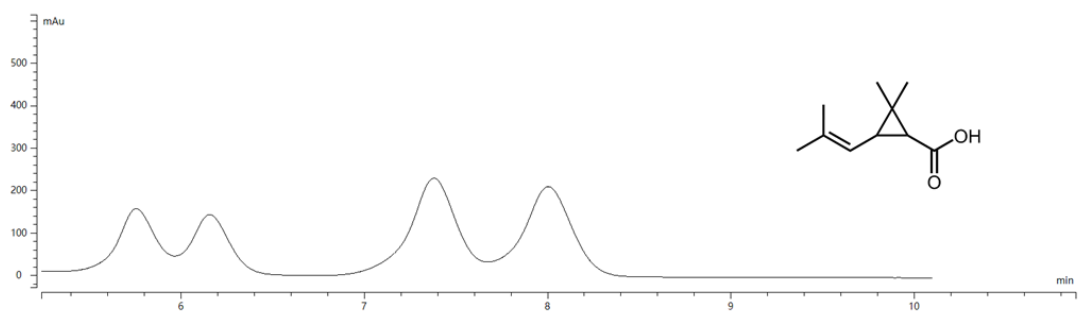
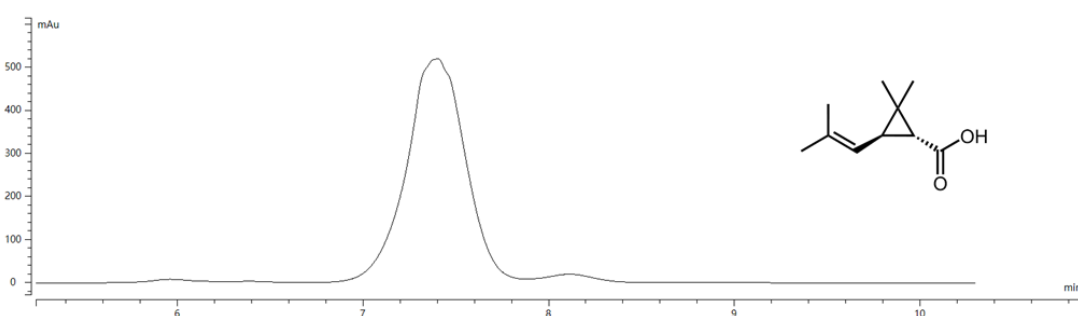
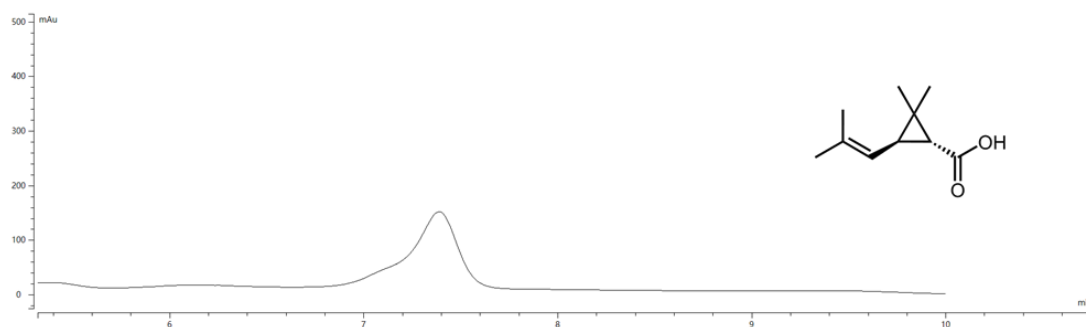
A**B****C**

Fig. S14 Chiral HPLC analysis of (±)-chrysanthemic acid standard, (1*R*, 3*R*)-chrysanthemic acid standard, and product **4**. (A) (±)-chrysanthemic acid standard; (B) (1*R*, 3*R*)-chrysanthemic acid standard; (C) Product **4** produced by the SINSCSAA strain.

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