

Electronic Supplementary Information

Ultrafast Fluorescent Detection of Mercury(II) Ions in Aqueous and Biological Systems Using
a Boronic Acid–Based Reaction Probe

Kishor Khadka^a, Sumita Subedi^a, Eun-Taex Oh^b, and Keun-Hyeung Lee^{a,*}

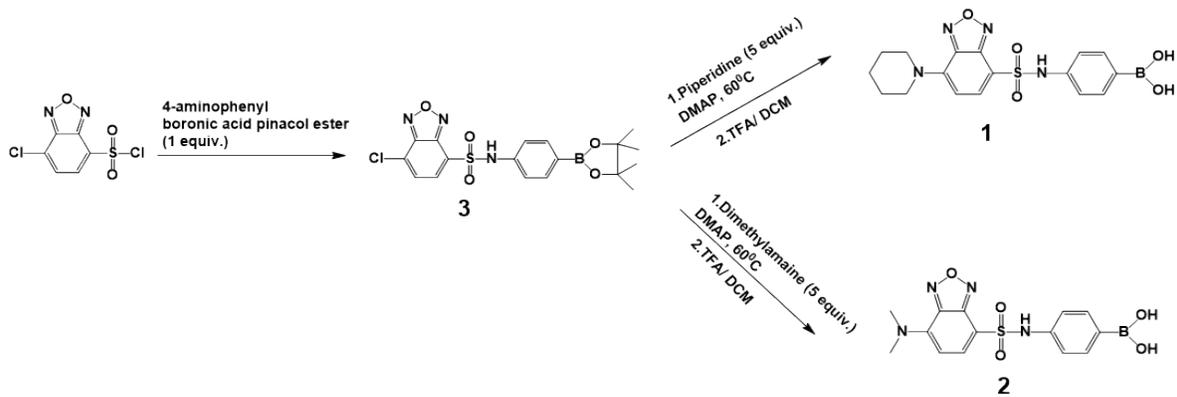
^aEducation and Research Center for Smart Energy Materials and Process, Department of
Chemistry and Chemical Engineering, Inha University, Incheon 402-751, South Korea

^bDepartment in Biomedical Sciences, College of Medicine, Inha University, Incheon 22212,
South Korea

E-mail: leekh@inha.ac.kr

Contents

1. Figures	S2-S21
Scheme S1. Synthesis scheme of 1 and 2	S3
Figure S1. HPLC of 1	S4
Figure S2. HPLC of 2	S5
Figure S3. HR-MS of 1	S6
Figure S4. ¹ H NMR of 1	S7
Figure S5. ¹³ C NMR of 1	S8
Figure S6. HR-MS of 2	S9
Figure S7. ¹ H NMR of 2	S10
Figure S8. ¹³ C NMR of 2	S11
Figure S9. HR-MS of 3	S12
Figure S10. ¹ H NMR of 3	S13
Figure S11. ¹³ C NMR of 3	S14
Figure S12. Fluorescent response of 2 toward various mercury salts	S15
Figure S13. C ₁₈ -TLC and HPLC analysis of probe 1 with Hg ²⁺	S16
Figure S14. FAB mass analysis of the product of probe 1 with Hg ²⁺	S17
Figure S15. Fluorescent detection of Hg(II) in the presence of other metal ions	S18
Figure S16. Cell toxicity study of probe 1	S19
Figure S17. Emission intensity of 1 at various pH for determining pK _a value	S20
Figure S18. Reaction kinetic study of probe 1 with Hg ²⁺	S21
Table S1. Recovery of spiked Hg ²⁺ in real natural water samples.	S22
Table S2. Detection properties of reported turn-on fluorescent chemodosimeters for Hg ²⁺	S23
References	S26



Scheme S1. Synthesis scheme of probe 1 and probe 2.

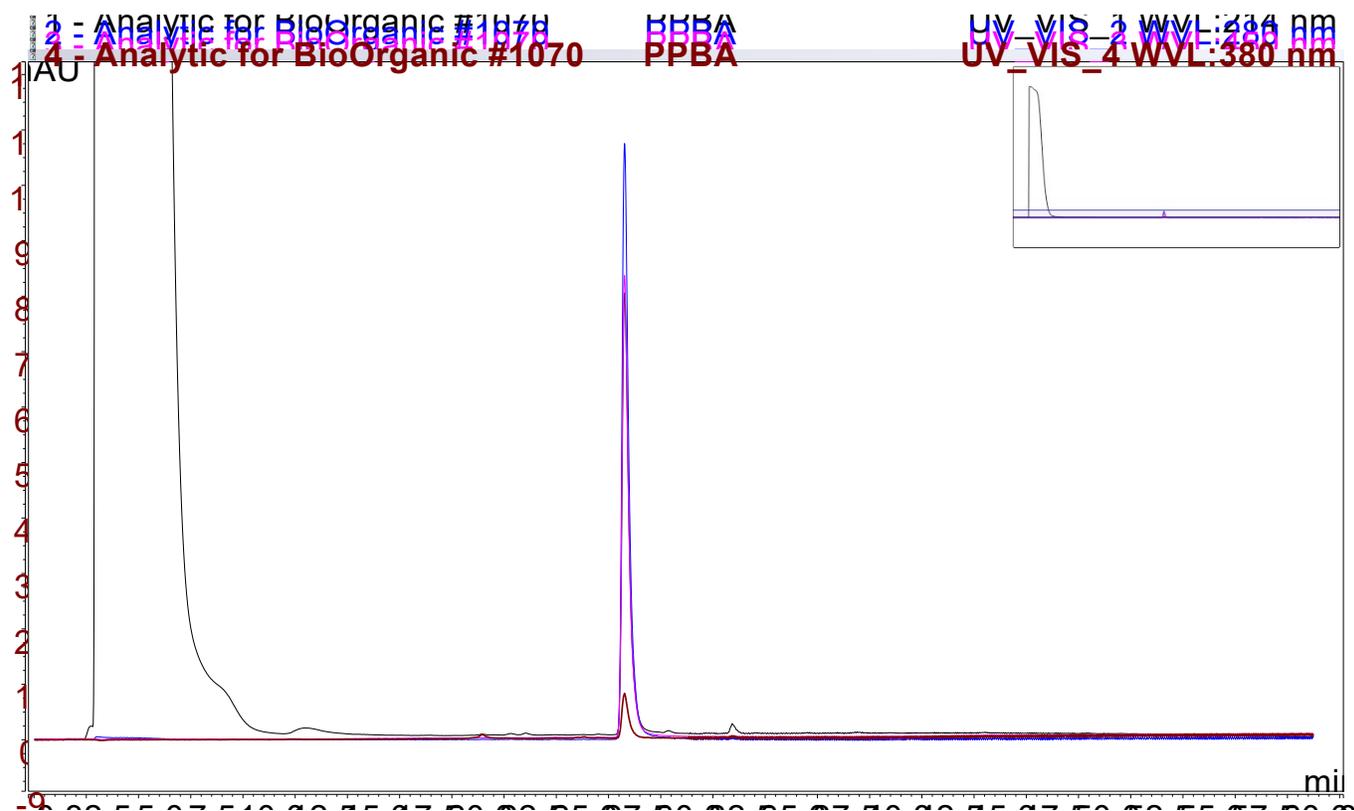


Figure S1. HPLC chromatogram of 1.

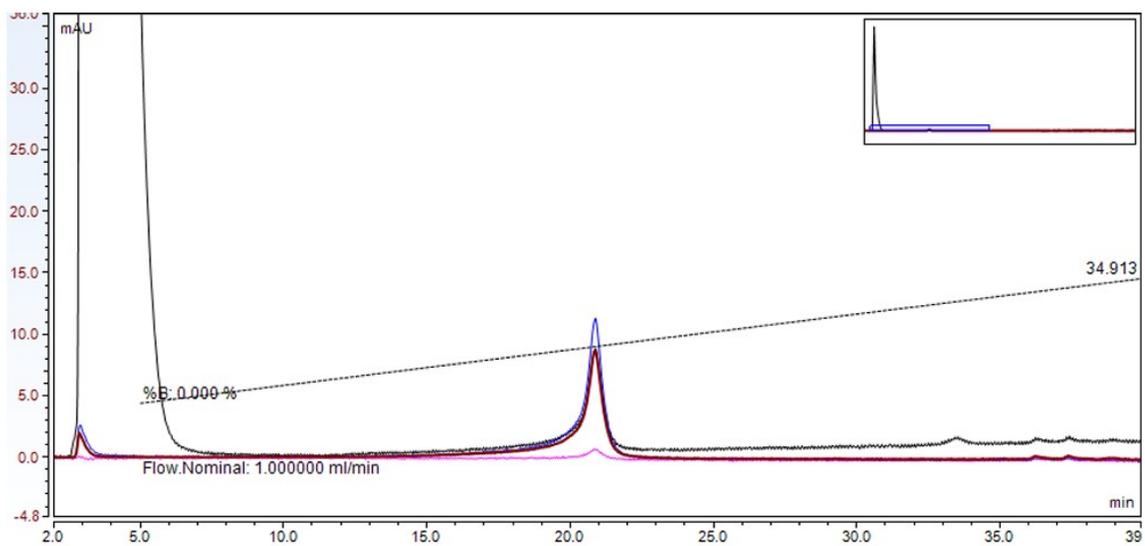


Figure S2. HPLC chromatogram of **2**.

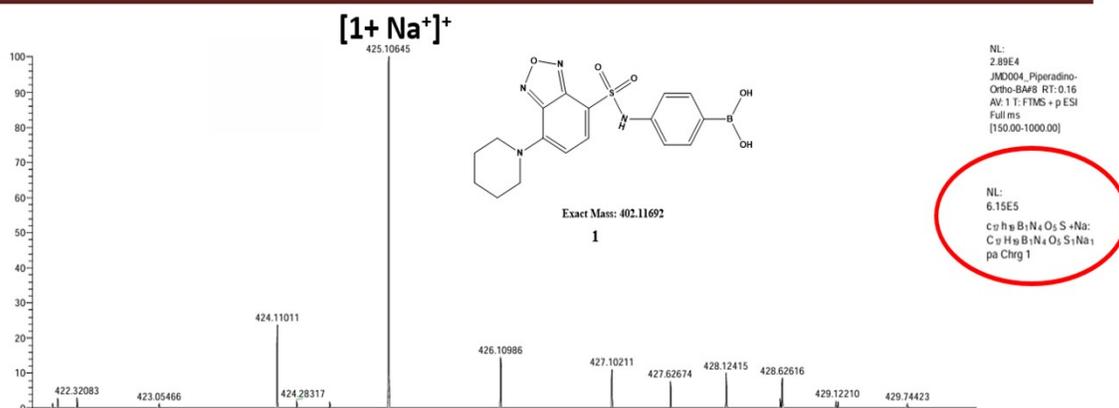


Figure S3. HR-mass spectrum of **1**.

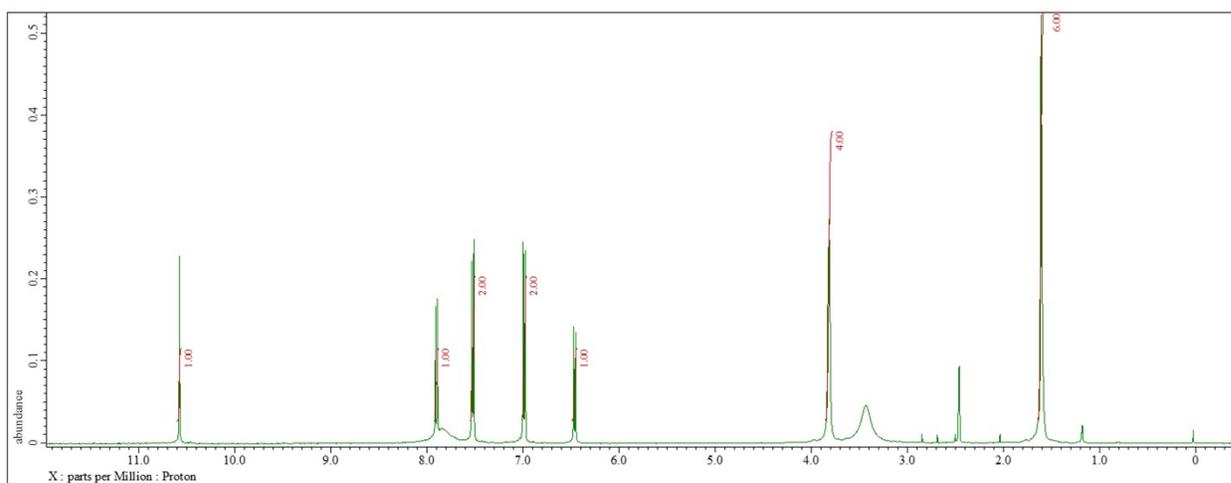


Figure S4. ^1H NMR of 1.

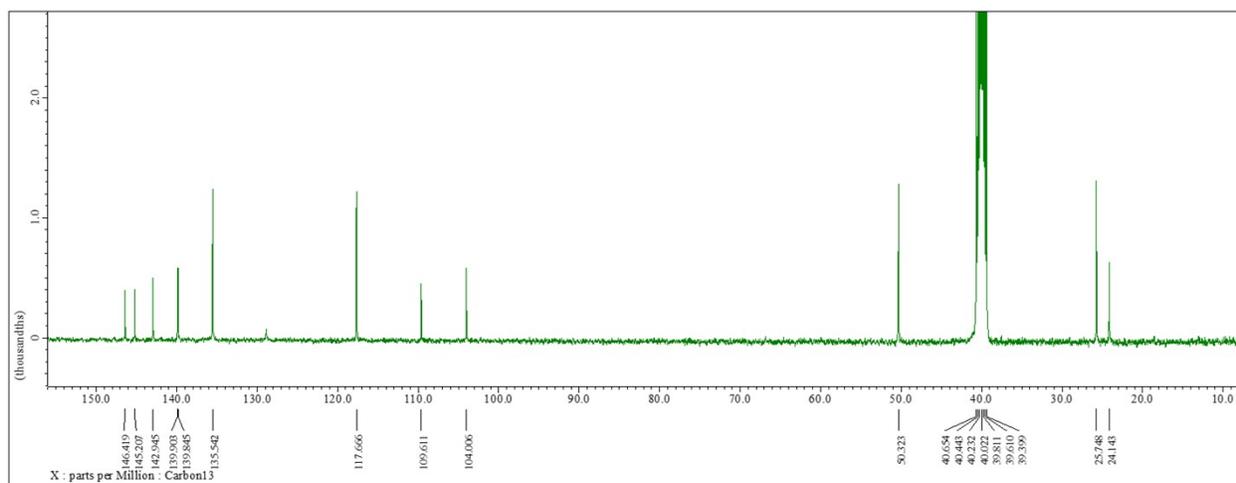


Figure S5. ¹³C NMR of 1.

*** [High Resolution] Orbitrap Mass Spectrometer (Model : LTQ Orbitrap XL / Company : ThermoFisher Scientific)

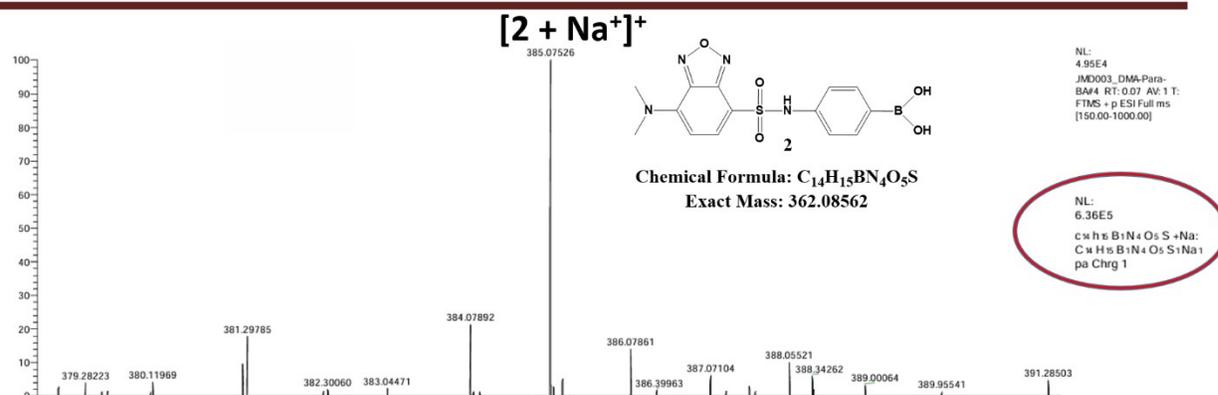


Figure S6. HR-mass spectrum of **2**.

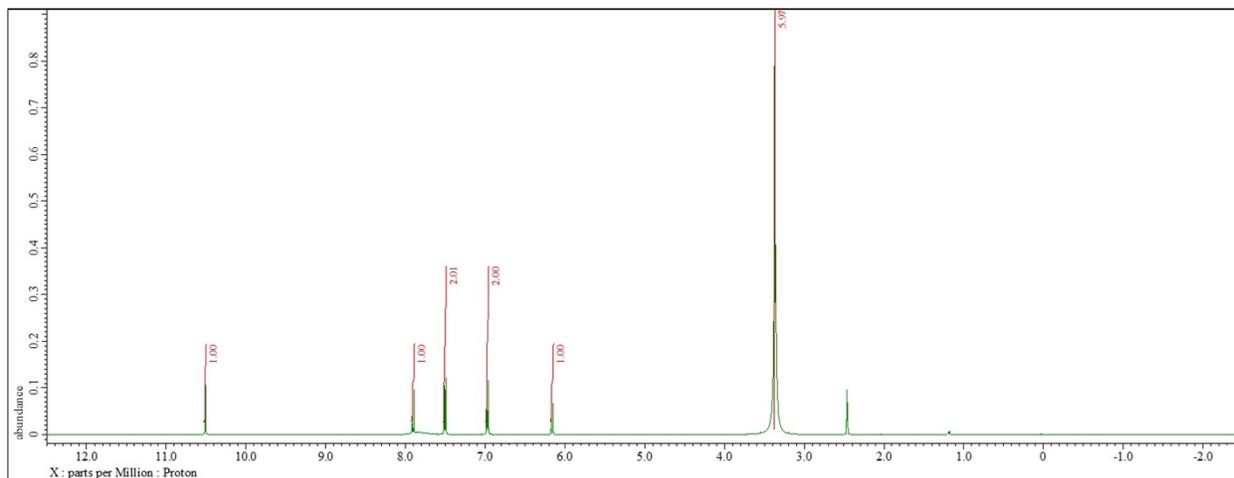


Figure S7. ^1H NMR of **2**.

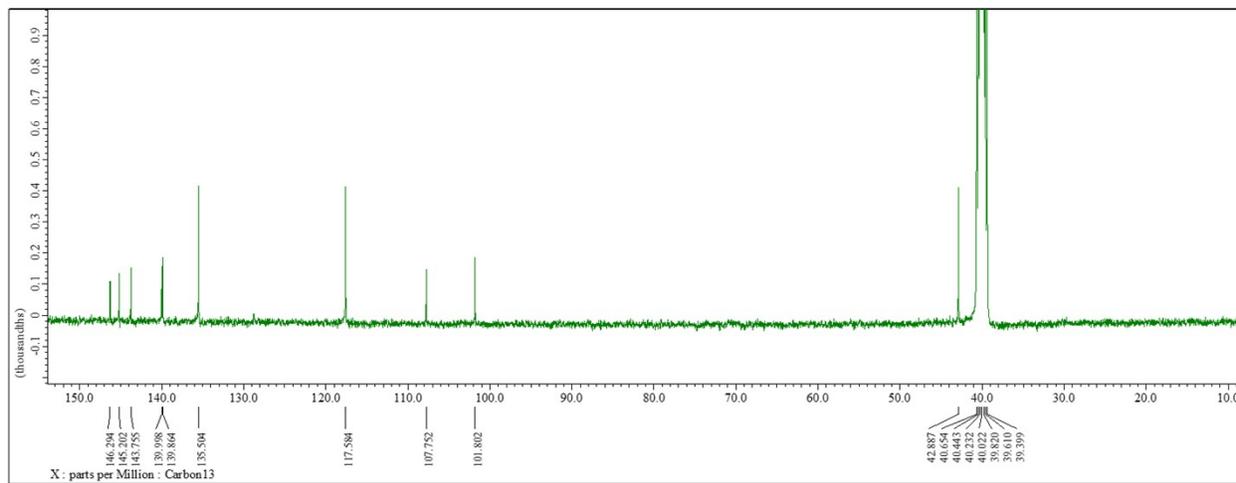
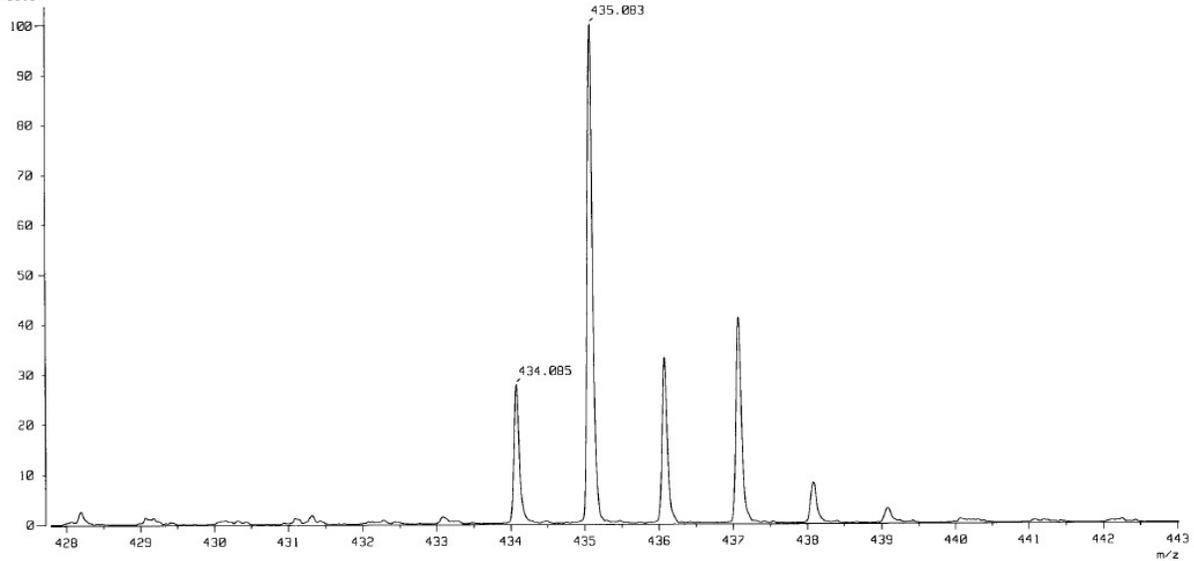


Figure S8. ^{13}C NMR of **2**.

[Mass Spectrum]
 Data : 26-YN-1281_Sample4 Date : 11-Mar-2026 10:05
 Sample: -
 Note: -
 Inlet : Direct Ion Mode : EI+
 Spectrum Type : Normal Ion [EF-Linear]
 RT : 0.84 min Scan#: 26
 BP : m/z 435.0830 Int. : 41.88
 Output m/z range : 427.7893 to 443.0119 Cut Level : 0.00 %
 456197



[Elemental Composition]

Data : 26-YN-1281_Sample4 Date : 11-Mar-2026 10:05
 Sample: -
 Note: -
 Inlet : Direct Ion Mode : EI+
 RT : 0.84 min Scan#: 26
 Elements : C 18/0, H 19/0, Cl 1/0, N 3/0, O 5/0, S 1/0, B 1/0
 Mass Tolerance : 1mmu
 Unsaturation (U.S.) : -0.5 - 1000.0

Observed m/z	Int%	Err [ppm / mmu]	U.S. Composition
420.0591	43.1	-0.4 / -0.2	12.5 C 17 H 16 Cl N 3 O 5 S B
421.0652	13.1		
422.0723	18.6		
423.1115	20.3		
434.0854	27.8		
435.0830	100.0	+0.6 / +0.3	12.0 C 18 H 19 Cl N 3 O 5 S B
436.0836	33.1		
437.0825	41.1		

Figure S9. HR-mass spectrum of **3**.

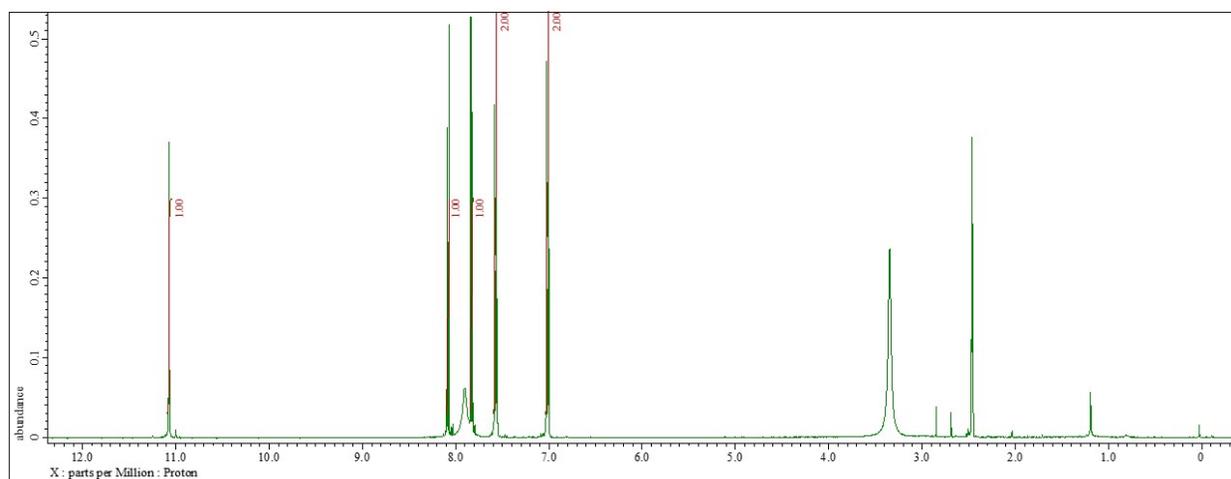


Figure S10. ^1H NMR of **3**.

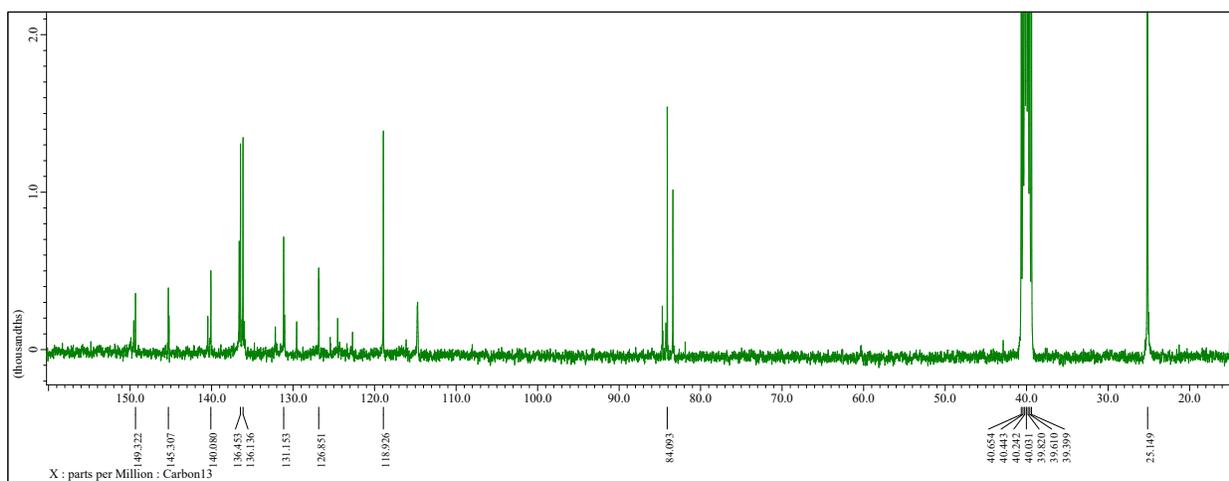


Figure S11. ^{13}C NMR of 3.

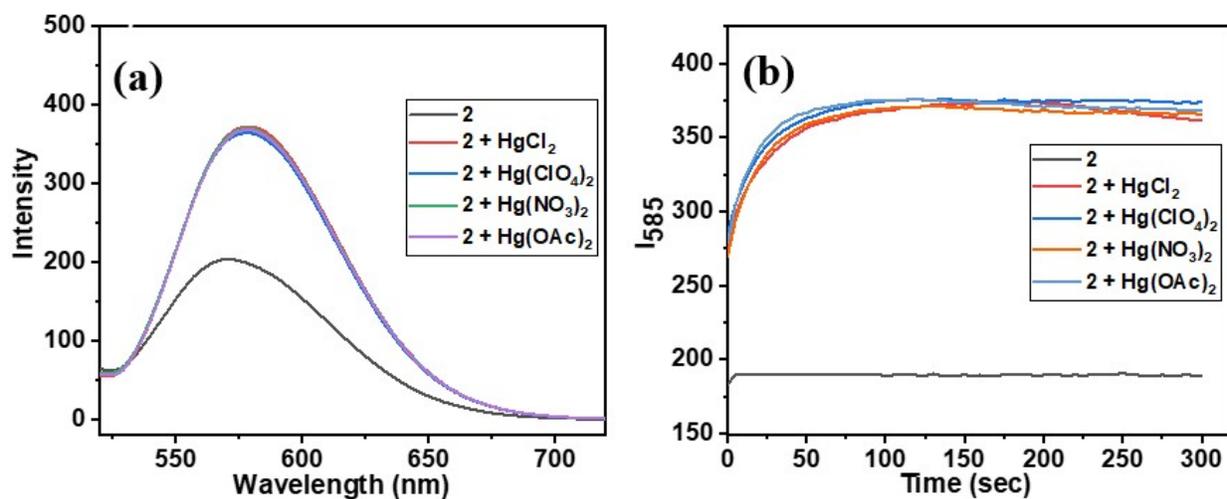


Figure S12. (a) Fluorescence emission spectra and (b) time-dependent emission intensity of probe **2** with various mercury salts ($5 \mu\text{M}$) after 2 min incubation in aqueous buffered solution (pH 7.4) containing 1% DMF.

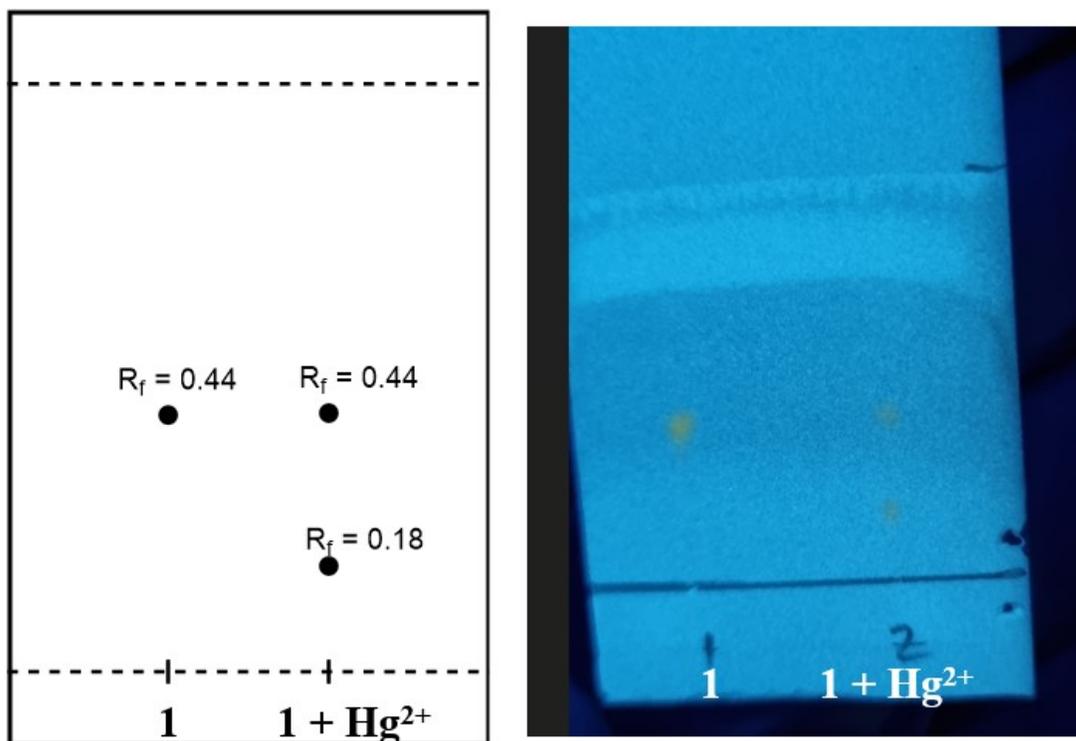


Figure S13. C_{18} -TLC analysis of probe **1** with $Hg(II)$ in $H_2O:CH_3CN$ (80:20)

[Mass Spectrum]
Data : 26-YN-1281_Sample5 Date : 11-Mar-2025 10:10
Sample : -
Note : -
Inlet : Direct Ion Mode : EI+
Spectrum Type : Normal Ion [EF-Linear]
RT : 1.74 min Scan# : 53
BP : m/z 594.1016 Int. : 78.82
Output m/z range : 564.0000 to 624.0000 Cut Level : 0.00 %

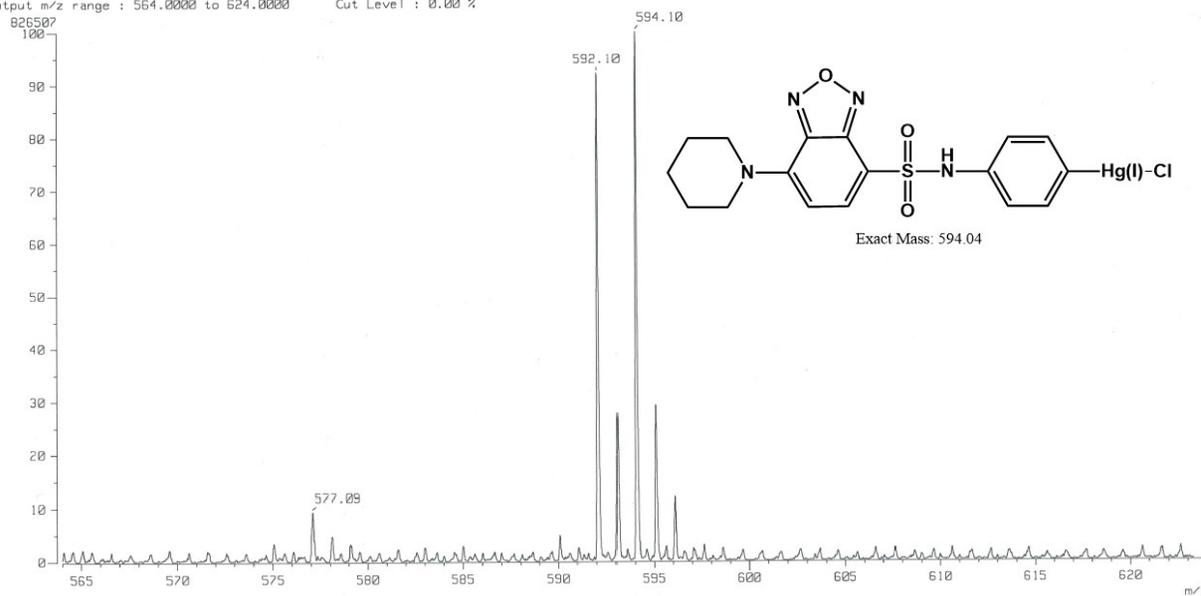


Figure S14. mass spectrometric analysis of the reaction product using FAB mass spectrometry.

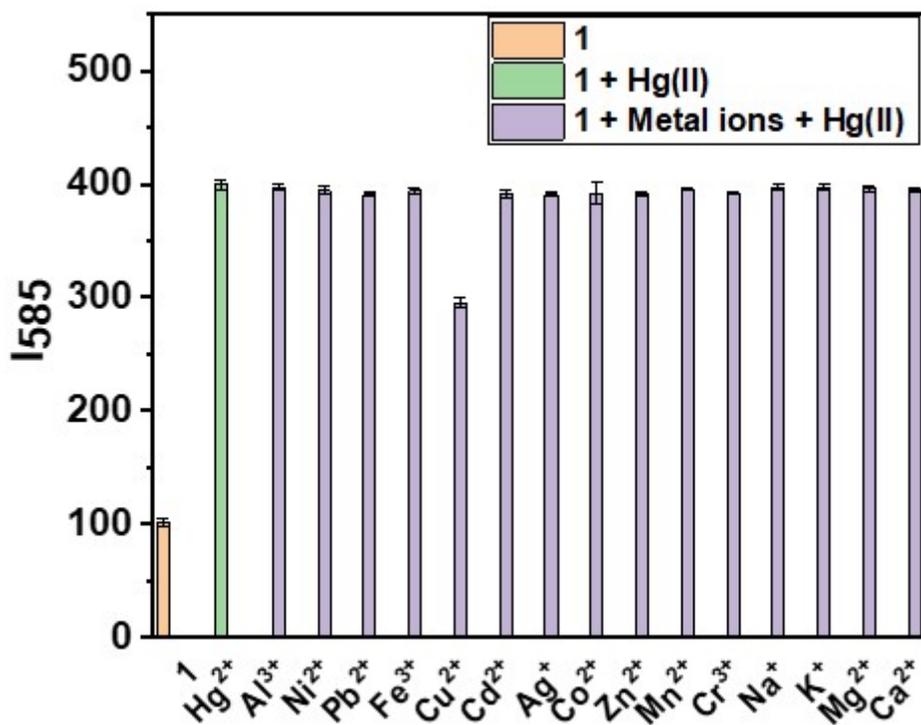


Figure S15. Fluorescence emission intensity of **1** with Hg²⁺ in the presence of coexisting metal ions in aqueous buffered solution (10 mM HEPES, pH 7.4) containing 1% DMF. The concentrations of probe **1**, heavy metal ions, and Group I and II metal ions were 5 μM, 10 μM, and 1000 μM, respectively.

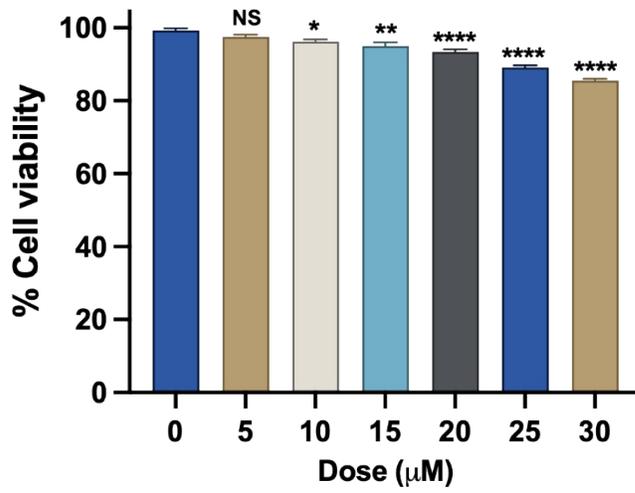


Figure S16. Cell toxicity study of probe 1 using an ATP-Glo assay.

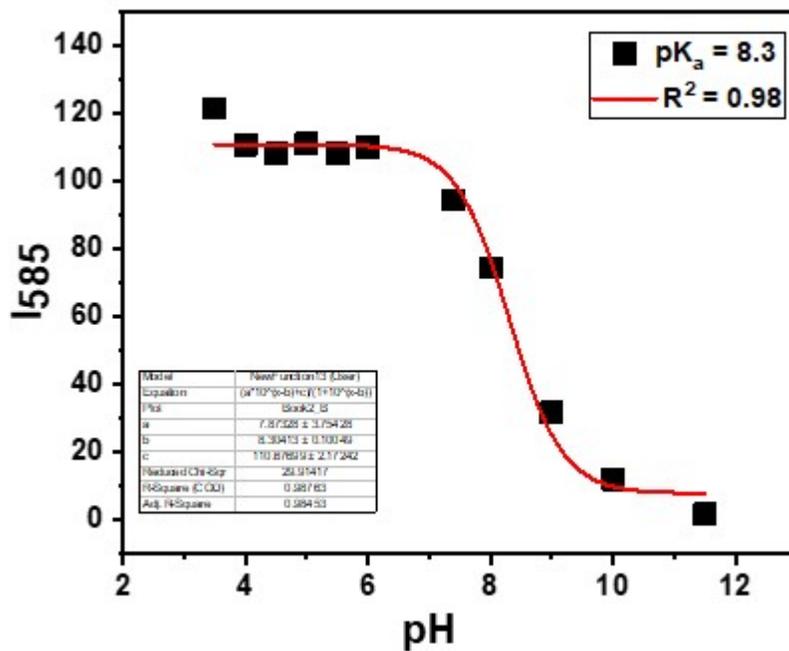
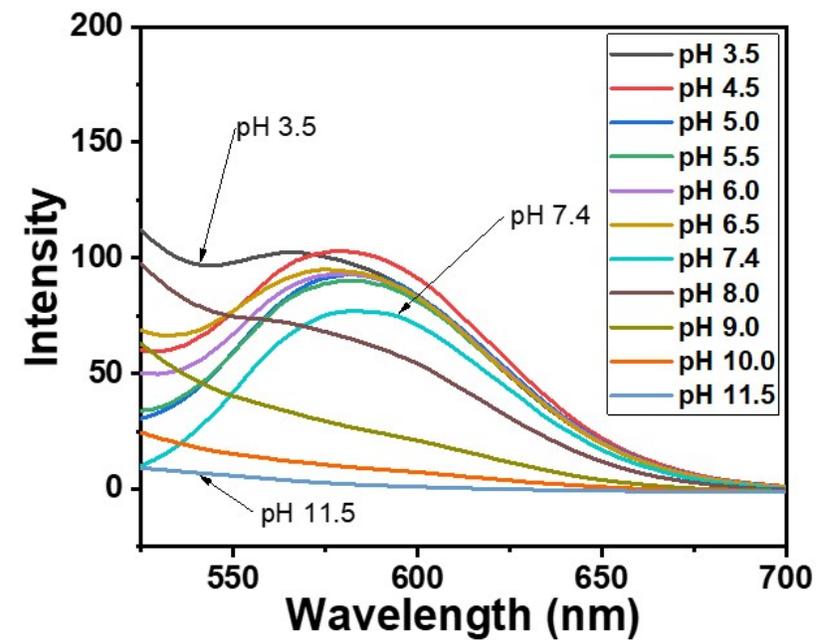


Figure S17. Emission intensity changes of probe 1 (5 μ M) at different at pHs for determination of pK_a value.

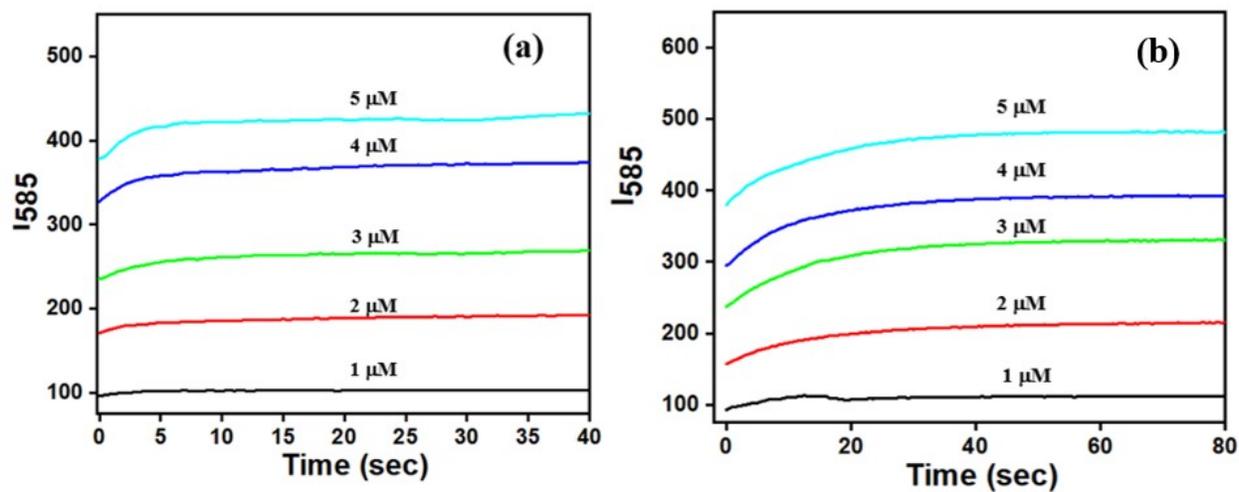
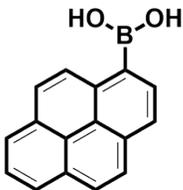
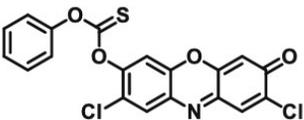
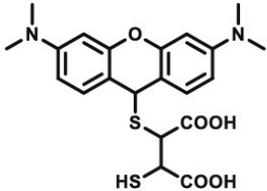
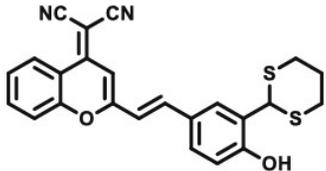


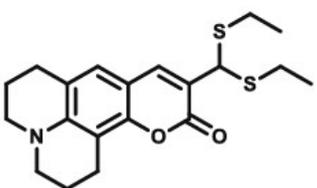
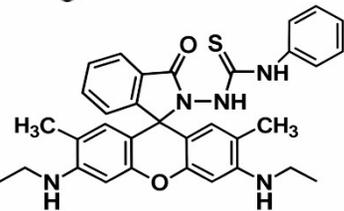
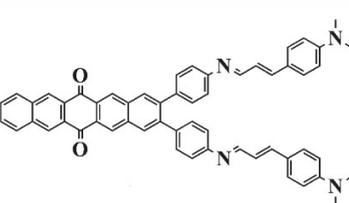
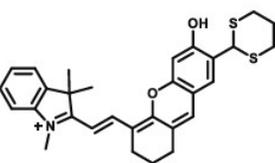
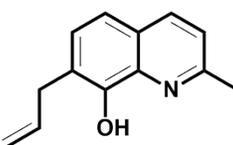
Figure S18. Time-dependent fluorescence intensity profiles of probe **1** (1, 2, 3, 4, and 5 μM) with Hg^{2+} (50 μM) in aqueous solution (10 mM HEPES, pH 7.4) including (a) 10% DMF and (b) 20% DMF.

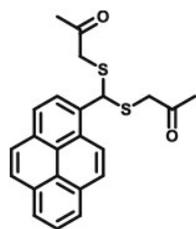
Table S1. Recovery of spiked Hg^{2+} in real natural water samples using probe 1.

Sample	Spiked Hg^{2+} (nM)	Measured concentration of Hg^{2+} using the probe (nM)	Recovery (%)
Tap water	0	0	100 ± 0.1
	200	186.06	93.1 ± 0.2
	400	393.48	98.4 ± 0.2
	600	557.68	92.9 ± 0.2
	800	799.67	99.9 ± 0.1
	1000	955.23	95.5 ± 0.3
Ground water	0	0	100 ± 0.1
	200	194.7109	97.3 ± 0.1
	400	393.4837	98.3 ± 0.2
	600	566.3296	94.3 ± 0.1
	800	808.3139	101.1 ± 0.3
	1000	972.5175	97.2 ± 0.2

Table S2. Detection properties of reported turn-on fluorescent chemodosimeters for Hg²⁺ ions.

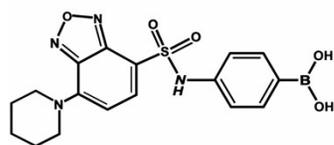
Probe Structure	$\lambda_{\text{ex}}/\lambda_{\text{em}}$ (nm)	Working solvent (v/v)	Saturation concentration of Hg ²⁺	Response time	LOD	Reference
	337/387	H ₂ O:CH ₃ OH (1:1)	1 equiv.	20 min	6.6 nM	1
	530/595	100% H ₂ O (HEPES Buffer, pH 7.4)	1 equiv.	30 min	0.49 nM	2
	490/570	100% H ₂ O (PBS Buffer, 7.4)	1 equiv.	~3 min	3 nM	3
	514/659	H ₂ O:DMSO (1:1) (PBS Buffer, pH 7.4)	15 equiv.	40 min	68 nM	4

	395/500	H ₂ O:DMSO (9:1) (HEPES Buffer, pH 7.4)	1 equiv.	30 min	90 nM	5
	500/556	H ₂ O:CH ₃ OH (20:80)	1 equiv.	5 min	10 nM	6
	380/462	H ₂ O:THF (0.5:9.5) (HEPES Buffer, pH 7.0)	200 equiv.	5 min	620 nM	7
	630/710	100% H ₂ O (HEPES buffer, pH 7.4)	1 equiv.	5 min	320 nM	8
	322/485	H ₂ O:CH ₃ CN (99.8:0.2)	1 equiv.	NA	6.2 nM	9



375/457 H₂O:CH₃CH₂OH
(1:2)
(PBS Buffer, pH
7.4)

1 equiv. 30 min 1.49 nM 10



440/585 H₂O:DMF
(99:1)
(HEPES Buffer,
pH 7.4)

1 equiv. 1~2 min 7.56 nM This Work

References

1. S. W. Lee, S. Y. Lee and S. H. Lee, *Tetrahedron Lett.*, 2019, 60, 151048.
2. Q. Duan, X. Lv, C. Liu, Z. Geng, F. Zhang, W. Sheng, Z. Wang, P. Jia, Z. Li, H. Zhu and B. Zhu, *Ind. Eng. Chem. Res.*, 2019, 58, 11–17.
3. A. Malek, K. Bera, S. Biswas, G. Perumal, A. K. Das, M. Doble, T. Thomas and E. Prasad, *Anal. Chem.*, 2019, 91, 3533–3538.
4. L. Huang, Z. Yang, Z. Zhou, Y. Li, S. Tang, W. Xiao, M. Hu, C. Peng, Y. Chen, B. Gu and H. Li, *Dyes Pigments*, 2019, 163, 118–125.
5. X. Cheng, S. Qu, L. Xiao, W. Li and P. He, *J. Photochem. Photobiol. A*, 2018, 364, 503–509.
6. Y. K. Yang, K. J. Yook and J. Tae, *J. Am. Chem. Soc.*, 2005, 127, 16760–16761.
7. V. Bhalla, Roopa, M. Kumar, P. R. Sharma and T. Kaur, *Dalton Trans.*, 2013, 42, 15063–15070.
8. Y. Wang, X. Hou, Z. Li, C. Liu, S. Hu, C. Li, Z. Xu and Y. Wang, *Dyes Pigments*, 2020, 173, 107951.
9. H. Jiang, W. Luo, J. Jiang, X. Jin, W. Dou, X. Tang, L. Yang, C. Chen, Z. Ju, X. Yao and W. Liu, *Sens. Actuators B Chem.*, 2014, 204, 68–73.
10. Y. Gao, T. Ma, Z. Ou, W. Cai, G. Yang, Y. Li, M. Xu and Q. Li, *Talanta*, 2018, 178, 663–669.