

Supporting Information for

**Asymmetric Oxidative Rearrangement of Tetrahydro- β -Carbolines
Catalyzed by Anionic Stereogenic-at-Cobalt(III) Complexes**

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1. Introduction

1.1. General Data

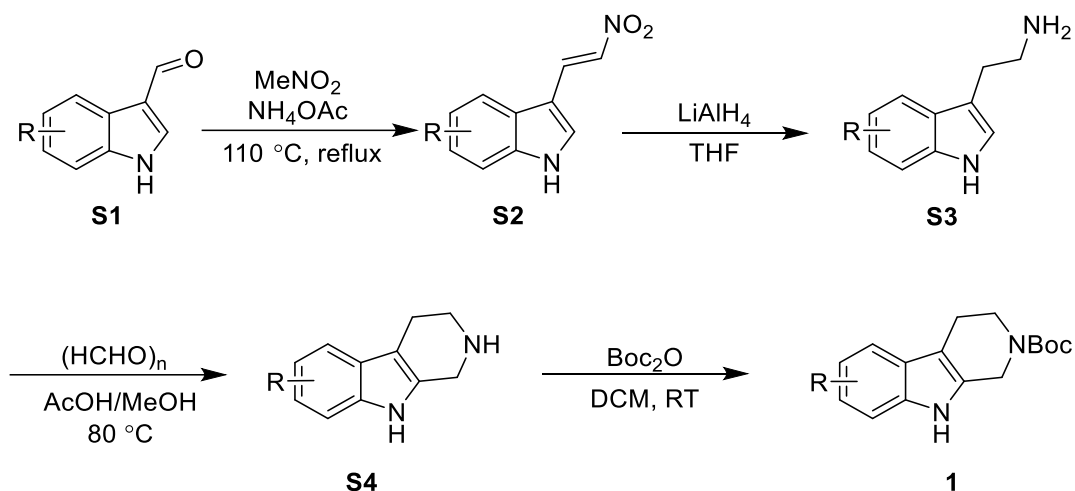
¹H NMR, ¹³C NMR, and ¹⁹F NMR spectra were recorded on an Agilent 600 NMR or Bruker-400 (or 500) MHz spectrometer at 600 MHz, 151 or 126 MHz, and 564 or 376 MHz using CDCl₃ as solvent, respectively. ¹H NMR data are reported as follows: δ, chemical shift; coupling constants (*J* are given in Hertz, Hz) and integration. Abbreviations to denote the multiplicity of a particular signal were s (singlet), d (doublet), t (triplet), q (quartet) and m (multiplet). Chemical shifts were reported in ppm from the tetramethylsilane with the solvent resonance as internal standard. Melting points were measured on a digital melting point apparatus and the temperature was uncorrected. High resolution mass spectrometric measurements (HRMS) were performed by the Waters Xevo G2-XS TOF (ESI Source). HPLC analysis was performed on Waters-Breeze (2487 Dual λ Absorbance Detector and 1525 Binary HPLC Pump, UV detection monitored at 254 nm). Chiralpak AD, AS, OJ, and OD columns were purchased from Daicel Chemical Industries, Ltd. Optical rotations were recorded on an Anton Paar MCP-100 polarimeter. Analytical-grade solvents for the column chromatography and commercially available reagents were used as received. *m*-Chloroperoxybenzoic acid was purified prior to use.

1.2. Materials

Analytical-grade solvents were used for column chromatography, and all commercially available reagents were used as received unless otherwise noted. *m*-CPBA was purified prior to use, and chloroform was distilled and dried before use. Catalysts (**Λ-1a** to **Λ-1e**) are known compounds and were synthesized according to the reported literature procedures.^[1] Indoles **1a–1g** and **1l–1w** are known compounds and were prepared following previously reported procedures,^[2–4] with their spectral data consistent with those reported in the literature. Indoles **1h–1k** and **1x** are new compounds and were synthesized according to the reported procedure.^[2–4]

2. General Procedure for the Preparation of Substrates

2.1 General Procedure for Preparation of Substrates 1a-1m



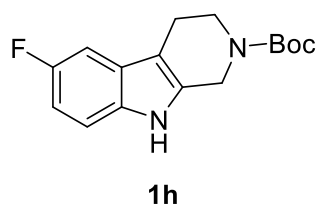
Step 1: Procedure for the synthesis of **S2**: To a solution of **S1** (1.0 equiv, 10 mmol) in nitromethane (15 mL) was added ammonium acetate (1.54 g, 2.0 equiv, 20 mmol). The reaction mixture was heated at 110 °C in an oil bath under reflux for 3 h. After cooling to room temperature, the organic layer was separated, and the aqueous phase was extracted with DCM (3 × 50 mL). The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The resulting crude nitroolefin **S2** was used directly in subsequent transformations without further purification.

Step 2: Procedure for the synthesis of **S3**: LiAlH₄ (1.9 g, 5.0 equiv, 50 mmol) was dissolved in anhydrous THF (to give a 2.5 M solution), and the corresponding nitroolefin **S2** was added dropwise at 0 °C. The reaction mixture was heated under reflux in an oil bath for 5 h and then allowed to cool to room temperature. The reaction was carefully quenched by the sequential addition of H₂O (1 mL) and 10% aqueous NaOH (2 mL) at 0 °C. The resulting mixture was filtered through Celite and washed with ethyl acetate (3 × 50 mL). The filtrate was concentrated under reduced pressure to afford the desired product **S3**.

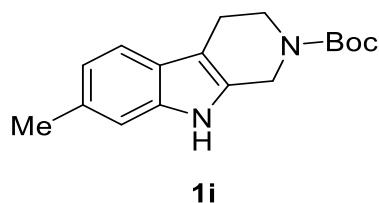
Step 3: Procedure for the synthesis of **S4**: To a solution of substituted tryptamine **S3** (1.0 equiv, 10 mmol) in a mixture of AcOH/MeOH (v/v = 2:5, 28 mL) was added paraformaldehyde (360 mg, 1.2 equiv, 12 mmol). The reaction mixture was heated at 80 °C for 3 h and then cooled to 0 °C. The mixture was basified to pH 9–10 by the sequential addition of saturated aqueous NaOH and NaHCO₃

solutions (NaOH to pH 4–5, followed by NaHCO₃ to pH 8, and finally a few drops of NaOH to adjust the pH to 9–10). The resulting mixture was extracted with DCM (3 × 30 mL). The combined organic layers were dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure to afford the crude amine **S4**.

Step 4: Procedure for the synthesis of **1**: At 0 °C, Boc₂O (2.18 g, 1.0 equiv, 10 mmol) was added to a solution of the crude amine **S4** in DCM (15 mL). The reaction mixture was allowed to warm to room temperature and stirred at this temperature, with the reaction progress monitored by TLC. Upon completion (≈2 h), saturated aqueous NaHCO₃ (15 mL) was added. The layers were separated, and the aqueous phase was extracted with ethyl acetate (3 × 30 mL). The combined organic layers were dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. Purification of the resulting crude residue by silica gel flash column chromatography (gradient elution with EtOAc in petroleum ether) afforded substrates **1a–1m**.

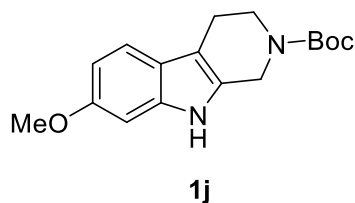


tert-Butyl 6-fluoro-1,3,4,9-tetrahydro-2H-pyrido[3,4-b]indole-2-carboxylate 1h: yield: 42% (1.218 g); (Flash column chromatography eluent, petrol ether/EtOAc = 3/1); yellow solid, m.p. 68.3–69.1 °C; ¹H NMR (600 MHz, CDCl₃) δ 8.76 (s, 1H), 7.23 – 7.19 (m, 1H), 7.11 (d, *J* = 9.4 Hz, 1H), 6.91 – 6.85 (m, 1H), 4.72 – 4.57 (m, 2H), 3.80 – 3.72 (m, 2H), 2.75 (t, *J* = 5.8 Hz, 2H), 1.55 (s, 9H); ¹³C NMR (151 MHz, CDCl₃) δ 158.0 (d, *J* = 234.8 Hz), 155.6, 132.8, 127.5, 111.5, 109.7 (d, *J* = 25.2 Hz), 103.2, 80.5, 60.6, 42.7, 41.4, 28.7, 21.5, 14.3; ¹⁹F NMR (565 MHz, CDCl₃) δ -124.90 ppm; **HRMS (ESI)** calculated for C₁₆H₂₀FN₂O₂ [M+H]⁺: 291.1509, found: 291.1510.

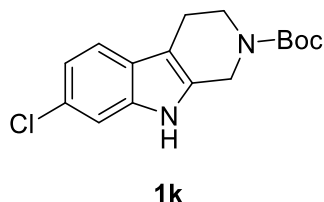


tert-Butyl 7-methyl-1,3,4,9-tetrahydro-2H-pyrido[3,4-b]indole-2-carboxylate 1i: yield: 45% (1.287 g); (Flash column chromatography eluent, petrol ether/EtOAc = 3/1); white solid. m.p. 103.7–

104.8 °C; $^1\text{H NMR}$ (600 MHz, CDCl_3) δ 8.15 (s, 1H), 7.38 – 7.34 (m, 1H), 7.09 (s, 1H), 6.94 (d, J = 8.0 Hz, 1H), 4.63 (s, 2H), 3.76 (s, 2H), 2.79 (s, 2H), 2.45 (s, 3H), 1.53 (s, 9H); $^{13}\text{C NMR}$ (151 MHz, CDCl_3) δ 155.5, 136.8, 131.5, 130.1, 125.0, 121.2, 117.6, 111.1, 80.2, 42.8, 41.9, 28.8, 28.2, 21.6; **HRMS (ESI)** calculated for $\text{C}_{17}\text{H}_{23}\text{N}_2\text{O}_2$ $[\text{M}+\text{H}]^+$: 287.1760, found: 287.1763.

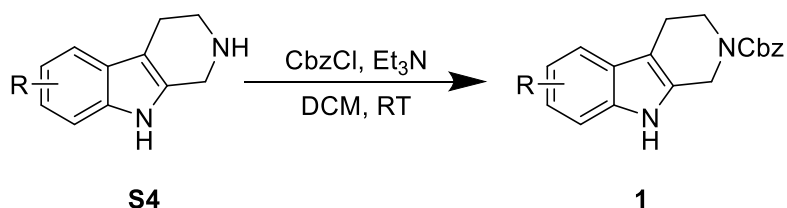


tert-Butyl 7-methoxy-1,3,4,9-tetrahydro-2H-pyrido[3,4-b]indole-2-carboxylate 1j: yield: 45% (1.431 g); (Flash column chromatography eluent, petrol ether/EtOAc = 3/1); white solid, m.p. 185.7–186.8 °C; $^1\text{H NMR}$ (600 MHz, CDCl_3) δ 8.18 (s, 1H), 7.34 (d, J = 8.6 Hz, 1H), 6.83 (d, J = 2.3 Hz, 1H), 6.79 – 6.73 (m, 1H), 4.66 – 4.54 (m, 2H), 3.83 (s, 3H), 3.78 – 3.72 (m, 2H), 2.78 – 2.73 (m, 2H), 1.53 (s, 9H); $^{13}\text{C NMR}$ (151 MHz, CDCl_3) δ 156.3, 137.1, 129.5, 121.6, 118.5, 109.0, 95.2, 80.2, 55.6, 42.7, 41.5, 28.6, 21.6.; **HRMS (ESI)** calculated for $\text{C}_{17}\text{H}_{22}\text{N}_2\text{NaO}_4$ $[\text{M}+\text{Na}]^+$: 325.1528, found: 325.1531.

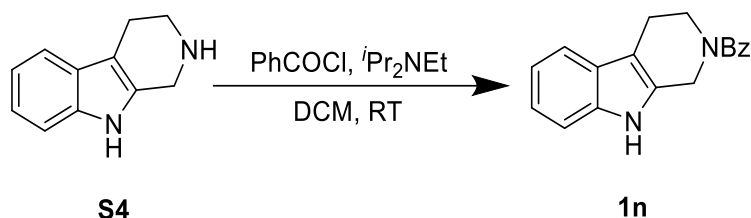


tert-Butyl 7-chloro-1,3,4,9-tetrahydro-2H-pyrido[3,4-b]indole-2-carboxylate 1k: yield: 45% (1.377 g); (Flash column chromatography eluent, petrol ether/EtOAc = 3/1); white solid, m.p. 196.7–197.8 °C; $^1\text{H NMR}$ (600 MHz, CDCl_3) δ 8.72 (s, 1H), 7.36 (d, J = 8.4 Hz, 1H), 7.28 (s, 1H), 7.11 – 7.01 (m, 1H), 4.65 (s, 2H), 3.76 (s, 2H), 2.76 (s, 2H), 1.54 (s, 9H); $^{13}\text{C NMR}$ (151 MHz, CDCl_3) δ 155.6, 136.7, 131.6, 127.4, 125.8, 120.1, 118.8, 111.0, 108.6, 80.5, 42.6, 41.3, 28.6, 21.5; **HRMS (ESI)** calculated for $\text{C}_{16}\text{H}_{20}\text{ClN}_2\text{O}_2$ $[\text{M}+\text{H}]^+$: 307.1213, found: 307.1216.

2.2 General Procedure for Preparation of Substrates 1n-1r

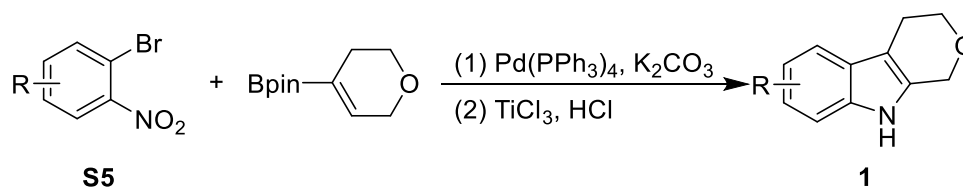


At 0 °C, to a solution of the crude amine **S4** in DCM (15 mL) was added Et₃N (1.4 mL, 1.0 equiv, 10 mmol) and CbzCl (1.5 mL, 1.05 equiv, 10.5 mmol). The resulting mixture was allowed to warm to room temperature and stirred at the same temperature. The reaction progress was monitored by TLC. Upon completion (~ 2 h), a solution of saturated aqueous solution of NaHCO₃ (15 mL) was added. The layers were separated, and the aqueous layer was extracted with ethyl acetate (30 mL × 3). The combined organic layers were dried over anhydrous Na₂SO₄, filtered, and concentrated. Purification of the resulting crude residue via silica gel flash column chromatography (gradient eluent: EtOAc in petrol ether) afforded the substrate **1o-1r**. The spectral data of **1o-1r** have been reported in previous literatures^[2-4].



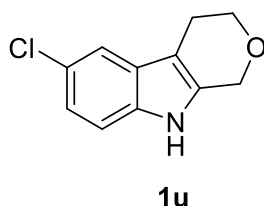
To a solution of **S4** (0.86 g, 5.0 mmol) in DCM (20 mL) was added *t*Pr₂NEt (0.97 g, 1.5 equiv, 7.5 mmol) and benzoyl chloride (0.89 g, 1.5 equiv, 7.5 mmol) at 0 °C under N₂. The resulting mixture was allowed to warm to room temperature and stirred at the same temperature. The reaction progress was monitored by TLC. Upon completion (~3 h), the solvent was evaporated to give the crude product, which was purified by flash column chromatography on silica gel (eluent: hexanes/DCM = 1:1 to 1:2) to afford pure **1n** as a brown solid. The spectral data of **1n** have been reported in previous literature^[2].

2.3 General Procedure for Preparation of Substrates 1s-1x



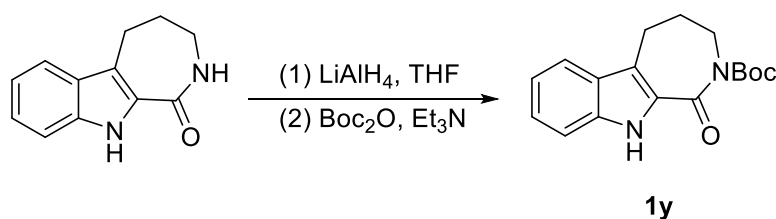
To a solution of **S5** (1.0 equiv, 10 mmol) and 1-cyclohexen-1-yl-boronic acid pinacol ester (2.5 g, 1.2 equiv, 12 mmol) in a mixed solvent of toluene, EtOH, and H₂O (v/v/v = 5:2:1, 64 mL) were added Pd(PPh₃)₄ (1.16 g, 10 mol%, 1 mmol) and K₂CO₃ (5.5 g, 4.0 equiv, 40 mmol). The resultant mixture was then purged with N₂ and heated to 90 °C. The reaction progress was monitored by TLC. Upon completion (~24 h), the reaction mixture was cooled to room temperature and diluted with water (20 mL) and ethyl acetate (20 mL). The layers were separated, and the aqueous layer was extracted with

ethyl acetate (20 mL × 3). The combined organic layers were washed with water, dried over Na₂SO₄, and concentrated. The residue was dissolved in acetonitrile (40 mL) and water (40 mL). A solution of TiCl₃ (15-20% in 30% hydrochloric acid, 40 mL) was added. The mixture was stirred at room temperature and the progress was monitored by TLC. Upon completion (~5 h), the mixture was extracted with ethyl acetate (20 mL × 3). The combined organic layers were washed with a saturated aqueous NaHCO₃ solution and brine, then dried over Na₂SO₄, and concentrated. The residue was purified by column chromatography on silica gel to afford pure **1s-1x**. The spectral data of **1s-1w** have been reported in previous literatures^[2-4].



6-Chloro-1,3,4,9-tetrahydropyrano[3,4-b]indole 1u: yield: 65% (1.346 g); (Flash column chromatography eluent, petrol ether/EtOAc = 3/1); yellow solid, m.p. 116.7-117.8 °C; ¹H NMR (600 MHz, CDCl₃) δ 7.81 (s, 1H), 7.24 – 7.19 (m, 1H), 7.17 – 7.11 (m, 1H), 6.93 – 6.86 (m, 1H), 4.82 – 4.78 (m, 2H), 4.06 – 4.00 (m, 2H), 2.84 – 2.75 (m, 2H); ¹³C NMR (151 MHz, CDCl₃) δ 134.7, 133.2, 132.2, 130.5, 128.4, 125.4, 121.9, 117.7, 112.0, 107.5, 75.2, 65.5, 63.6, 29.8, 25.0, 22.1; **HRMS (ESI)** calculated for C₁₁H₁₁ClNO [M+H]⁺: 208.0529, found: 208.0521.

2.4 General Procedure for Preparation of Substrates 1y

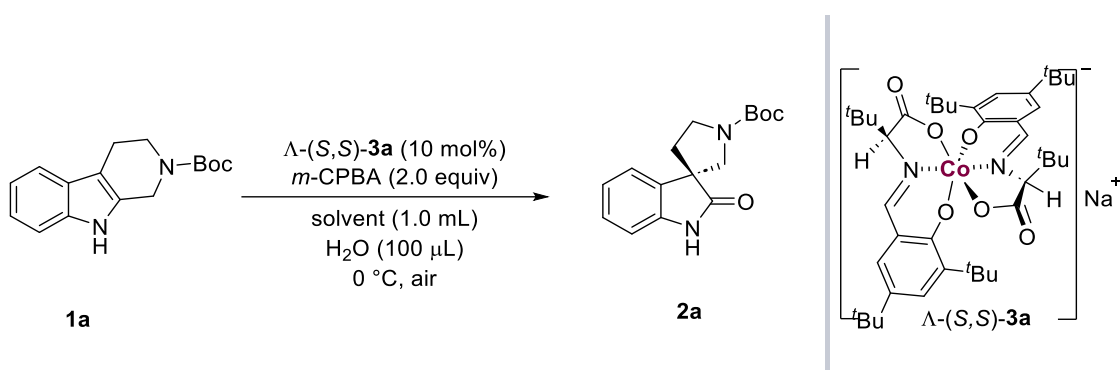


Under N₂ at room temperature, to a solution of 3,4,5,10-tetrahydroazepino[3,4 b]indol-1(2H)-one^[5] (1.0 g, 5.0 mmol) in anhydrous THF (40 mL) was added LiAlH₄ (1.0 g, 5.0 equiv, 25.0 mmol). The mixture was heated to 90 °C and stirred for 6 h before it was cooled to 0 °C and treated successively with water (1 mL), a saturated aqueous NaOH solution (1 mL), and water (3.0 mL). This mixture was stirred for a few minutes and filtered. The filtrate was concentrated in vacuo to give the crude amine. The crude amine was redissolved in DCM (20 mL). To this solution were added Boc₂O (1.09 g, 1.0

equiv, 5.0 mmol) and Et₃N (505 mg, 1.0 equiv, 5.0 mmol). The mixture was stirred at room temperature, and the reaction progress was monitored by TLC. Upon completion (~ 2 h), the solvent was evaporated, and the residue was purified by column chromatography on silica gel (eluent: hexanes/ethyl acetate = 5:1) to afford the pure indole **1y** as a white solid (428 mg, 30% yield for two steps). Due to the substitution pattern of the amide functionality, this compound is a mixture of two conformational isomers. The spectral data of **1y** have been reported in previous literatures^[2].

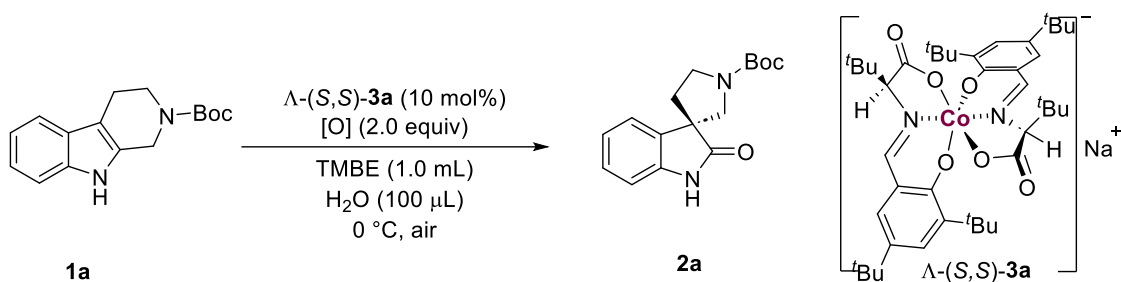
3. Optimization of reaction conditions

Table S1. Optimization for solvent^a.



entry	solvent	yield (%) ^b	e.r. ^c
1	THF	40	55.5:45.5
2	CHCl ₃	62	69:31
3	Et ₂ O	59	67.5:32.5
4	EtOAc	60	61.5:38.5
5	DCM	42	74:26
6	CH ₃ CN	43	63:37
7	TBME	60	82.5:17.5
8	1,4-dioxane	63	67.5:32.5
9	Tol	20	61:39
10	Acetone	40	65:35

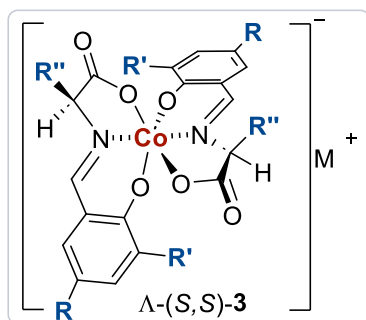
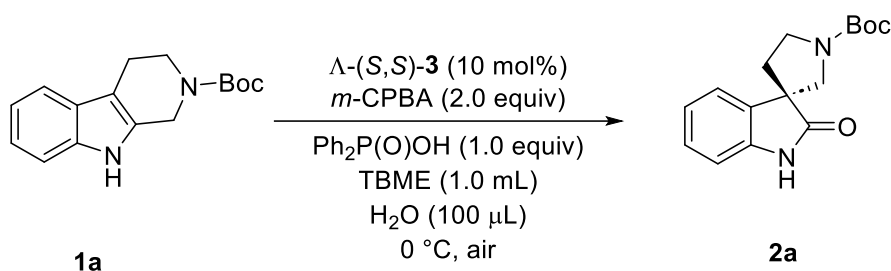
^a The reactions were carried out with **1a** (0.1 mmol), H₂O (100 μ L), Λ -(S,S)-**3a** (10 mol%), and *m*-CPBA (0.2 mmol) in solvent (1.0 mL) at 0 °C. ^b Isolated yields were based on **1a**. ^c The e.r. values were determined by chiral stationary HPLC.

Table S2. Optimization for oxidant reagent^a.

entry	oxidant	yield (%) ^b	e.r. ^c
1	NCS	83	53.5:46.5
2	NBS	78	76:24
3	NIS	82	68:32
4	I ₂	80	56.5:43.5
5	<i>m</i> -CPBA	60	82.5:17.5

^a The reactions were carried out with **1a** (0.1 mmol), H₂O (100 μ L), Δ -(S,S)-**3a** (10 mol%), and [O] (0.2 mmol) in TBME (1.0 mL) at 0 °C. ^b Isolated yields were based on **1a**. ^c The e.r. values were determined by chiral stationary HPLC.

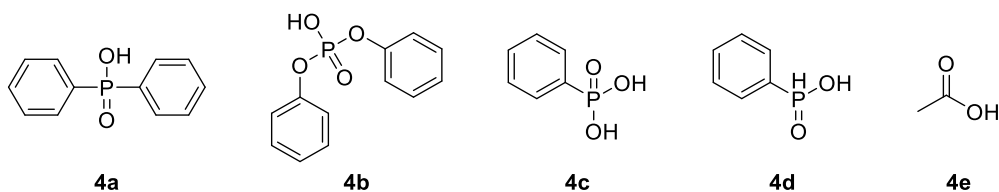
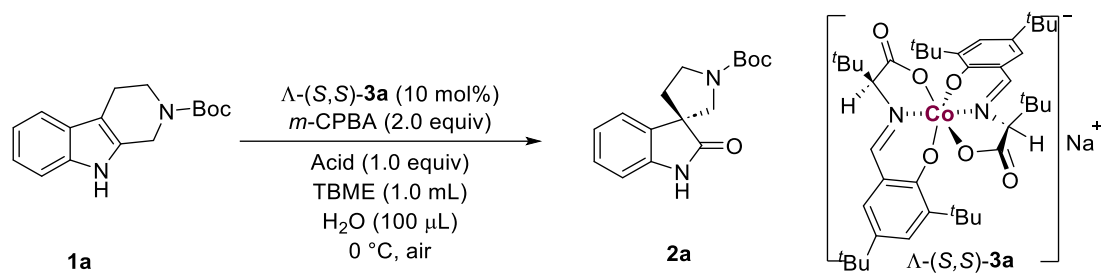
Table S3. Optimization for chiral catalyst^a.



- 3a**, R = R' = R'' = ^tBu, M = Na;
3b, R = R' = R'' = ^tBu, M = H;
3c, R = R' = ^tBu, R'' = ⁱPr, M = Na;
3d, R = R' = ^tBu, R'' = ⁱPr, M = H;
3e, R = R' = ^tAmyl, R'' = ⁱPr, M = Na;
3f, R = R' = ^tAmyl, R'' = ^tBu, M = Na;
3g, R = R' = ^tAmyl, R'' = ^tBu, M = H;
3h, R = R' = ^tAmyl, R'' = ^tBu, M = Cs;

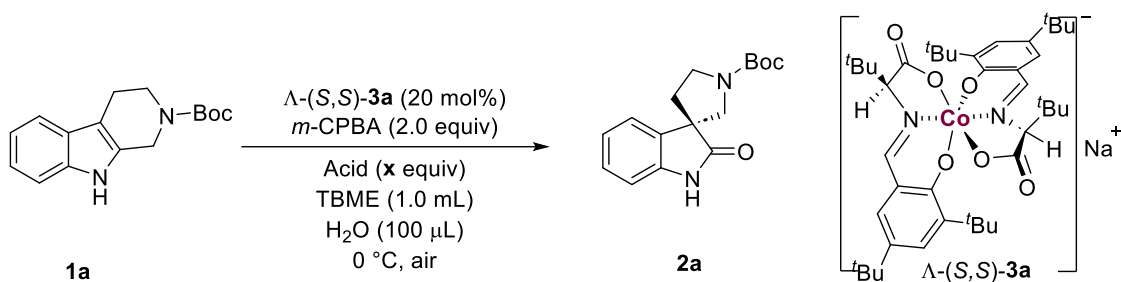
entry	3	yield (%) ^b	e.r. ^c
1	Λ -(<i>S,S</i>)- 3a	56	90:10
2	Λ -(<i>S,S</i>)- 3b	65	83.5:16.5
3	Λ -(<i>S,S</i>)- 3c	52	89:11
4	Λ -(<i>S,S</i>)- 3d	65	83.5:16.5
5	Λ -(<i>S,S</i>)- 3e	36	86:14
6	Λ -(<i>S,S</i>)- 3f	58	86:14
7	Λ -(<i>S,S</i>)- 3g	58	85.5:14.5
8	Λ -(<i>S,S</i>)- 3h	58	78.5:21.5

^a The reactions were carried out with **1a** (0.1 mmol), H₂O (100 μL), Ph₂P(O)OH (0.1 mmol), Λ -(*S,S*)-**3** (10 mol%), and *m*-CPBA (0.2 mmol) in TBME (1.0 mL) at 0 °C. ^b Isolated yields were based on **1a**. ^c The e.r. values were determined by chiral stationary HPLC.

Table S4. Optimization for acid^a.

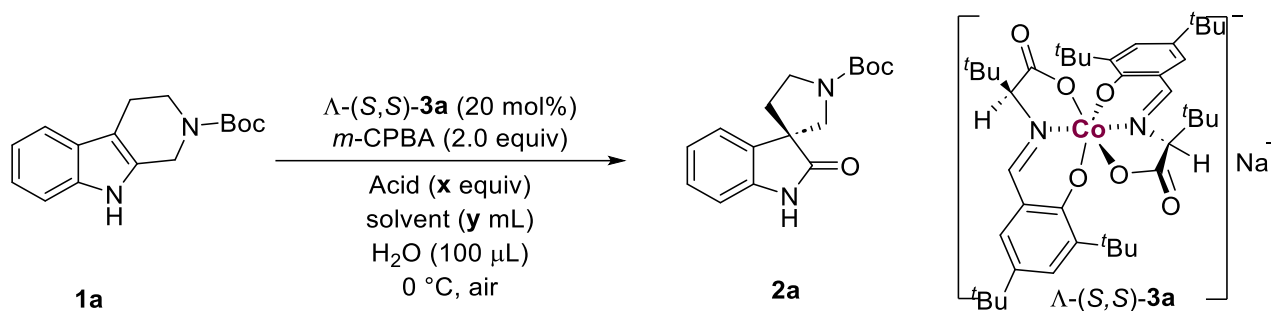
entry	acid	yield (%) ^b	e.r. ^c
1	4a	52	90:10
2	4b	40	61:39
3	4c	59	55:45
4	4d	32	67:33
5	4e	56	90:10

^aThe reactions were carried out with **1a** (0.1 mmol), H₂O (100 μ L), $\Delta\text{-(S, S)-3a}$ (10 mol%), acid (0.1 mmol) and *m*-CPBA (0.2 mmol) in TBME (1.0 mL) at 0 °C. ^bIsolated yields were based on **1a**. ^cThe e.r. values were determined by chiral stationary HPLC.

Table S5. Optimization for the equivalent of chiral catalysts and acid^a.

entry	Acid	x	yield (%) ^b	e.r. ^c
1	4a	0.2	65	92:8
2	4a	0.5	63	92:8
3	4a	0.7	65	94:6
4	4a	1.0	65	92:8
5	4e	1.0	60	91.5:8.5
6	4e	5.0	63	92:8
7	4e	10.0	68	94:6
8	4e	15.0	65	91.5:8.5

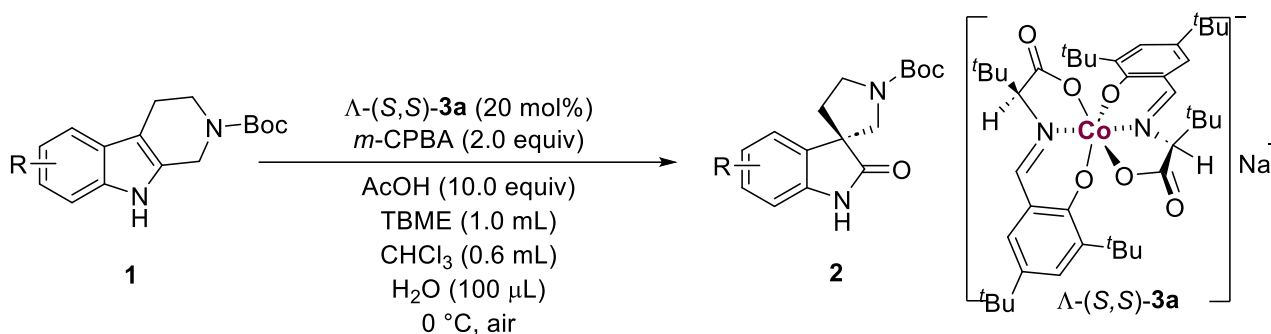
^a The reactions were carried out with **1a** (0.1 mmol), H₂O (100 μL), $\Lambda\text{-(S, S)-3a}$ (20 mol%), acid (x mmol), and *m*-CPBA (0.2 mmol) in TBME (1.0 mL) at 0 °C. ^b Isolated yields were based on **1a**. ^c The e.r. values were determined by chiral stationary HPLC.

Table S6. Optimization for mixed solvent^a.

entry	Acid	x	Solvent	y	yield (%) ^b	e.r. ^c
1	4a	0.7	TBME/Tol (5:1)	1.2	62	91:9
2	4a	0.7	TBME/DCM (5:1)	1.2	60	90:10
3	4a	0.7	TBME/CHCl ₃ (5:1)	1.2	63	91:9
4	4e	10.0	TBME/Tol (5:1)	1.2	70	92.5:7.5
5	4e	10.0	TBME/DCM (5:1)	1.2	73	92.5:7.5
6	4e	10.0	TBME/CHCl ₃ (5:1)	1.2	66	93:7
7	4e	10.0	TBME/CHCl ₃ (5:2)	1.4	65	93.5:6.5
8	4e	10.0	TBME/CHCl ₃ (5:3)	1.6	68	94.5:5.5

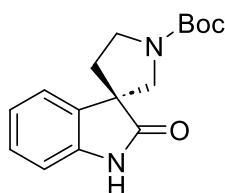
^a The reactions were carried out with **1a** (0.1 mmol), H₂O (100 μL), Δ-(S, S)-**3a** (20 mol%), acid (x mmol), and *m*-CPBA (0.2 mmol) in TBME/Solvent (y mL) at 0 °C. ^b Isolated yields were based on **1a**. ^c The e.r. values were determined by chiral stationary HPLC.

4. Experimental Procedures of Asymmetric Reaction and Characterization Data



General Procedure:

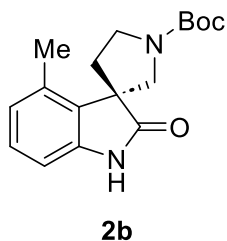
At room temperature, a 10 mL vial was charged with Λ -(*S,S*)-**3a** (16 mg, 20 mol%), H₂O (100 μ L), AcOH (60 μ L), and indole **1** (0.10 mmol), after which a mixed solvent of TBME/CHCl₃ (1.6 mL, v/v = 5:3) was added. The resulting mixture was cooled to 0 °C and stirred for 20 min, followed by the addition of *m*-CPBA (34.5 mg, 2.0 equiv, 0.20 mmol). The reaction mixture was stirred at 0 °C, and the progress of the reaction was monitored by TLC. Upon completion, the reaction was quenched by the addition of saturated aqueous Na₂S₂O₃ solution (3 mL). The mixture was extracted with ethyl acetate (3 \times 10 mL). The combined organic layers were washed with saturated aqueous NaHCO₃ solution and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. Purification of the residue by silica gel column chromatography afforded the pure product. Note: Compounds **2a–2m** were observed as a mixture of slowly interconverting rotamers in solution due to restricted rotation around the N–C(O) bond of the Boc carbamate. Accordingly, duplicated and/or broadened signals are observed in the ¹H and ¹³C NMR spectra recorded at room temperature.



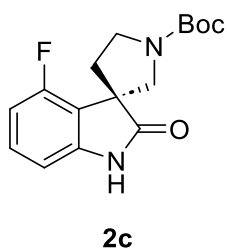
2a

tert-Butyl (*R*)-2-oxospiro[indoline-3,3'-pyrrolidine]-1'-carboxylate **2a:** yield: 68% (19.6 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; brown oil; $[\alpha]_D^{20} = -21.2$ (c 0.13 CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ 8.38/8.28 (s, 1H), 7.26 – 7.15 (m, 2H), 7.09 – 7.01 (m, 1H), 6.97 – 6.89 (m, 1H), 3.88 – 3.75 (m, 2H), 3.74 – 3.66 (m, 1H), 3.66 – 3.54 (m, 1H), 2.45 –

2.37 (m, 1H), 2.12 – 2.04 (m, 1H), 1.51/1.46 (s, 9H); ^{13}C NMR (151 MHz, CDCl_3) δ 180.2, 154.6, 140.2, 133.2, 128.5, 123.0, 110.2, 80.0, 54.3, 53.0, 45.3, 36.5, 35.7, 28.6; **HRMS (ESI)** calculated for $\text{C}_{16}\text{H}_{20}\text{N}_2\text{NaO}_3$ $[\text{M}+\text{Na}]^+$: 311.1472, found: 311.1475; **Enantiomeric ratio**: 94.5:5.5, determined by HPLC (Daicel Chirapak AD, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, $T = 30^\circ\text{C}$, $\lambda = 254$ nm): $t_{\text{R}} = 8.80$ min (minor), $t_{\text{R}} = 12.25$ min (major). The absolute configuration of **2a** was assigned by comparison of its optical rotation with the reported literature data^[2].

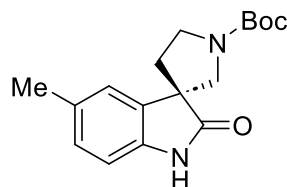


tert-Butyl (R)-4-methyl-2-oxospiro[indoline-3,3'-pyrrolidine]-1'-carboxylate 2b: yield: 60% (18.1 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_{\text{D}}^{20} = -20.8$ (c 0.08 CH_2Cl_2); ^1H NMR (600 MHz, CDCl_3) δ 8.58/8.46 (s, 1H), 7.12 (t, $J = 7.8$ Hz, 1H), 6.86 – 6.80 (m, 1H), 6.78 – 6.70 (m, 1H), 3.89 – 3.71 (m, 4H), 2.46 – 2.38 (m, 1H), 2.35 (s, 3H), 2.26 – 2.16 (m, 1H), 1.50/1.46 (s, 9H); ^{13}C NMR (151 MHz, CDCl_3) δ 182.2, 154.2, 141.0, 134.6, 128.5, 125.5, 107.9, 79.9, 52.0, 45.7, 34.0, 29.8, 28.6, 18.0; **HRMS (ESI)** calculated for $\text{C}_{17}\text{H}_{22}\text{N}_2\text{NaO}_3$ $[\text{M}+\text{Na}]^+$: 325.1528, found: 325.1530; **Enantiomeric ratio**: 80.5:19.5, determined by HPLC (Daicel Chirapak AD, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, $T = 30^\circ\text{C}$, $\lambda = 254$ nm): $t_{\text{R}} = 8.95$ min (minor), $t_{\text{R}} = 11.95$ min (major).



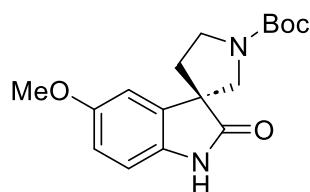
tert-Butyl (R)-4-fluoro-2-oxospiro[indoline-3,3'-pyrrolidine]-1'-carboxylate 2c: yield: 62% (19.0 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_{\text{D}}^{20} = -15.3$ (c 0.12 CH_2Cl_2); ^1H NMR (600 MHz, CDCl_3) δ 7.87/7.80 (s, 1H), 7.23 – 7.17 (m, 1H), 6.78 – 6.64 (m, 2H), 3.88 – 3.83 (m, 1H), 3.83 – 3.76 (m, 2H), 3.76 – 3.71 (m, 1H), 2.43 – 2.37 (m, 1H), 2.37 – 2.32 (m, 1H), 1.50/1.46 (s, 9H); ^{13}C NMR (151 MHz, CDCl_3) δ 180.4, 158.9 (d, $J = 247.5$ Hz), 154.4, 142.6, 130.5, 110.5 (d, $J = 21.2$ Hz), 106.3 (d, $J = 3.8$ Hz), 79.9, 53.0, 52.5, 45.5, 34.9,

29.9; ^{19}F NMR (376 MHz, CDCl_3) δ -118.99 ppm; **HRMS (ESI)** calculated for $\text{C}_{16}\text{H}_{19}\text{FN}_2\text{NaO}_3$ $[\text{M}+\text{Na}]^+$: 329.1277, found: 329.1281; **Enantiomeric ratio**: 94.5:5.5, determined by HPLC (Daicel Chirapak AD, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, $T = 30\text{ }^\circ\text{C}$, $\lambda = 254\text{ nm}$): $t_{\text{R}} = 8.06\text{ min}$ (minor), $t_{\text{R}} = 13.57\text{ min}$ (major).



2d

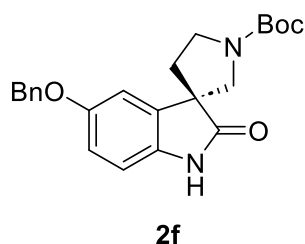
tert-Butyl (R)-5-methyl-2-oxospiro[indoline-3,3'-pyrrolidine]-1'-carboxylate 2d: yield: 62% (18.7 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_{\text{D}}^{20} = -21.4$ (c 0.18 CH_2Cl_2); ^1H NMR (600 MHz, CDCl_3) δ 8.75/8.65 (s, 1H), 7.09 – 6.94 (m, 2H), 6.87 – 6.77 (m, 1H), 3.88 – 3.75 (m, 2H), 3.73 – 3.65 (m, 1H), 3.65 – 3.53 (m, 1H), 2.44 – 2.39 (m, 1H), 2.32 (s, 3H), 2.10 – 2.02 (m, 1H), 1.52/1.46 (s, 9H); ^{13}C NMR (151 MHz, CDCl_3) δ 180.5, 154.7, 137.8, 133.4, 132.7, 128.8, 123.6, 109.9, 80.0, 54.6, 53.6, 45.6, 36.5, 28.7, 21.3; **HRMS (ESI)** calculated for $\text{C}_{17}\text{H}_{22}\text{N}_2\text{NaO}_3$ $[\text{M}+\text{Na}]^+$: 325.1528, found: 325.1533; **Enantiomeric ratio**: 93:7, determined by HPLC (Daicel Chirapak AD, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, $T = 30\text{ }^\circ\text{C}$, $\lambda = 254\text{ nm}$): $t_{\text{R}} = 7.95\text{ min}$ (minor), $t_{\text{R}} = 12.23\text{ min}$ (major).



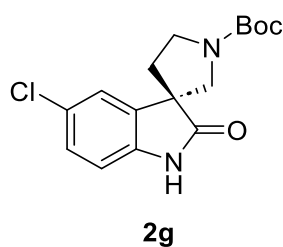
2e

tert-Butyl (R)-5-methoxy-2-oxospiro[indoline-3,3'-pyrrolidine]-1'-carboxylate 2e: yield: 65% (20.7 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_{\text{D}}^{20} = -38.8$ (c 0.092 CH_2Cl_2); ^1H NMR (600 MHz, CDCl_3) δ 8.78/8.70 (s, 1H), 6.87 – 6.82 (m, 1H), 6.80 – 6.75 (m, 2H), 3.89 – 3.81 (m, 1H), 3.77 (s, 3H), 3.76 – 3.69 (m, 2H), 3.64 – 3.52 (m, 1H), 2.43 – 2.38 (m, 1H), 2.08 – 2.02 (m, 1H), 1.54/1.45 (s, 9H); ^{13}C NMR (151 MHz, CDCl_3) δ 180.1, 156.4, 154.5, 134.6, 133.5, 112.9, 110.5, 110.2, 80.0, 56.0, 54.5, 45.3, 36.5, 35.7, 28.6; **HRMS (ESI)** calculated for $\text{C}_{17}\text{H}_{22}\text{N}_2\text{NaO}_4$ $[\text{M}+\text{Na}]^+$: 341.1477, found: 341.1477; **Enantiomeric ratio**: 91:9, determined by HPLC (Daicel Chirapak AD, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, $T = 30\text{ }^\circ\text{C}$, $\lambda = 254\text{ nm}$): $t_{\text{R}} = 7.95\text{ min}$ (minor), $t_{\text{R}} = 12.23\text{ min}$ (major).

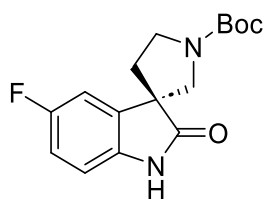
= 30 °C, λ = 254 nm): t_R = 12.30 min (minor), t_R = 18.42 min (major).



tert-Butyl (R)-5-(benzyloxy)-2-oxospiro[indoline-3,3'-pyrrolidine]-1'-carboxylate 2f: yield: 58% (22.9 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_D^{20}$ = -27.3 (c 0.10 CH₂Cl₂); **¹H NMR** (600 MHz, CDCl₃) δ 8.04/7.96 (s, 1H), 7.47 – 7.23 (m, 5H), 6.93 – 6.75 (m, 3H), 5.01 (s, 2H), 3.82 – 3.66 (m, 4H), 2.45 – 2.37 (m, 1H), 2.07 – 2.00 (m, 1H), 1.51/1.46 (s, 9H); **¹³C NMR** (151 MHz, CDCl₃) δ 178.0, 155.5, 154.5, 136.9, 134.6, 133.9, 128.7, 128.2, 127.7, 114.0, 111.3, 110.5, 80.0, 71.0, 54.5, 53.0, 45.3, 35.6, 29.7; **HRMS (ESI)** calculated for C₂₃H₂₆N₂NaO₄ [M+Na]⁺: 417.1790, found: 417.1795; **Enantiomeric ratio:** 89:11, determined by HPLC (Daicel Chirapak AD, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, T = 30 °C, λ = 254 nm): t_R = 13.62 min (minor), t_R = 14.64 min (major).

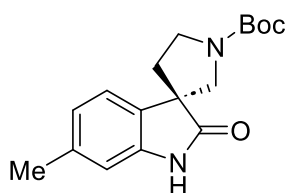


tert-Butyl (R)-5-chloro-2-oxospiro[indoline-3,3'-pyrrolidine]-1'-carboxylate 2g: yield: 68% (21.9 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_D^{20}$ = -16.5 (c 0.21 CH₂Cl₂); **¹H NMR** (600 MHz, CDCl₃) δ 8.20/8.14 (s, 1H), 7.25 – 7.11 (m, 2H), 6.88 – 6.81 (m, 1H), 3.90 – 3.81 (m, 1H), 3.76 – 3.69 (m, 2H), 3.65 – 3.53 (m, 1H), 2.45 – 2.39 (m, 1H), 2.07 (m, 1H), 1.52/1.47 (s, 9H); **¹³C NMR** (151 MHz, CDCl₃) δ 179.9, 154.5, 138.9, 134.8, 134.5, 128.5, 123.4, 111.3, 80.3, 54.4, 53.9, 45.4, 35.6, 28.6; **HRMS (ESI)** calculated for C₁₆H₁₉ClN₂NaO₃ [M+Na]⁺: 345.0982, found: 345.0986; **Enantiomeric ratio:** 92.5:7.5, determined by HPLC (Daicel Chirapak AD, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, T = 30 °C, λ = 254 nm): t_R = 7.70 min (minor), t_R = 13.57 min (major).



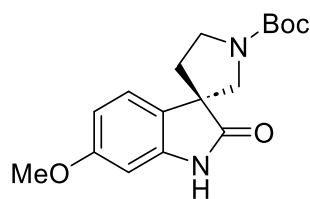
2h

tert-Butyl (R)-5-fluoro-2-oxospiro[indoline-3,3'-pyrrolidine]-1'-carboxylate 2h: yield: 63% (19.1 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_D^{20} = -9.0$ (c 0.21 CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ 8.43/8.37 (s, 1H), 6.98 – 6.82 (m, 3H), 3.91 – 3.75 (m, 2H), 3.74 – 3.70 (m, 1H), 3.65 – 3.53 (m, 1H), 2.46 – 2.39 (m, 1H), 2.10 – 2.04 (m, 1H), 1.51/1.46 (s, 9H); ¹³C NMR (151 MHz, CDCl₃) δ 180.2, 159.5 (d, *J* = 242.0 Hz), 154.5, 136.2, 115.0 (d, *J* = 23.1 Hz), 110.9 (d, *J* = 7.8 Hz), 80.2, 59.6, 54.2 (d, *J* = 48.0 Hz), 45.3, 38.3, 28.6; ¹⁹F NMR (565 MHz, CDCl₃) δ -119.58 ppm; **HRMS (ESI)** calculated for C₁₆H₁₉FN₂NaO₃ [M+Na]⁺: 329.1277, found: 329.1281; **Enantiomeric ratio**: 94:6, determined by HPLC (Daicel Chirapak AD, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, T = 30 °C, λ = 254 nm): t_R = 8.08 min (minor), t_R = 14.03 min (major).



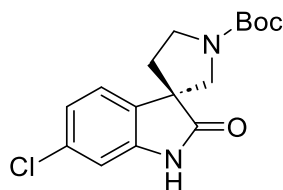
2i

tert-Butyl (R)-6-methyl-2-oxospiro[indoline-3,3'-pyrrolidine]-1'-carboxylate 2i: yield: 60% (18.1 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_D^{20} = -9.5$ (c 0.25 CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ 8.75/8.65 (s, 1H), 7.09 – 6.94 (m, 2H), 6.87 – 6.77 (m, 1H), 3.88 – 3.75 (m, 2H), 3.73 – 3.65 (m, 1H), 3.65 – 3.53 (m, 1H), 2.44 – 2.39 (m, 1H), 2.32 (s, 3H), 2.10 – 2.02 (m, 1H), 1.52/1.46 (s, 9H); ¹³C NMR (151 MHz, CDCl₃) δ 180.9, 154.6, 140.4, 138.8, 130.3, 123.6, 122.6, 111.1, 80.0, 54.6, 53.3, 45.6, 36.6, 28.7, 21.7; **HRMS (ESI)** calculated for C₁₇H₂₂N₂NaO₃ [M+Na]⁺: 325.1528, found: 325.1533; **Enantiomeric ratio**: 92:8, determined by HPLC (Daicel Chirapak AD, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, T = 30 °C, λ = 254 nm): t_R = 10.47 min (minor), t_R = 19.04 min (major).



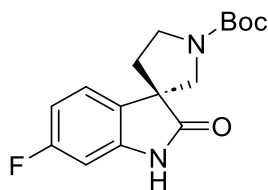
2j

tert-Butyl (R)-6-methoxy-2-oxospiro[indoline-3,3'-pyrrolidine]-1'-carboxylate 2j: yield: 60% (19.1 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_D^{20} = -12.5$ (c 0.15 CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ 8.75/8.65 (s, 1H), 7.10 – 7.02 (m, 1H), 6.58 – 6.50 (m, 2H), 3.90 – 3.81 (m, 1H), 3.79 (s, 3H), 3.75 – 3.65 (m, 2H), 3.60 – 3.49 (m, 1H), 2.42 – 2.32 (m, 1H), 2.08 – 2.00 (m, 1H), 1.50/1.45 (s, 9H); ¹³C NMR (151 MHz, CDCl₃) δ 180.5, 160.4, 154.6, 141.3, 125.1, 107.6, 97.6, 80.0, 55.7, 54.1, 53.0, 52.1, 45.6, 36.6, 29.8; HRMS (ESI) calculated for C₁₇H₂₂N₂NaO₄ [M+Na]⁺: 341.1477, found: 341.1481; **Enantiomeric ratio**: 80:20, determined by HPLC (Daicel Chirapak AD, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, T = 30 °C, λ = 254 nm): t_R = 13.42 min (minor), t_R = 21.21 min (major).



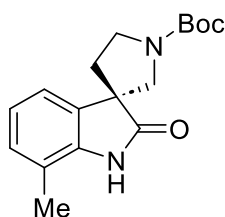
2k

tert-Butyl (R)-6-chloro-2-oxospiro[indoline-3,3'-pyrrolidine]-1'-carboxylate 2k: yield: 60% (19.4 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_D^{20} = -11.0$ (c 0.14 CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ 8.31/8.25 (s, 1H), 7.13 – 6.98 (m, 2H), 6.94 (s, 1H), 3.88 – 3.78 (m, 1H), 3.78 – 3.64 (m, 2H), 3.63 – 3.50 (m, 1H), 2.44 – 2.37 (m, 1H), 2.07 – 2.02 (m, 1H), 1.51/1.46 (s, 9H); ¹³C NMR (151 MHz, CDCl₃) δ 180.3, 154.6, 141.4, 134.3, 131.5, 123.8, 123.0, 110.9, 80.3, 54.4, 53.2, 45.2, 35.6, 28.6; HRMS (ESI) calculated for C₁₆H₁₉ClN₂NaO₃ [M+Na]⁺: 345.0982, found: 345.0980; **Enantiomeric ratio**: 90:10, determined by HPLC (Daicel Chirapak AD, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, T = 30 °C, λ = 254 nm): t_R = 9.97 min (minor), t_R = 21.17 min (major).



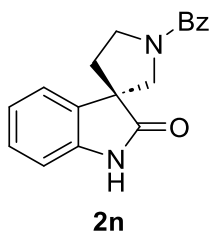
2l

tert-Butyl (R)-6-fluoro-2-oxospiro[indoline-3,3'-pyrrolidine]-1'-carboxylate 2l: yield: 62% (19.0 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_D^{20} = -14.8$ (c 0.19 CH₂Cl₂); **¹H NMR** (600 MHz, CDCl₃) δ 8.44/8.37 (s, 1H), 7.15 – 7.06 (m, 1H), 6.79 – 6.57 (m, 2H), 3.88 – 3.78 (m, 1H), 3.77 – 3.64 (m, 2H), 3.62 – 3.51 (m, 1H), 2.45 – 2.36 (m, 1H), 2.07 – 2.01 (m, 1H), 1.51/1.46 (s, 9H); **¹³C NMR** (151 MHz, CDCl₃) δ 180.7, 163.0 (d, *J* = 245.6 Hz), 154.6, 141.7, 128.3 (d, *J* = 72.7 Hz), 123.9, 109.3 (d, *J* = 22.6 Hz), 98.9 (d, *J* = 27.1 Hz), 80.2, 54.5, 52.2, 45.2, 35.7, 28.6; **¹⁹F NMR** (376 MHz, CDCl₃) δ -111.68 ppm; **HRMS (ESI)** calculated for C₁₆H₁₉FN₂NaO₃ [M+Na]⁺: 329.1277, found: 329.1280; **Enantiomeric ratio:** 89:11, determined by HPLC (Daicel Chirapak AD, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, T = 30 °C, λ = 254 nm): t_R = 8.92 min (minor), t_R = 19.49 min (major)

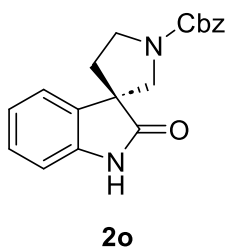


2m

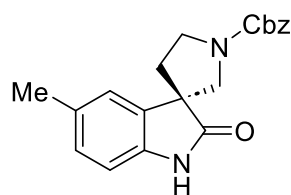
tert-Butyl (R)-7-methyl-2-oxospiro[indoline-3,3'-pyrrolidine]-1'-carboxylate 2m: yield: 60% (18.1 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_D^{20} = -9.3$ (c 0.13 CH₂Cl₂); **¹H NMR** (600 MHz, CDCl₃) δ 8.59/8.53 (s, 1H), 7.10 – 6.94 (m, 3H), 3.87 – 3.75 (m, 2H), 3.73 – 3.66 (m, 1H), 3.64 – 3.53 (m, 1H), 2.45 – 2.38 (m, 1H), 2.29 (s, 3H), 2.13 – 2.05 (m, 1H), 1.51/1.45 (s, 9H); **¹³C NMR** (151 MHz, CDCl₃) δ 180.8, 154.6, 139.0, 132.8, 129.8, 123.1, 120.2, 119.7, 80.0, 54.6, 54.0, 45.6, 36.6, 28.7, 16.6; **HRMS (ESI)** calculated for C₁₇H₂₂N₂NaO₃ [M+Na]⁺: 325.1528, found: 325.1533; **Enantiomeric ratio:** 92:8, determined by HPLC (Daicel Chirapak AD, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, T = 30 °C, λ = 254 nm): t_R = 6.78 min (minor), t_R = 7.27 min (major).



(R)-1'-Benzoylspiro[indoline-3,3'-pyrrolidin]-2-one 2n: yield: 58% (17.0 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_{\text{D}}^{20} = -5.6$ (c 0.06 CH_2Cl_2); $^1\text{H NMR}$ (600 MHz, CDCl_3) δ 9.18 (s, 1H), 7.81 – 7.78 (m, 1H), 7.67 – 7.63 (m, 2H), 7.49 – 7.45 (m, 1H), 7.42 – 7.37 (m, 4H), 7.18 – 7.14 (m, 1H), 3.81 (q, $J = 6.5$ Hz, 2H), 3.45 (t, $J = 6.8$ Hz, 2H), 2.10 (s, 1H), 2.04 (s, 1H); $^{13}\text{C NMR}$ (151 MHz, CDCl_3) δ 180.8, 168.0, 137.7, 134.4, 132.9, 131.7, 130.1, 128.7, 127.9, 127.0, 121.6, 112.7, 46.2, 41.6, 29.8, 24.0; **HRMS (ESI)** calculated for $\text{C}_{18}\text{H}_{16}\text{N}_2\text{NaO}_2$ $[\text{M}+\text{Na}]^+$: 315.1109, found: 315.1112; **Enantiomeric ratio**: 86:14, determined by HPLC (Daicel Chirapak OD, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, $T = 30$ °C, $\lambda = 254$ nm): $t_{\text{R}} = 23.62$ min (minor), $t_{\text{R}} = 28.59$ min (major).

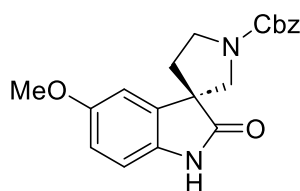


Benzyl (R)-2-oxospiro[indoline-3,3'-pyrrolidine]-1'-carboxylate 2o: yield: 63% (20.3 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_{\text{D}}^{20} = -11.6$ (c 0.17 CH_2Cl_2); $^1\text{H NMR}$ (600 MHz, CDCl_3) δ 8.98/8.94 (s, 1H), 7.44 – 7.40 (m, 1H), 7.40 – 7.37 (m, 1H), 7.35 – 7.27 (m, 3H), 7.25 – 7.21 (m, 1H), 7.17 – 7.12 (m, 1H), 7.06 – 7.00 (m, 1H), 6.96 – 6.92 (m, 1H), 5.21 – 5.15 (m, 2H), 3.95 – 3.89 (m, 1H), 3.85 – 3.79 (m, 2H), 3.71 – 3.63 (m, 1H), 2.47 – 2.41 (m, 1H), 2.15 – 2.10 (m, 1H); $^{13}\text{C NMR}$ (151 MHz, CDCl_3) δ 180.3, 178.2, 154.9, 140.3, 136.8, 132.7, 128.6, 128.1, 127.9, 123.1, 122.8, 110.3, 67.2, 60.6, 54.1, 53.4, 45.9, 36.4, 29.7; **HRMS (ESI)** calculated for $\text{C}_{19}\text{H}_{18}\text{N}_2\text{NaO}_3$ $[\text{M}+\text{Na}]^+$: 345.1215, found: 345.1218; **Enantiomeric ratio**: 90:10, determined by HPLC (Daicel Chirapak OD, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, $T = 30$ °C, $\lambda = 254$ nm): $t_{\text{R}} = 15.42$ min (minor), $t_{\text{R}} = 21.02$ min (major).



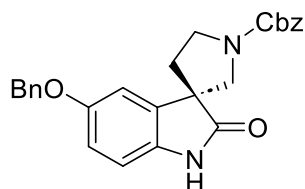
2p

Benzyl (*R*)-5-methyl-2-oxospiro[indoline-3,3'-pyrrolidine]-1'-carboxylate 2p: yield: 52% (17.5 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_D^{20} = -15.9$ (c 0.28 CH₂Cl₂); $^1\text{H NMR}$ (600 MHz, CDCl₃) δ 8.88/8.83 (s, 1H), 7.50 – 7.37 (m, 2H), 7.37 – 7.25 (m, 3H), 7.03 (d, $J = 7.9$ Hz, 1H), 6.99 – 6.92 (m, 1H), 6.85 – 6.79 (m, 1H), 5.26 – 5.16 (m, 2H), 3.97 – 3.90 (m, 1H), 3.86 – 3.78 (m, 2H), 3.71 – 3.62 (m, 1H), 2.45 – 2.40 (m, 1H), 2.30 (s, 3H), 2.12 – 2.07 (m, 1H); $^{13}\text{C NMR}$ (151 MHz, CDCl₃) δ 180.1, 178.1, 154.9, 137.7, 136.7, 132.9, 132.6, 128.9, 128.6, 128.1, 127.9, 123.6, 110.0, 67.2, 54.5, 53.5, 45.9, 36.4, 29.7, 21.2; **HRMS (ESI)** calculated for C₂₀H₂₀N₂NaO₃ [M+Na]⁺: 359.1372, found: 359.1375; **Enantiomeric ratio:** 90:10, determined by HPLC (Daicel Chirapak OD, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, T = 30 °C, $\lambda = 254$ nm): $t_R = 13.44$ min (minor), $t_R = 18.55$ min (major).



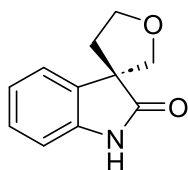
2q

Benzyl (*R*)-5-methoxy-2-oxospiro[indoline-3,3'-pyrrolidine]-1'-carboxylate 2q: yield: 68% (24.0 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_D^{20} = -32.4$ (c 0.24 CH₂Cl₂); $^1\text{H NMR}$ (600 MHz, CDCl₃) δ 8.45/8.42 (s, 1H), 7.48 – 7.18 (m, 5H), 6.87 – 6.80 (m, 1H), 6.79 – 6.69 (m, 2H), 5.23 – 5.11 (m, 2H), 3.91 (m, 1H), 3.87 – 3.77 (m, 2H), 3.75 (s, 3H), 3.70 – 3.62 (m, 1H), 2.47 – 2.39 (m, 1H), 2.13 – 2.06 (m, 1H); $^{13}\text{C NMR}$ (151 MHz, CDCl₃) δ 179.8, 156.4, 154.9, 136.9, 136.7, 134.0, 133.4, 128.7, 128.1, 127.9, 112.8, 110.6, 110.2, 67.2, 55.9, 54.2, 53.6, 45.4, 36.5, 29.8; **HRMS (ESI)** calculated for C₂₀H₂₀N₂NaO₄ [M+Na]⁺: 375.1321, found: 375.1324; **Enantiomeric ratio:** 89.5:10.5, determined by HPLC (Daicel Chirapak OD, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, T = 30 °C, $\lambda = 254$ nm): $t_R = 18.67$ min (minor), $t_R = 27.89$ min (major).



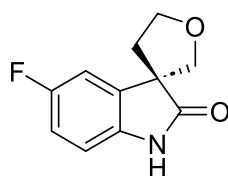
2r

Benzyl (R)-5-(benzyloxy)-2-oxospiro[indoline-3,3'-pyrrolidine]-1'-carboxylate 2r: yield: 62% (26.6 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_D^{20} = -17.6$ (c 0.14 CH₂Cl₂); **¹H NMR** (600 MHz, CDCl₃) δ 8.63/8.58 (s, 1H), 7.38 – 7.30 (m, 5H), 7.29 (s, 1H), 7.28 – 7.27 (m, 1H), 7.26 – 7.17 (m, 3H), 6.80 – 6.69 (m, 3H), 5.14 – 5.06 (m, 2H), 4.92 (s, 2H), 3.90 – 3.77 (m, 1H), 3.77 – 3.65 (m, 2H), 3.63 – 3.54 (m, 1H), 2.40 – 2.31 (m, 1H), 2.05 – 1.95 (m, 1H); **¹³C NMR** (151 MHz, CDCl₃) δ 179.8, 177.7, 155.5, 154.8, 136.9, 134.1, 133.9, 133.6, 128.8, 128.6, 128.2, 127.9, 127.7, 114.1, 111.4, 110.5, 71.0, 67.2, 54.5, 45.8, 35.6, 29.7; **HRMS (ESI)** calculated for C₂₆H₂₄N₂NaO₄ [M+Na]⁺: 451.1634, found: 451.1634; **Enantiomeric ratio:** 83.5:16.5, determined by HPLC (Daicel Chirapak OD, isopropanol / hexane = 20/80, flow rate = 1.0 mL/min, T = 30 °C, λ = 254 nm): t_R = 12.93 min (minor), t_R = 17.19 min (major).



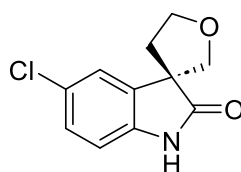
2s

(R)-4,5-Dihydro-2H-spiro[furan-3,3'-indolin]-2'-one 2s: yield: 65% (12.3 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_D^{20} = +14.0$ (c 0.10 CH₂Cl₂); **¹H NMR** (600 MHz, CDCl₃) δ 8.83 (s, 1H), 7.29 (d, *J* = 7.4 Hz, 1H), 7.23 (t, *J* = 7.7 Hz, 1H), 7.06 (t, *J* = 7.5 Hz, 1H), 6.94 (d, *J* = 7.7 Hz, 1H), 4.25 – 4.19 (m, 2H), 4.08 (d, *J* = 8.6 Hz, 1H), 3.96 (d, *J* = 8.5 Hz, 1H), 2.58 – 2.52 (m, 1H), 2.22 – 2.16 (m, 1H); **¹³C NMR** (151 MHz, CDCl₃) δ 181.5, 140.4, 134.2, 128.3, 123.1, 123.0, 110.1, 69.2, 54.9, 38.8; **HRMS (ESI)** calculated for C₁₁H₁₂NO₂ [M+H]⁺: 190.0868, found: 190.0872; **Enantiomeric ratio:** 92.5:7.5, determined by HPLC (Daicel Chirapak AS, isopropanol / hexane = 40/60, flow rate = 1.0 mL/min, T = 30 °C, λ = 254 nm): t_R = 38.85 min (minor), t_R = 52.37 min (major).



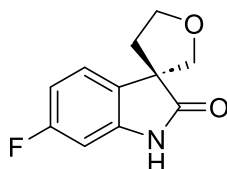
2t

(R)-5'-Fluoro-4,5-dihydro-2H-spiro[furan-3,3'-indolin]-2'-one 2t: yield: 62% (12.8 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_D^{20} = +16.6$ (c 0.18 CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ 9.38 (s, 1H), 7.04 – 7.00 (m, 1H), 6.95 – 6.90 (m, 1H), 6.90 – 6.86 (m, 1H), 4.24 – 4.18 (m, 2H), 4.05 (d, *J* = 8.6 Hz, 1H), 3.95 (d, *J* = 8.6 Hz, 1H), 2.59 – 2.52 (m, 1H), 2.21 – 2.15 (m, 1H); ¹³C NMR (151 MHz, CDCl₃) δ 181.3, 159.6 (d, *J* = 241.1 Hz), 136.2, 135.9 (d, *J* = 8.0 Hz), 114.6 (d, *J* = 23.7 Hz), 111.1 (d, *J* = 25.0 Hz), 110.7 (d, *J* = 8.1 Hz), 69.08, 55.50, 38.65; ¹⁹F NMR (565 MHz, CDCl₃) δ -119.66 ppm; **HRMS (ESI)** calculated for C₁₁H₁₁FNO₂ [M+H]⁺: 208.0774, found: 208.0777; **Enantiomeric ratio:** 90:10, determined by HPLC (Daicel Chirapak OJ, isopropanol / hexane = 20/80, flow rate = 1.0 mL/min, T = 30 °C, λ = 254 nm): t_R = 6.52 min (major), t_R = 7.78 min (minor).



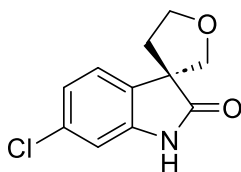
2u

(R)-5'-Chloro-4,5-dihydro-2H-spiro[furan-3,3'-indolin]-2'-one 2u: yield: 60% (13.4 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_D^{20} = +26.7$ (c 0.08 CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ 9.17 (s, 1H), 7.28 – 7.26 (m, 1H), 7.23 – 7.18 (m, 1H), 6.88 (d, *J* = 8.3 Hz, 1H), 4.25 – 4.16 (m, 2H), 4.04 (d, *J* = 8.6 Hz, 1H), 3.95 (d, *J* = 8.6 Hz, 1H), 2.59 – 2.53 (m, 1H), 2.20 – 2.15 (m, 1H); ¹³C NMR (151 MHz, CDCl₃) δ 180.9, 138.8, 136.0, 132.2, 128.2, 123.6, 111.1, 69.1, 55.2, 38.7; **HRMS (ESI)** calculated for C₁₁H₁₁ClNO₂ [M+H]⁺: 224.0478, found: 224.0482; **Enantiomeric ratio:** 87:13, determined by HPLC (Daicel Chirapak OJ, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, T = 30 °C, λ = 254 nm): t_R = 9.77 min (major), t_R = 12.77 min (minor).



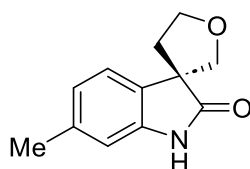
2v

(R)-6'-Fluoro-4,5-dihydro-2H-spiro[furan-3,3'-indolin]-2'-one 2v: yield: 60% (12.6 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_D^{20} = +12.6$ (c 0.09 CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ 9.38 (s, 1H), 7.23 – 7.18 (m, 1H), 6.76 – 6.69 (m, 2H), 4.23 – 4.18 (m, 2H), 4.05 (d, *J* = 8.6 Hz, 1H), 3.93 (d, *J* = 8.6 Hz, 1H), 2.57 – 2.51 (m, 1H), 2.19 – 2.13 (m, 1H); ¹³C NMR (151 MHz, CDCl₃) δ 181.9, 162.9 (d, *J* = 245.2 Hz), 141.7 (d, *J* = 11.8 Hz), 129.5 (d, *J* = 3.1 Hz), 124.0 (d, *J* = 9.6 Hz), 109.4 (d, *J* = 22.5 Hz), 98.9 (d, *J* = 27.4 Hz), 69.1, 54.5, 38.7; ¹⁹F NMR (565 MHz, CDCl₃) δ -112.18 ppm; **HRMS (ESI)** calculated for C₁₁H₁₁FNO₂ [M+H]⁺: 208.0774, found: 208.0773; **Enantiomeric ratio:** 82:18, determined by HPLC (Daicel Chirapak OJ, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, T = 30 °C, λ = 254 nm): t_R = 10.72 min (major), t_R = 12.23 min (minor).



2w

(R)-6'-Chloro-4,5-dihydro-2H-spiro[furan-3,3'-indolin]-2'-one 2w: yield: 65% (14.5 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_D^{20} = +11.8$ (c 0.10 CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ 9.48 (s, 1H), 7.19 (d, *J* = 8.0 Hz, 1H), 7.05 – 7.01 (m, 1H), 6.99 – 6.95 (m, 1H), 4.23 – 4.19 (m, 2H), 4.04 (d, *J* = 8.6 Hz, 1H), 3.93 (d, *J* = 8.6 Hz, 1H), 2.57 – 2.51 (m, 1H), 2.19 – 2.13 (m, 1H); ¹³C NMR (151 MHz, CDCl₃) δ 181.5, 141.5, 134.0, 132.6, 123.9, 123.1, 110.8, 77.0, 69.1, 54.7, 38.7; **HRMS (ESI)** calculated for C₁₁H₁₁ClNO₂ [M+H]⁺: 224.0478, found: 224.0482; **Enantiomeric ratio:** 85:15, determined by HPLC (Daicel Chirapak OJ, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, T = 30 °C, λ = 254 nm): t_R = 12.24 min (major), t_R = 14.97 min (minor).

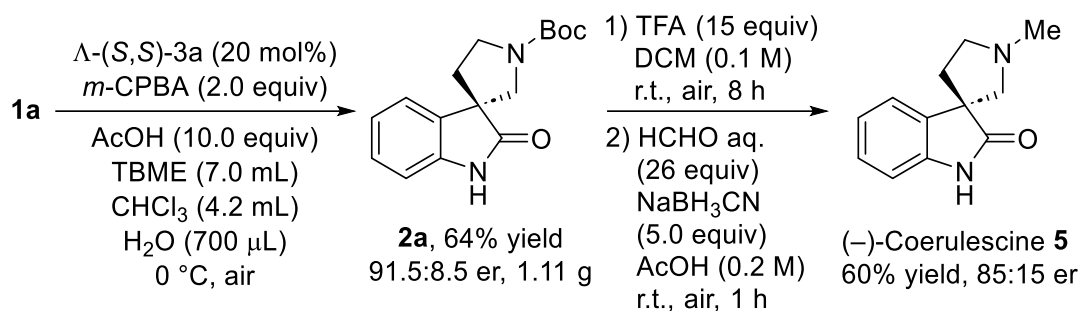


2x

(R)-6'-Methyl-4,5-dihydro-2H-spiro[furan-3,3'-indolin]-2'-one 2x: yield: 60% (12.2 mg); flash column chromatography eluent, petroleum ether/ethyl acetate = 2/1; colorless oil; $[\alpha]_D^{20} = +3.2$ (c 0.18 CH₂Cl₂); **¹H NMR** (600 MHz, CDCl₃) δ 7.92 (s, 1H), 7.16 (d, $J = 7.6$ Hz, 1H), 6.87 (d, $J = 7.6$ Hz, 1H), 6.74 (s, 1H), 4.22 – 4.18 (m, 2H), 4.05 (d, $J = 8.5$ Hz, 1H), 3.93 (d, $J = 8.5$ Hz, 1H), 2.55 – 2.50 (m, 1H), 2.35 (s, 3H), 2.18 – 2.13 (m, 1H); **¹³C NMR** (151 MHz, CDCl₃) δ 181.9, 140.5, 138.5, 131.2, 123.6, 122.7, 111.0, 69.2, 54.7, 38.8, 21.7; **HRMS (ESI)** calculated for C₁₂H₁₄NO₂ [M+H]⁺: 204.1025, found: 204.1027; **Enantiomeric ratio:** 86.5:13.5, determined by HPLC (Daicel Chirapak AD, isopropanol / hexane = 10/90, flow rate = 1.0 mL/min, T = 30 °C, $\lambda = 254$ nm): $t_R = 9.69$ min (minor), $t_R = 10.32$ min (major).

5. Synthetic Applications and Mechanistic Experiments

5.1 Gram-scale reaction and the synthesis of (-)-Coerulescine

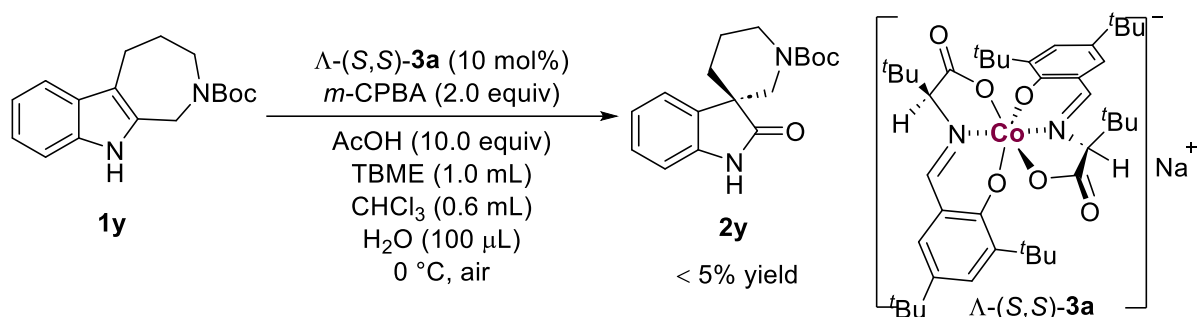


In a 50 mL tube equipped with a magnetic stirring bar, indole **1a** (6.0 mmol, 1.0 equiv), Δ -(*S,S*)-**3a** (960 mg, 20 mol%), H₂O (700 μ L), AcOH (60 mmol, 10.0 equiv), and a mixed solvent of TBME/CHCl₃ (11.2 mL, v/v = 5:3) were combined. The resulting mixture was cooled to 0 °C and stirred for 20 min, followed by the addition of *m*-CPBA (2.42 g, 12 mmol, 2.0 equiv). The reaction mixture was stirred at the same temperature, and the reaction progress was monitored by TLC. Upon completion, the reaction was quenched by the addition of saturated aqueous Na₂S₂O₃ solution. The mixture was extracted with ethyl acetate (3 \times 15 mL). The combined organic layers were washed with saturated aqueous NaHCO₃ solution and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. Purification of the residue by silica gel column chromatography afforded the pure product **2a** in 64% yield (1.11 g, 91.5:8.5 er).

To a solution of **2a** (57.6 mg, 0.20 mmol, 1.0 equiv) in anhydrous DCM (0.1 M) was added TFA (0.2 mL, 15 equiv). The reaction mixture was stirred at room temperature for 8 h, then diluted with DCM (5 mL) and H₂O (1 mL). The mixture was basified to pH 9–10 with 2 N aqueous NaOH and extracted with DCM. The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, and concentrated to afford the crude amine. The crude amine was dissolved in AcOH (0.2 M), and aqueous formaldehyde solution (37 wt%, 0.40 mL) and NaBH₃CN (63 mg, 5.0 equiv, 1.0 mmol) were added sequentially. The resulting mixture was stirred at room temperature for 1 h, then diluted with ethyl acetate (2 mL) and basified to pH 9–10 with 2 N aqueous NaOH. The mixture was extracted with ethyl acetate, and the combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, and concentrated. Purification of the residue by preparative TLC (eluent: DCM/MeOH = 9:1) afforded (-)-Coerulescine **5** as a colorless wax in 60% overall yield (24.2 mg, 85:15 er).

(-)-**Coerulescine 5**: yield: 60% (24.2 mg); flash column chromatography eluent, DCM/MeOH = 9/1; colorless oil; $[\alpha]_D^{20} = +12.6$ (c 0.1 CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃) δ 8.29 (s, 1H), 7.40 (d, *J* = 7.4 Hz, 1H), 7.23 – 7.14 (m, 1H), 7.08 – 7.01 (m, 1H), 6.88 (d, *J* = 7.7 Hz, 1H), 3.05 – 2.99 (m, 1H), 2.91 – 2.84 (m, 2H), 2.83 – 2.76 (m, 1H), 2.46 (s, 3H), 2.44 – 2.36 (m, 1H), 2.14 – 2.08 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 182.8, 140.2, 136.2, 127.9, 123.5, 123.0, 109.6, 66.5, 56.9, 53.8, 42.0, 38.1; HRMS (ESI) calculated for C₁₂H₁₅N₂O [M+H]⁺: 203.1184, found: 203.1189; **Enantiomeric ratio**: 85:15, determined by HPLC (Daicel Chirapak AS, isopropanol / hexane = 30/70, flow rate = 1.0 mL/min, T = 30 °C, λ = 254 nm): t_R = 8.96 min (major), t_R = 14.01 min (minor).

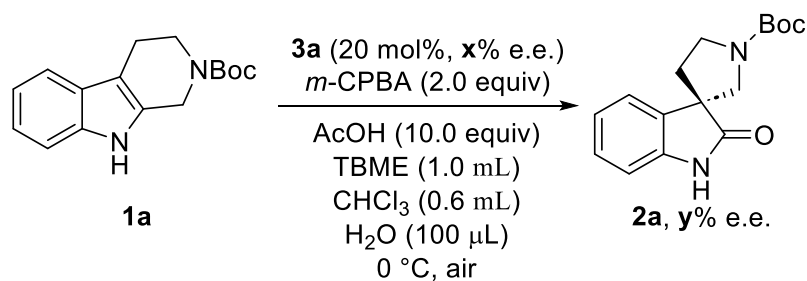
5.2 Synthesis of seven-membered ring substrates **2y**



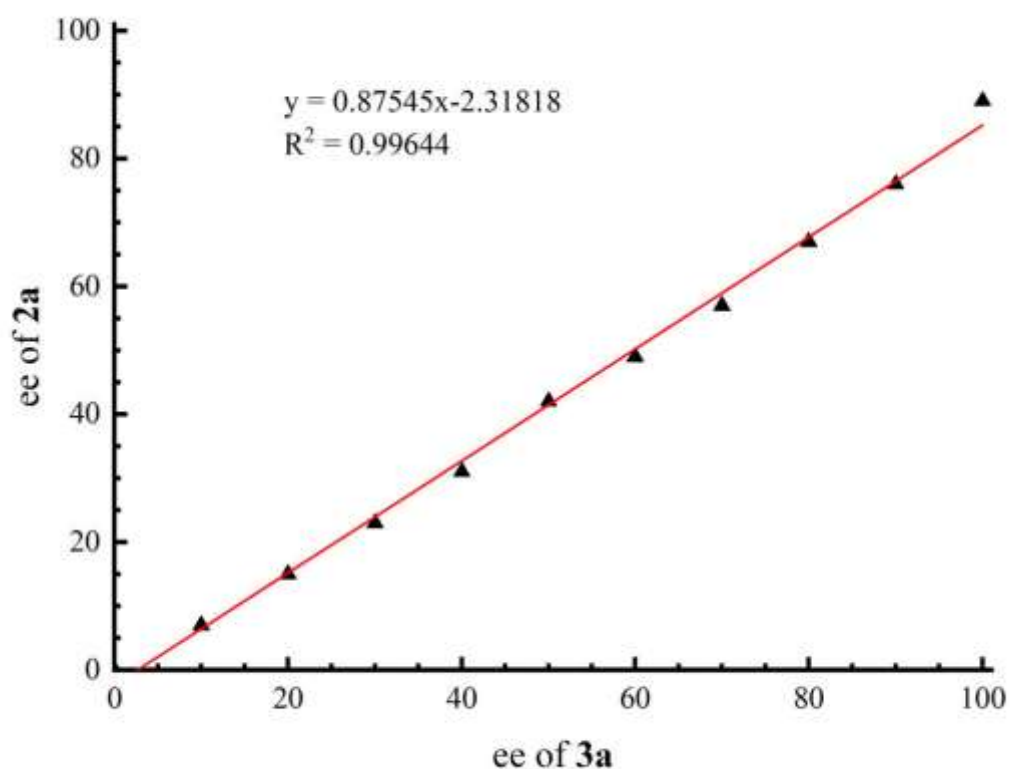
At room temperature, a 10 mL vial was charged with Δ -(*S,S*)-**3a** (16 mg, 20 mol%), H₂O (100 μ L), AcOH (60 μ L), and indole **1y** (0.10 mmol), after which a mixed solvent of TBME/CHCl₃ (1.6 mL, v/v = 5:3) was added. The resulting mixture was cooled to 0 °C and stirred for 20 min, followed by the addition of *m*-CPBA (34.5 mg, 2.0 equiv, 0.20 mmol). The reaction mixture was stirred at 0 °C, and the progress of the reaction was monitored by TLC. Unfortunately, according to TLC monitoring, only a trace amount of the desired product was observed, corresponding to a yield of less than 5%.

5.3 Non-Linear Effects

The standard reaction of **1a** showed a linear correlation between the enantiomeric excess (ee) of the product and that of the catalyst, which is consistent with the proposed enantiodetermining transition state involving a single catalyst molecule acting in a bifunctional manner.

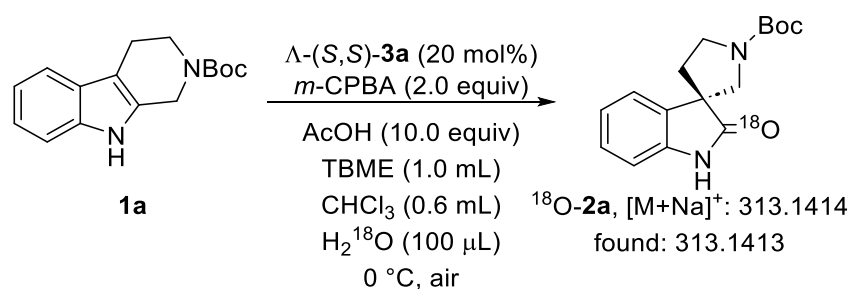


entry	Δ - 3a (mg)	Λ - 3a (mg)	ee of 3a (%)	yield of 2a (%)	ee of 2a (%)
1	2.5	47.5	90	62	76
2	5.0	45.0	80	60	67
3	7.5	42.5	70	60	57
4	10.0	40.0	60	62	49
5	12.5	37.5	50	64	42
6	15.0	35.0	40	60	31
7	17.5	32.5	30	58	23
8	20.0	30.0	20	60	15
9	22.5	27.5	10	60	7
10	25	25	0	62	0
11	--	50	100	62	89
12	50	--	--	60	89



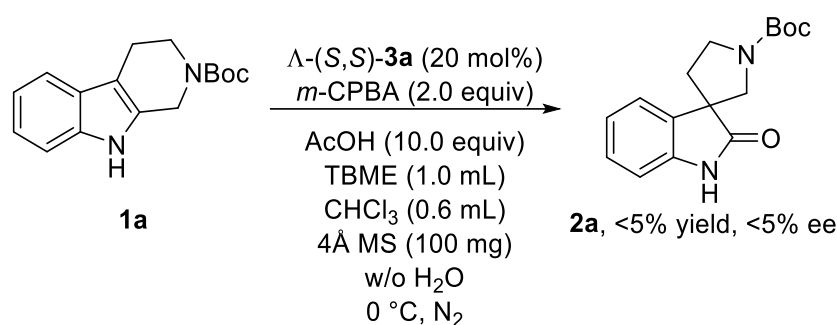
At room temperature, a 10 mL vial was charged with scalemic catalyst **3a** (16 mg, 20 mol%), H₂O (100 μ L), AcOH (60 μ L), and indole **1** (0.10 mmol), after which a mixed solvent of TBME/CHCl₃ (1.6 mL, v/v = 5:3) was added. The resulting mixture was cooled to 0 °C and stirred for 20 min, followed by the addition of *m*-CPBA (34.5 mg, 2.0 equiv, 0.20 mmol). The reaction mixture was stirred at 0 °C, and the reaction progress was monitored by TLC. Upon completion, the reaction was quenched by the addition of saturated aqueous Na₂S₂O₃ solution. The mixture was extracted with ethyl acetate. The combined organic layers were washed with saturated aqueous NaHCO₃ solution and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. Purification of the residue by silica gel column chromatography afforded the pure product.

5.4 Isotopic labeling experiment

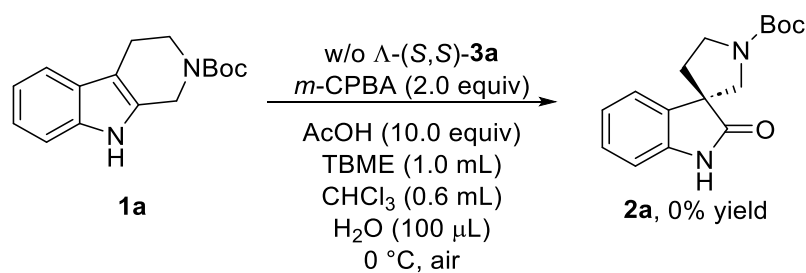


In a 10 mL tube equipped with a magnetic stirring bar, indole **1a** (0.10 mmol, 1.0 equiv), Λ -(*S,S*)-**3a** (16 mg, 20 mol%), H₂¹⁸O (100 μ L), AcOH (60 μ L, 10.0 equiv), and dry TBME (1.0 mL) were combined under a nitrogen atmosphere. The resulting mixture was cooled to 0 °C and stirred for 20 min, followed by the addition of *m*-CPBA (34.5 mg, 2.0 equiv, 0.20 mmol). The reaction mixture was stirred at the same temperature, and the reaction progress was monitored by TLC. Upon completion, the reaction was quenched by the addition of saturated aqueous Na₂S₂O₃ solution. The mixture was extracted with ethyl acetate. The combined organic layers were washed with saturated aqueous NaHCO₃ solution and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. Purification of the residue by silica gel column chromatography afforded the pure product ¹⁸O-**2a**.

5.5 Control Experiment



In a 10-mL tube equipped with a magnetic stirring bar was charged with indole **1a** (0.1 mmol, 1.0 equiv), Λ -(*S,S*)-**3a** (16 mg, 20 mol%), AcOH (60 μ L, 10.0 equiv), 4 Å MS (100 mg) and TBME/CHCl₃ (1.6 mL, v/v = 5:3) under the nitrogen atmosphere. The mixture was cooled to 0 °C and stirred for 20 min followed by addition of *m*-CPBA (34.5 mg, 2.0 equiv, 0.2 mmol). The mixture was stirred at the same temperature, and the reaction progress was monitored by TLC. The reaction was quenched by addition of a saturated aqueous Na₂S₂O₃ solution. Next, the mixture was extracted with ethyl acetate. The combined organic layers were washed by saturated aqueous NaHCO₃ solution and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated. The residue was subjected to silica gel column chromatography to afford the pure product.

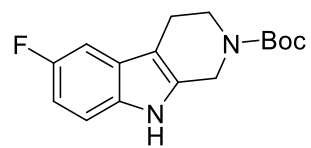


In a 10-mL tube equipped with a magnetic stirring bar was charged with indole **1a** (0.1 mmol, 1.0 equiv), AcOH (60 μ L, 10.0 equiv), and TBME/CHCl₃ (1.6 mL, v/v = 5:3) under the nitrogen atmosphere. The mixture was cooled to 0 °C and stirred for 20 min followed by addition of *m*-CPBA (34.5 mg, 2.0 equiv, 0.2 mmol). The mixture was stirred at the same temperature, and the reaction progress was monitored by TLC and no target product was detected.

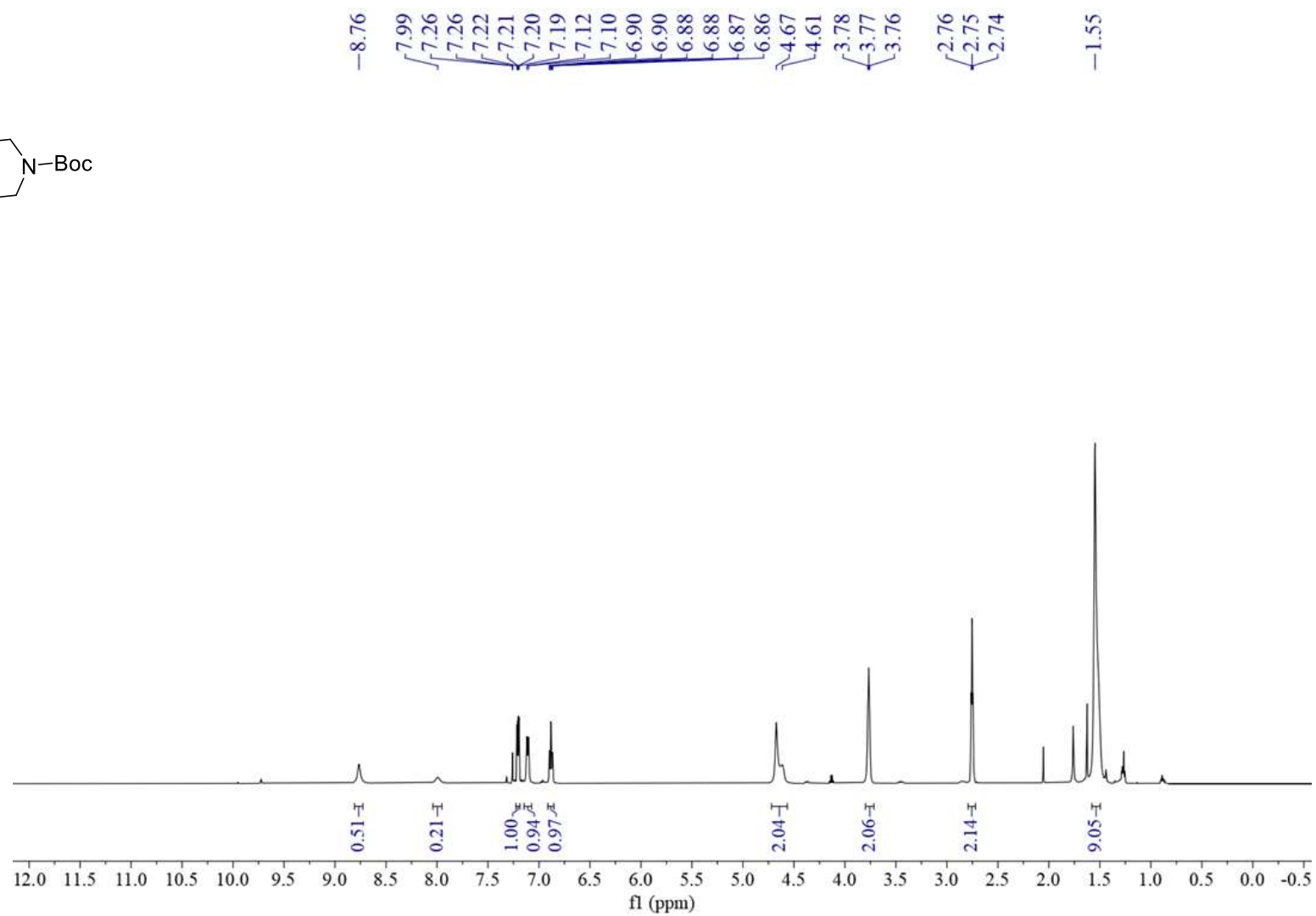
6. References

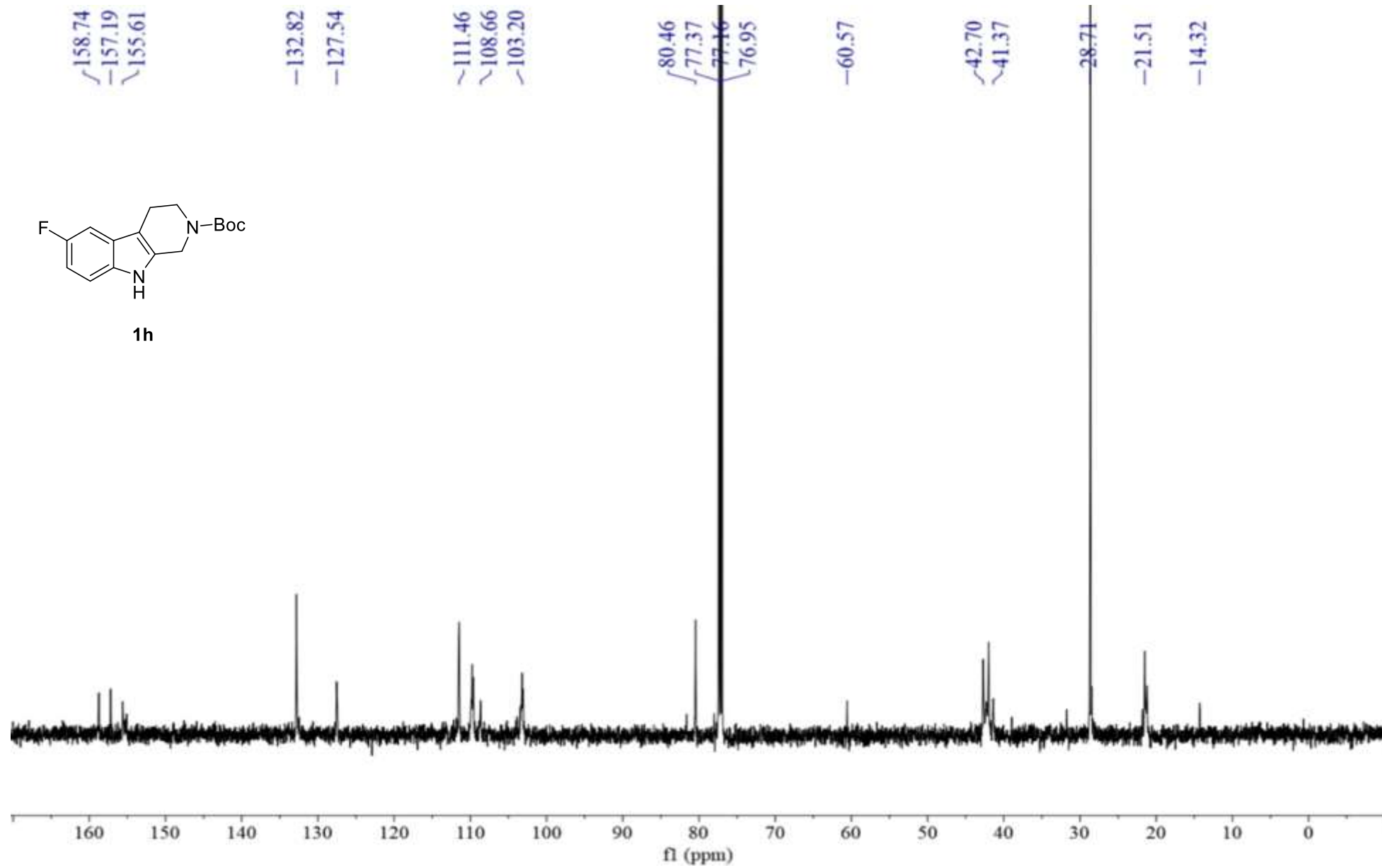
- [1] (a) J. Yu, H.-J. Jiang, Y. Zhou, S.-W. Luo and L.-Z. Gong, *Angew. Chem. Int. Ed.*, 2015, **54**, 11209–11213; (b) H.-J. Jiang, K. Liu, J. Yu, L. Zhang and L.-Z. Gong, *Angew. Chem. Int. Ed.*, 2017, **56**, 11931–11935; (c) H.-J. Jiang, X.-M. Zhong, J. Yu, Y. Zhang, X. Zhang, Y.-D. Wu and L.-Z. Gong, *Angew. Chem. Int. Ed.*, 2019, **58**, 1803–1807; (d) T.-T. Sun, K. Liu, S.-X. Zhang, C.-R. Wang, C.-Z. Yao and J. Yu, *Synlett*, 2021, **32**, 701–707.
- [2] C. Qian, P. Li and J. Sun, *Angew. Chem. Int. Ed.*, 2021, **60**, 5871–5875.
- [3] H. Gao, X. Yang and L. Shi, *Org. Chem. Front.*, 2025, **12**, 3848–3855.
- [4] X. Tan, Z. Zhou, M. Shao and J. Sun, *Angew. Chem. Int. Ed.*, 2025, e202510078.
- [5] B. Jackson and J. Hester, *J. Org. Chem.*, 1970, **35**, 875–883.

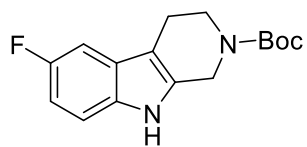
7. NMR Spectra and HPLC Chromatograms



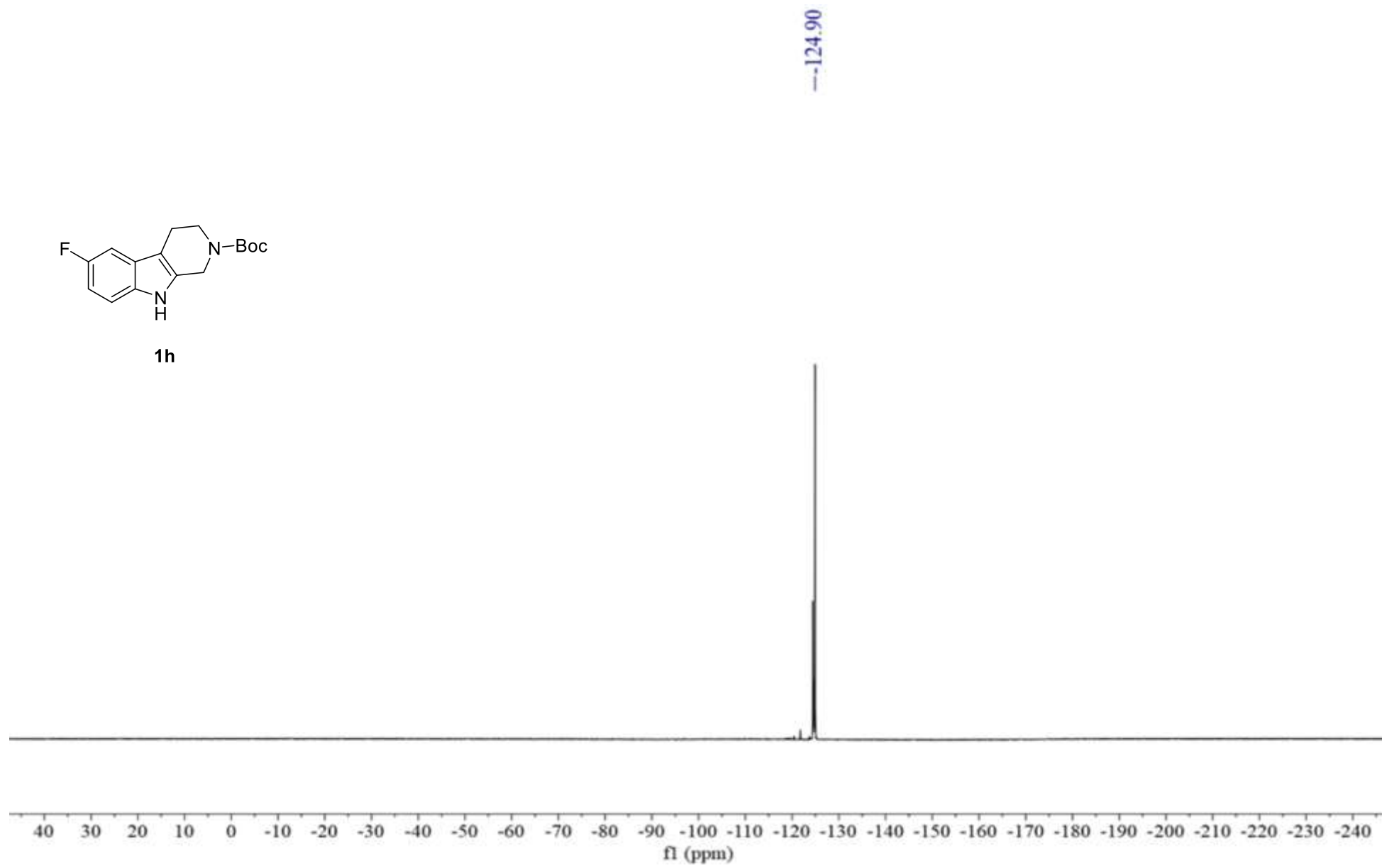
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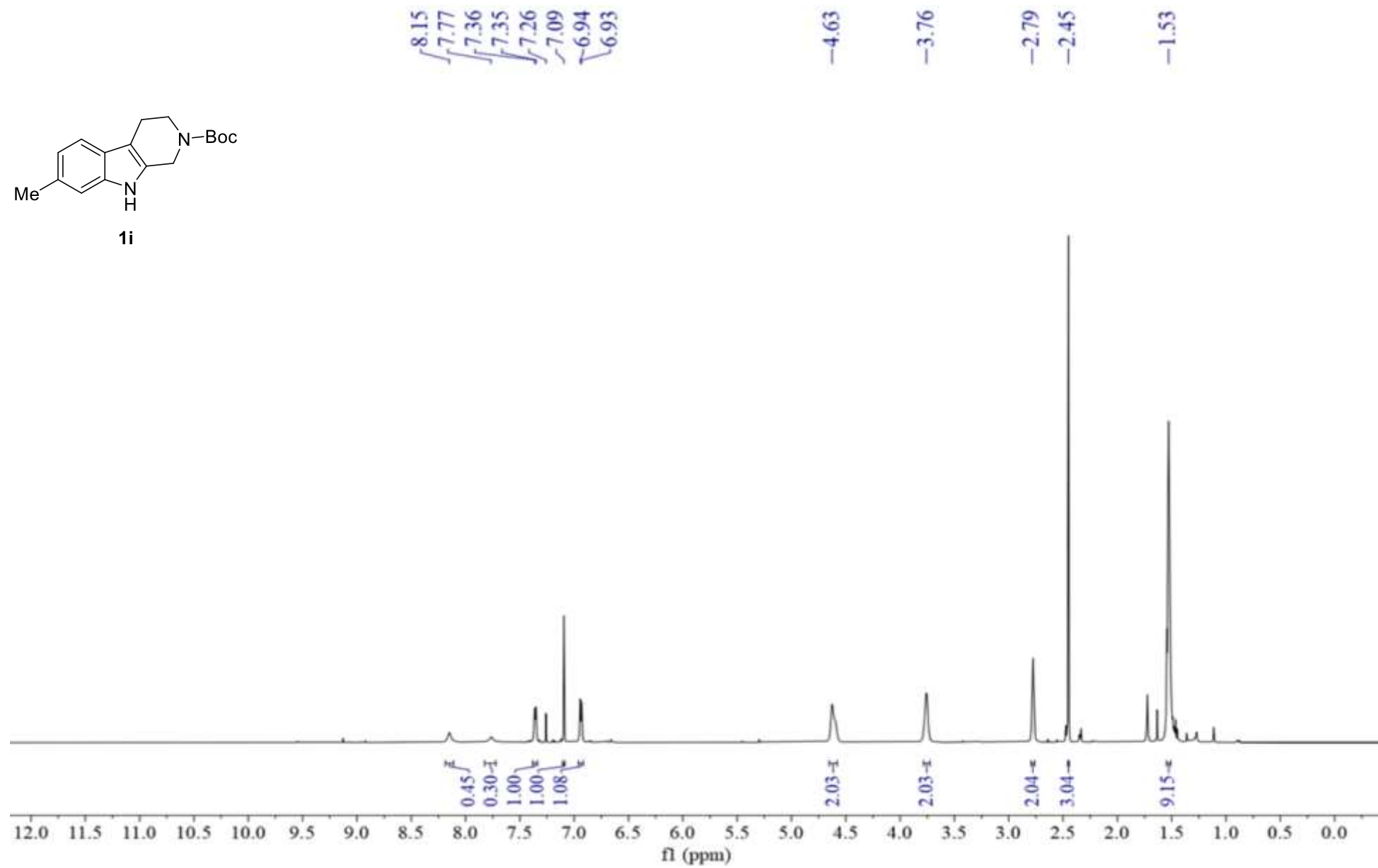
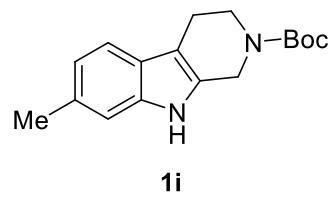


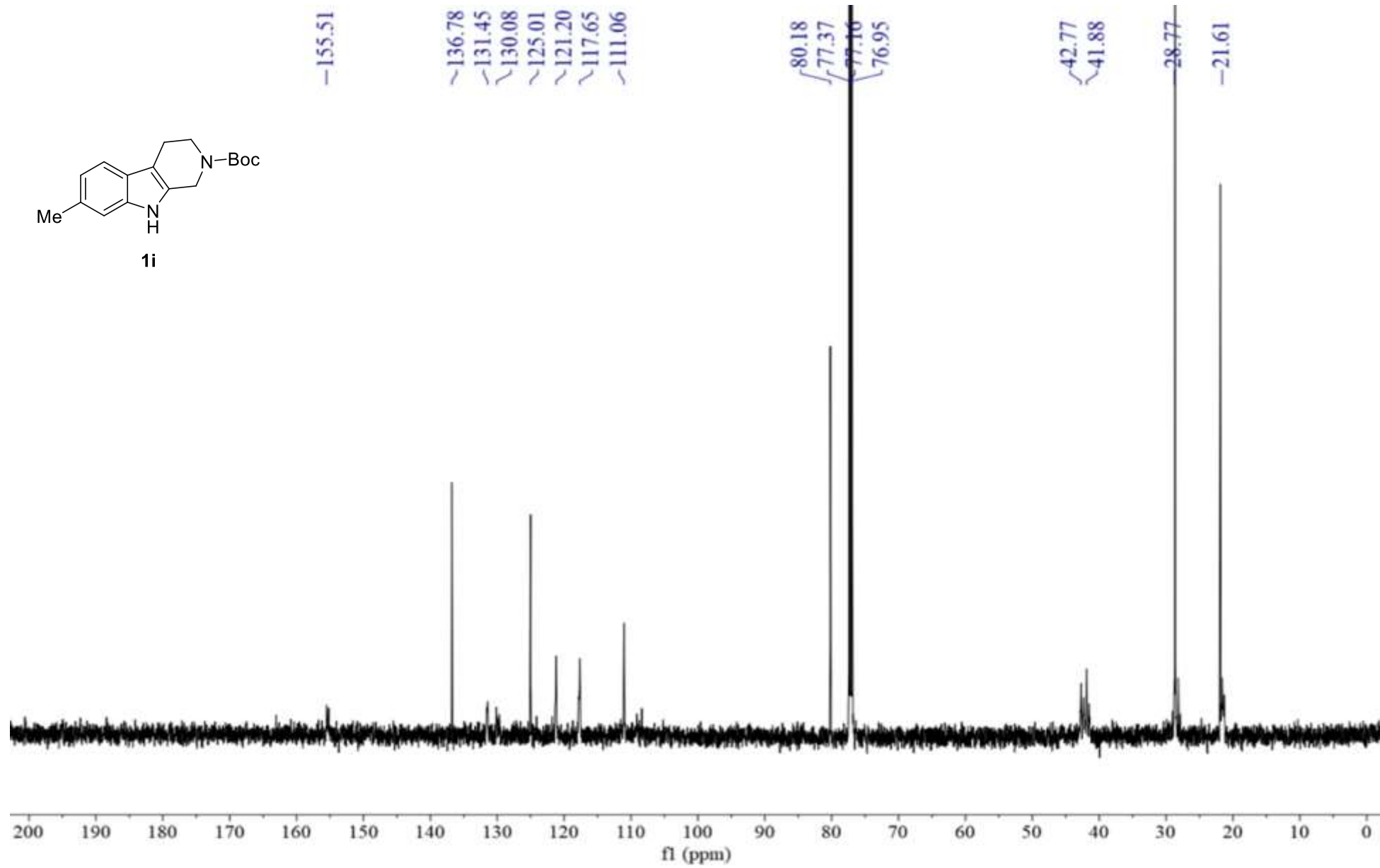


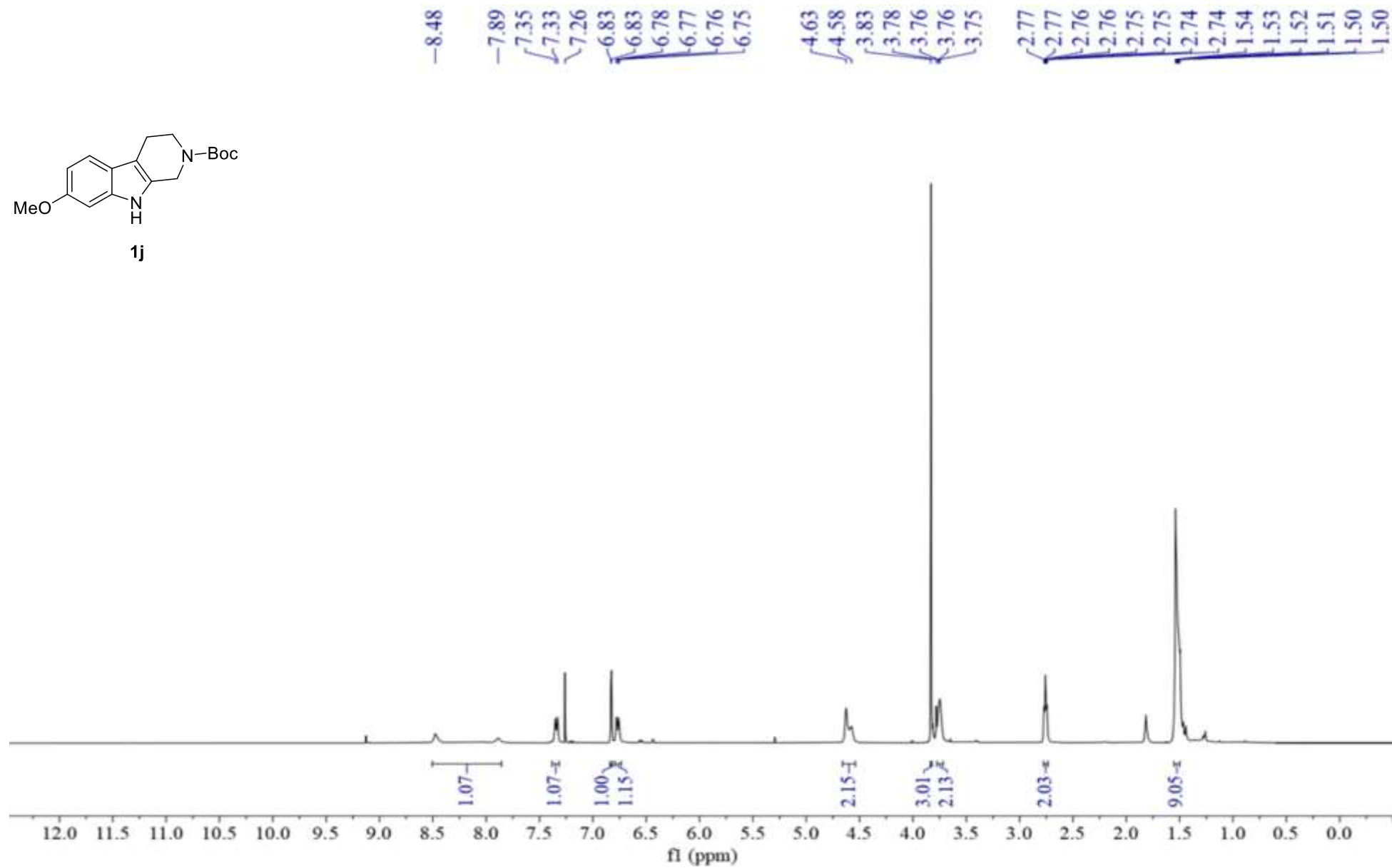
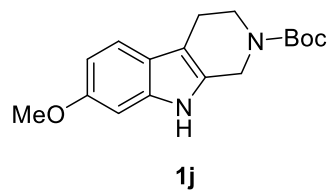


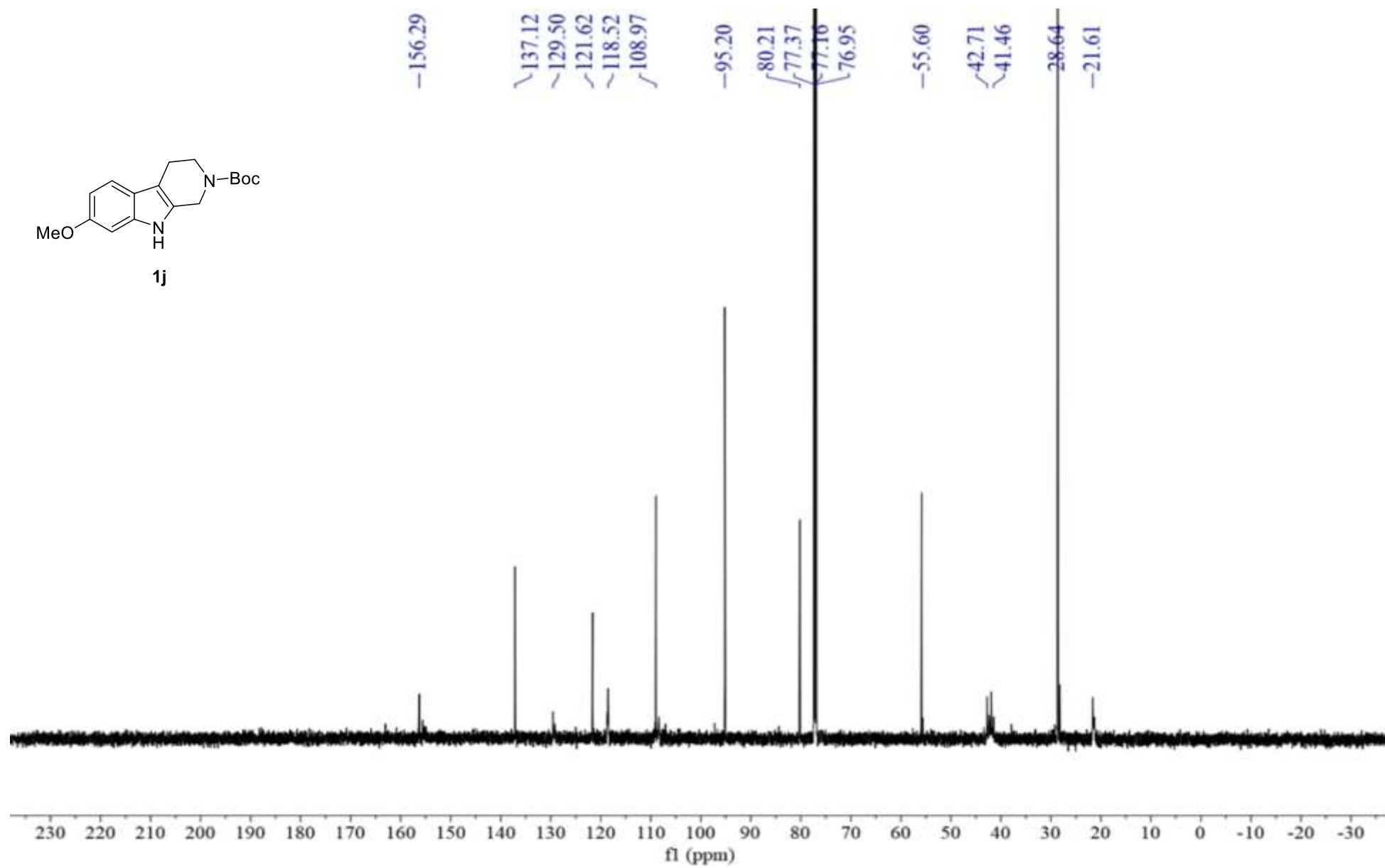
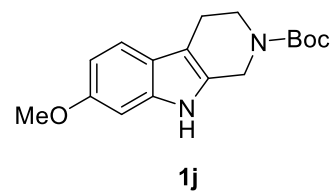
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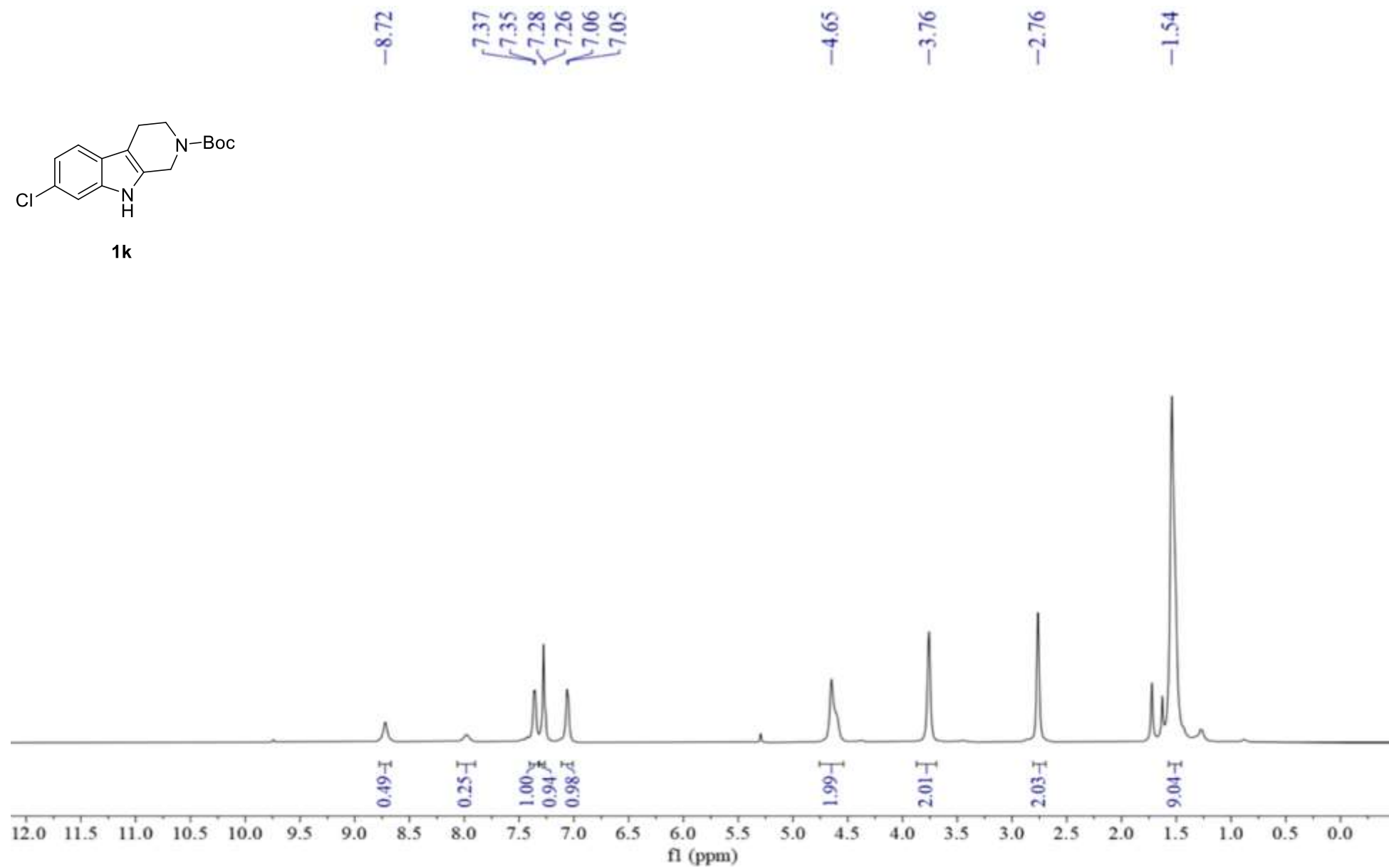
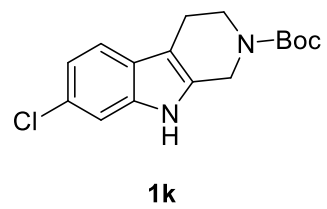


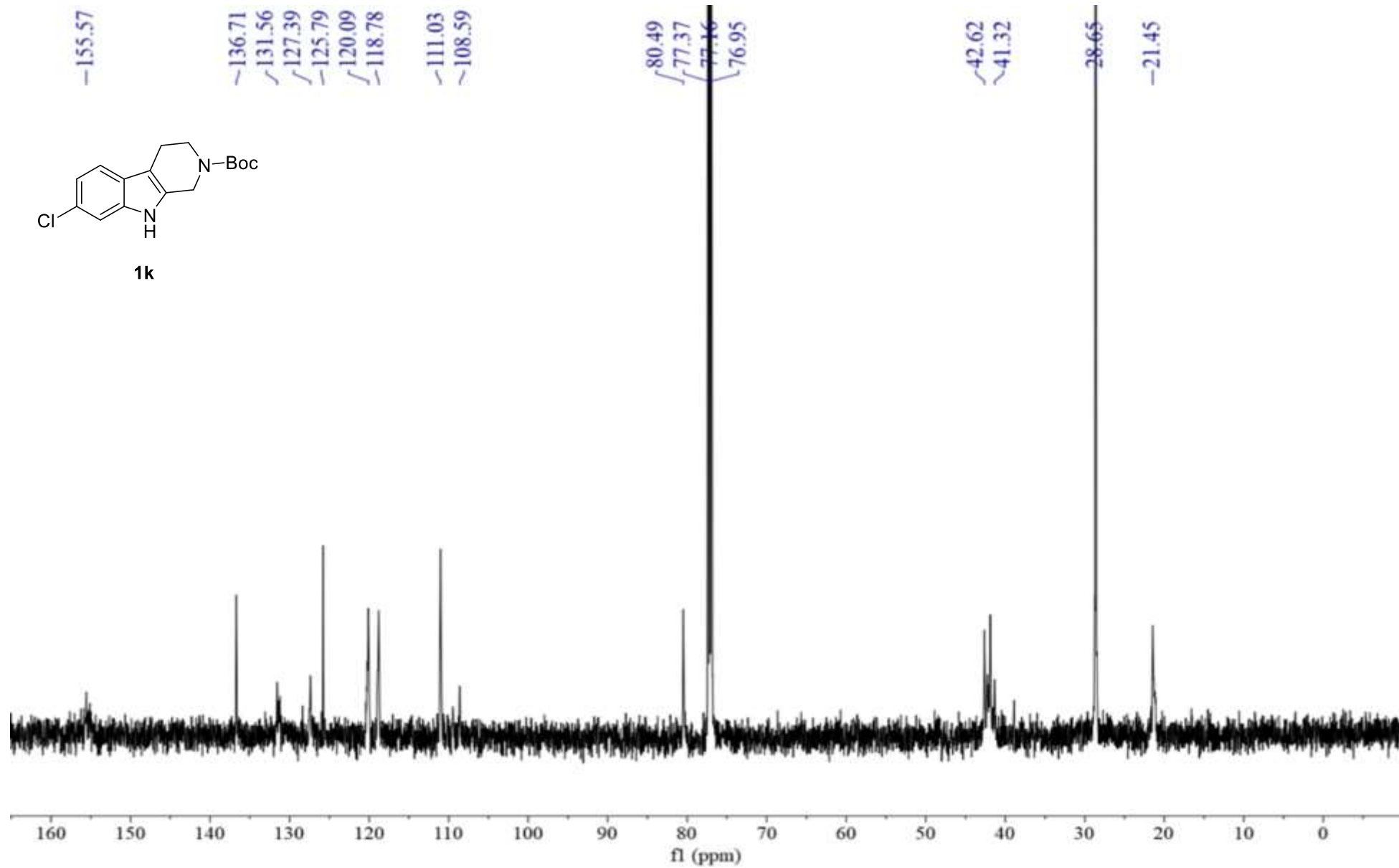


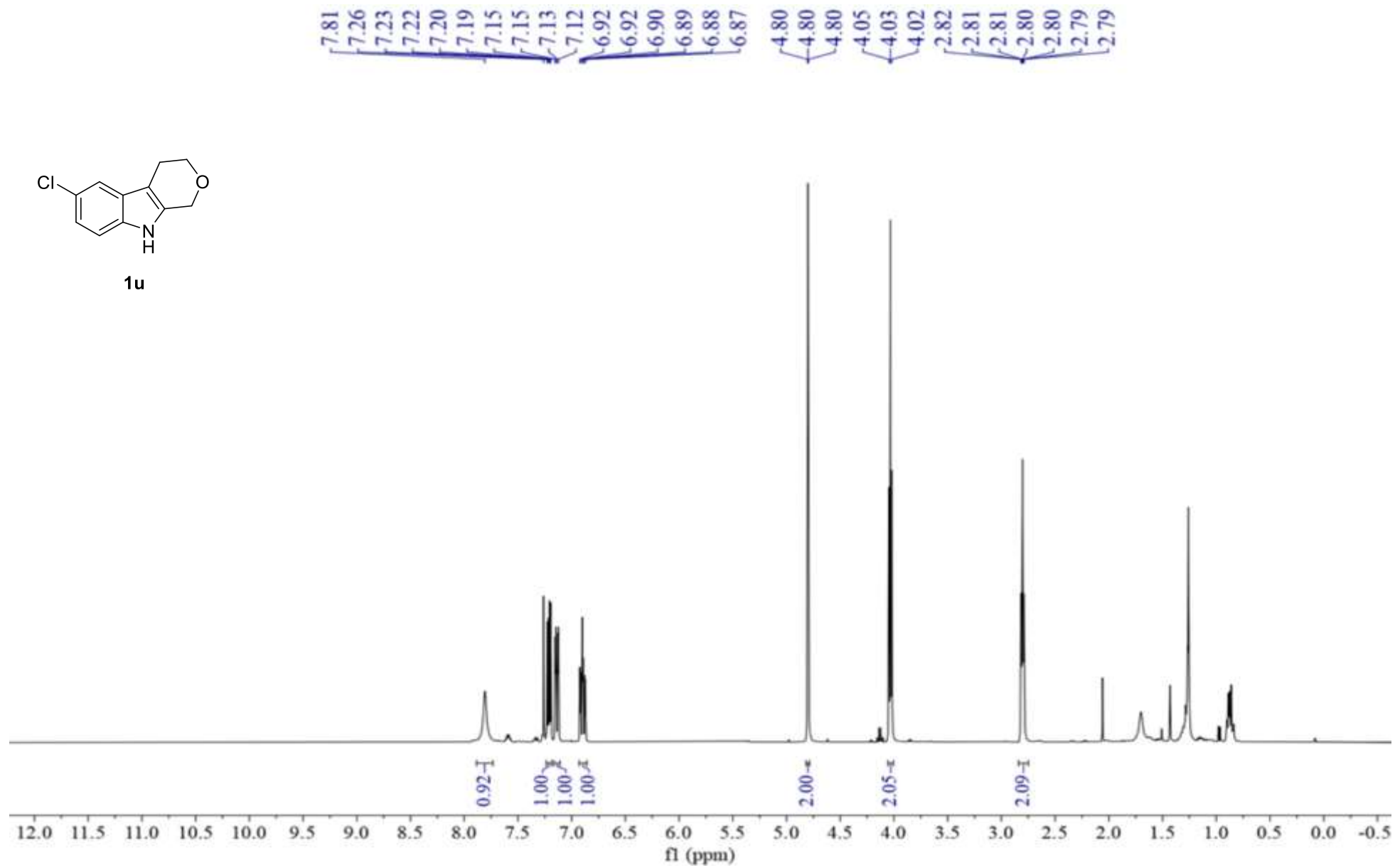
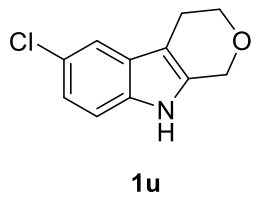


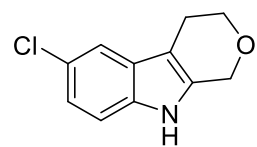




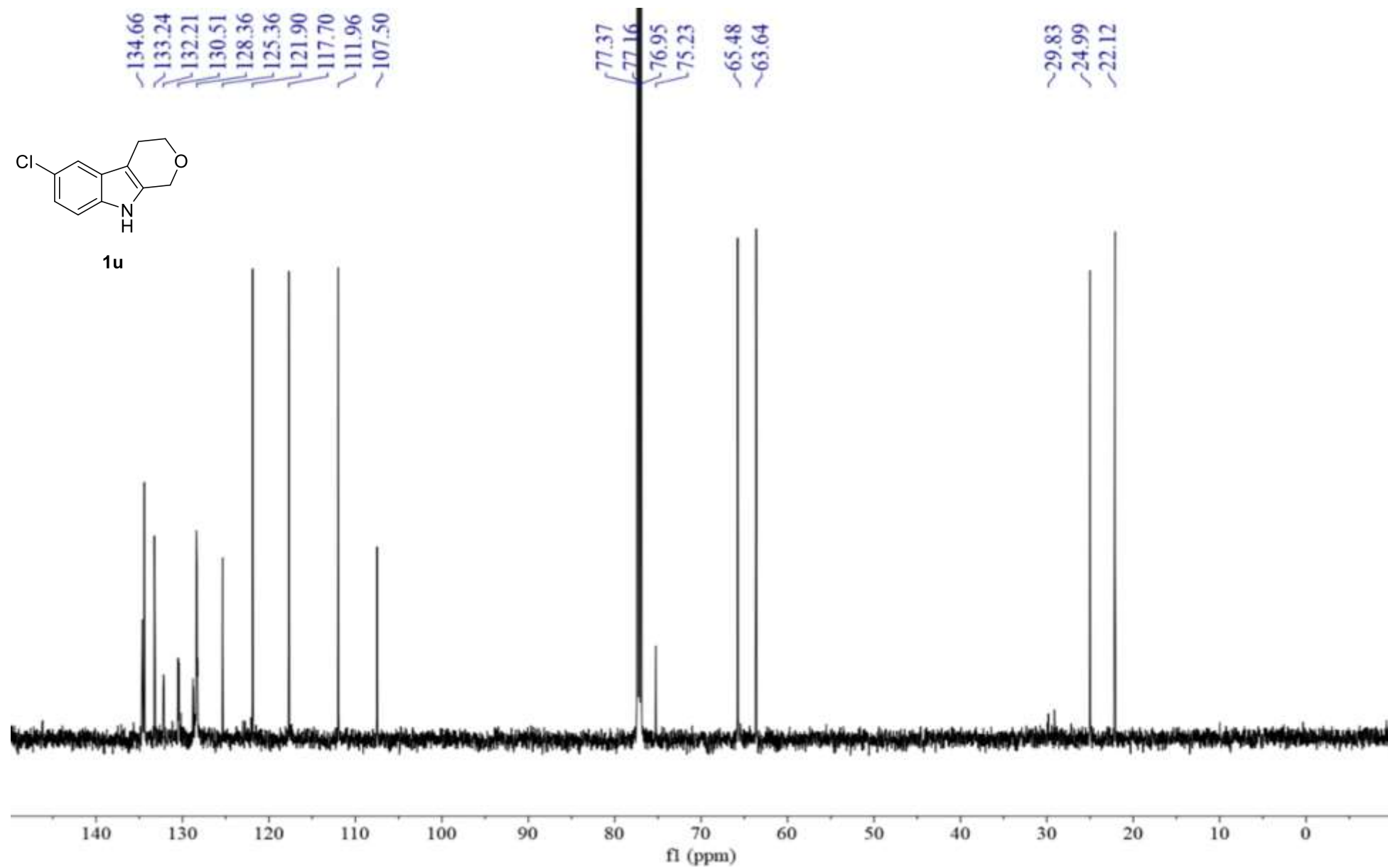


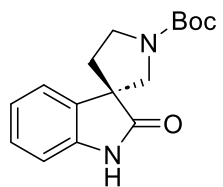




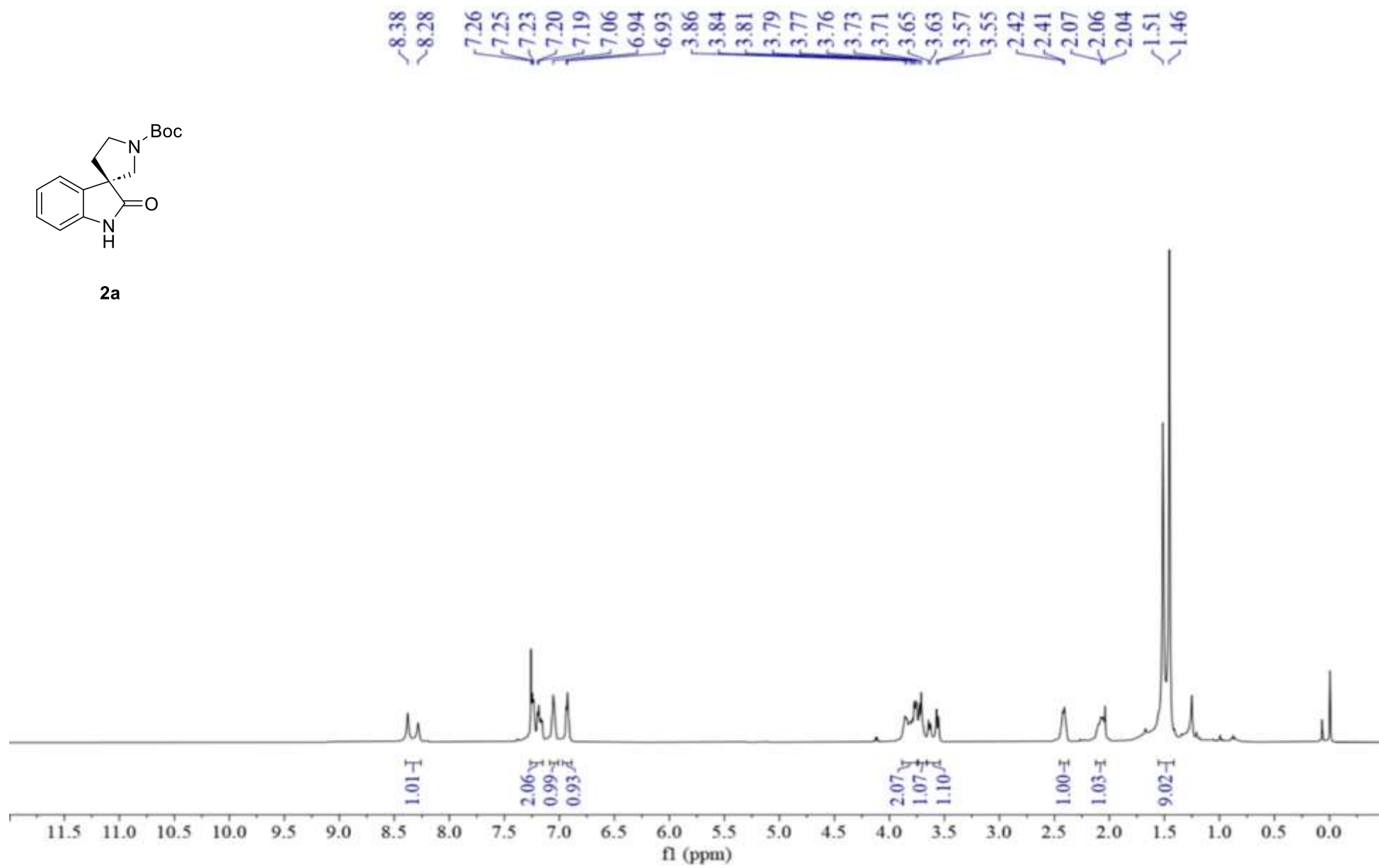


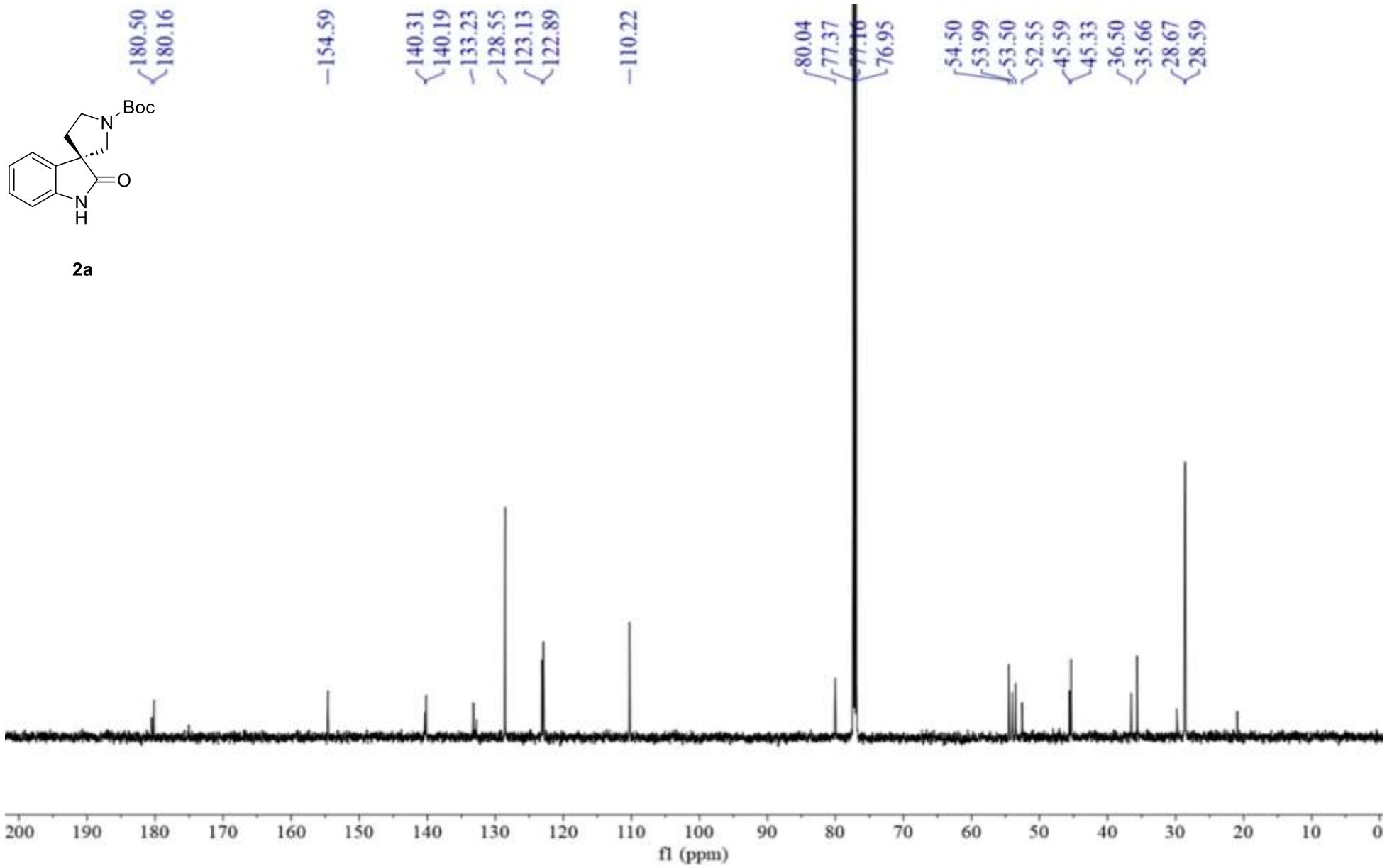
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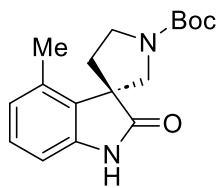




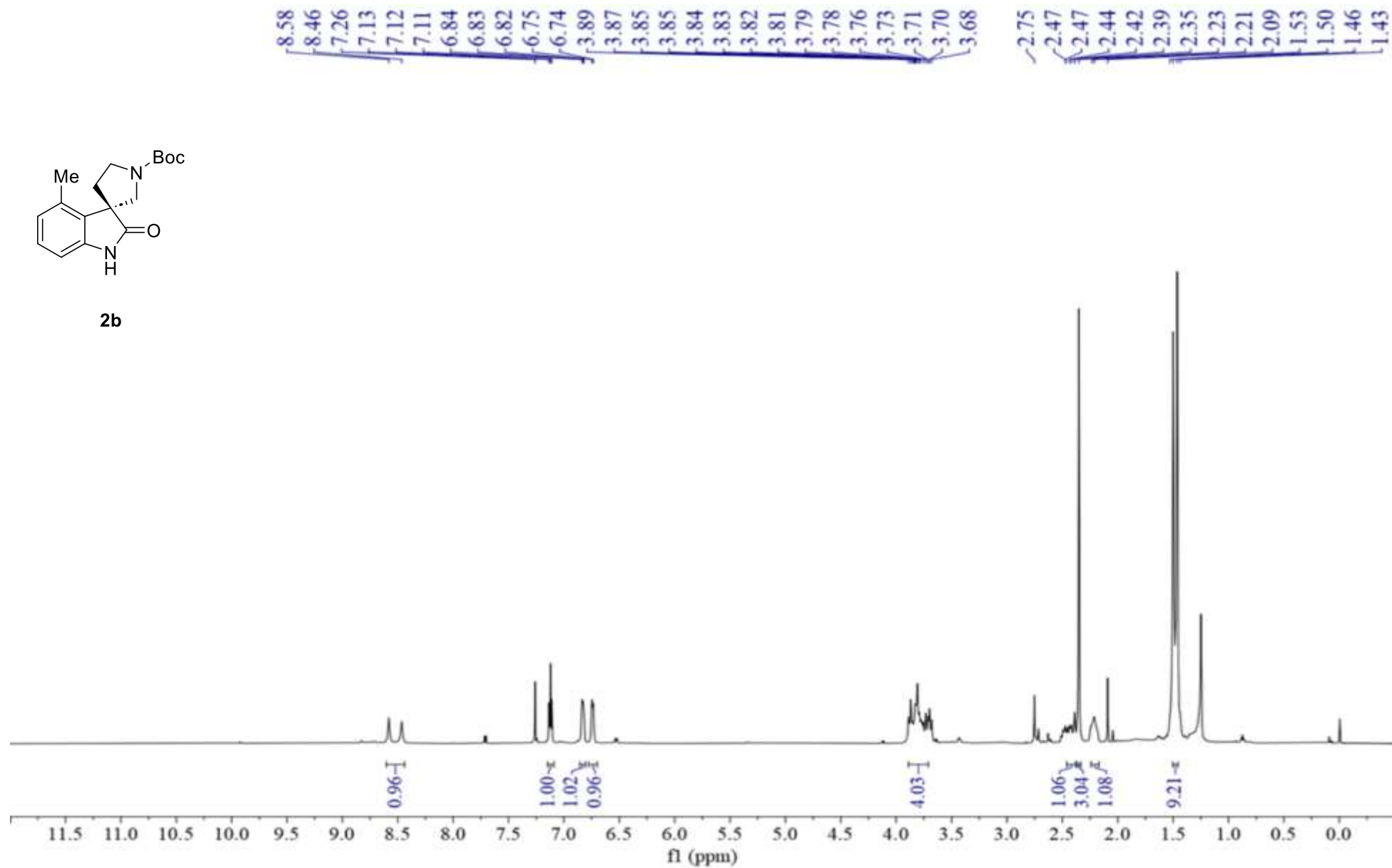
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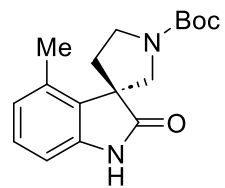




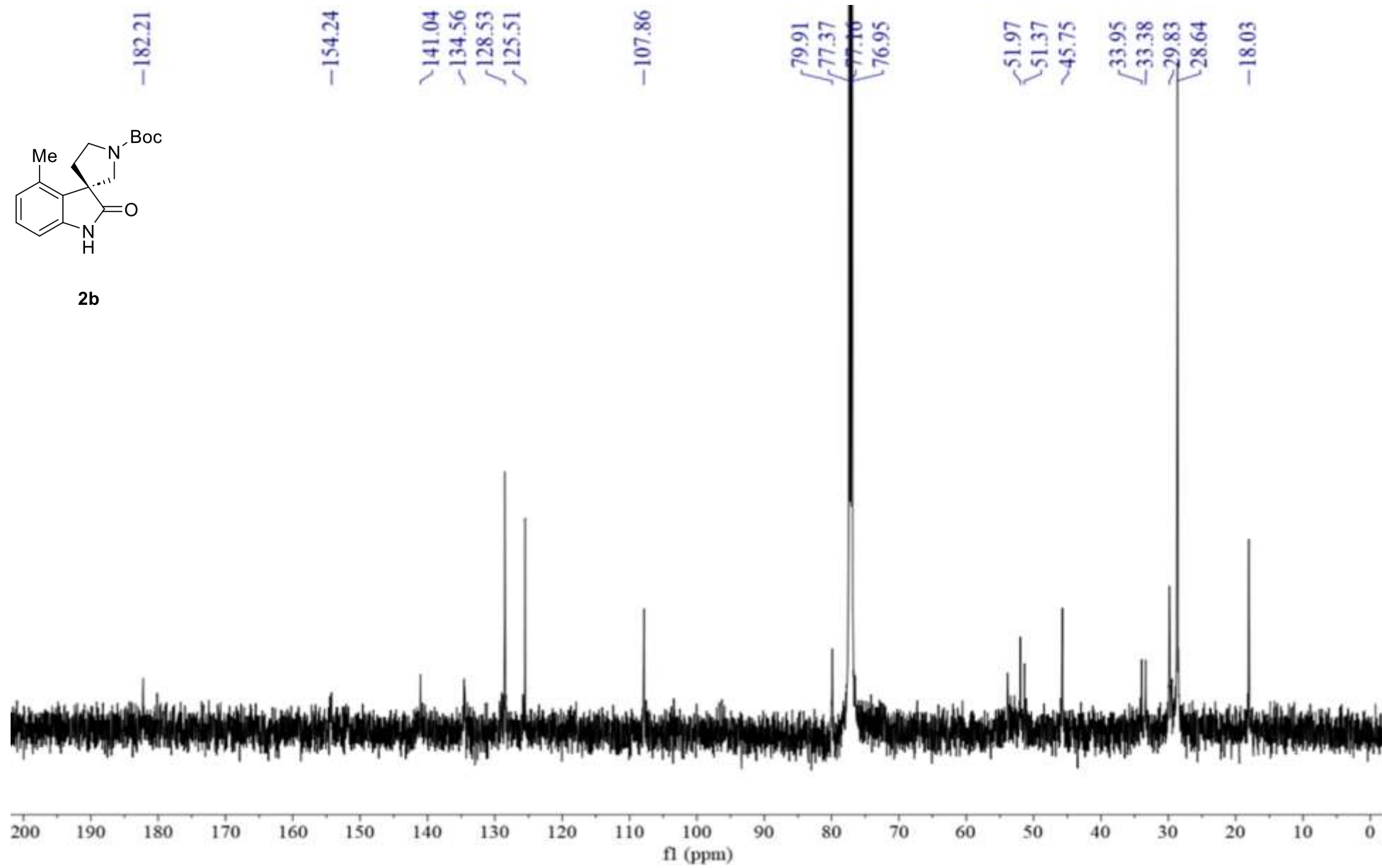


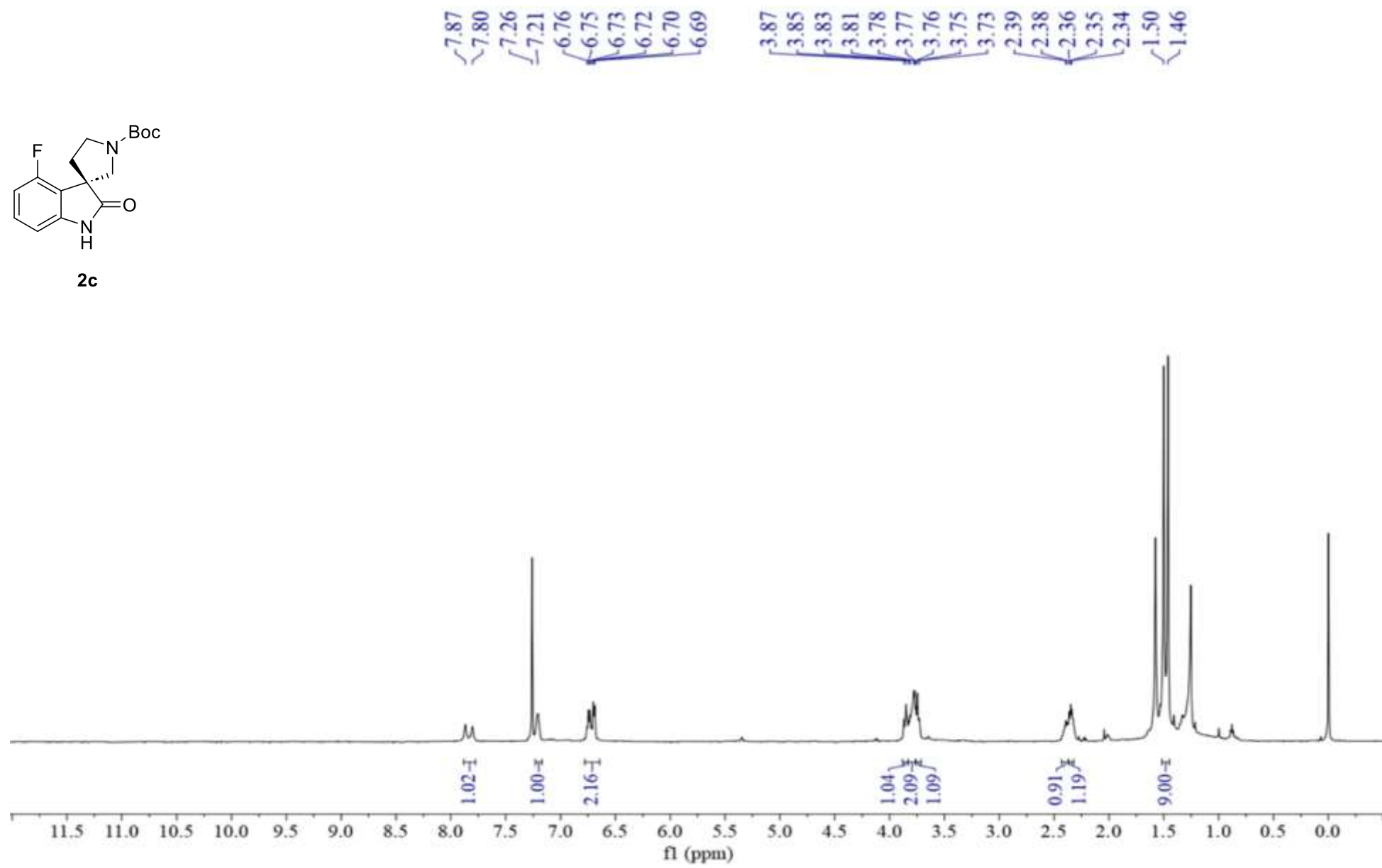
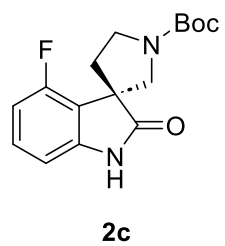
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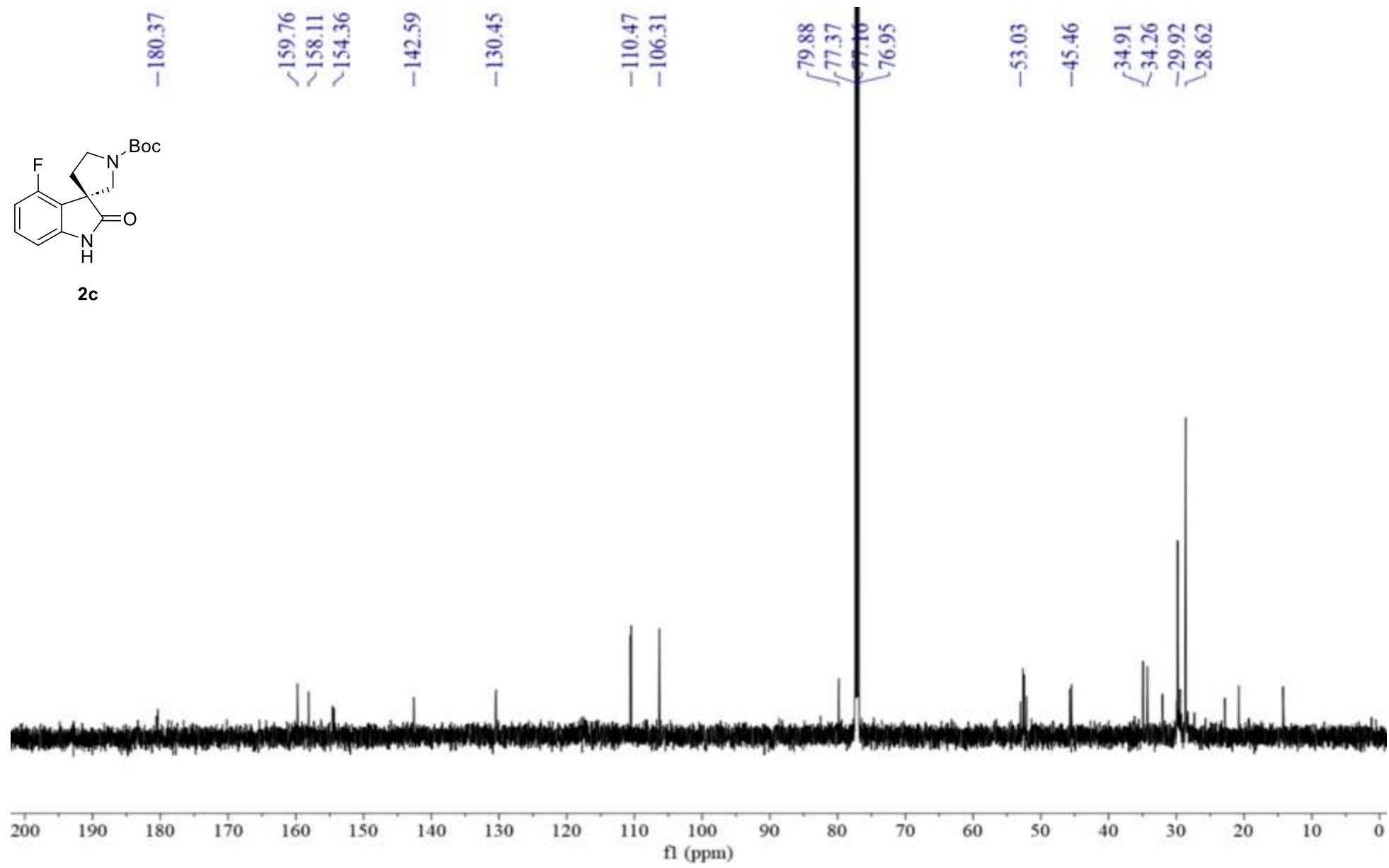
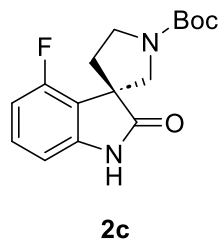


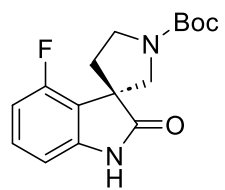


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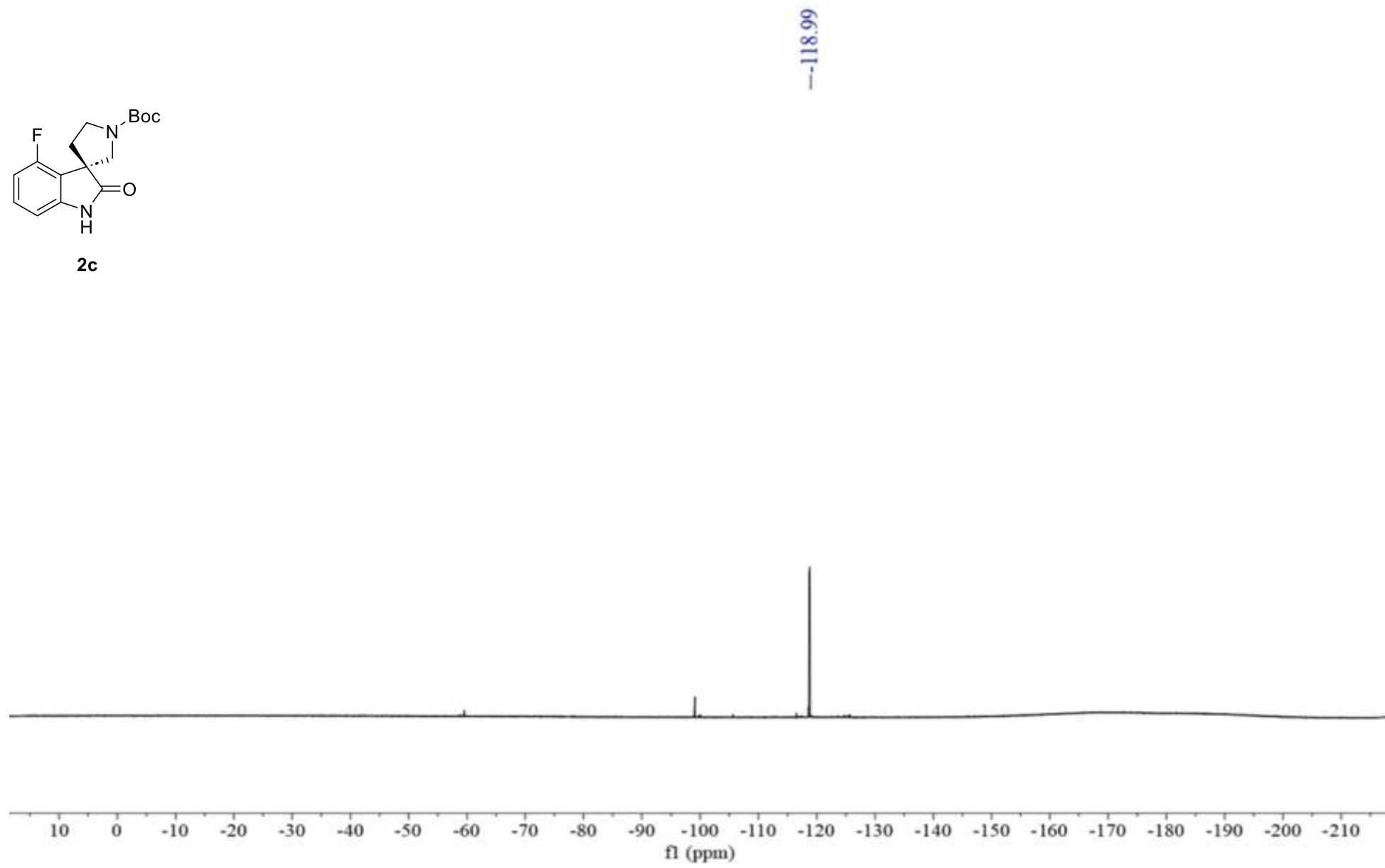


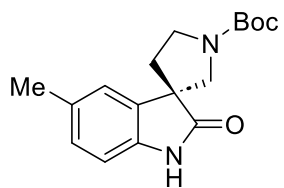




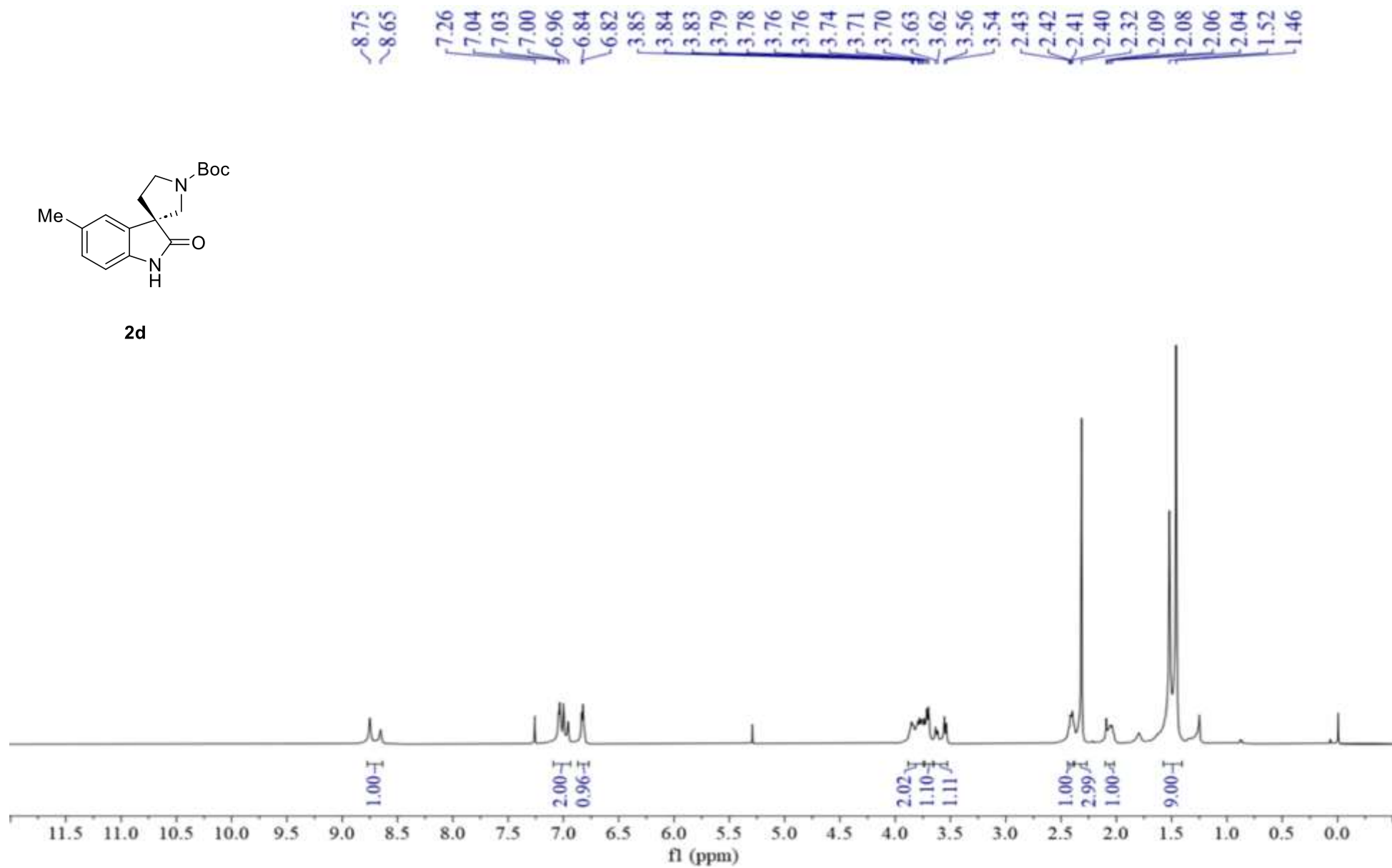


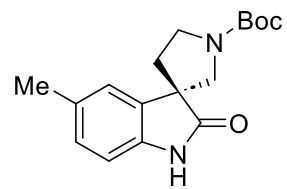
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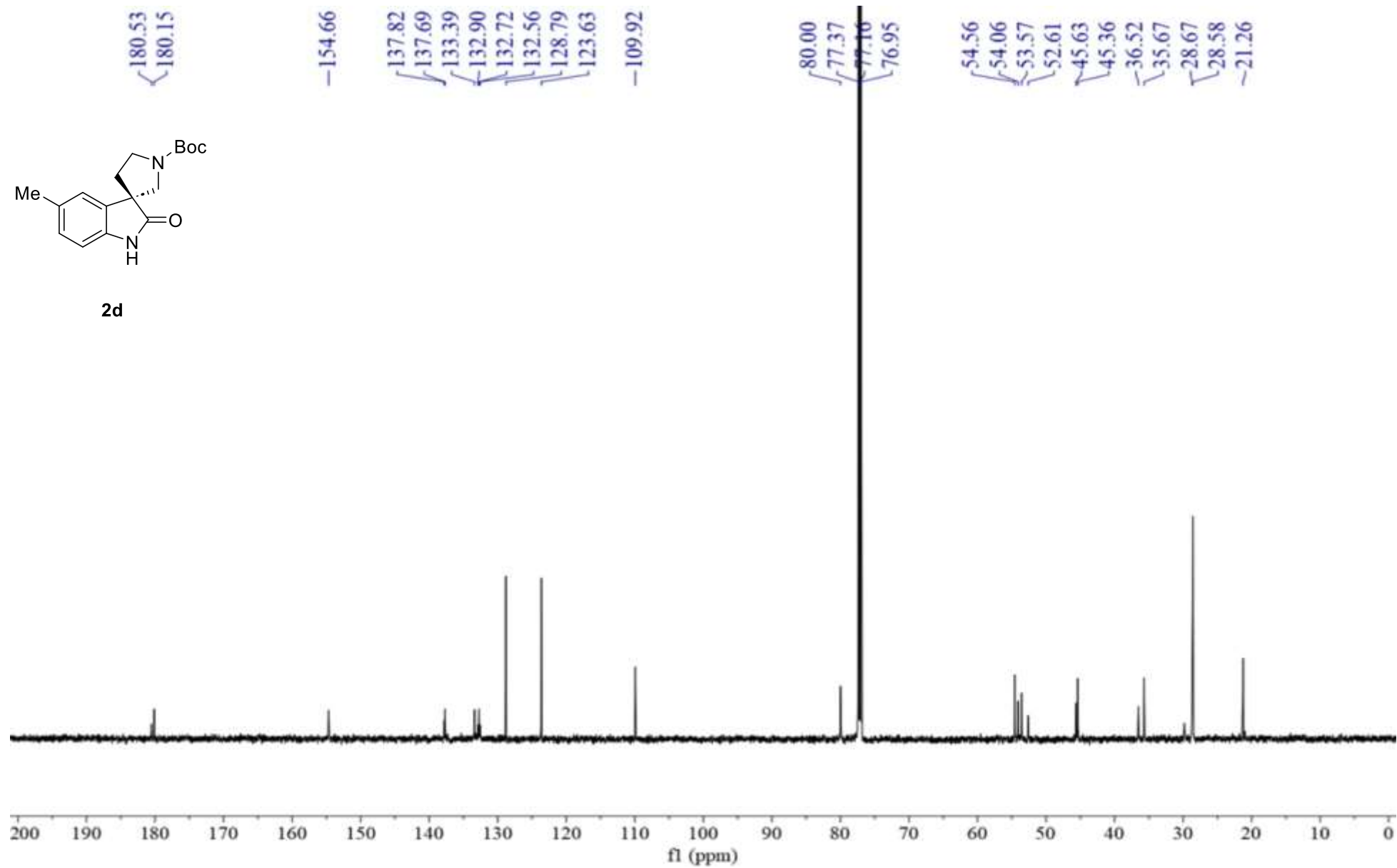


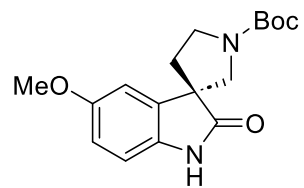
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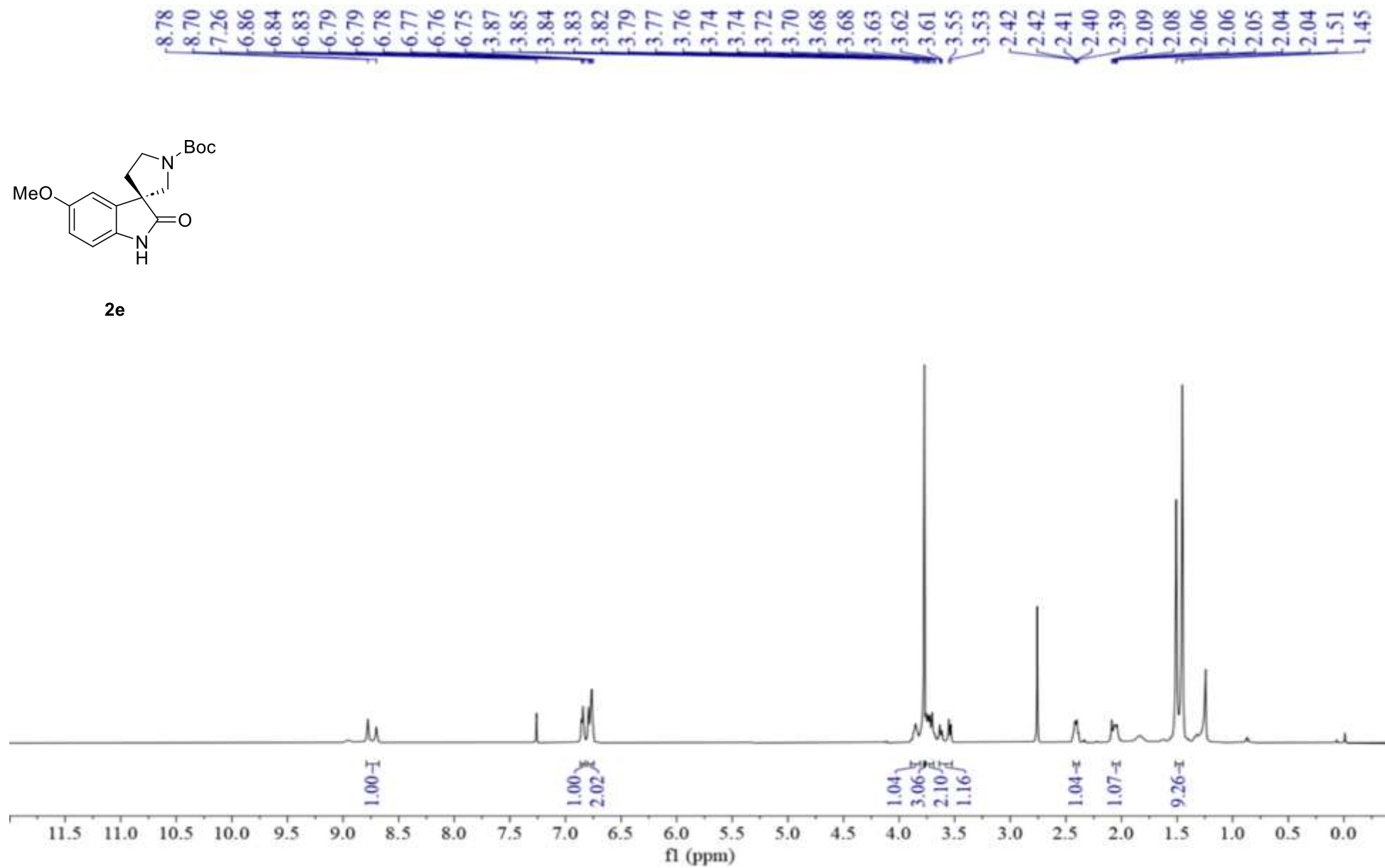


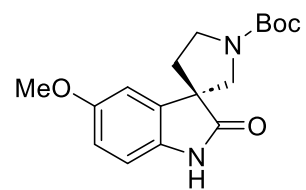
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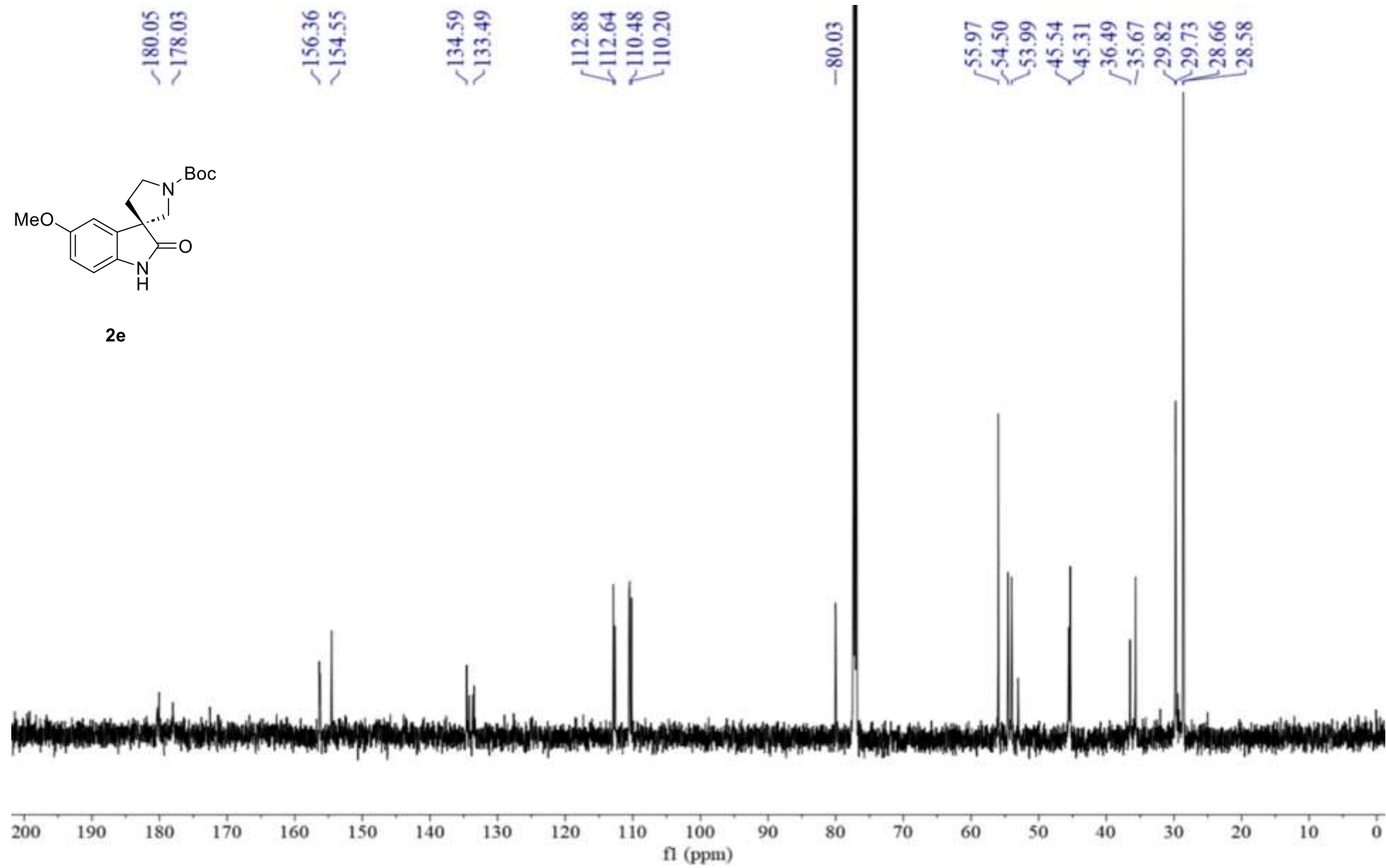


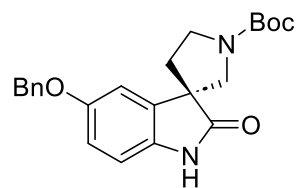
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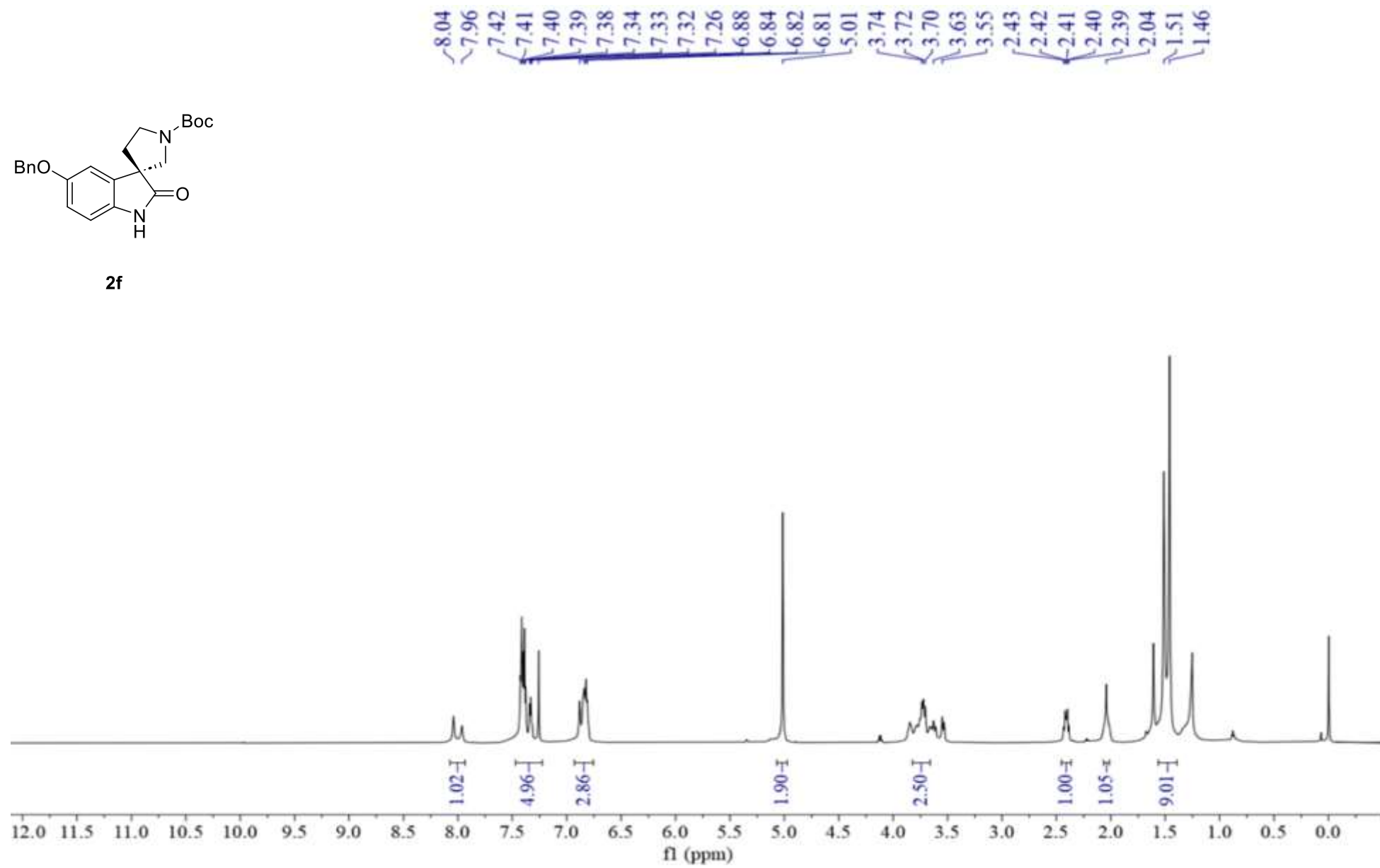


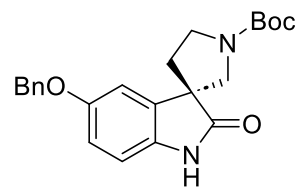
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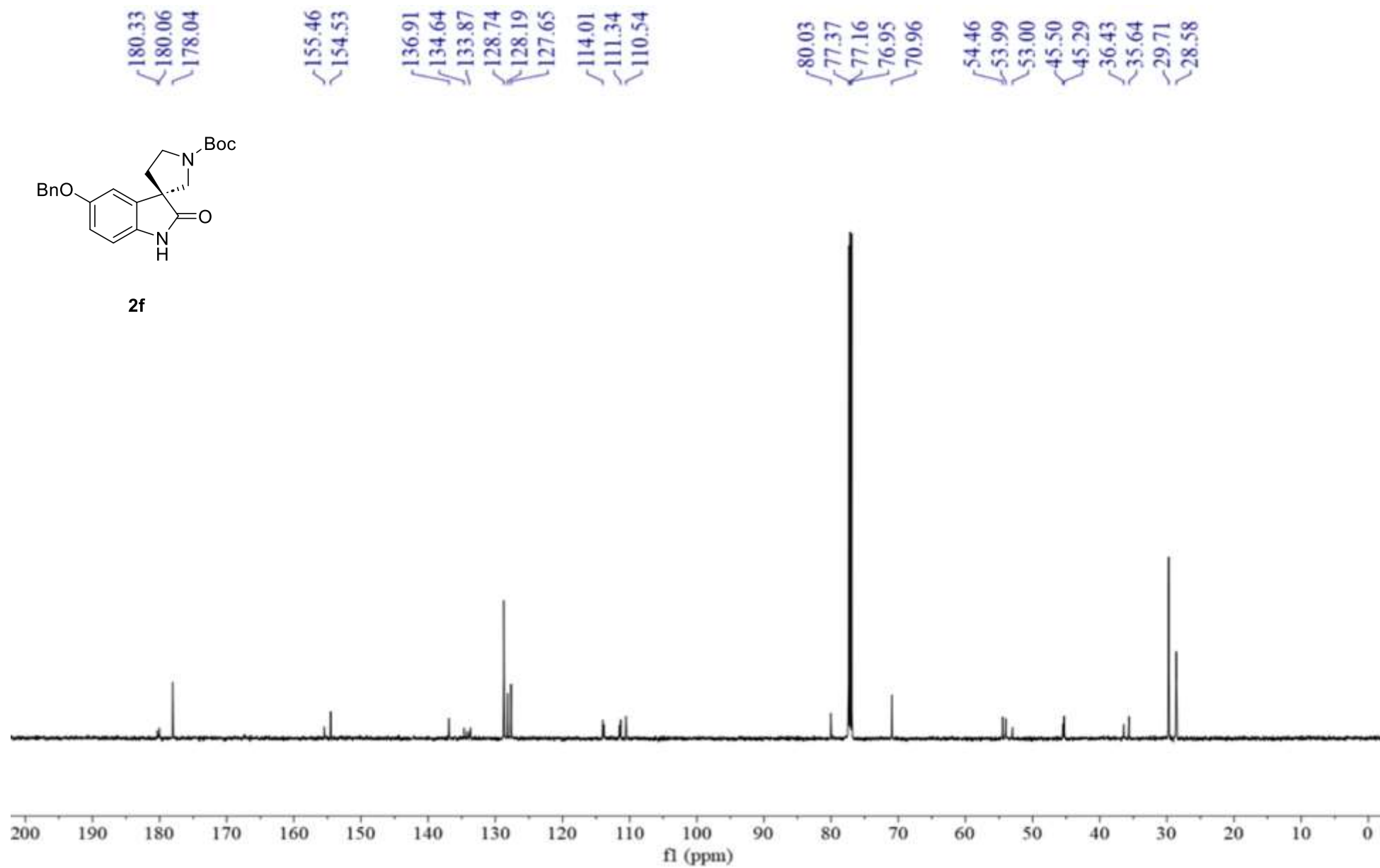


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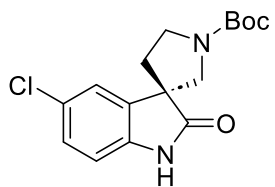




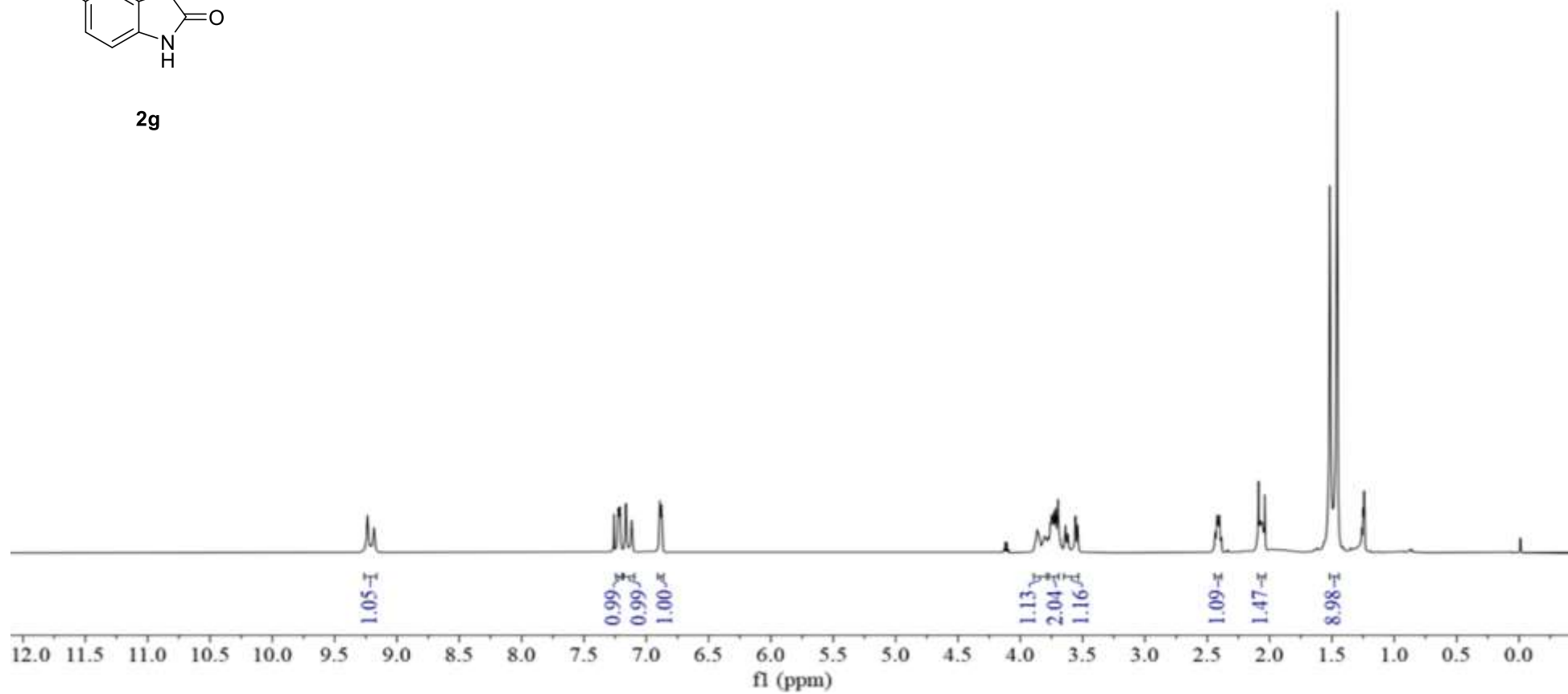
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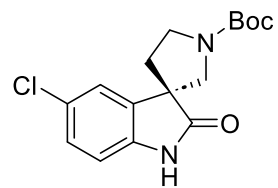


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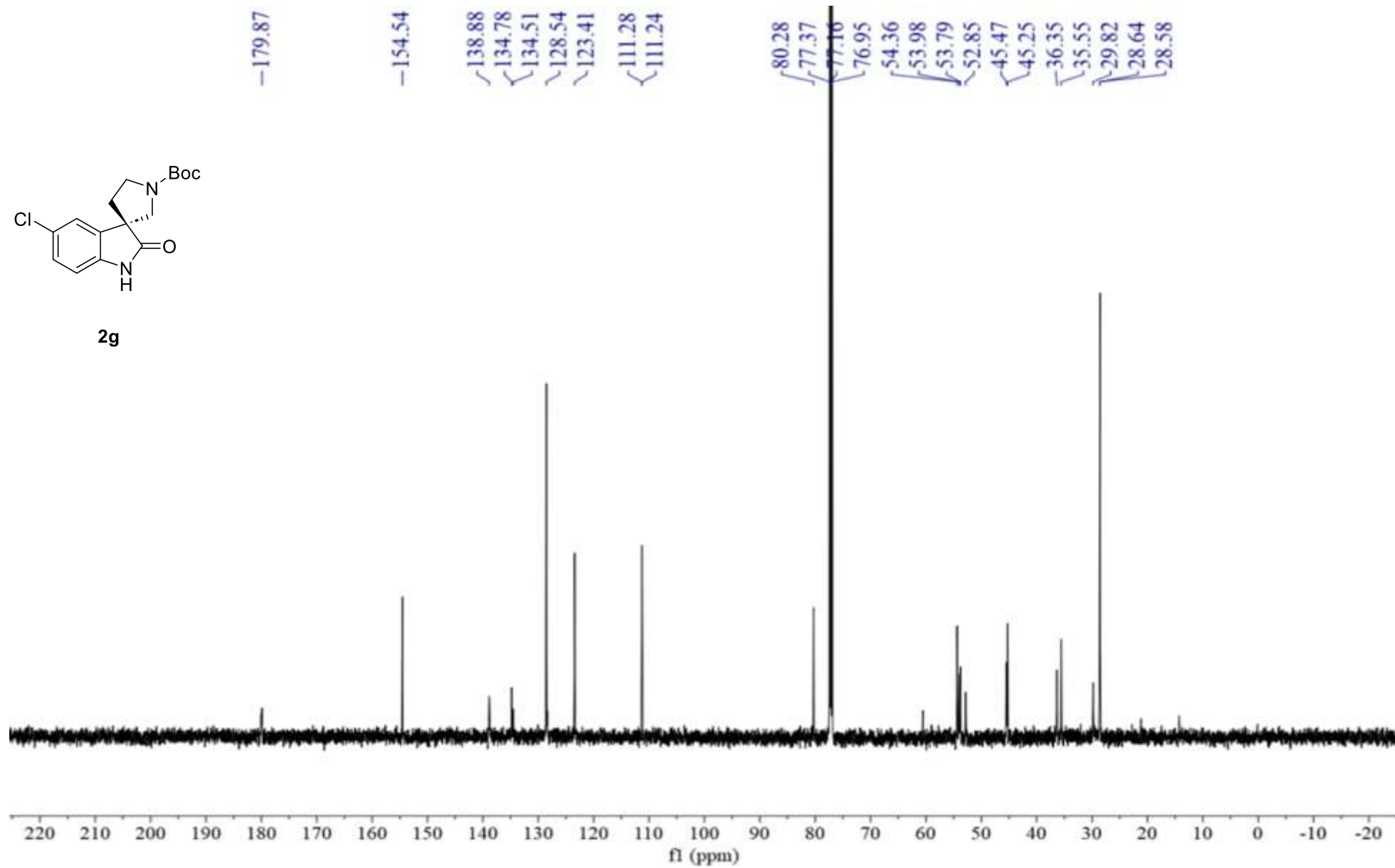


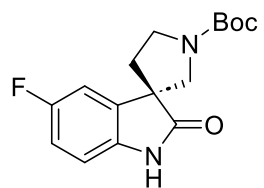
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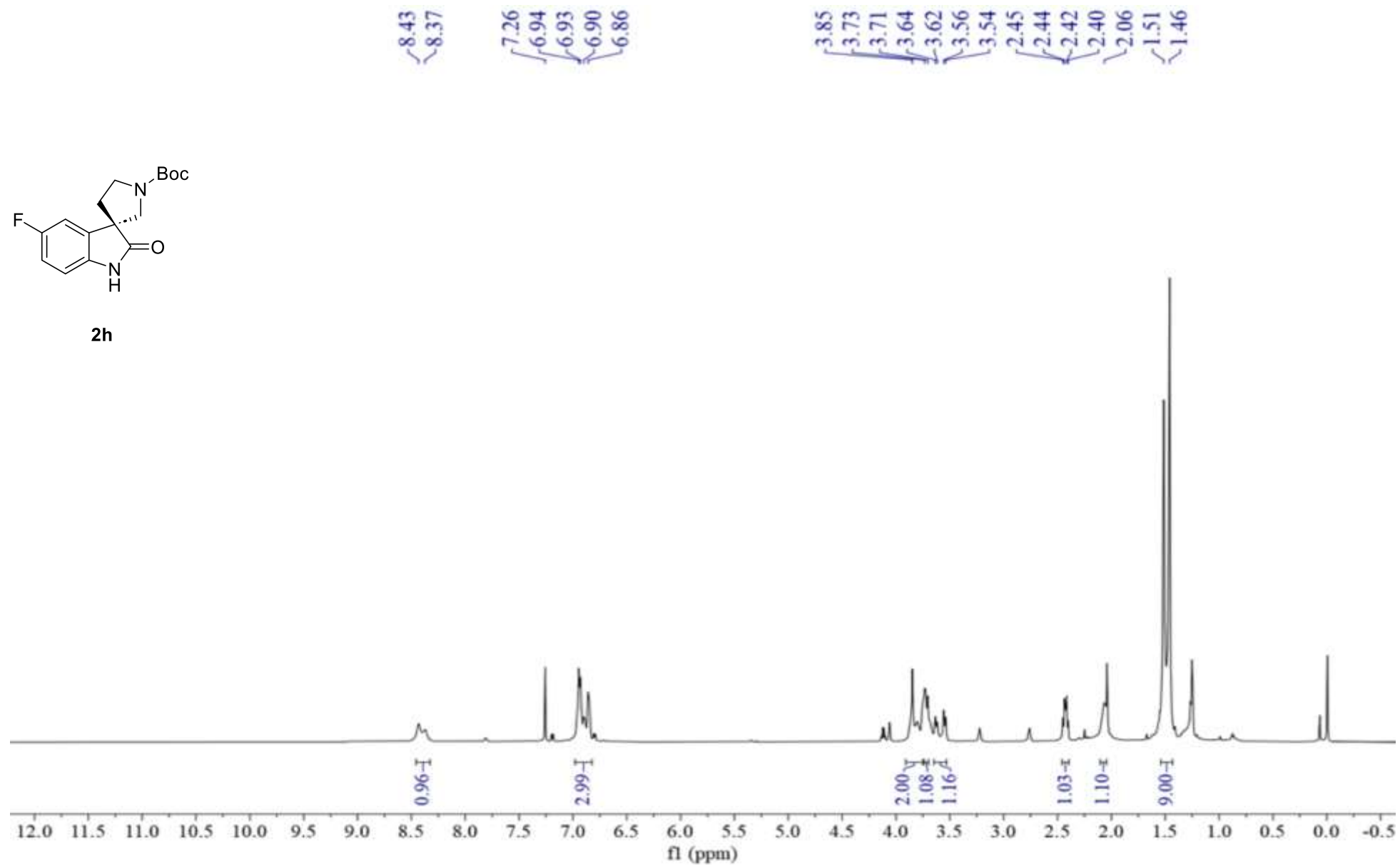


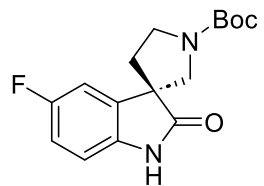
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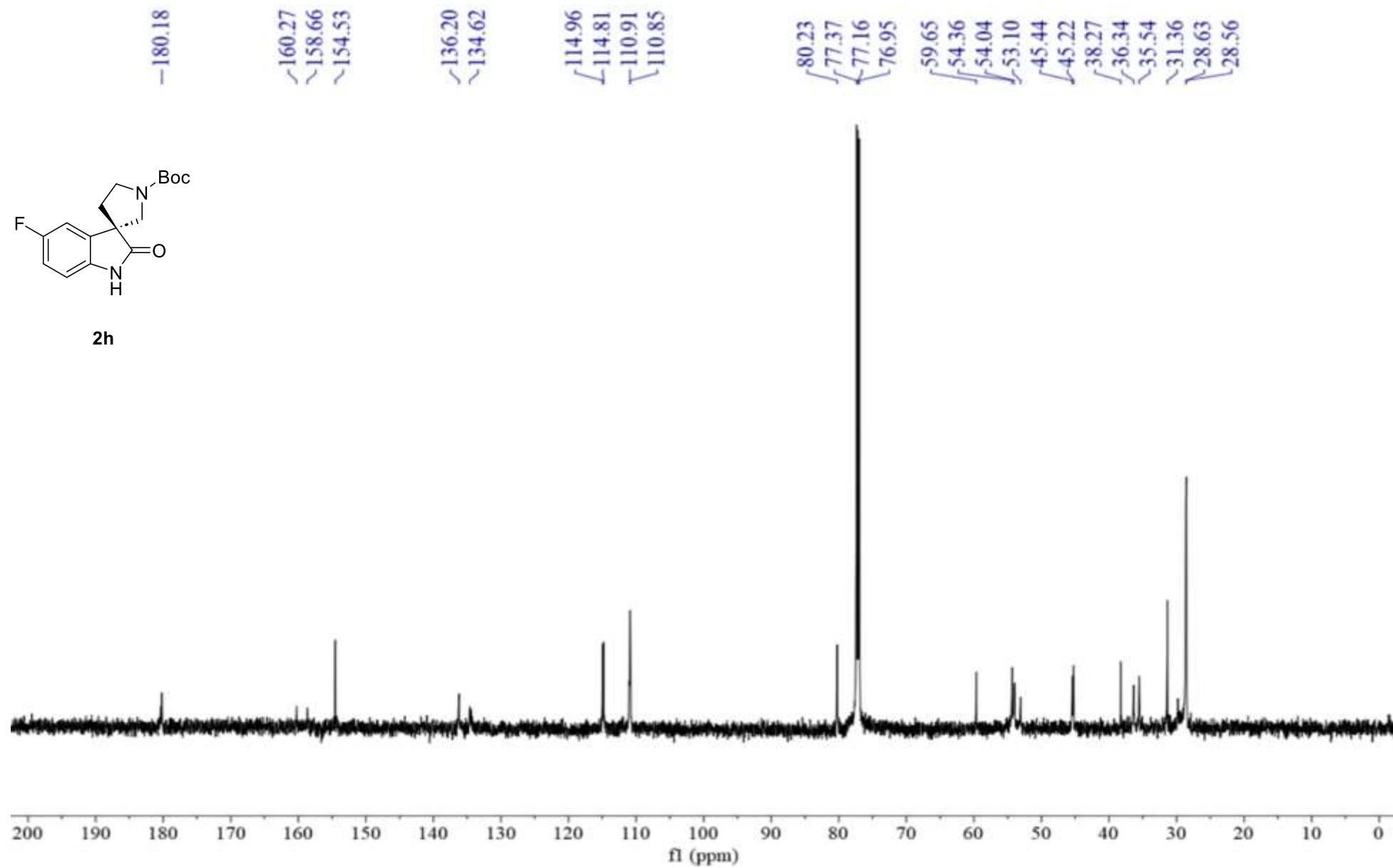


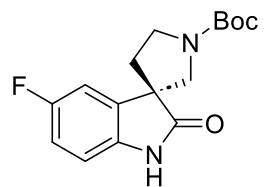
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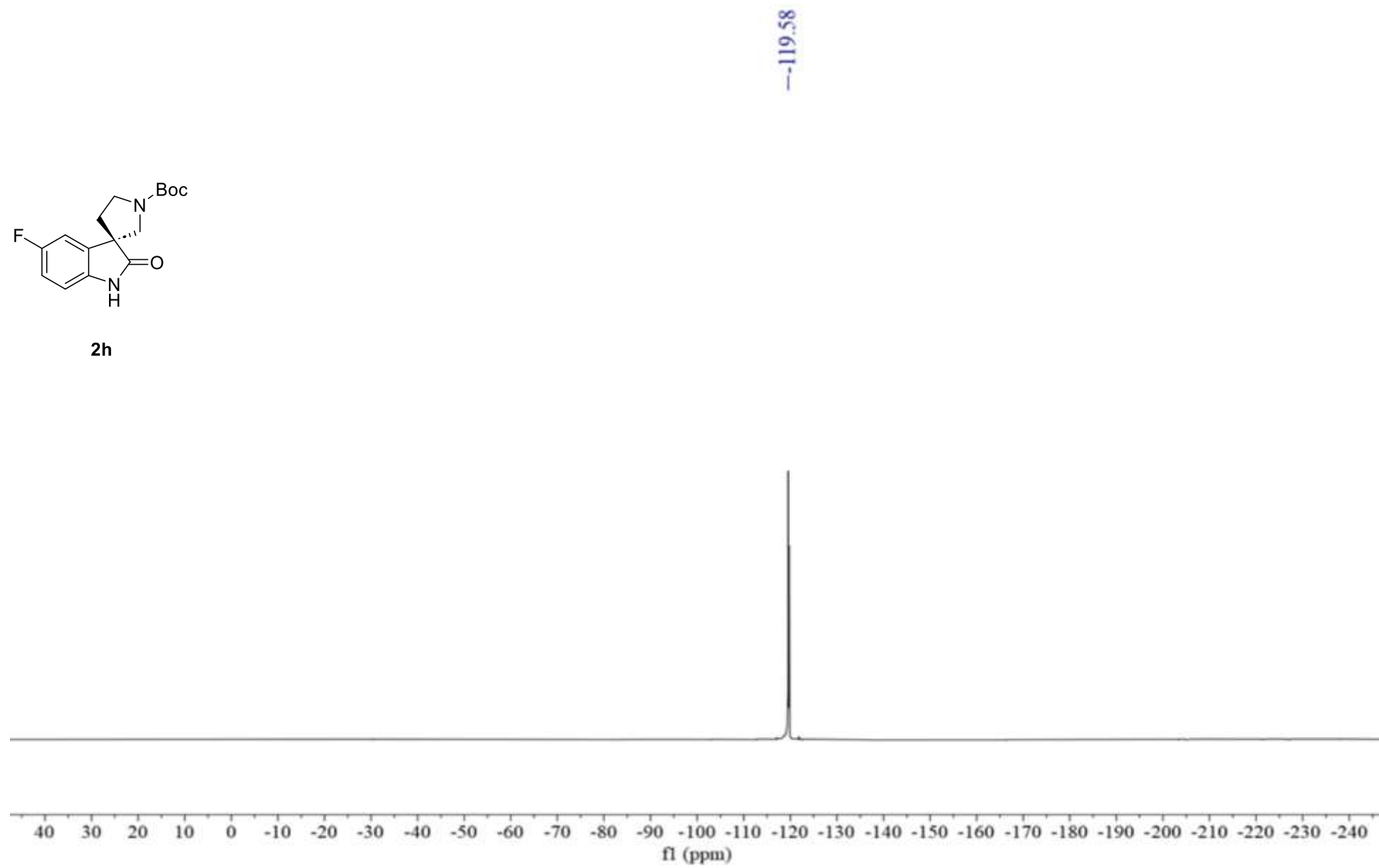


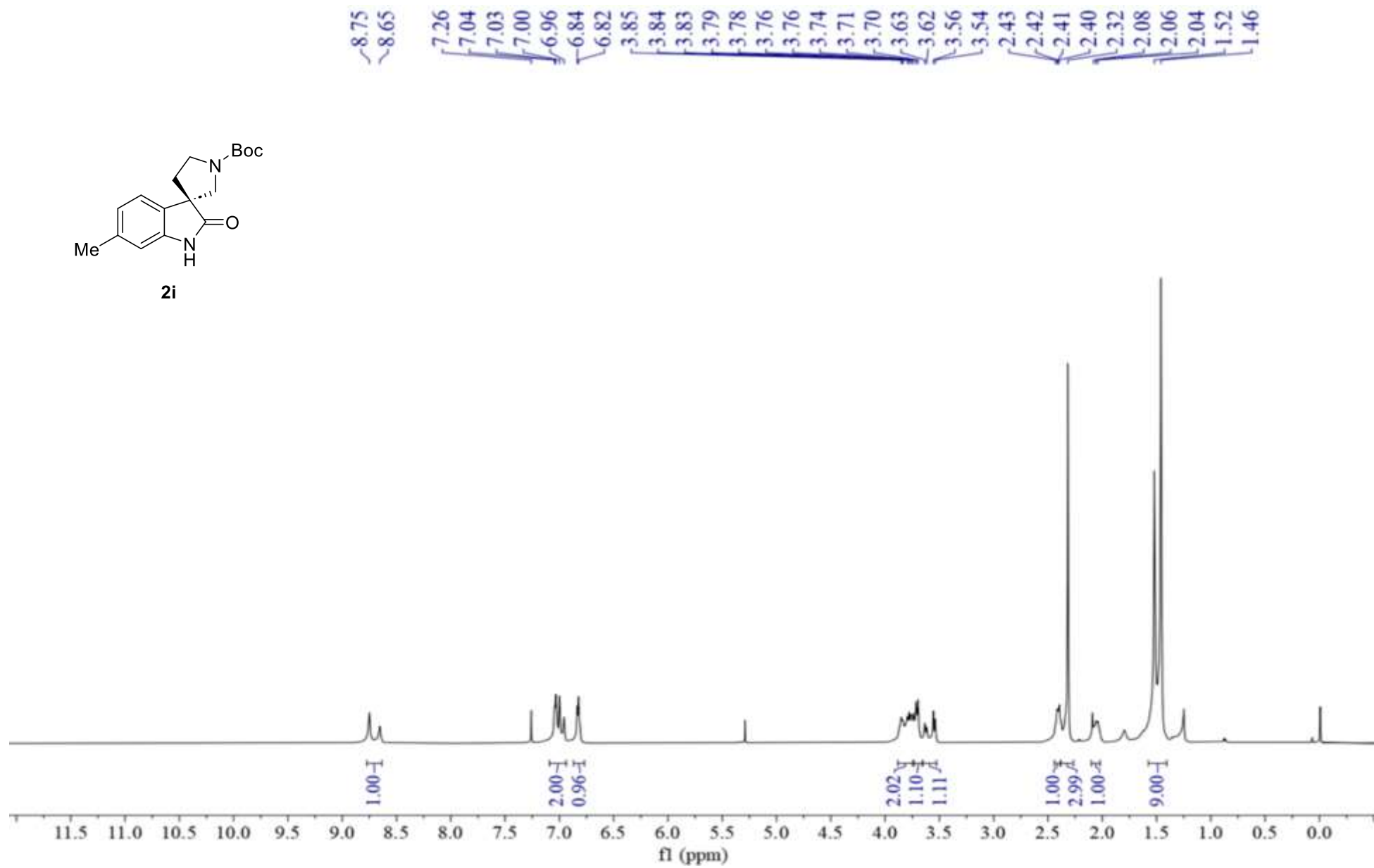
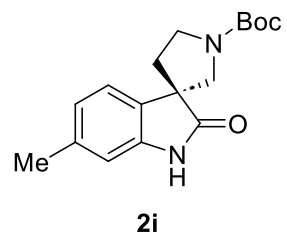
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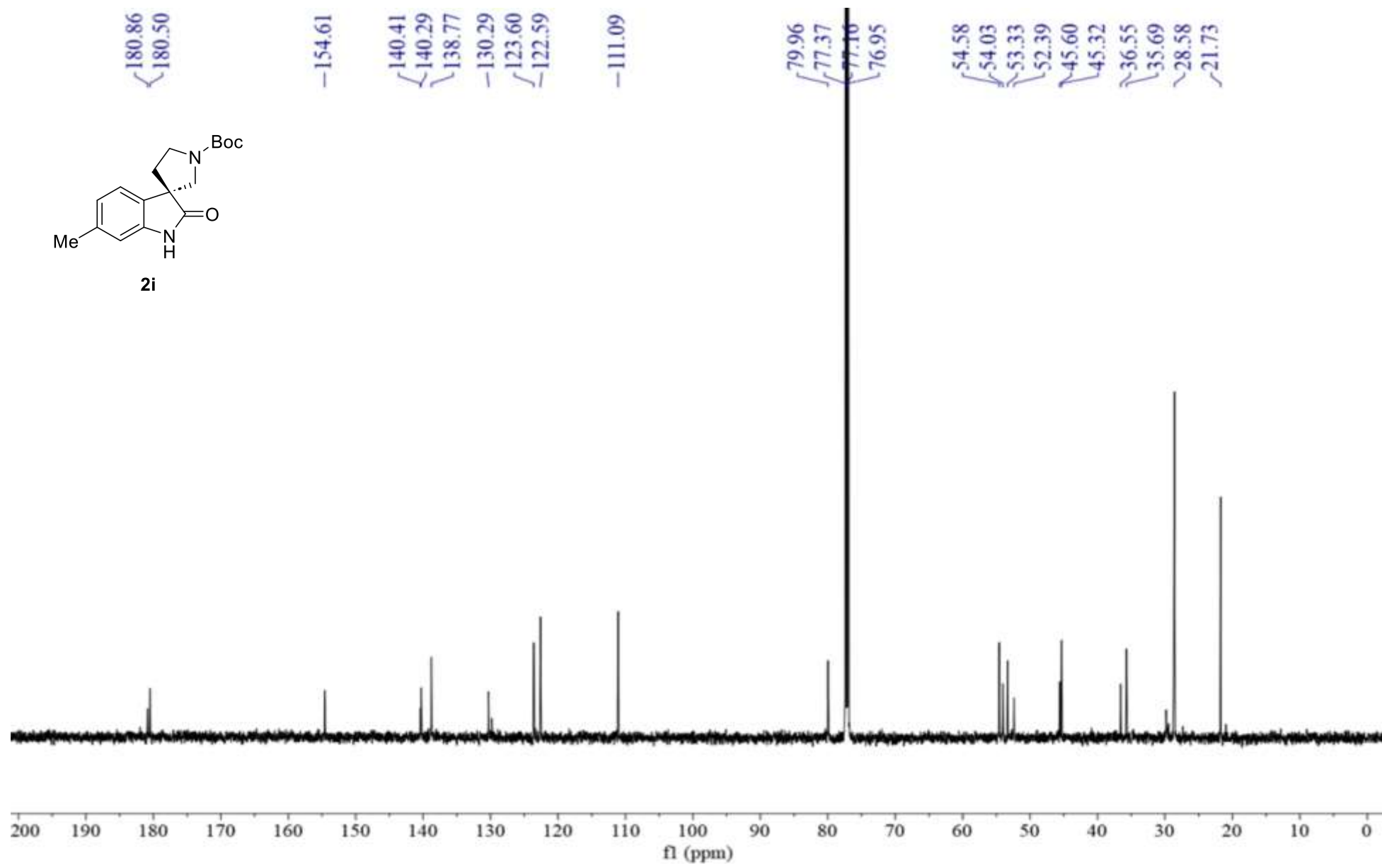


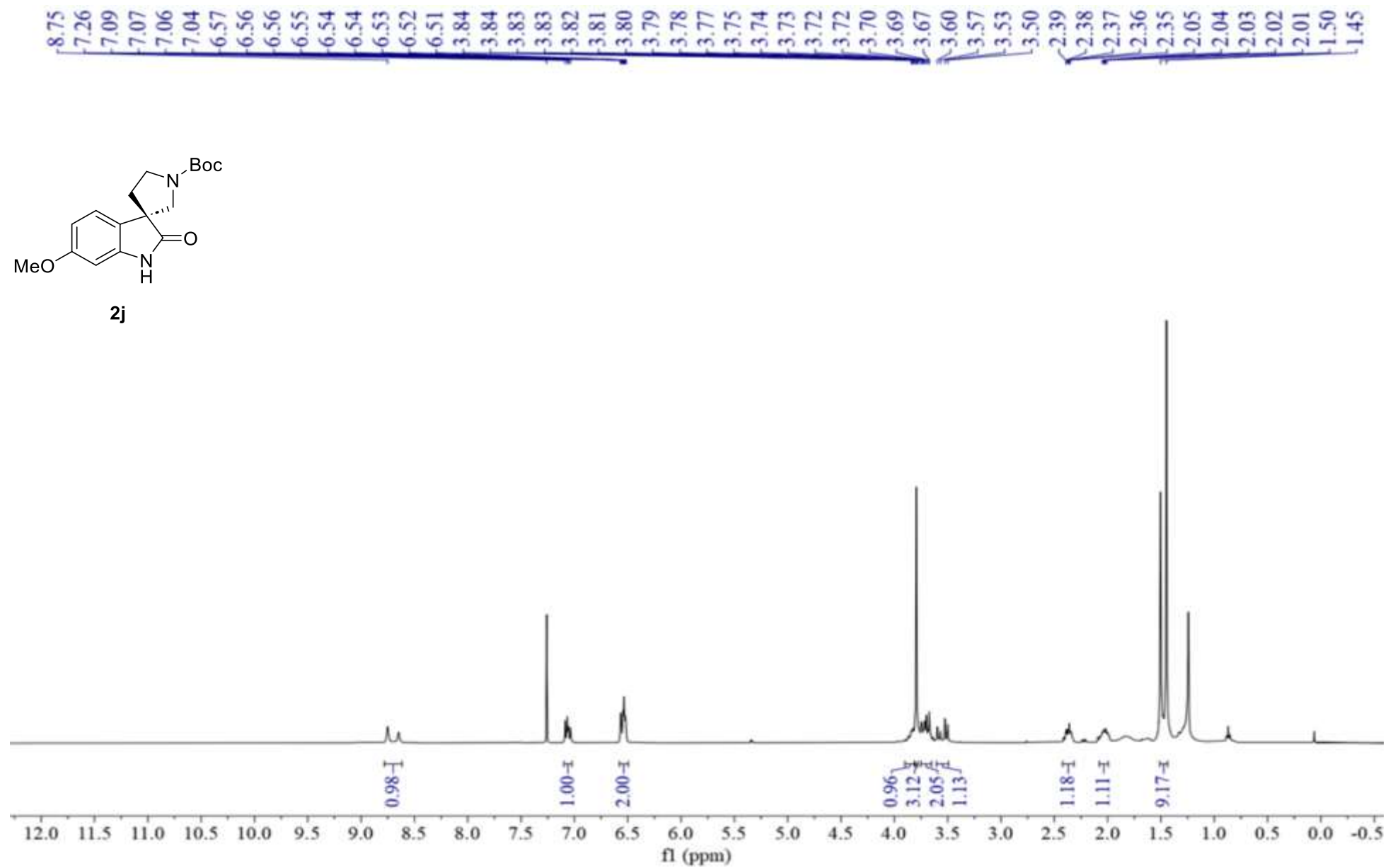
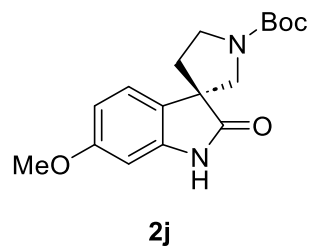


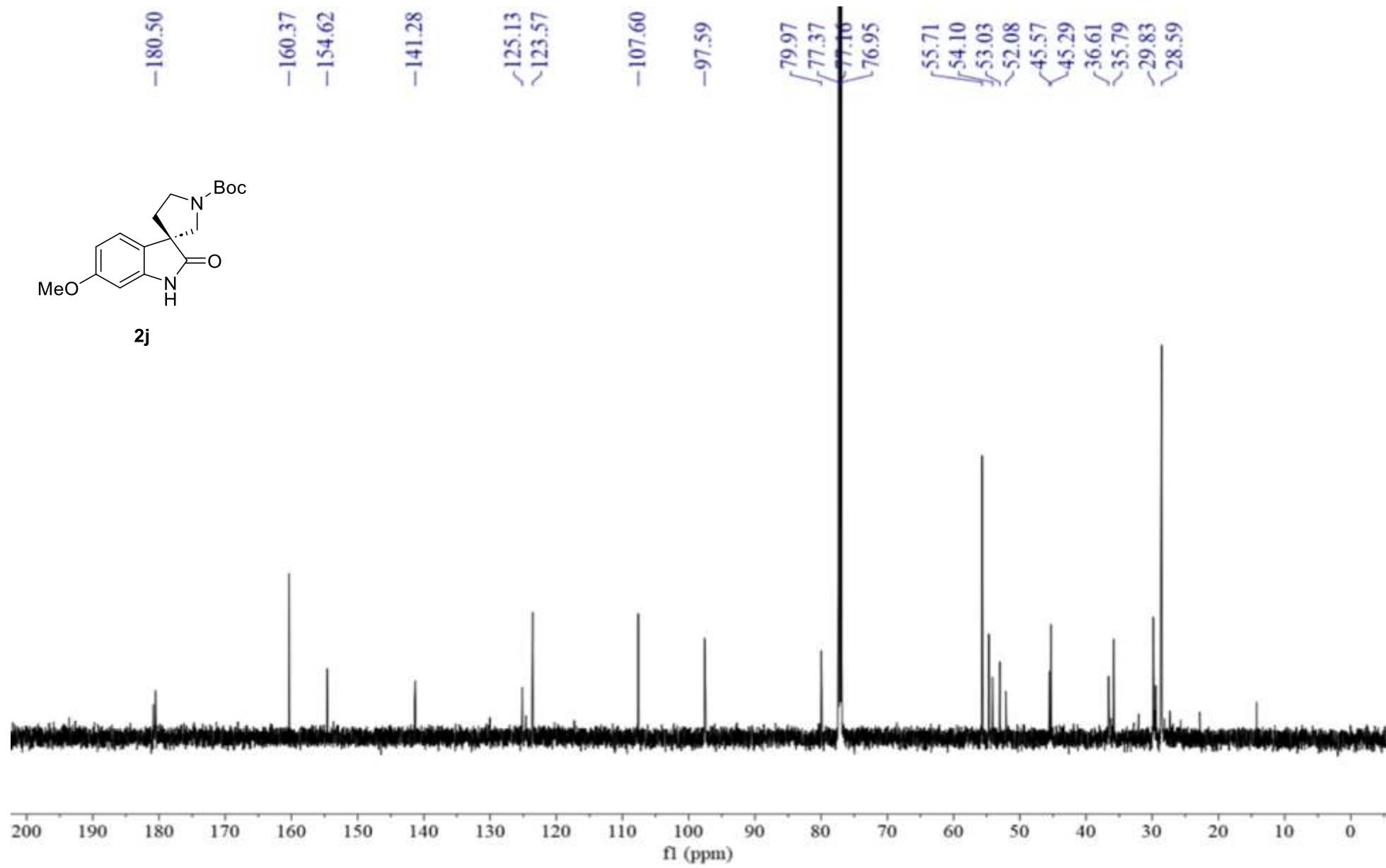
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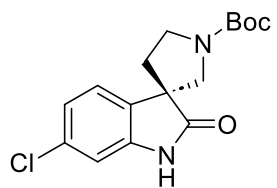




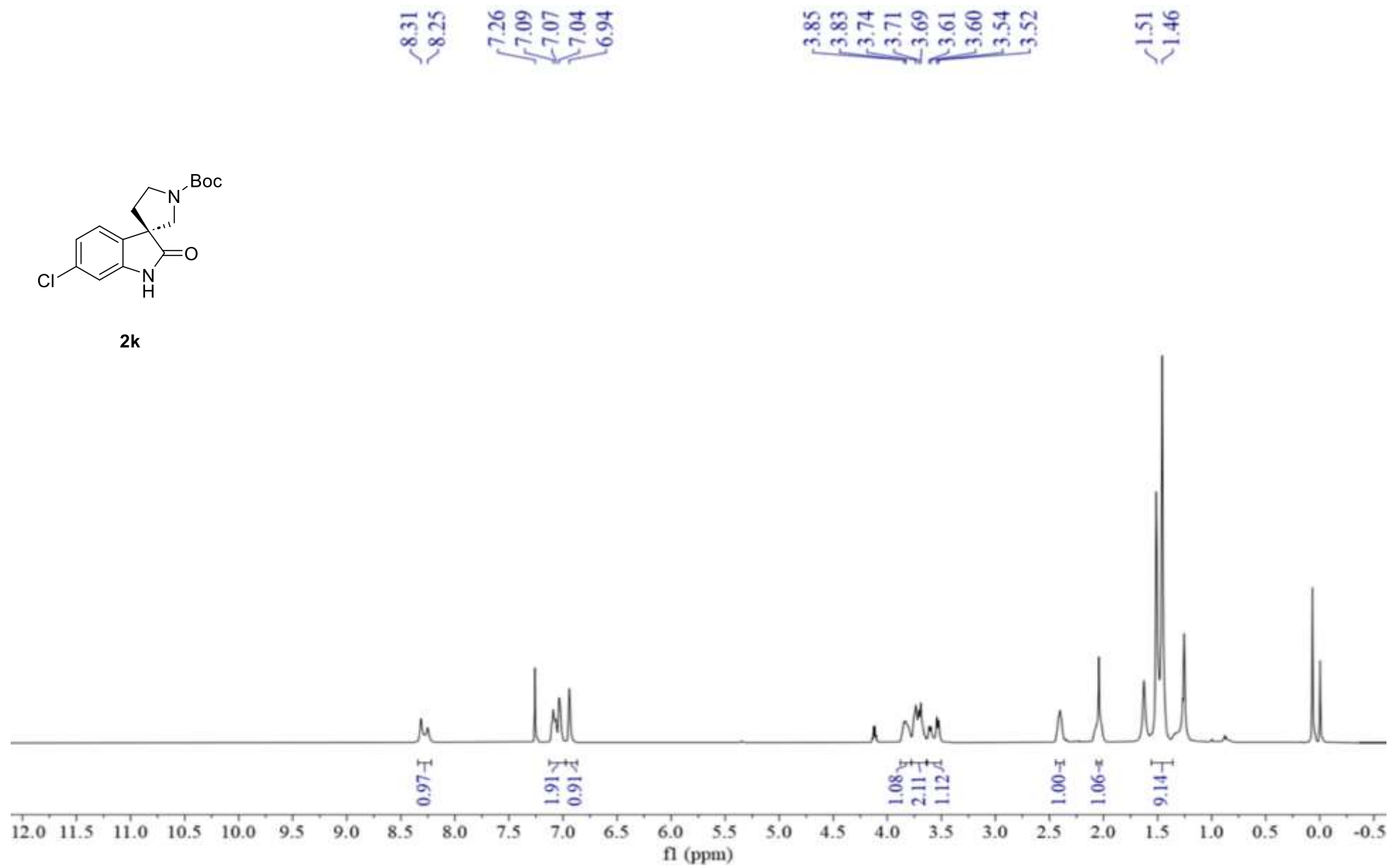


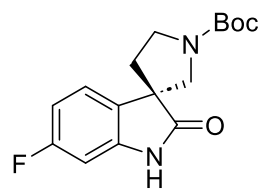




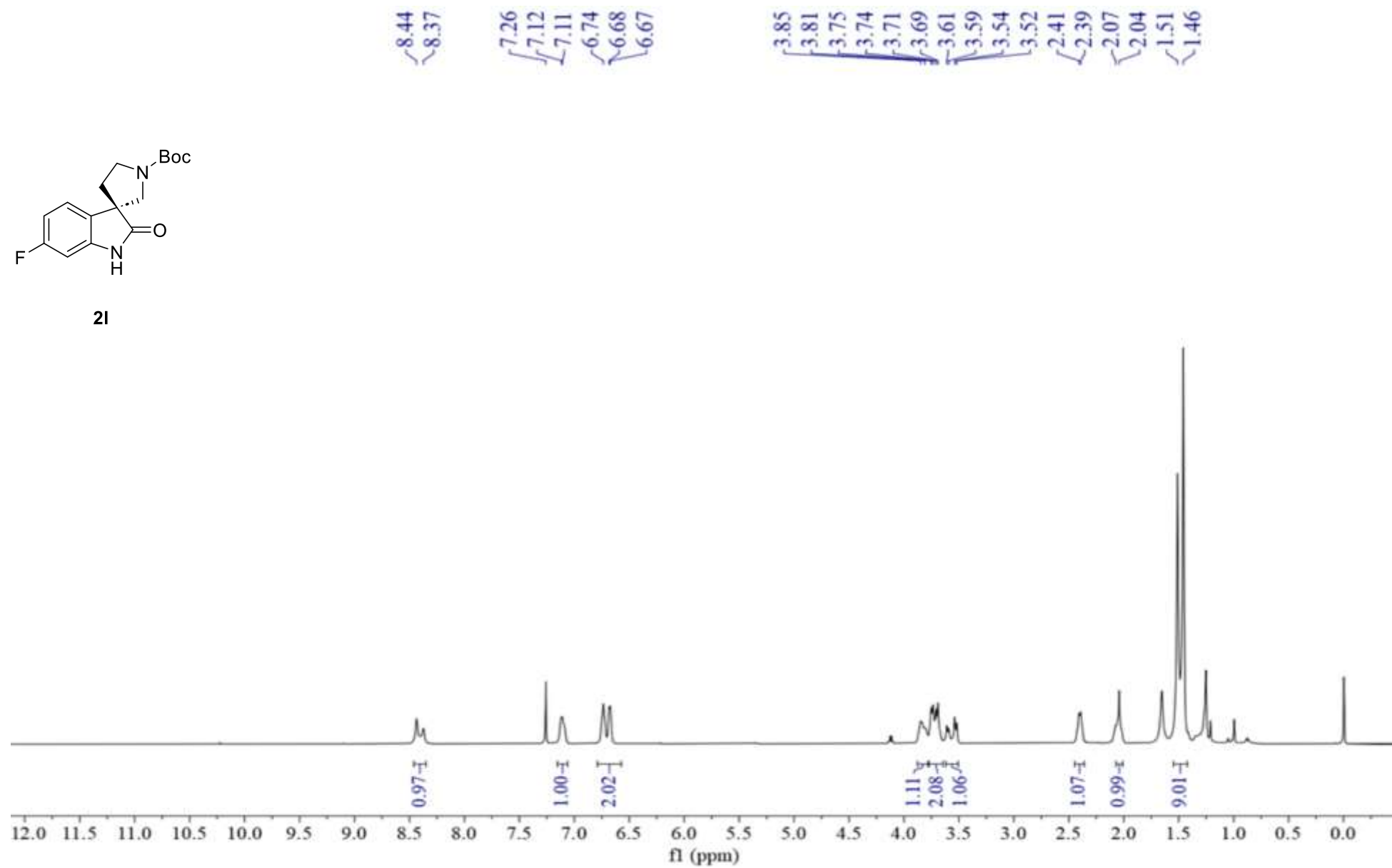


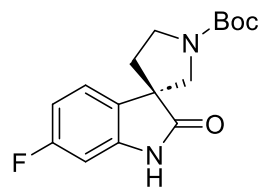
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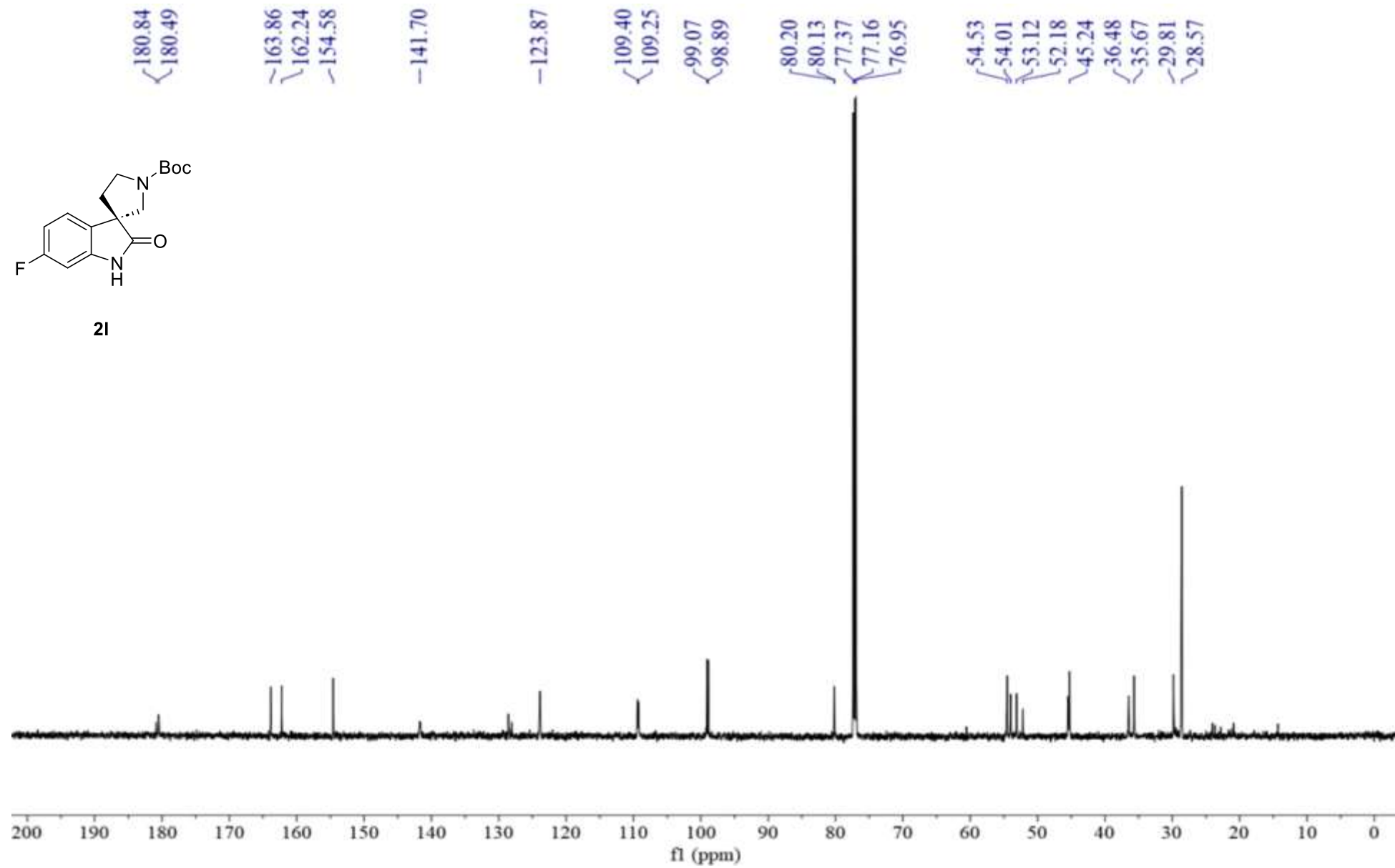


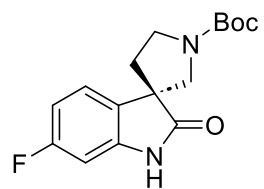
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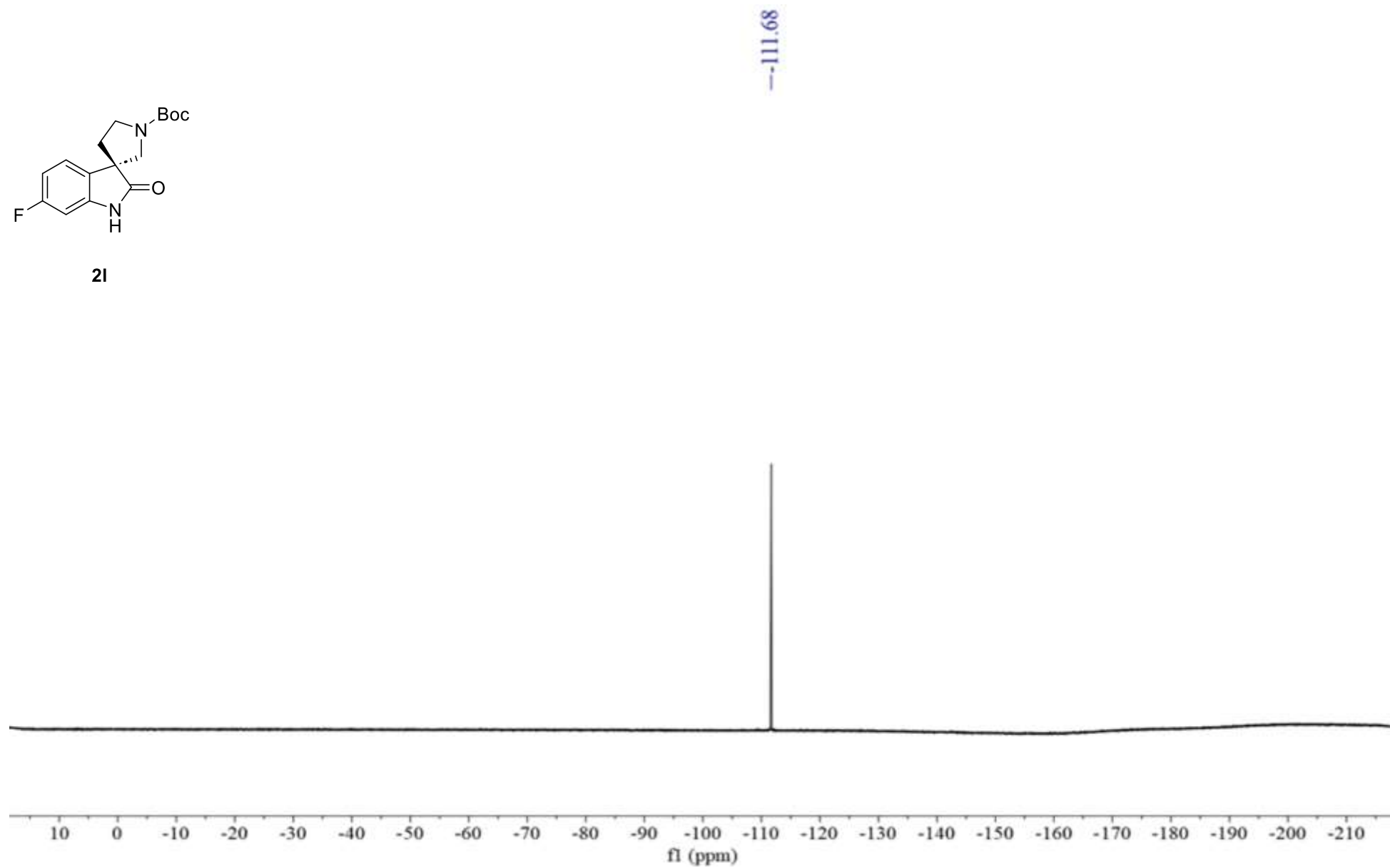


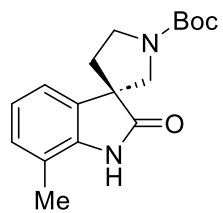
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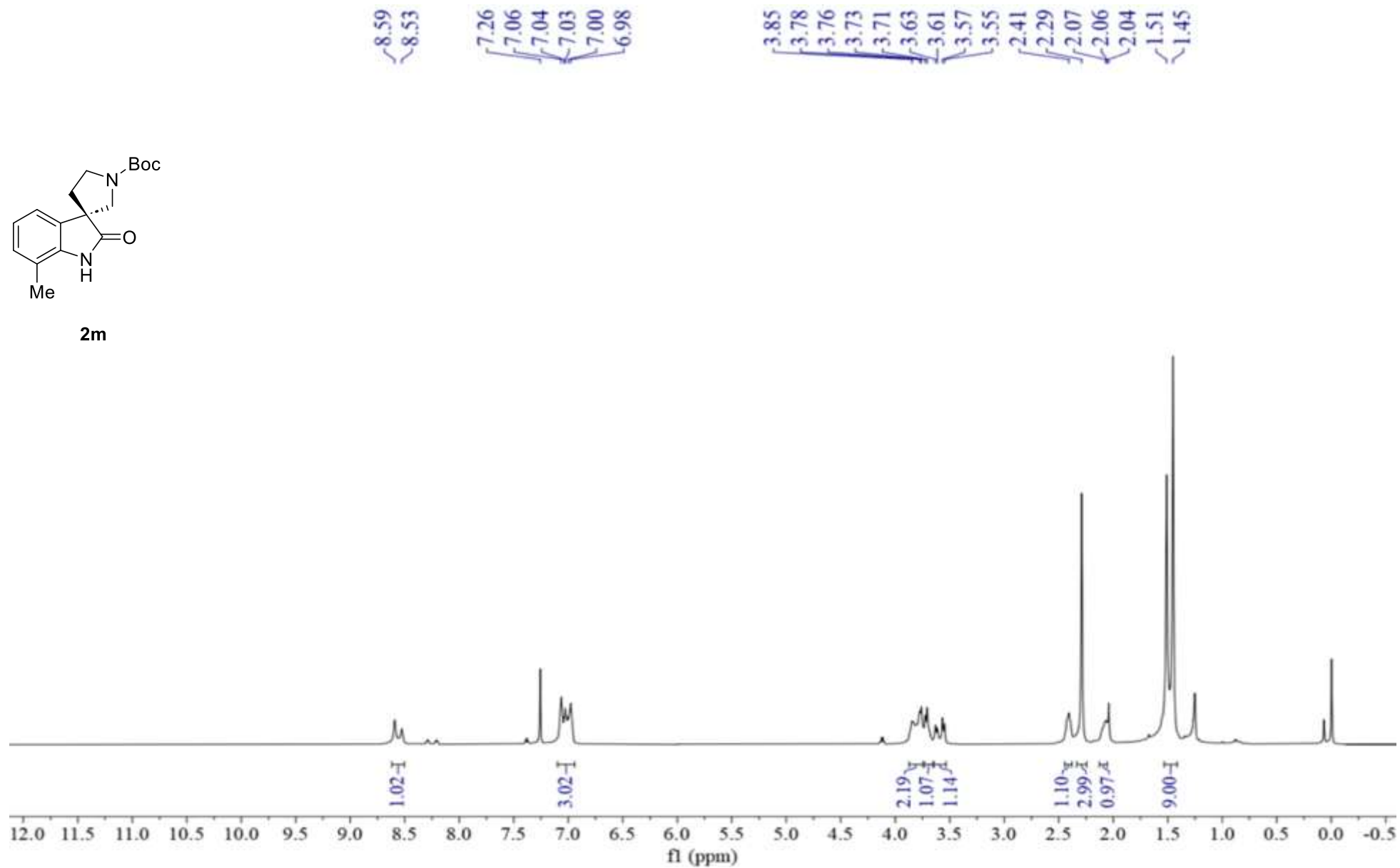


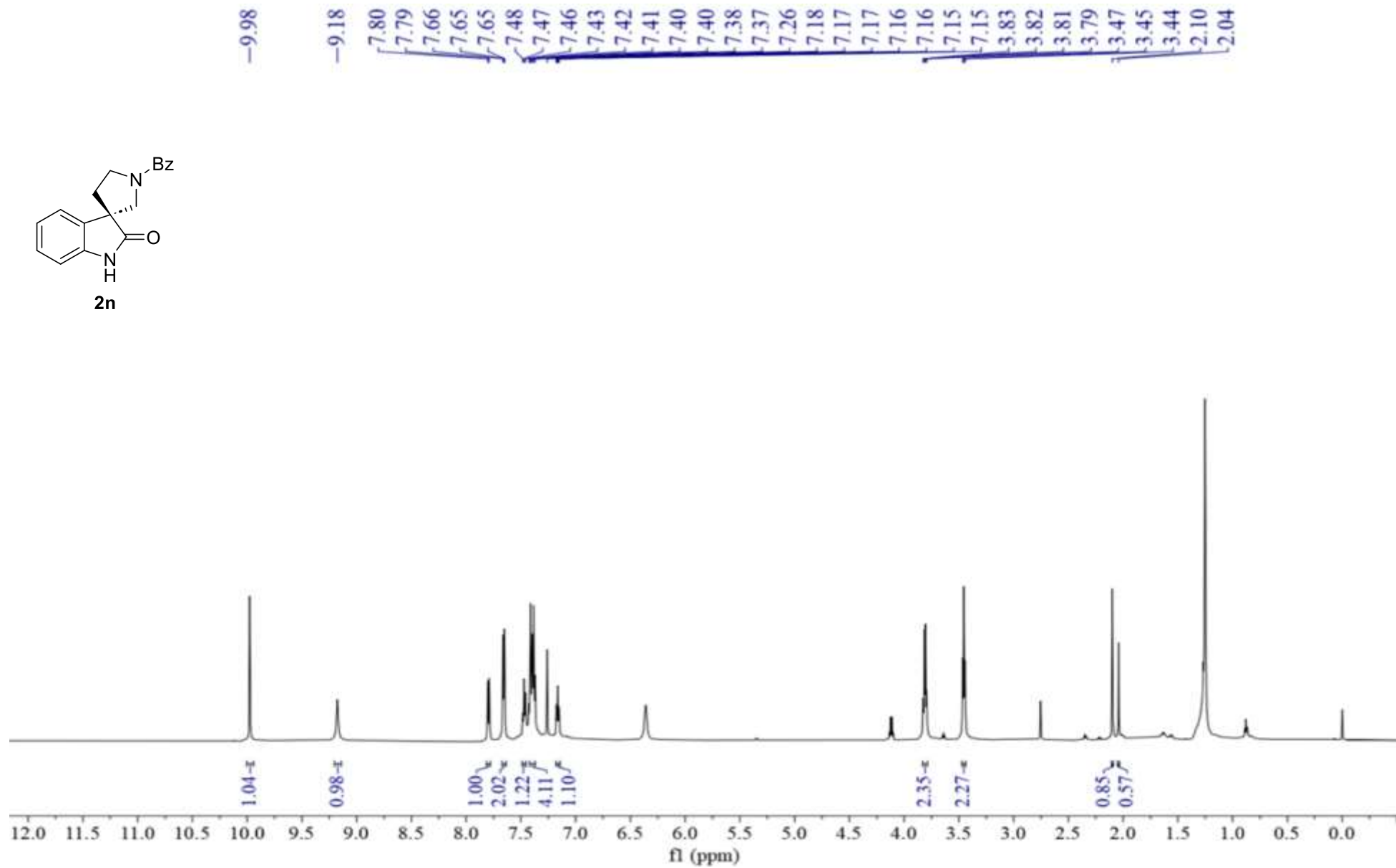
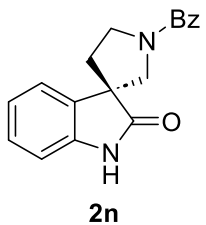
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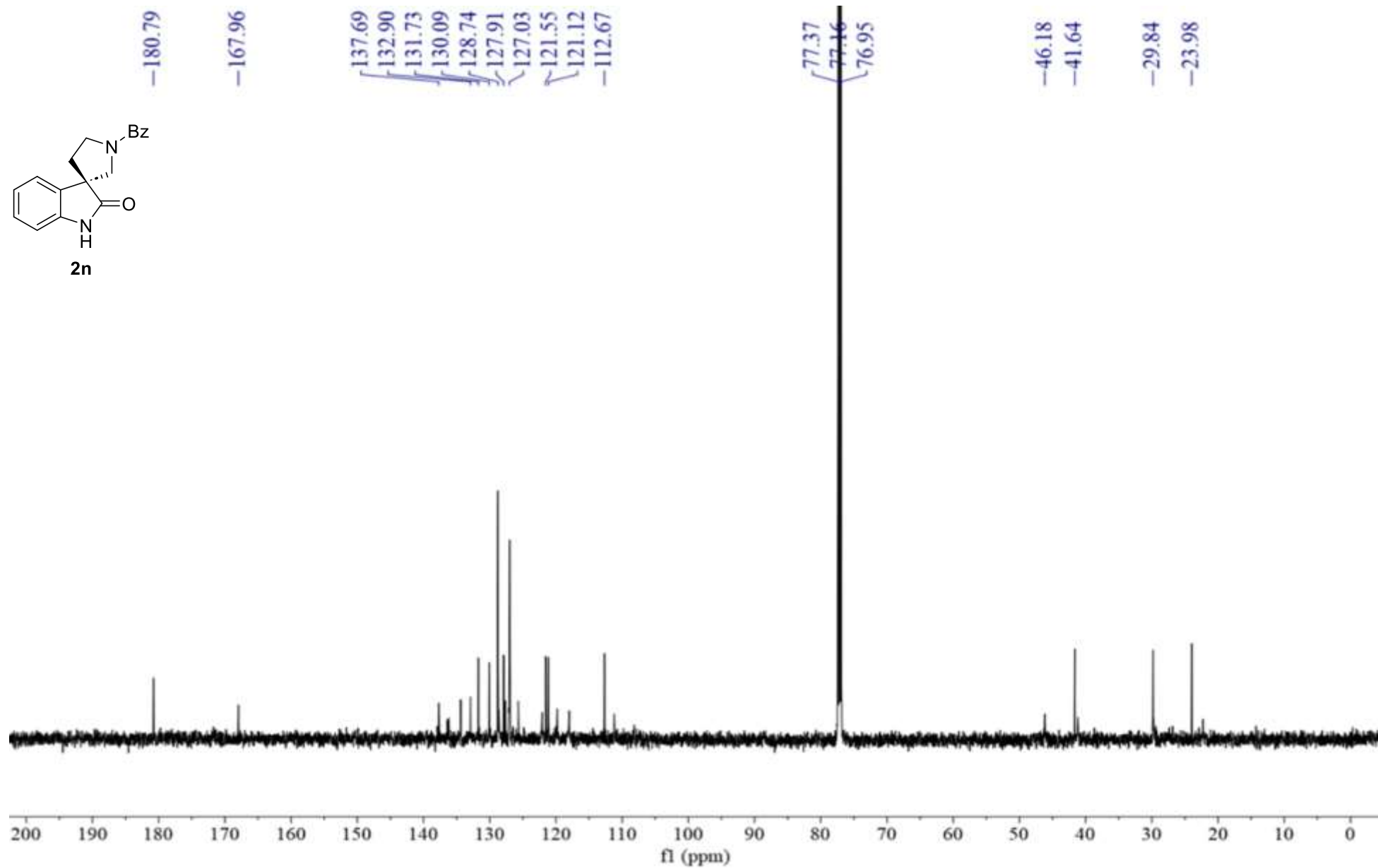
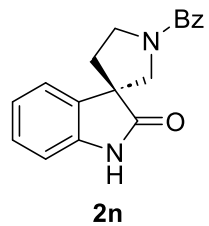


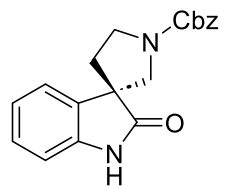


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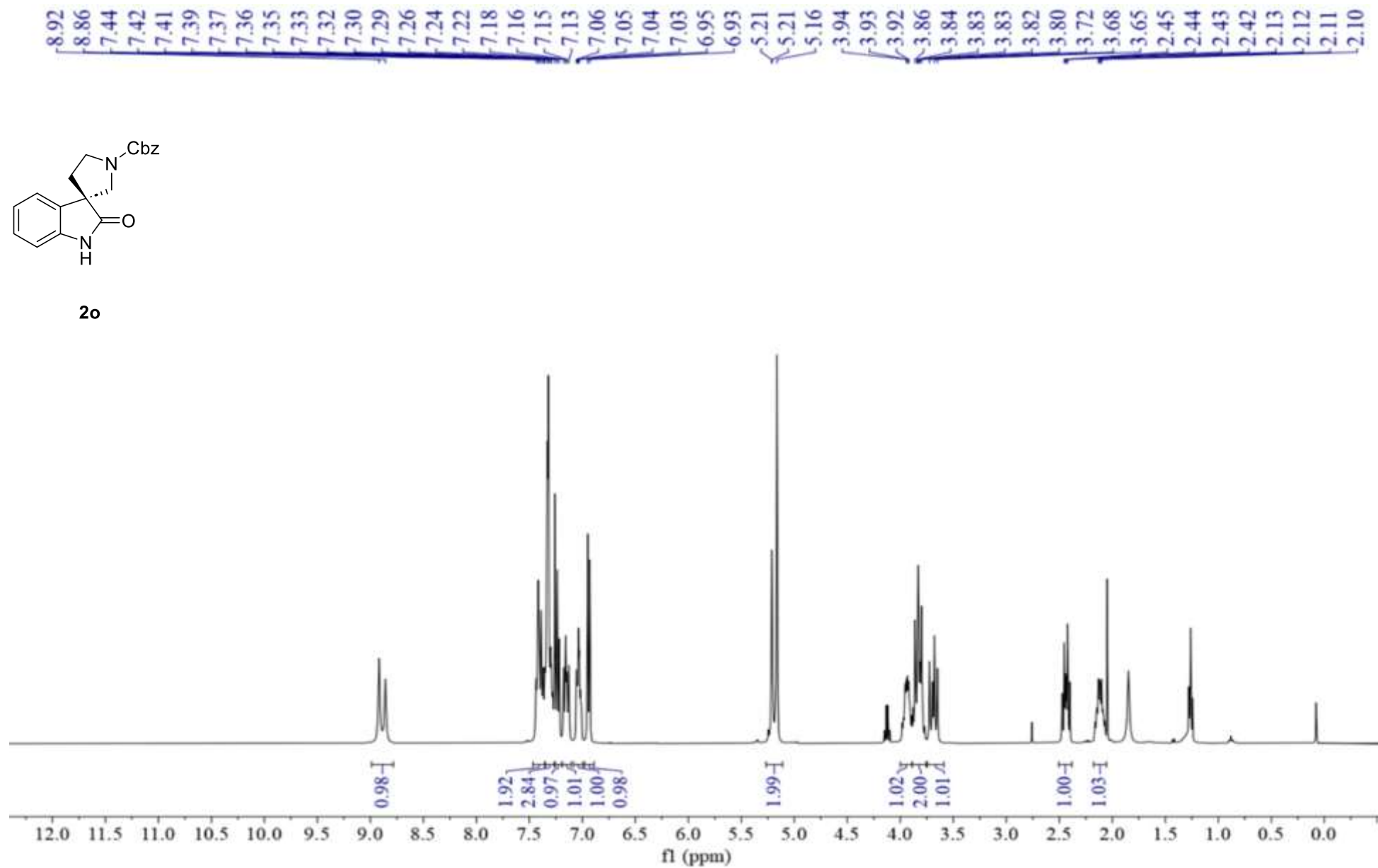


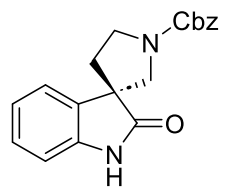




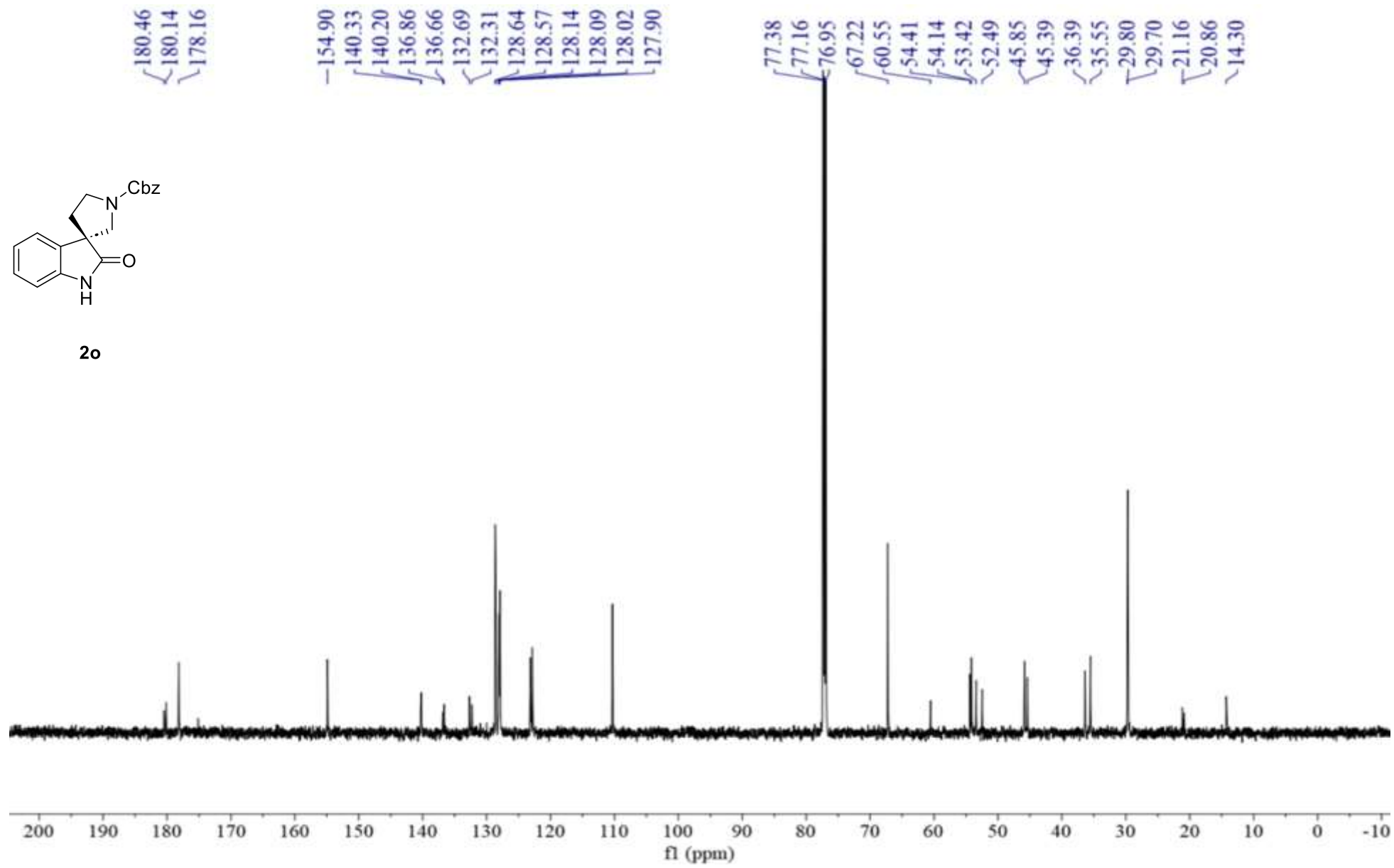


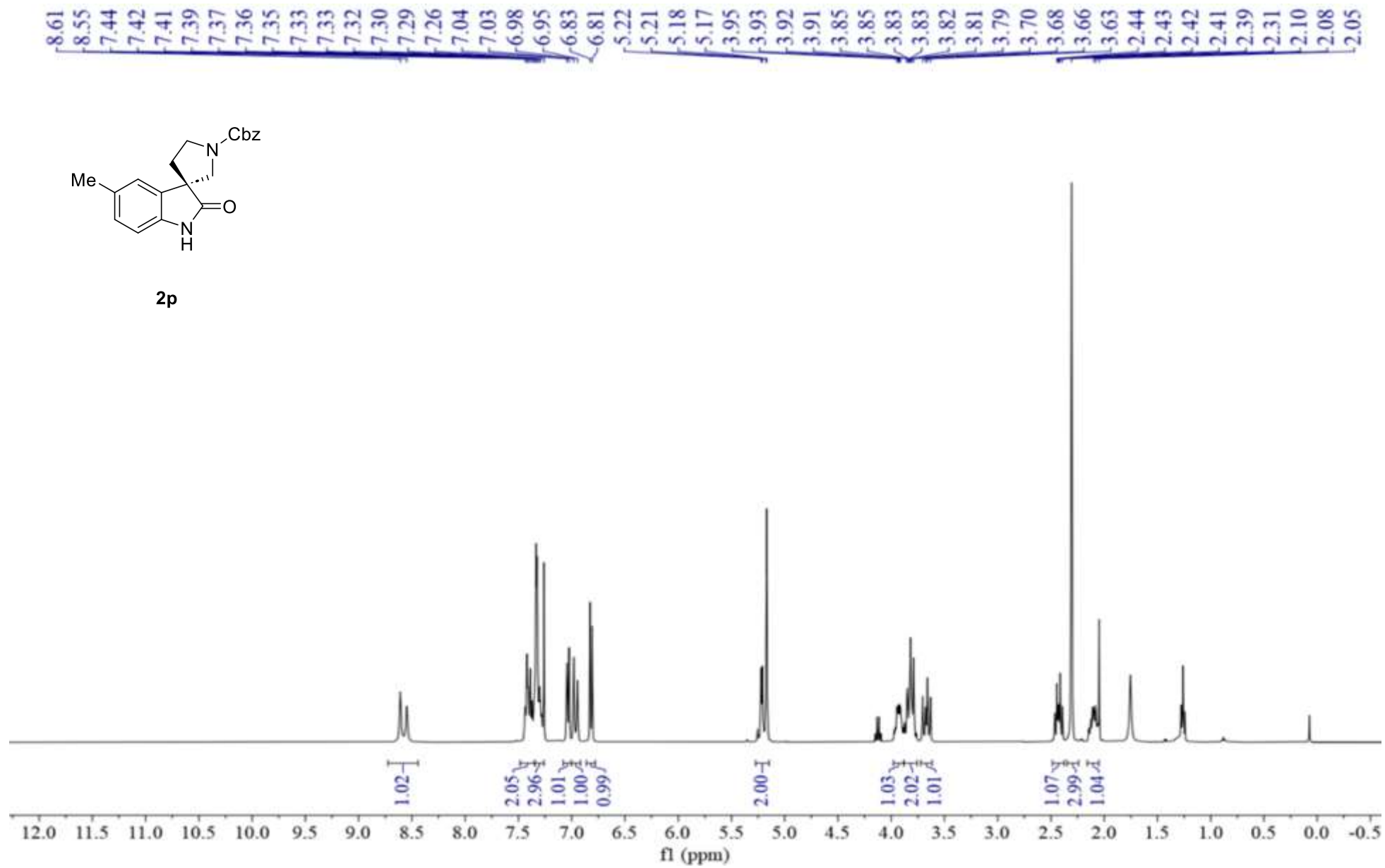
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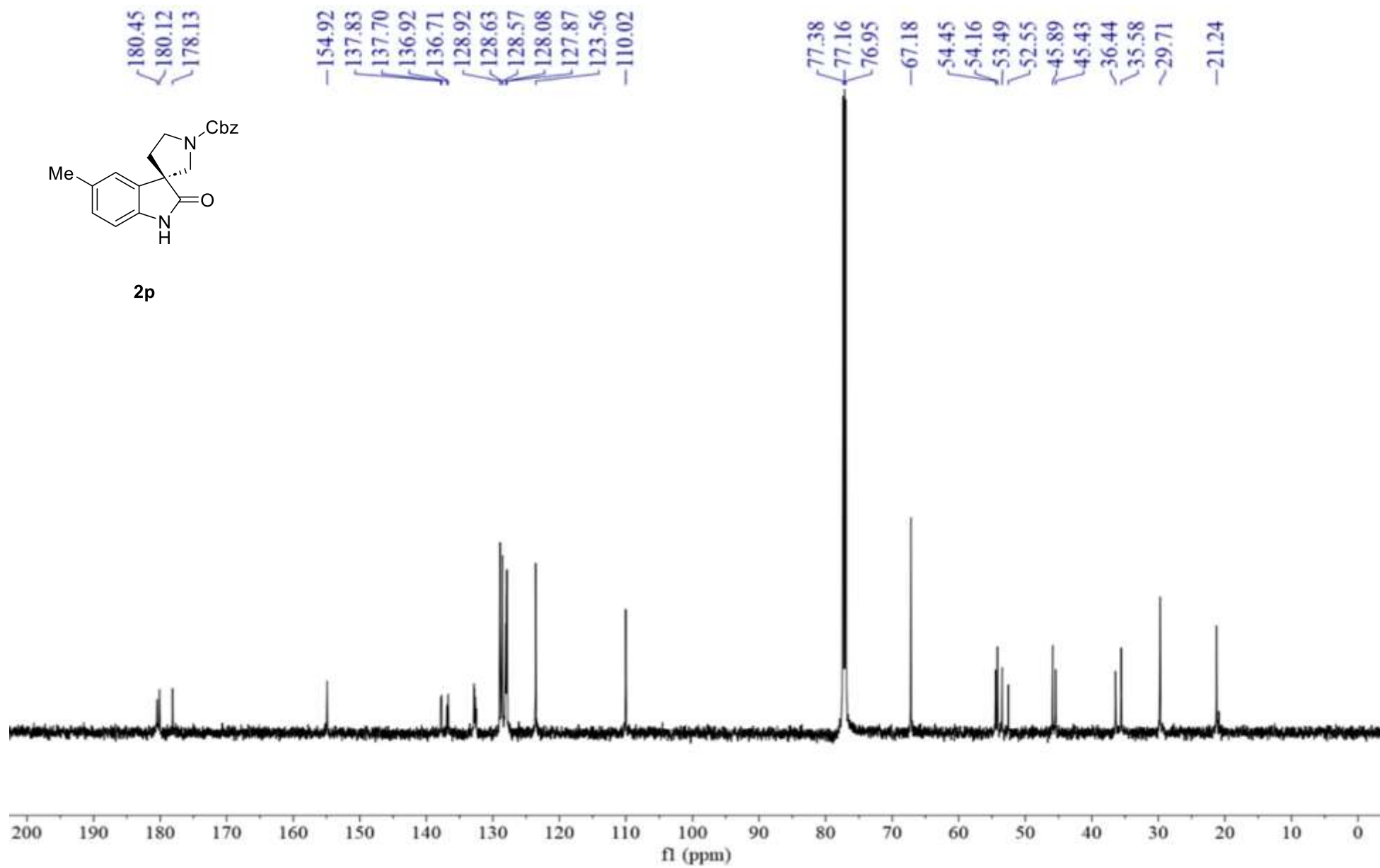


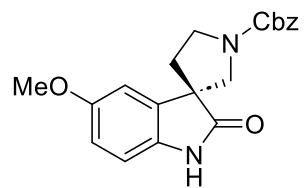


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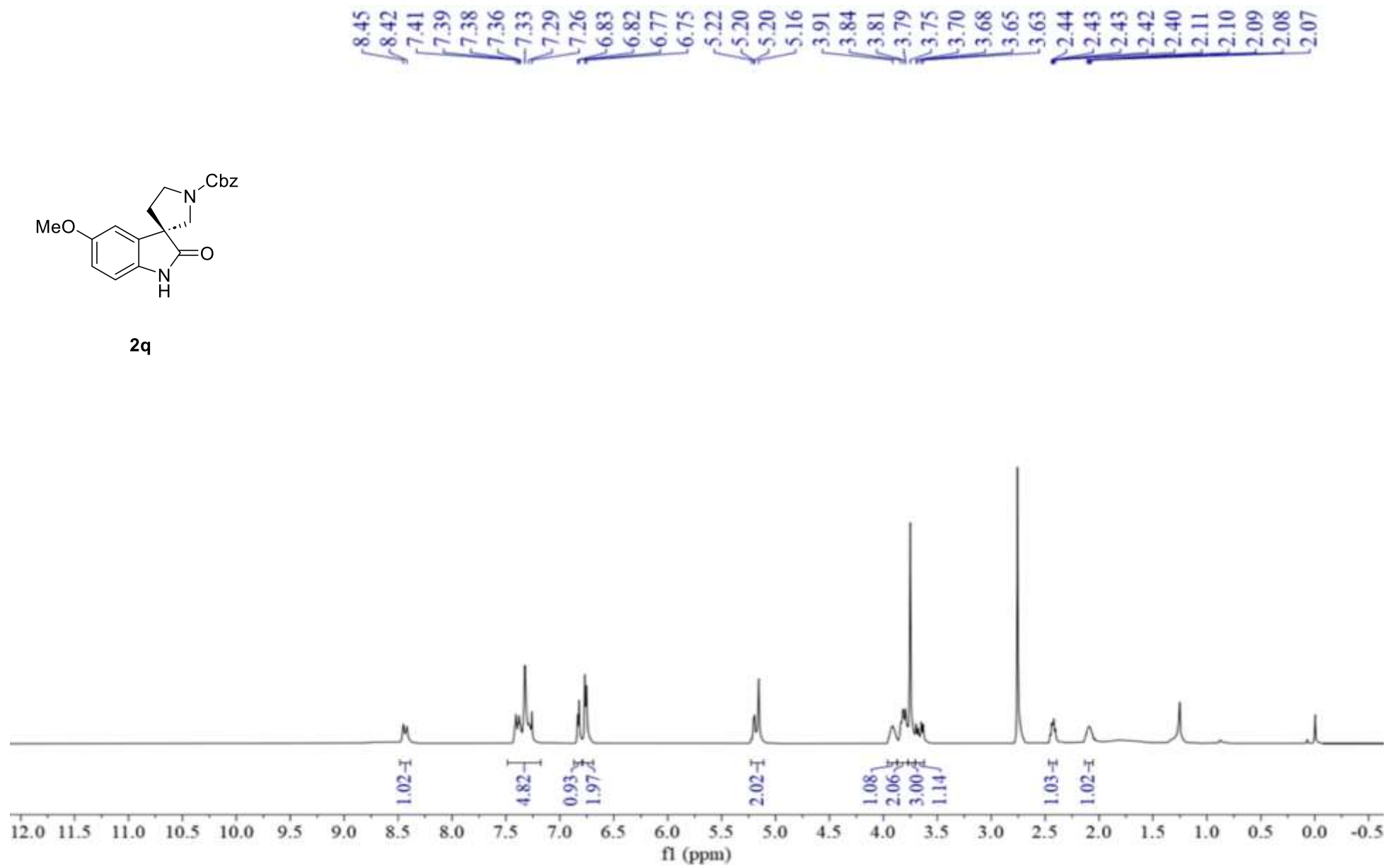


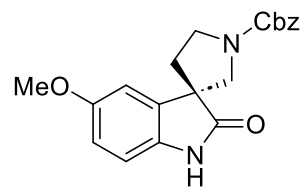




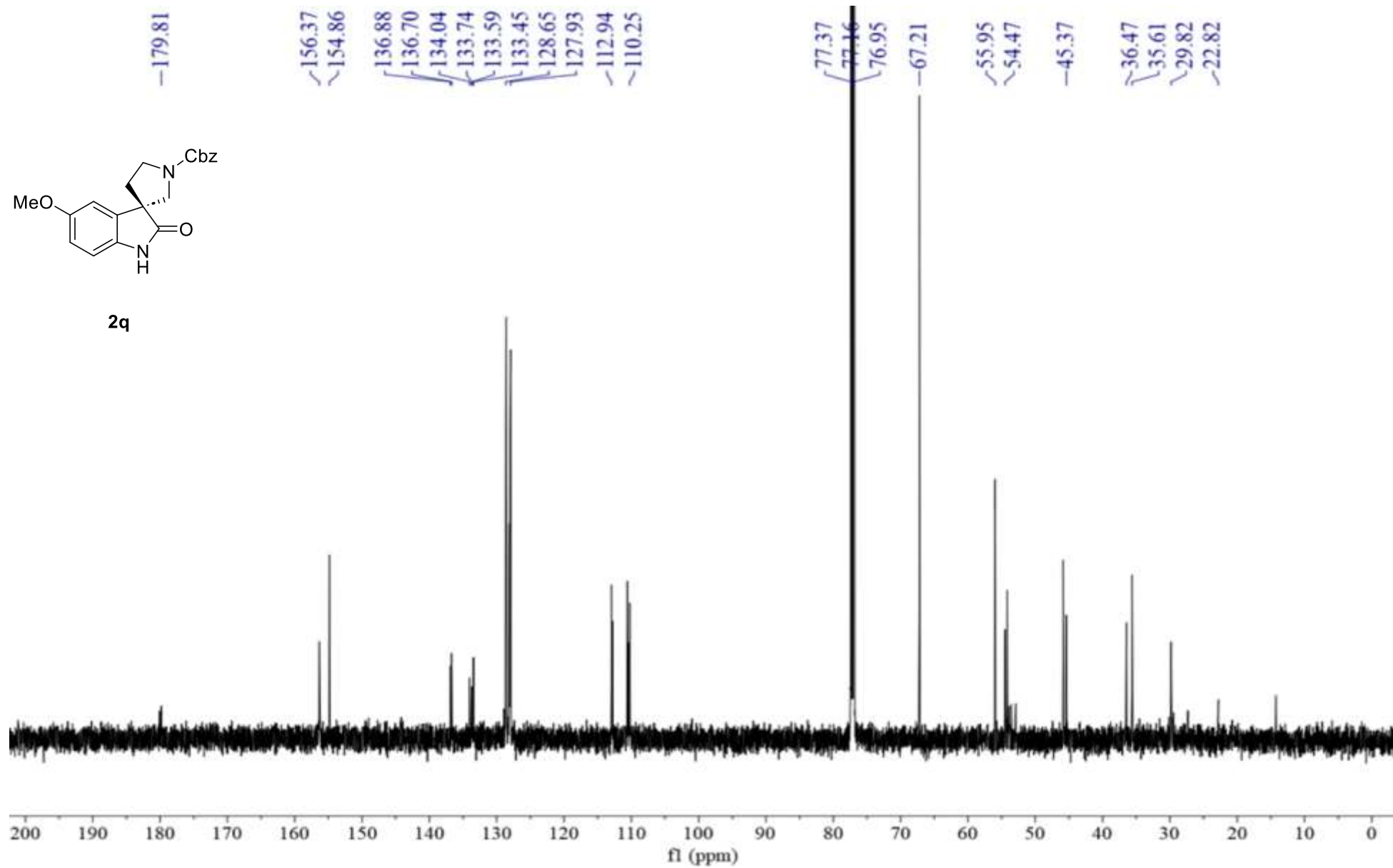


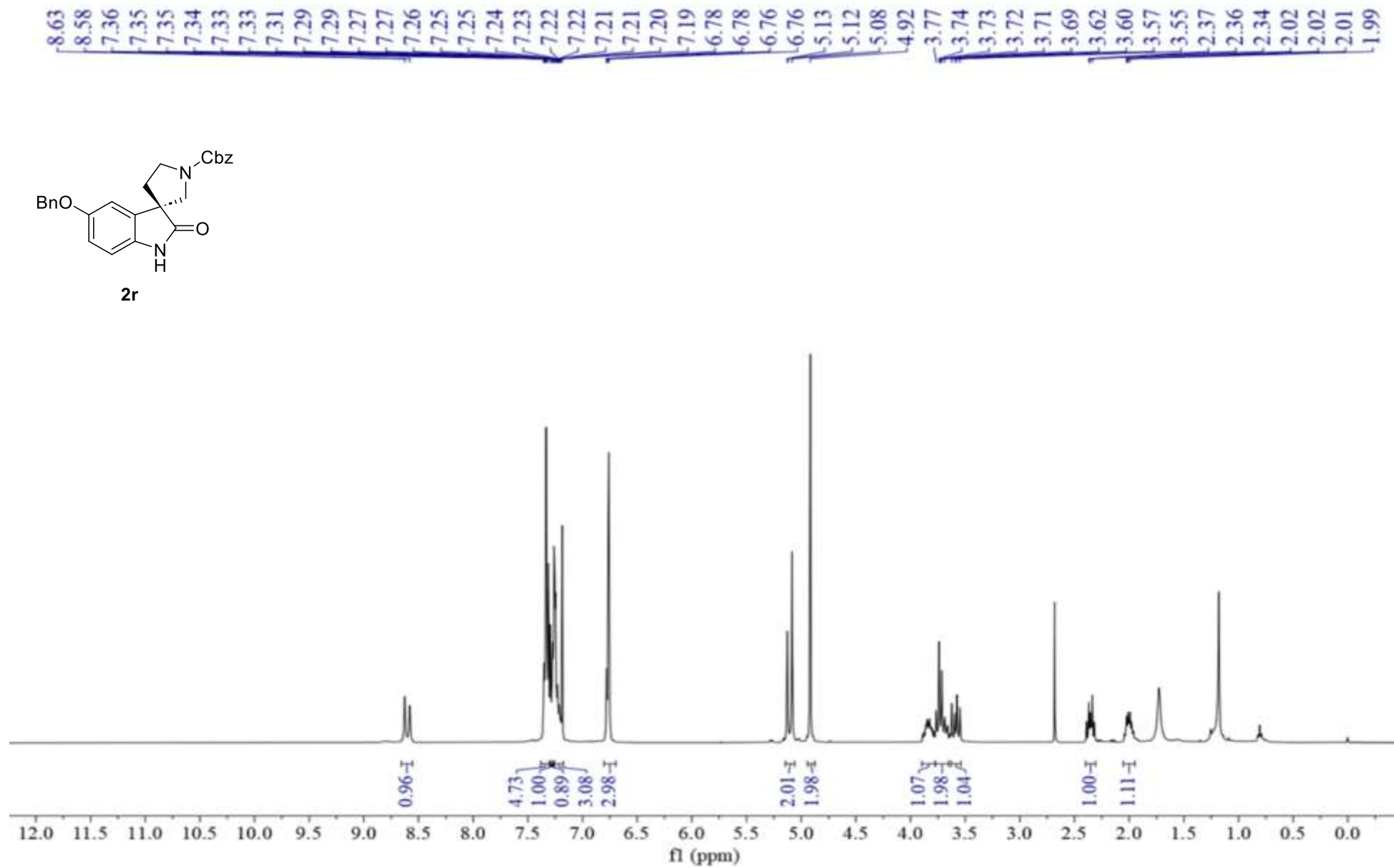
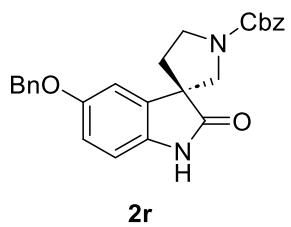
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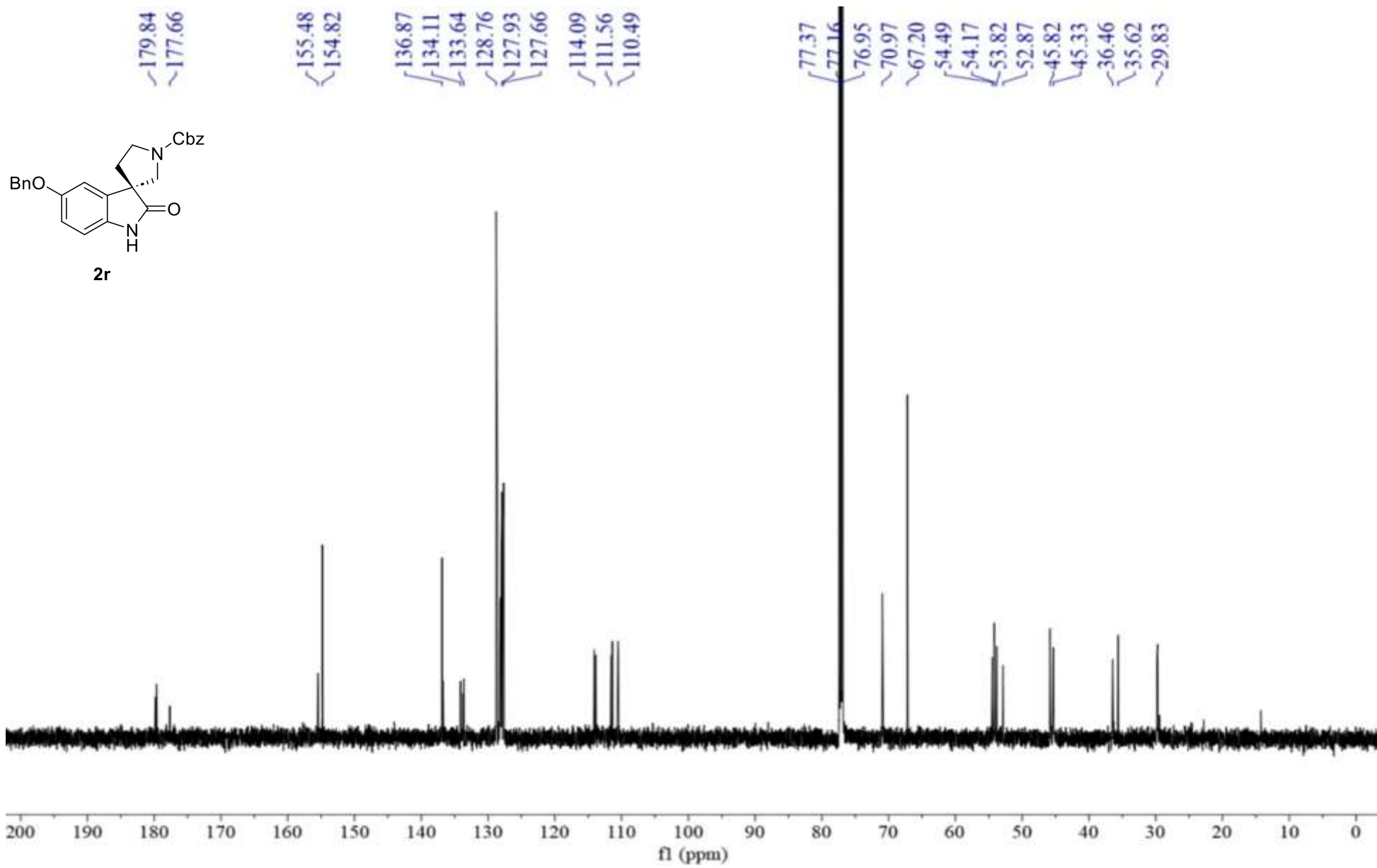


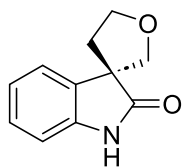


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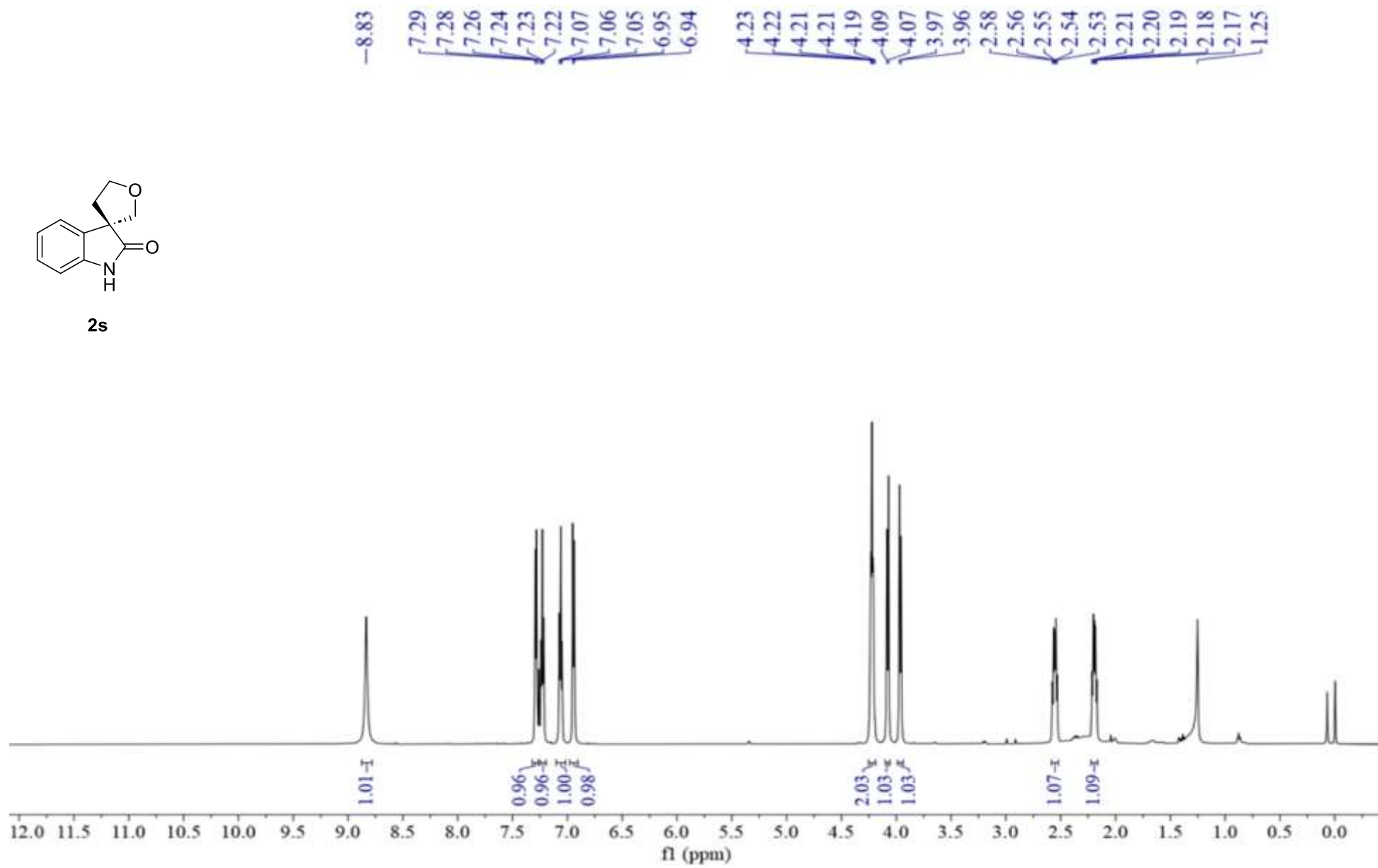


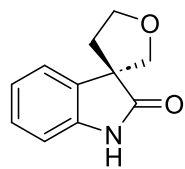




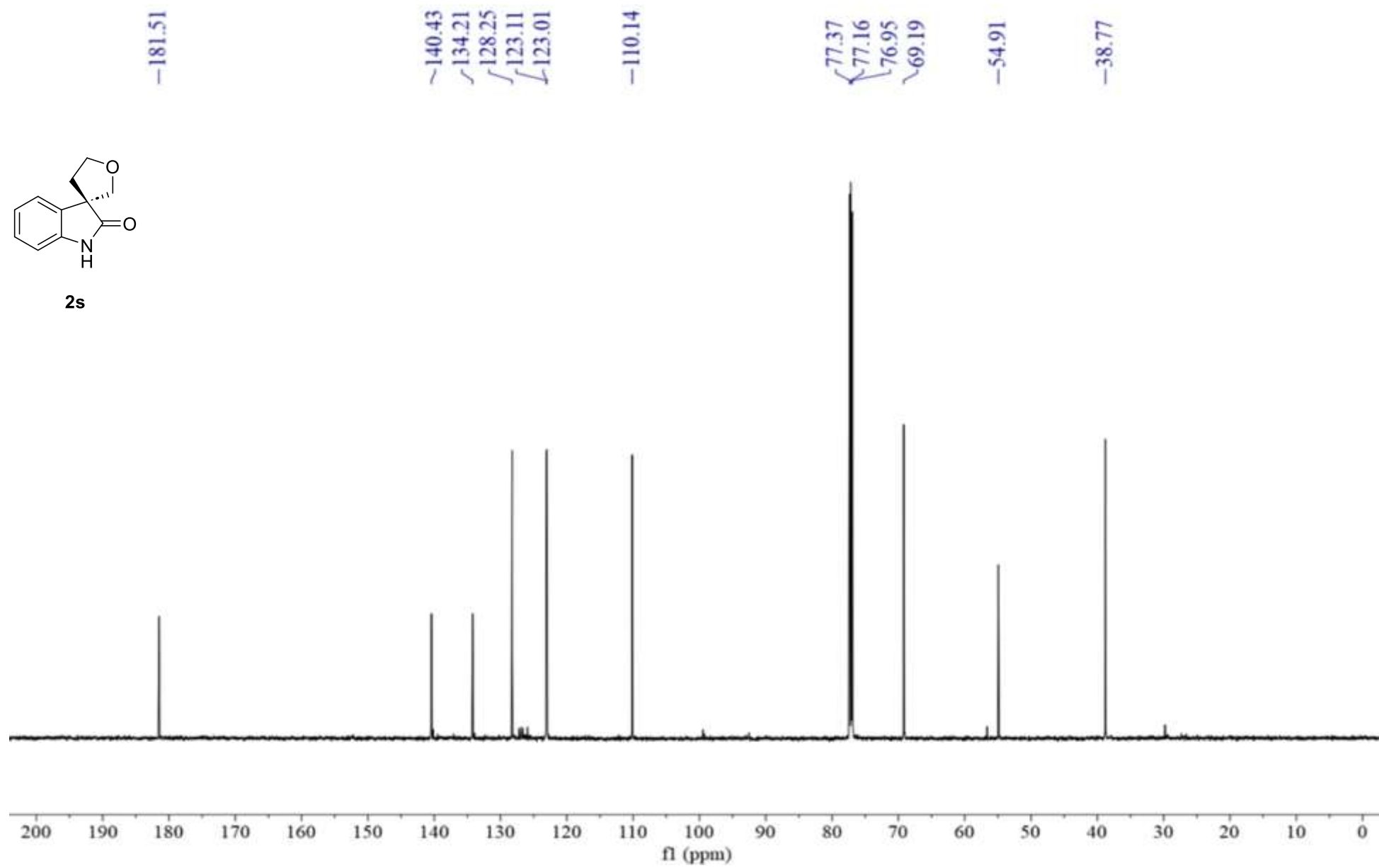


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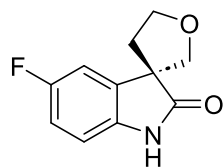




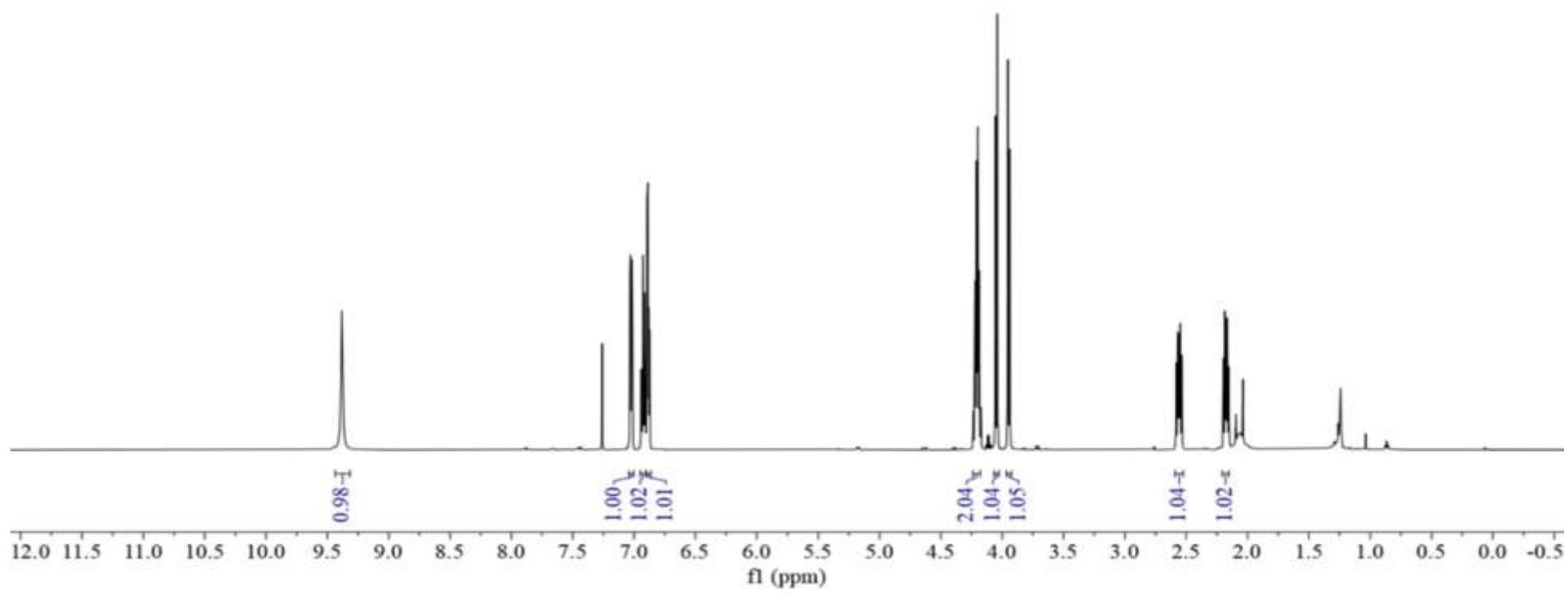
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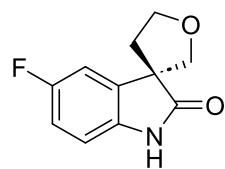


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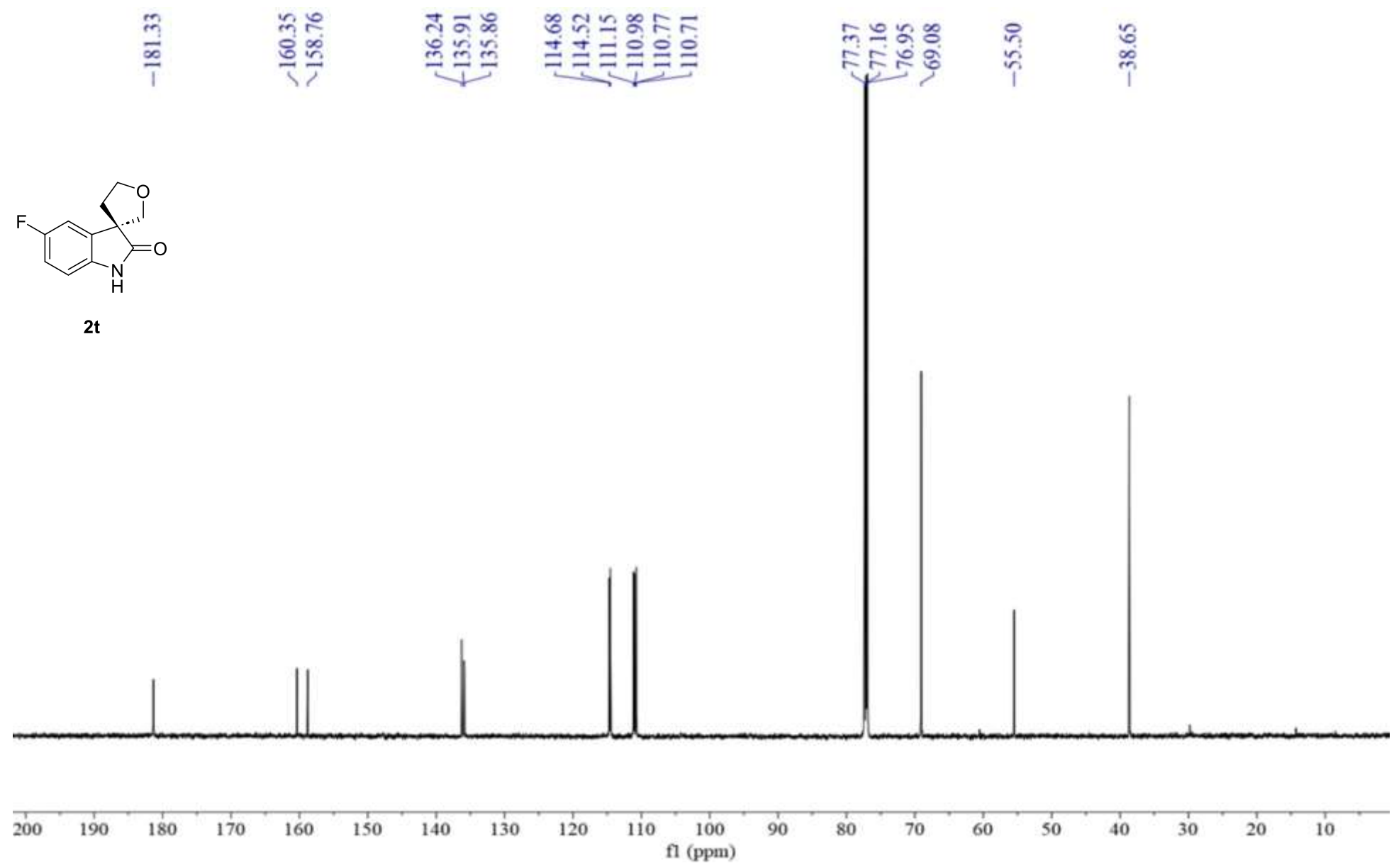


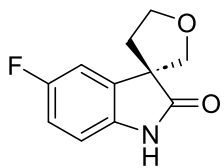
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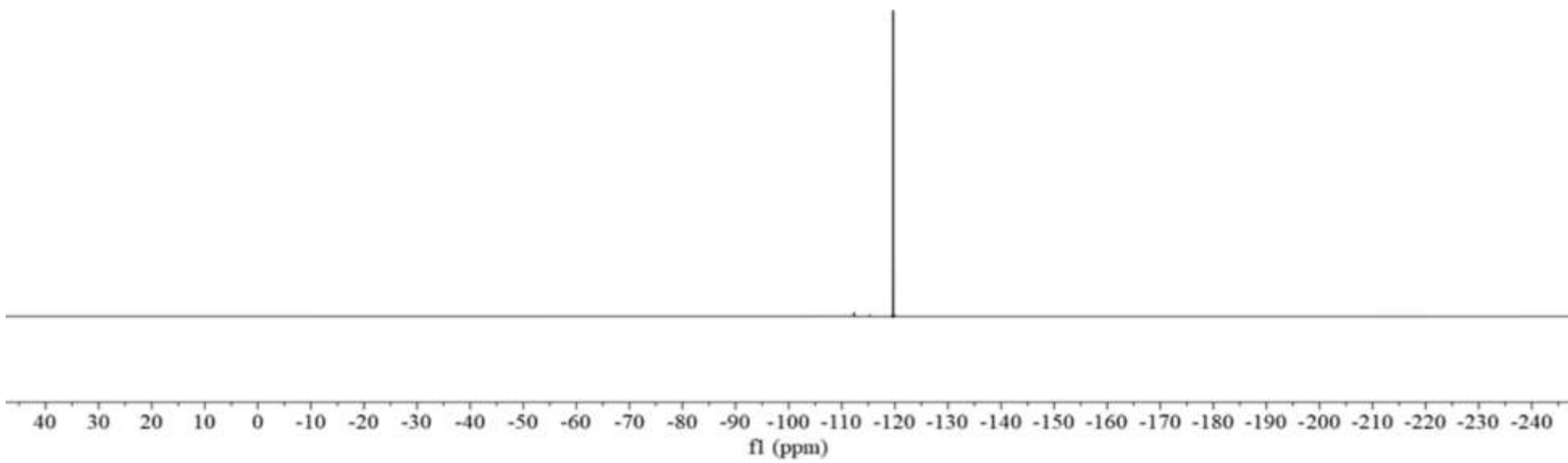
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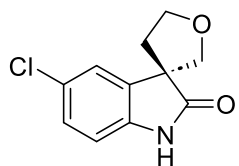




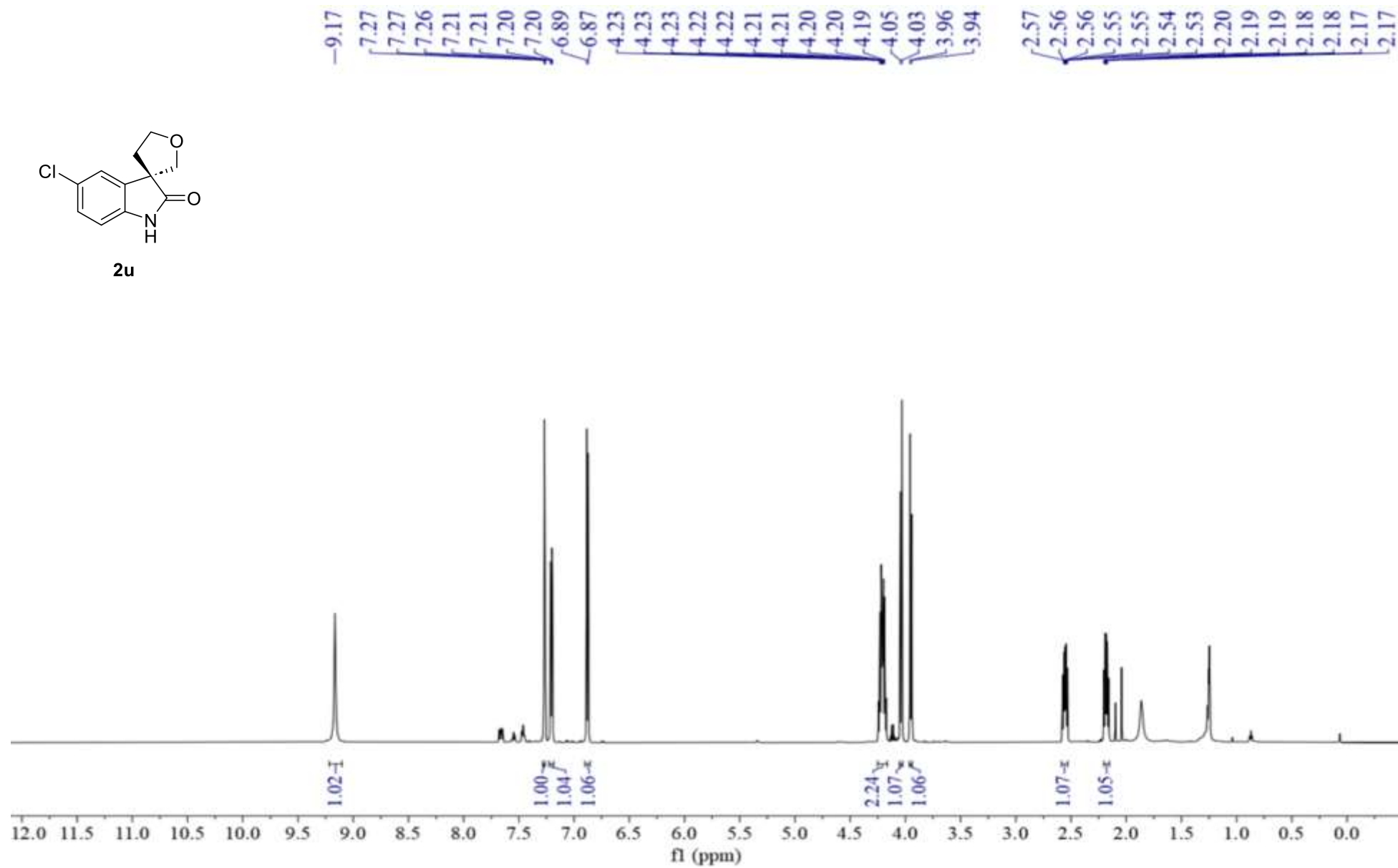
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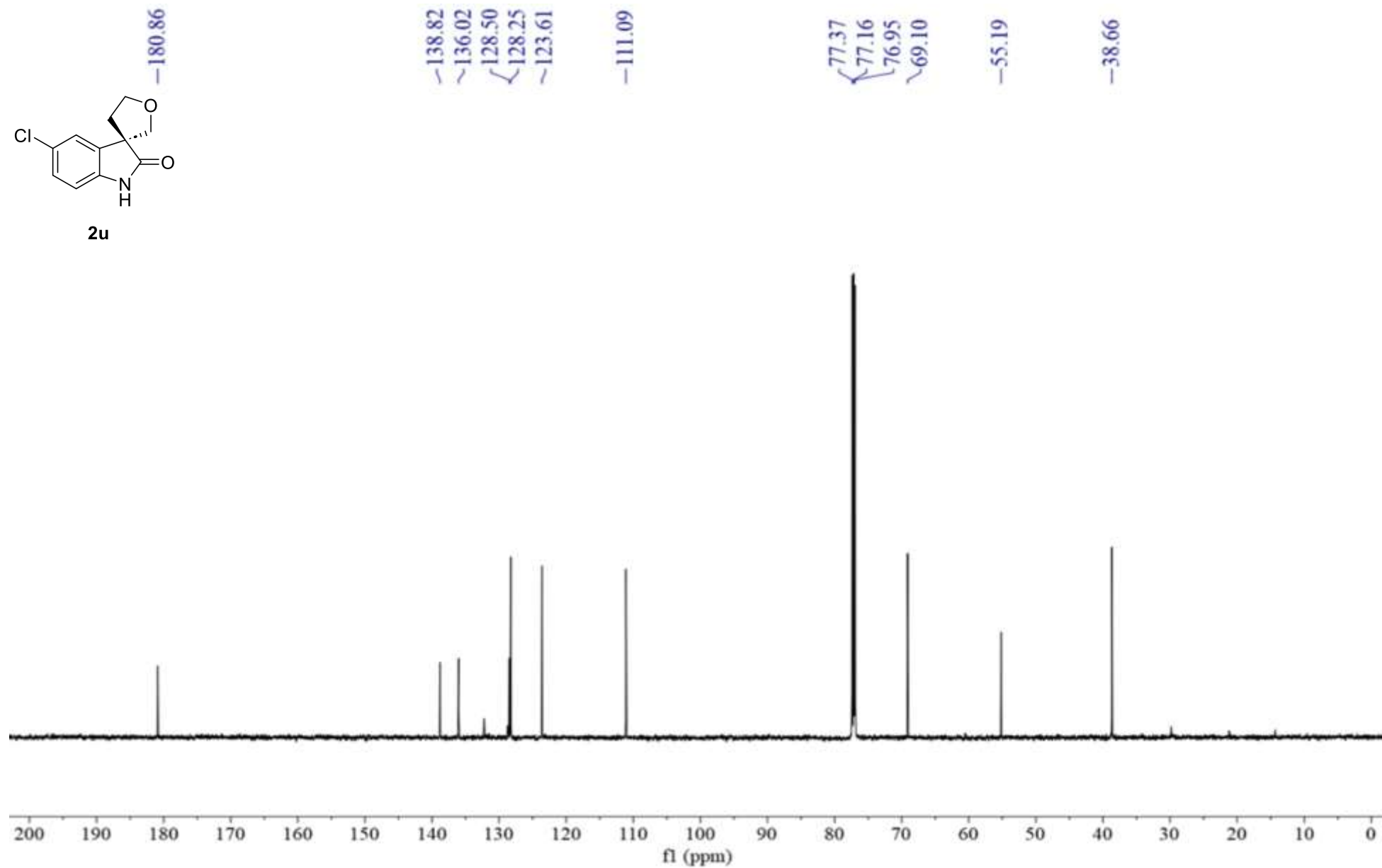
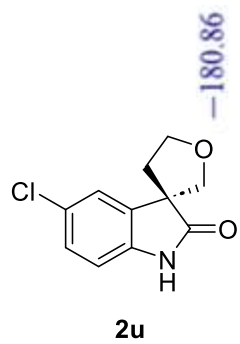
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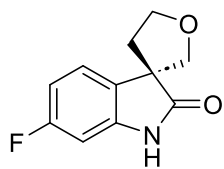




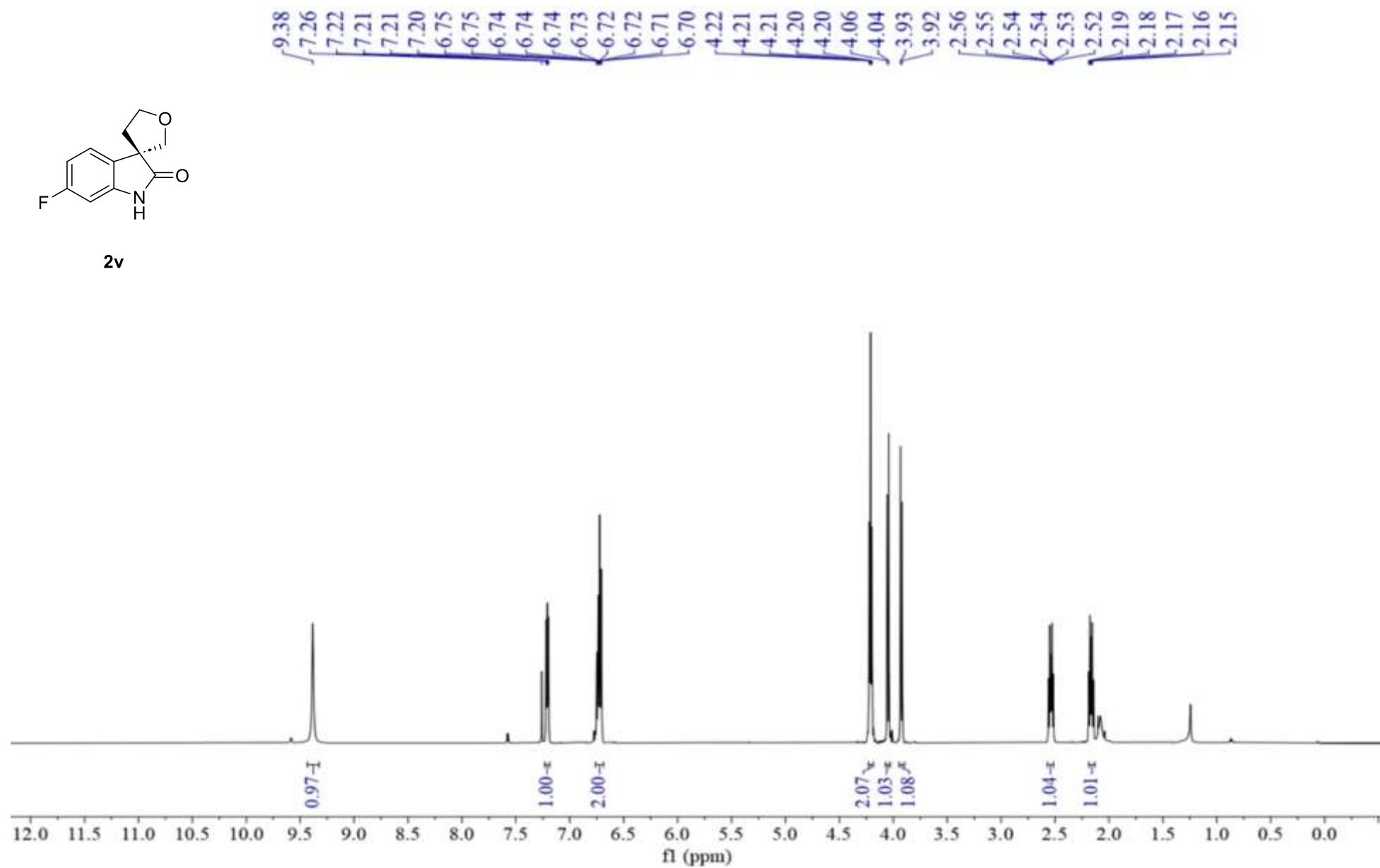
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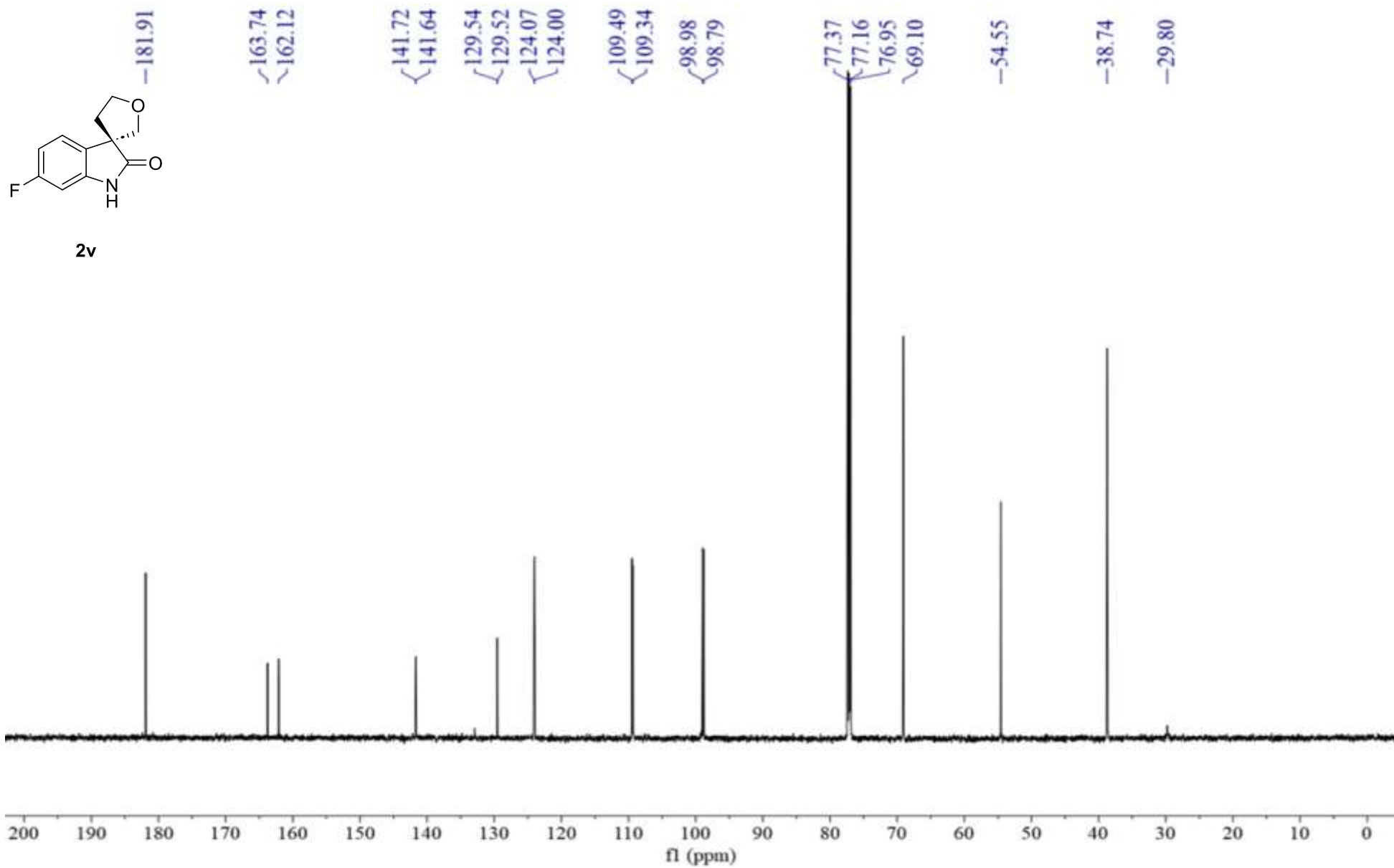


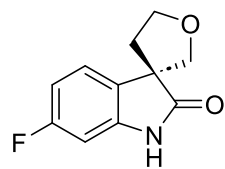




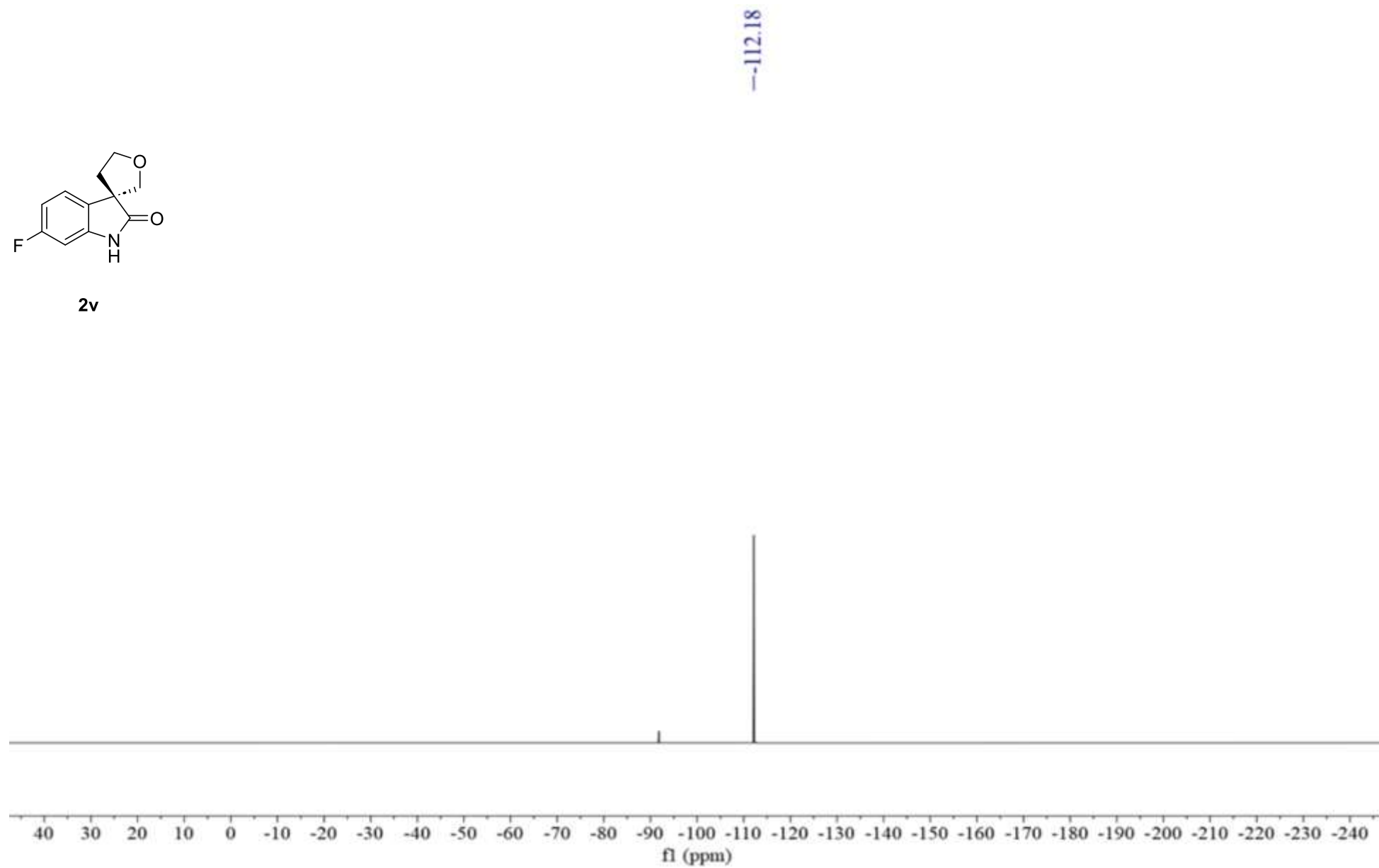
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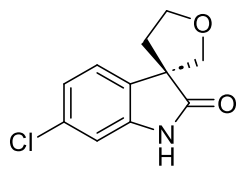




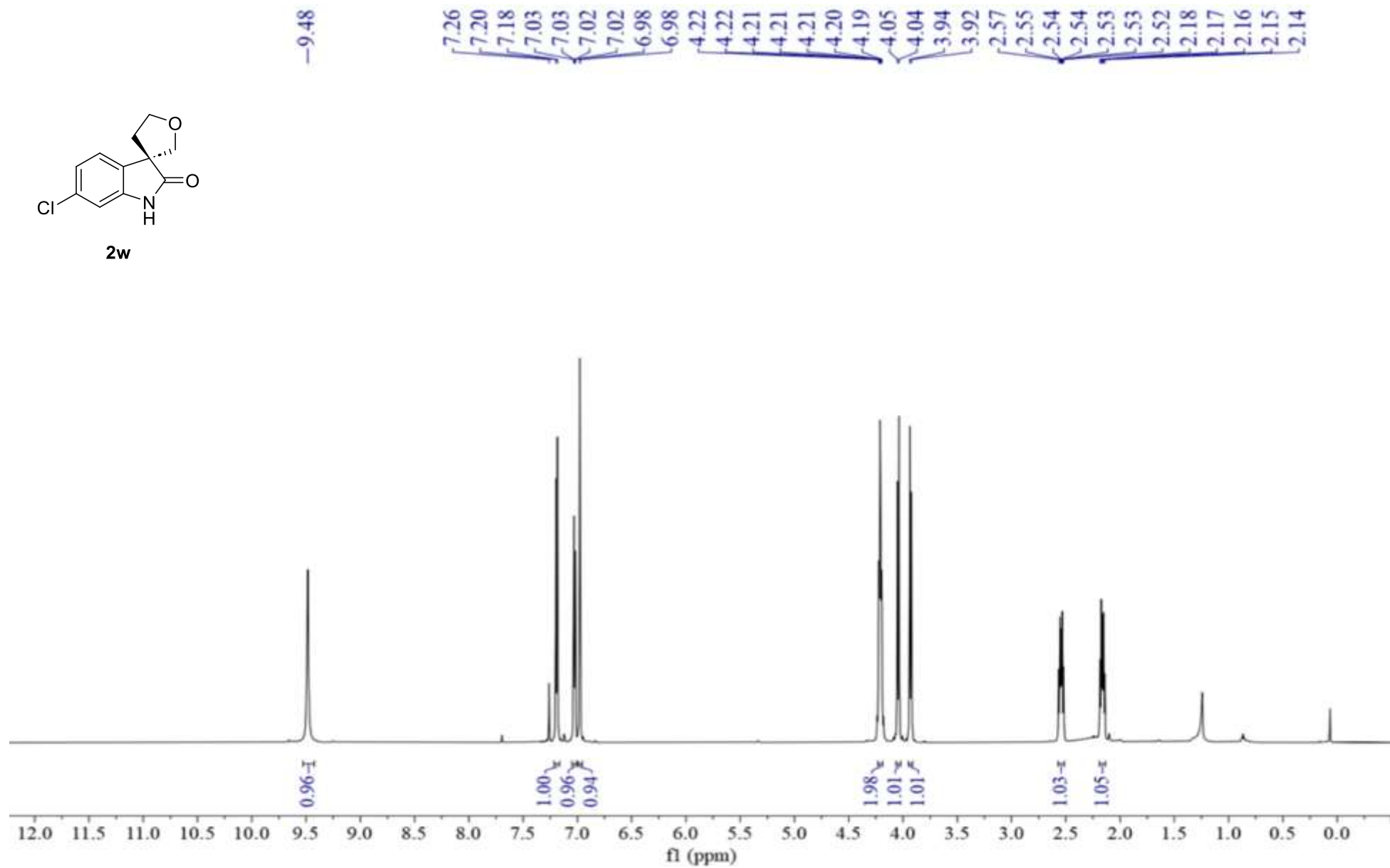


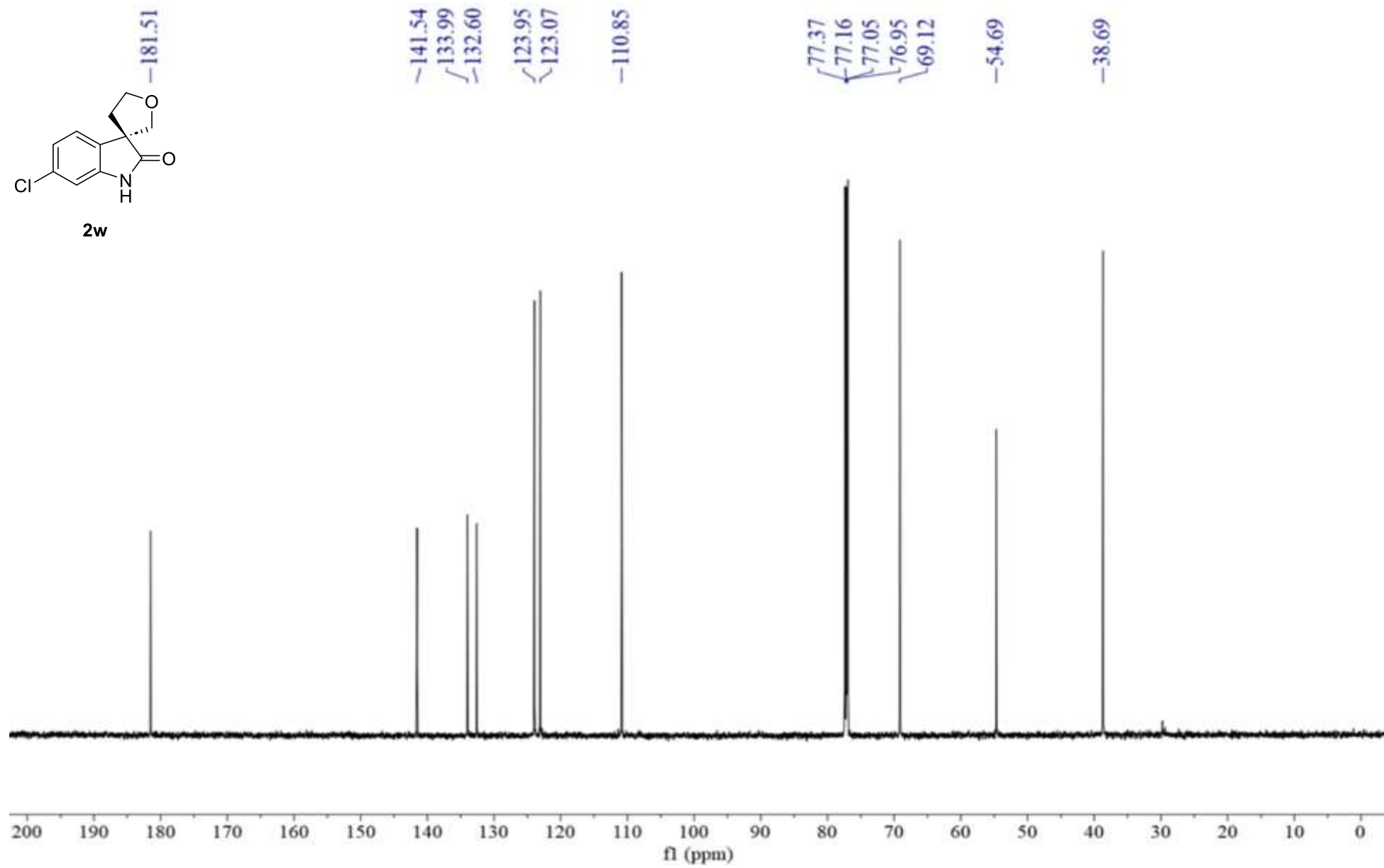
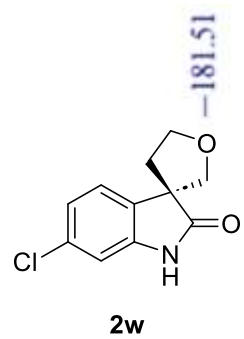
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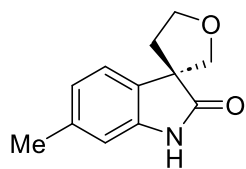




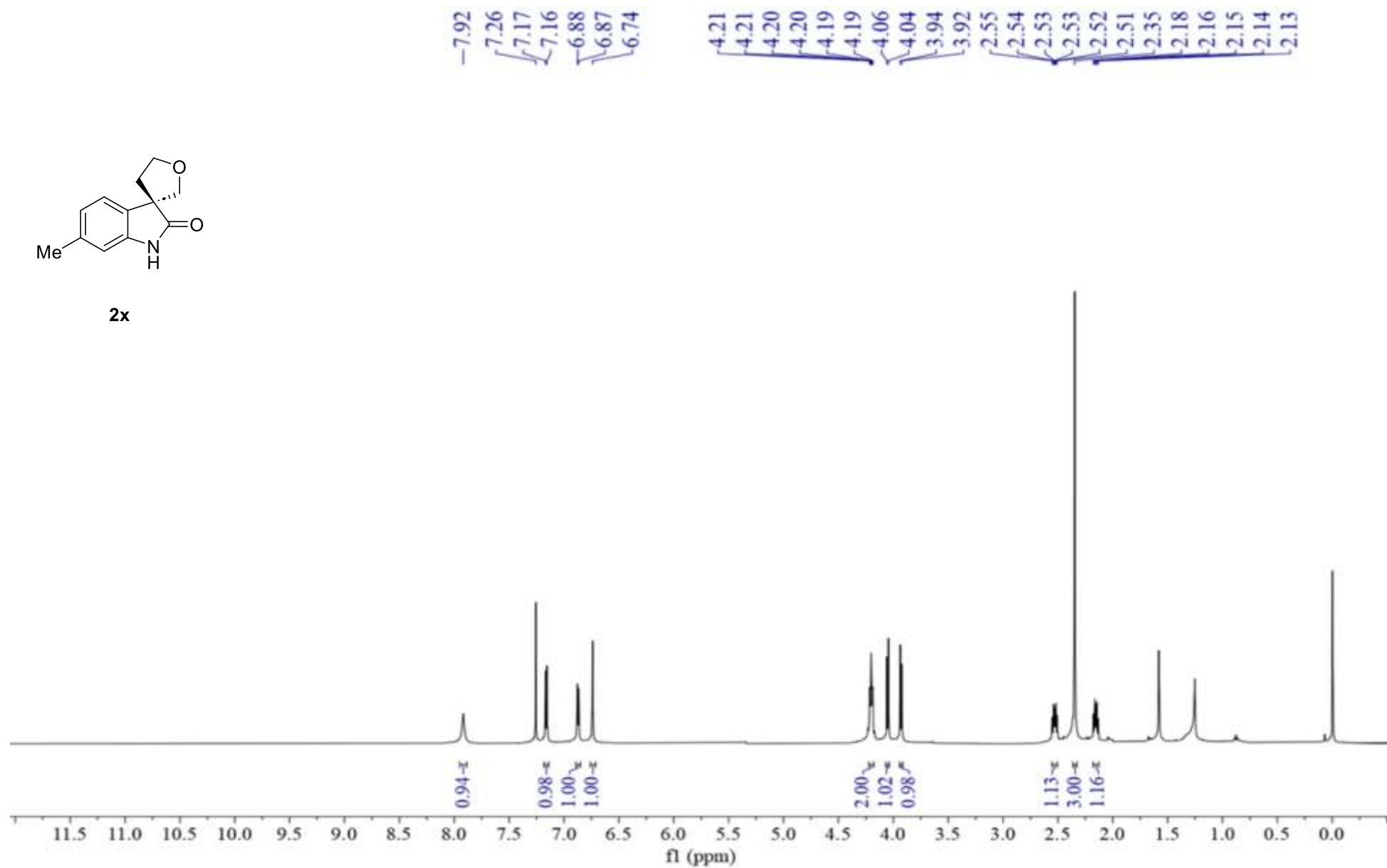
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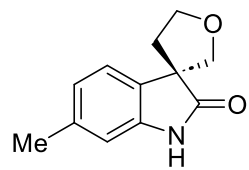




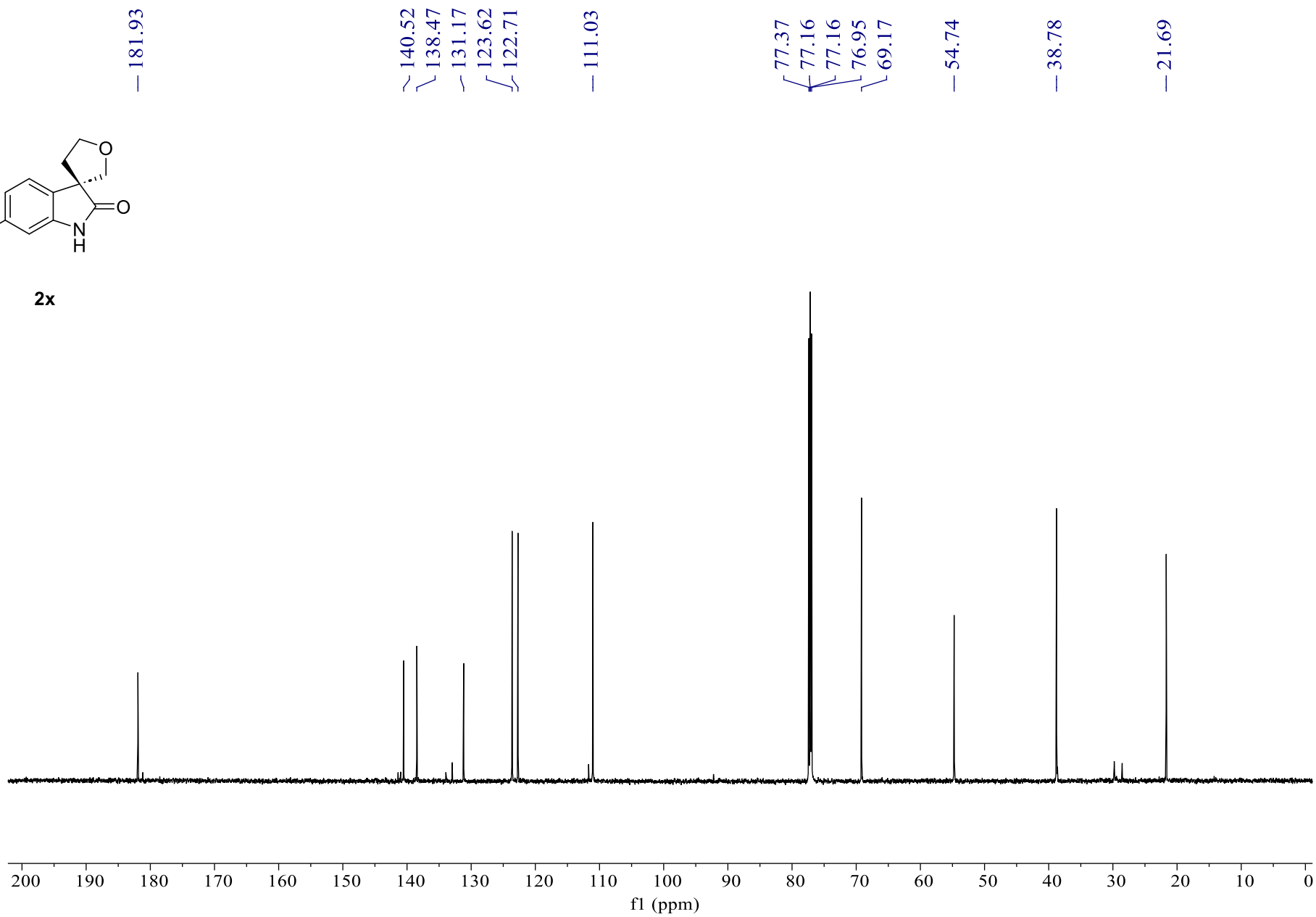


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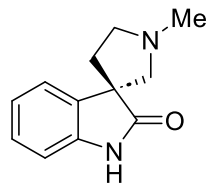




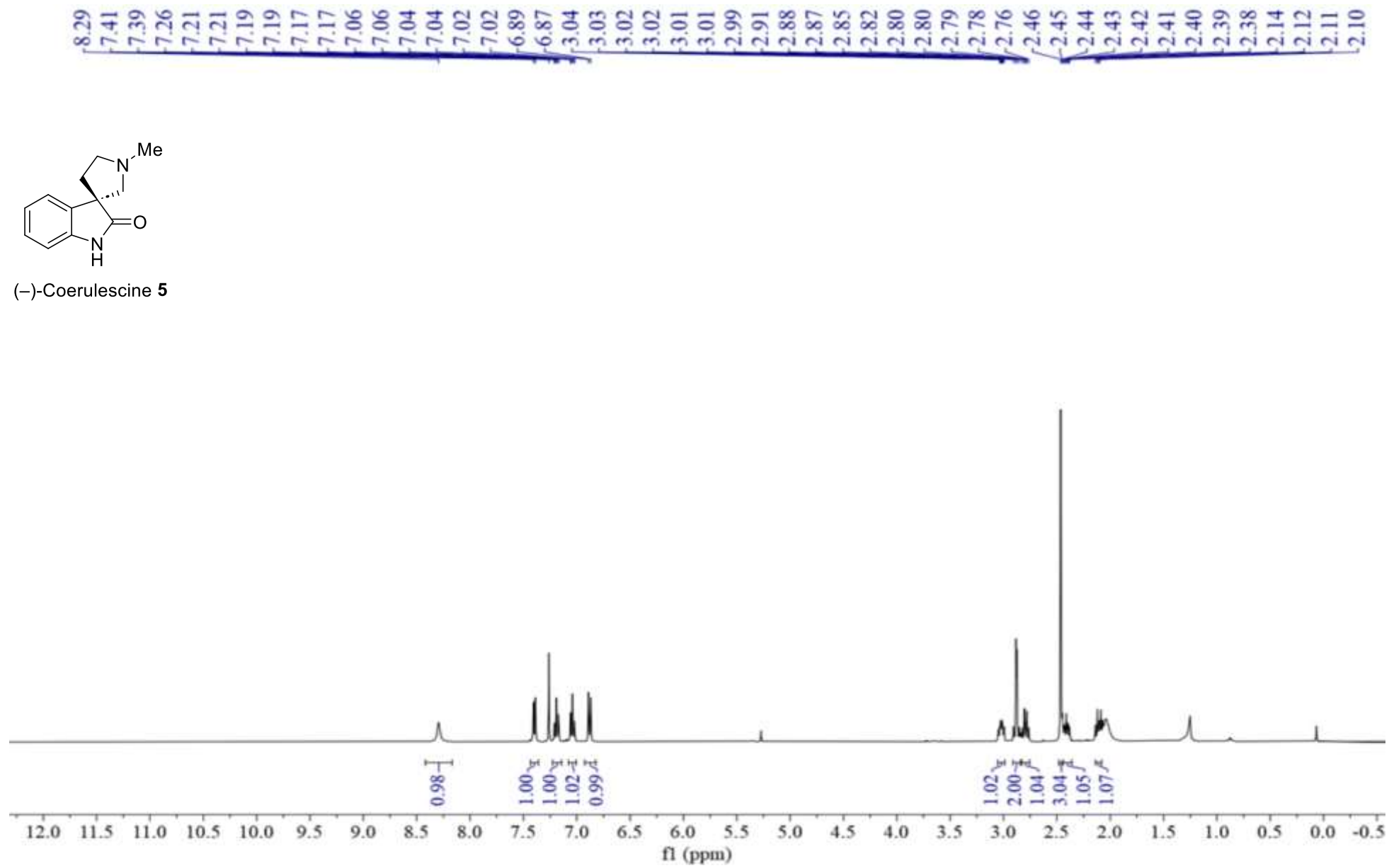
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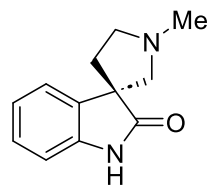


S-98

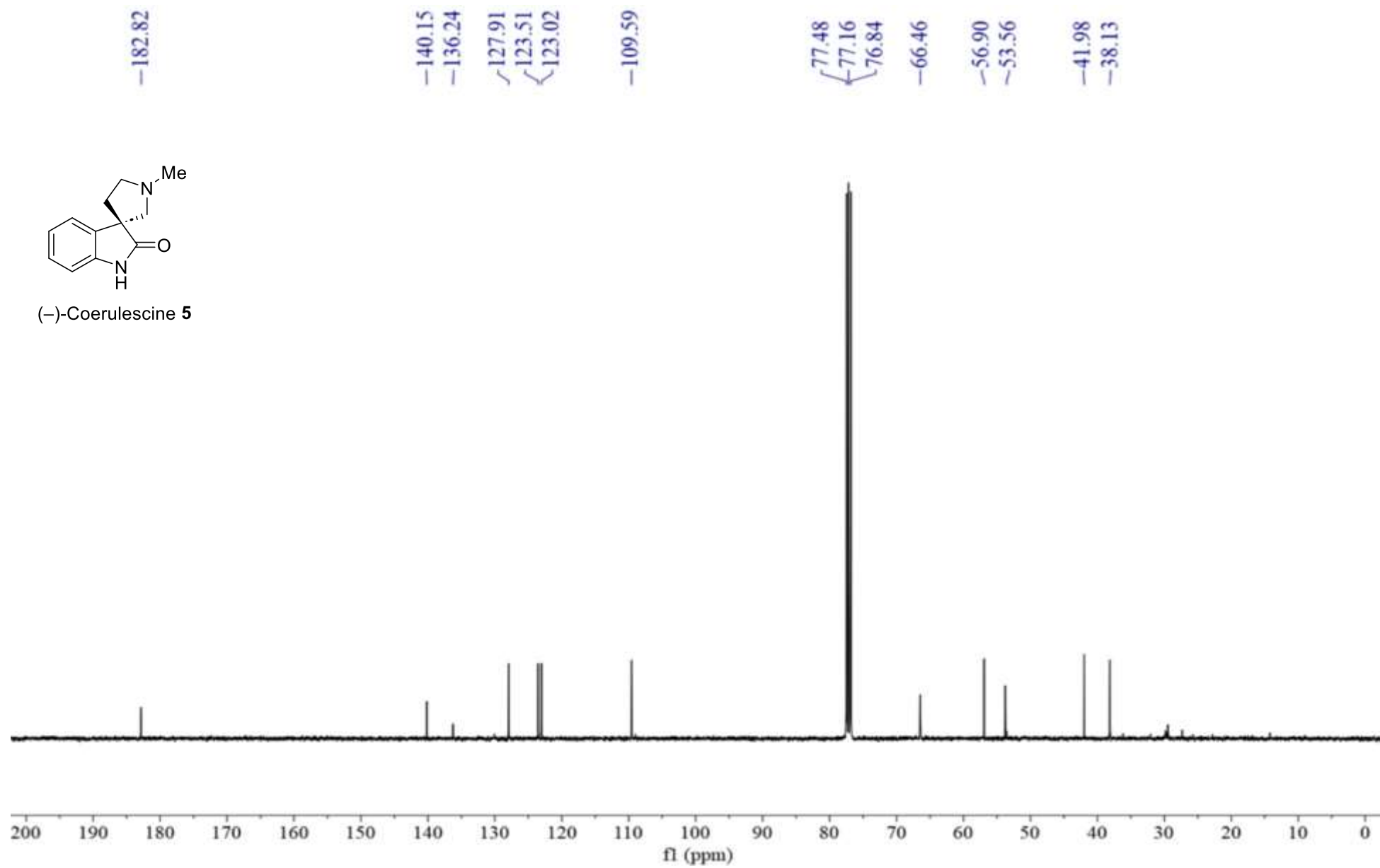


(-)-Coerulescine 5

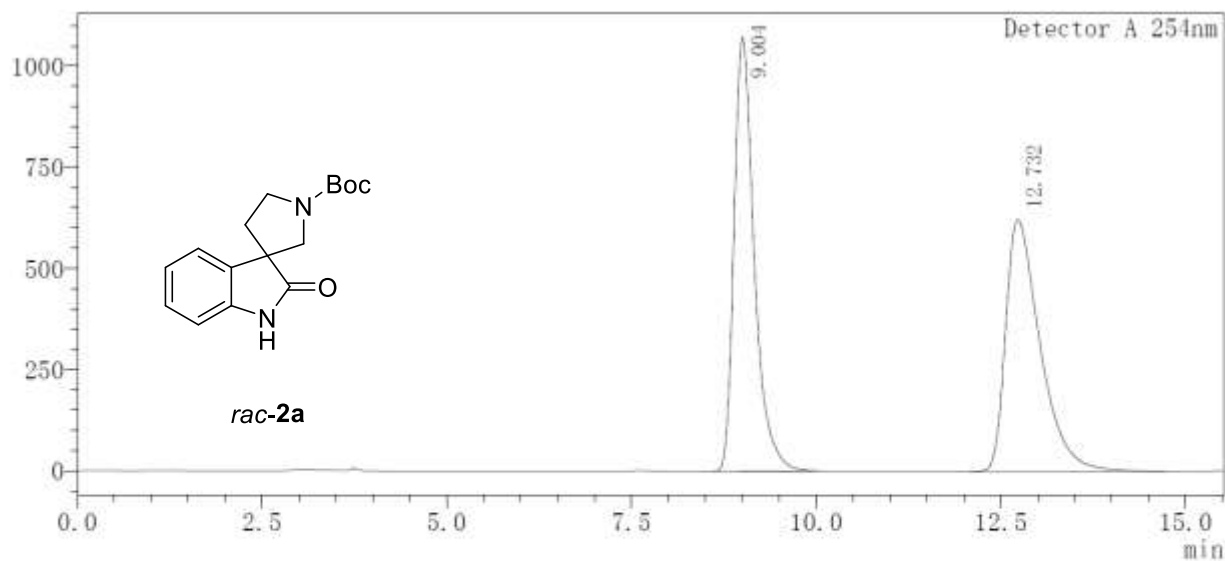




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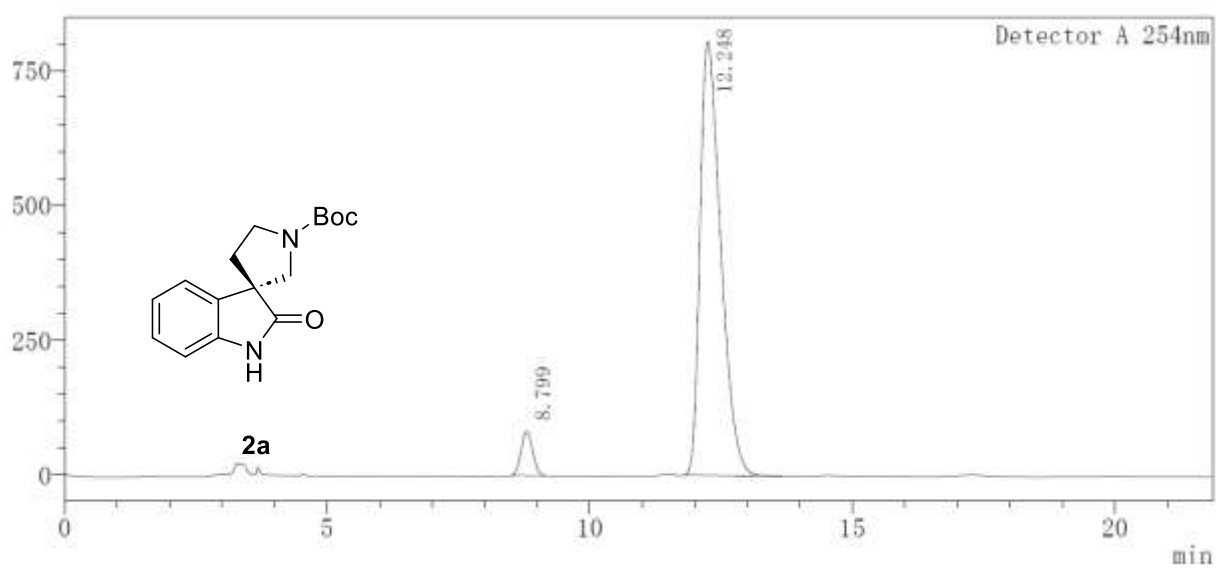
HPLC Chromatograms



<Peak Table>

Detector A 254nm

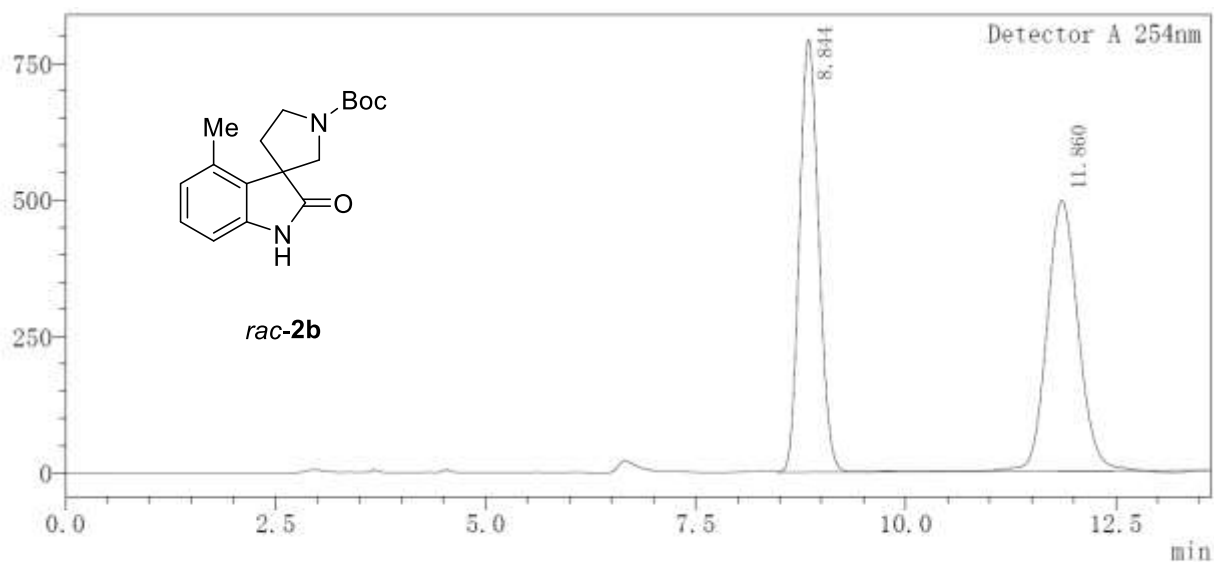
Peak	RetTime	Area	Height	Area%
1	9.004	20411978	1072021	50.315
2	12.732	20156246	621994	49.685
Total		40568224	1694015	



<Peak Table>

Detector A 254nm

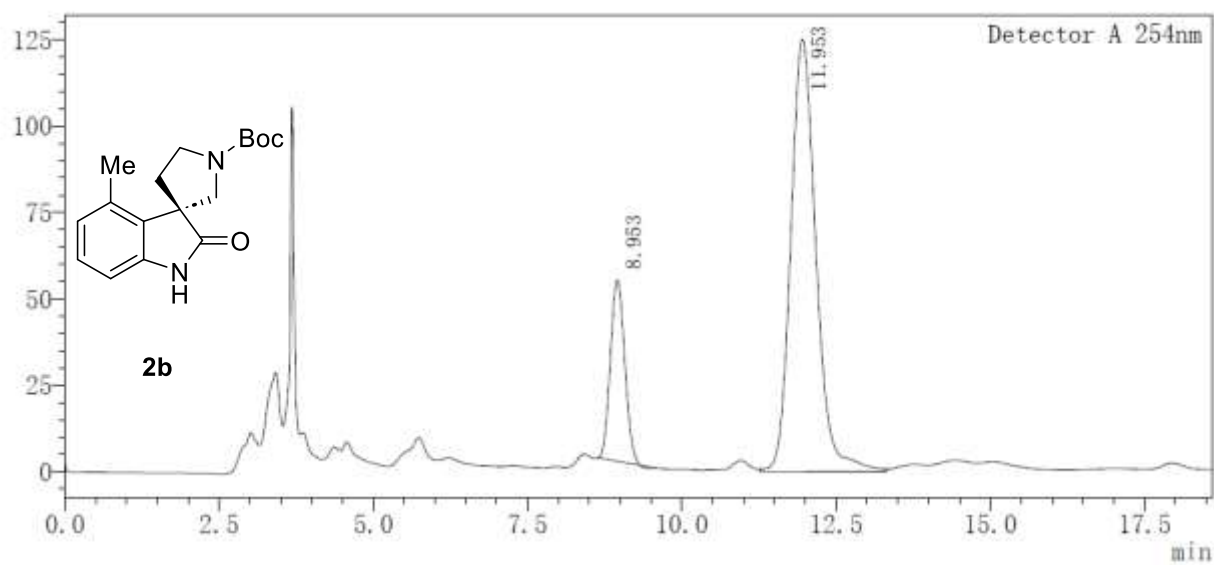
Peak	RetTime	Area	Height	Area%
1	8.799	1253049	82099	5.374
2	12.248	22063723	805135	94.626
Total		23316772	887234	



<Peak Table>

Detector A 254nm

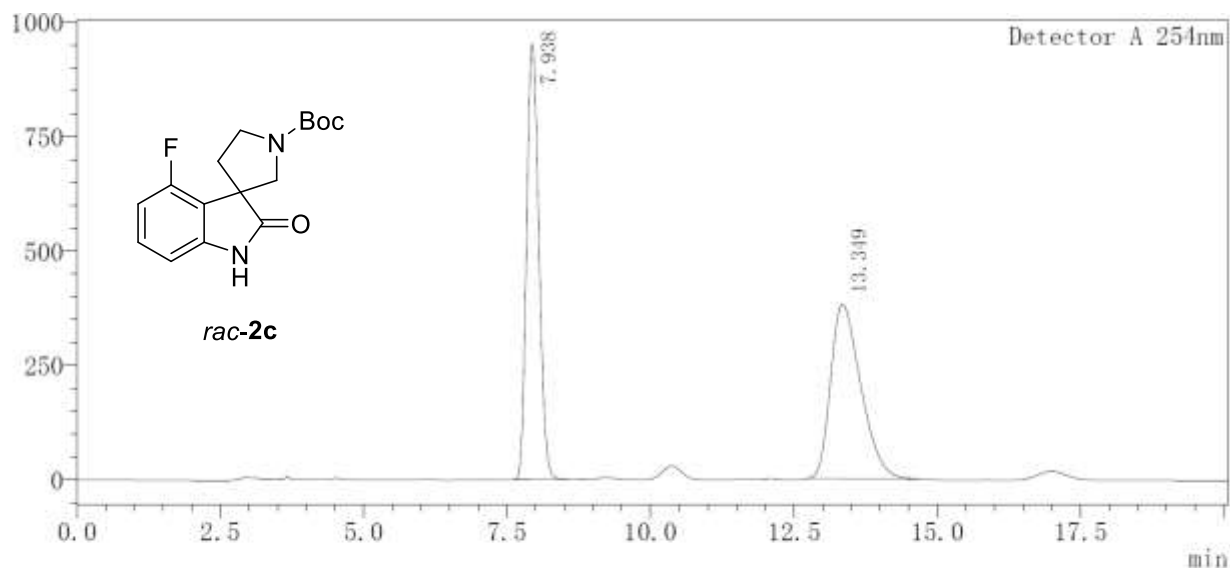
Peak	RetTime	Area	Height	Area%
1	8.844	12727383	793260	49.995
2	11.860	12729806	495963	50.005
Total		25457189	1289223	



<Peak Table>

Detector A 254nm

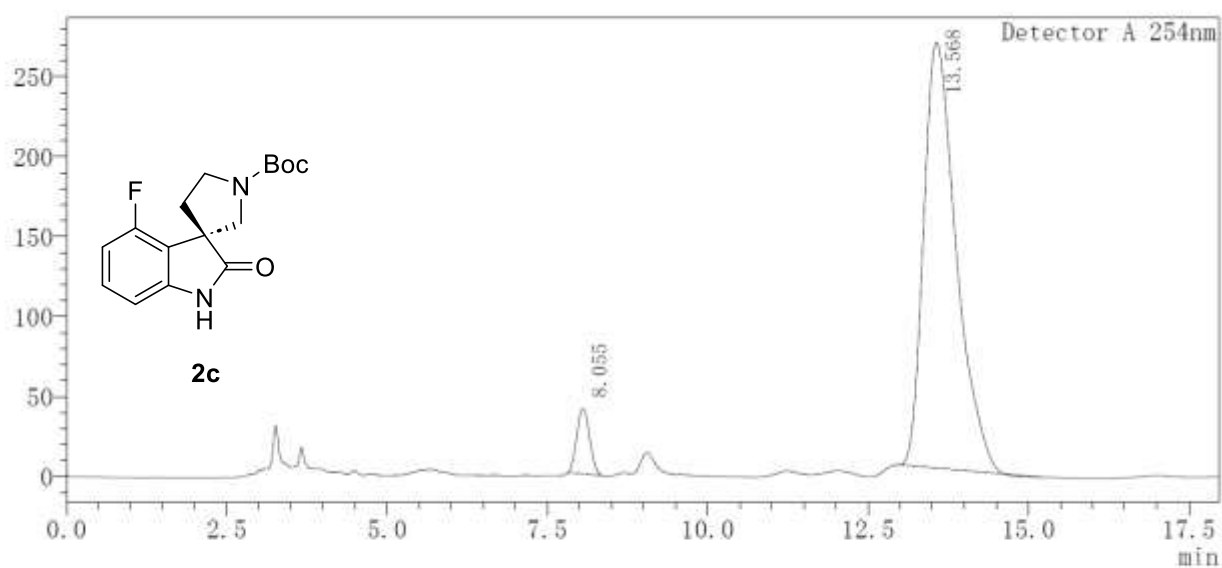
Peak	RetTime	Area	Height	Area%
1	8.953	843272	52433	19.204
2	11.953	3547836	124922	80.796
Total		4391107	177355	



<Peak Table>

Detector A 254nm

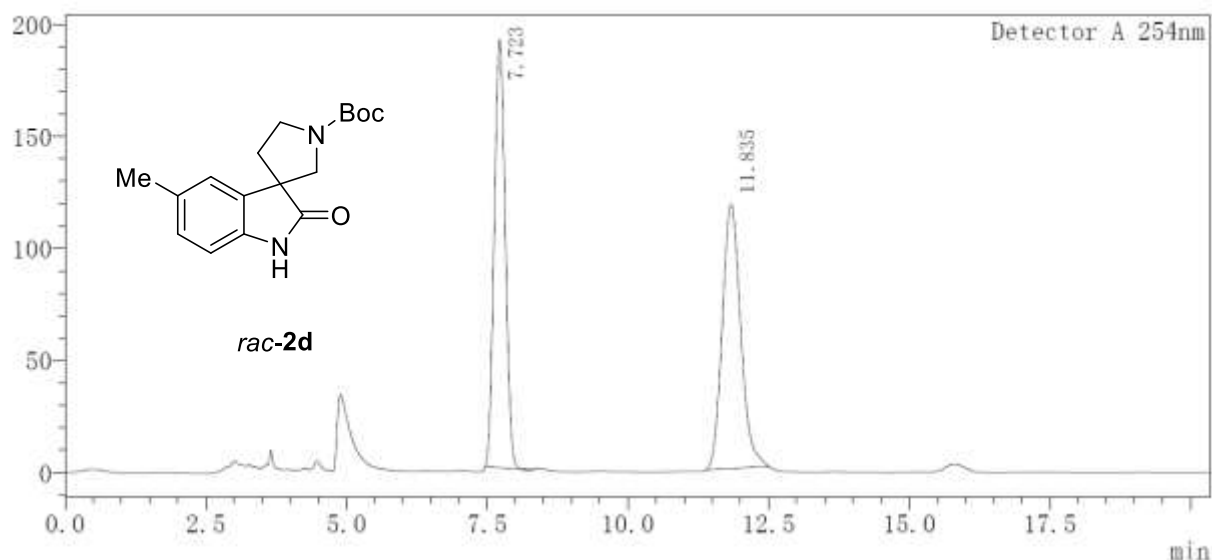
Peak	RetTime	Area	Height	Area%			
1	7.938	14111989	950901	50.326			
2	13.349	13929392	381566	49.674			
Total		28041381	1332466				



<Peak Table>

Detector A 254nm

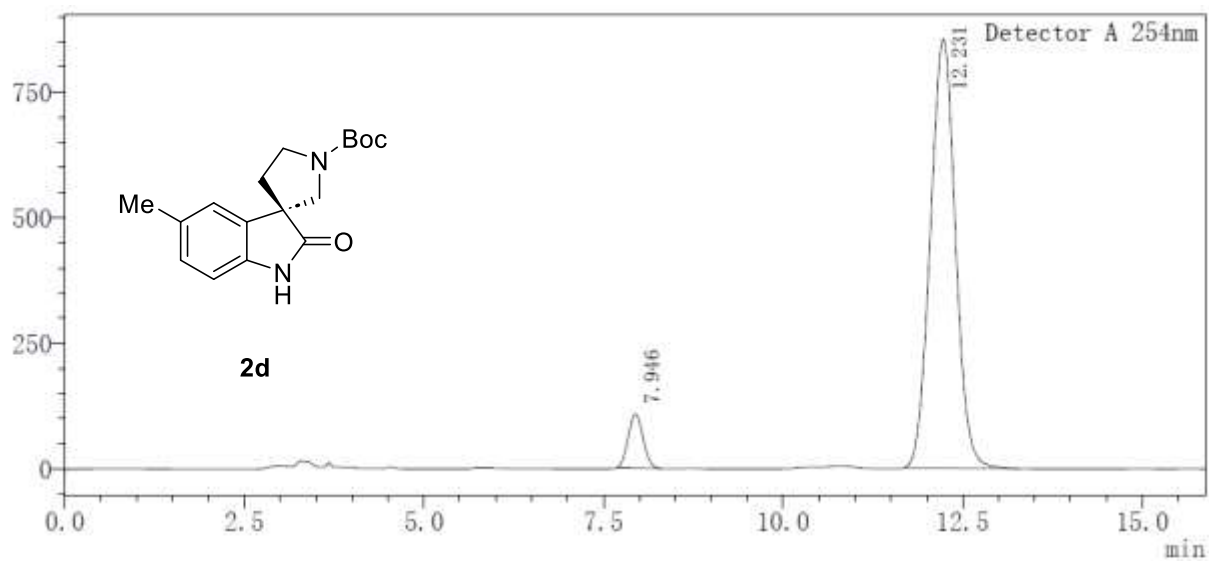
Peak	RetTime	Area	Height	Area%			
1	8.055	566588	40926	5.690			
2	13.568	9390946	266248	94.310			
Total		9957534	307174				



<Peak Table>

Detector A 254nm

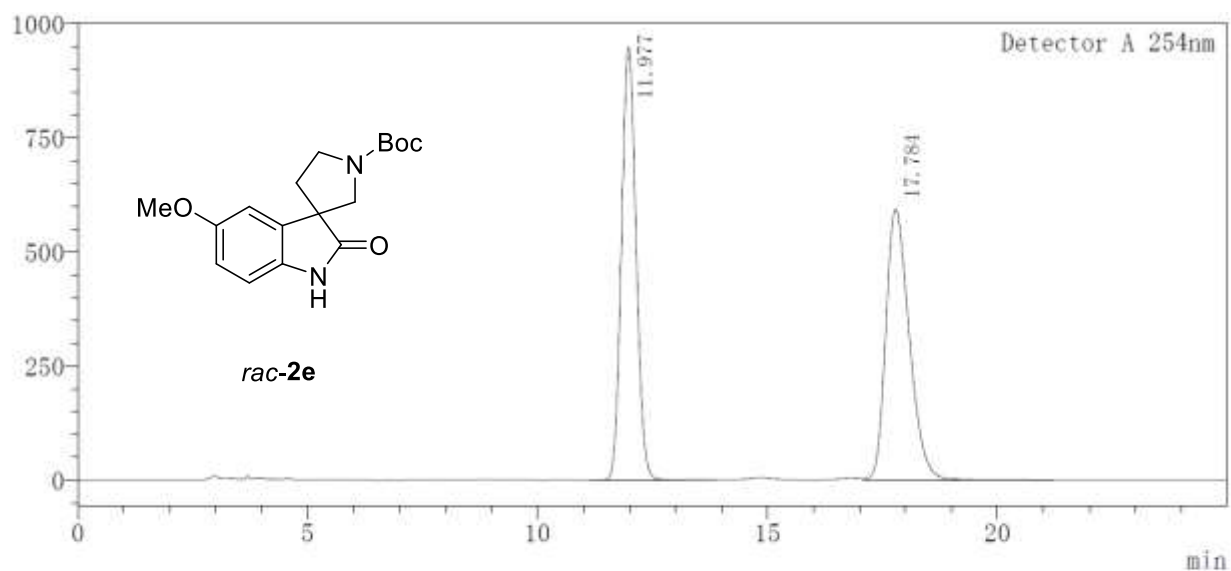
Peak	RetTime	Area	Height	Area%			
1	7.723	2595549	190942	49.155			
2	11.835	2684769	117777	50.845			
Total		5280318	308719				



<Peak Table>

Detector A 254nm

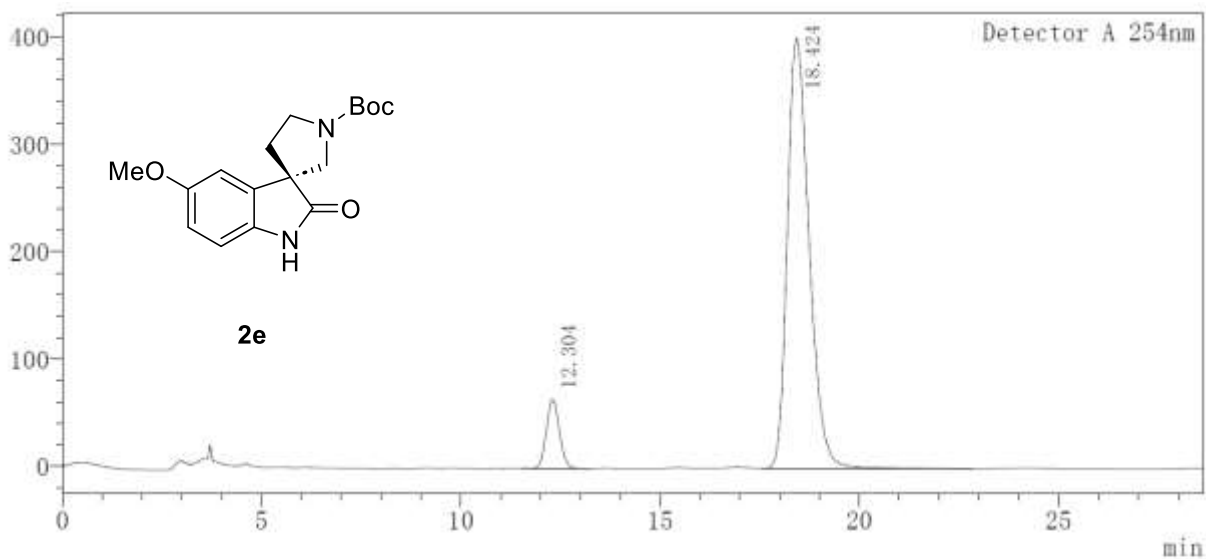
Peak	RetTime	Area	Area	Area%			
1	7.946	1540455	106785	6.915			
2	12.231	20735028	854051	93.085			
Total		22275483	960836				



<Peak Table>

Detector A 254nm

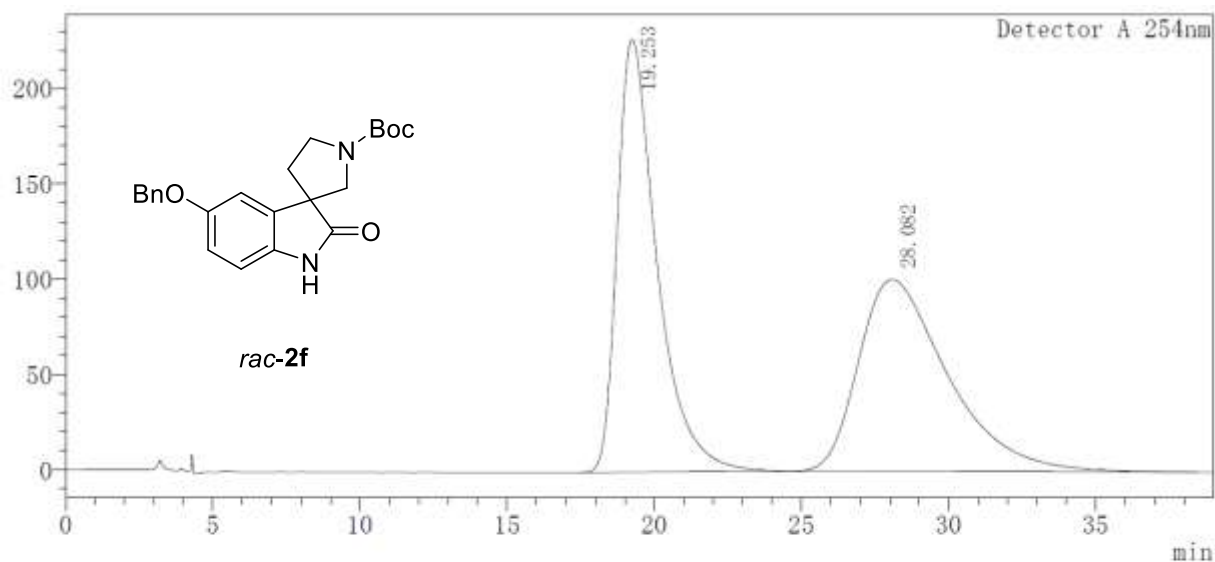
Peak	RetTime	Area	Height	Area%			
1	11.977	20942931	948641	49.860			
2	17.784	21060146	593319	50.140			
Total		42003077	1541960				



<Peak Table>

Detector A 254nm

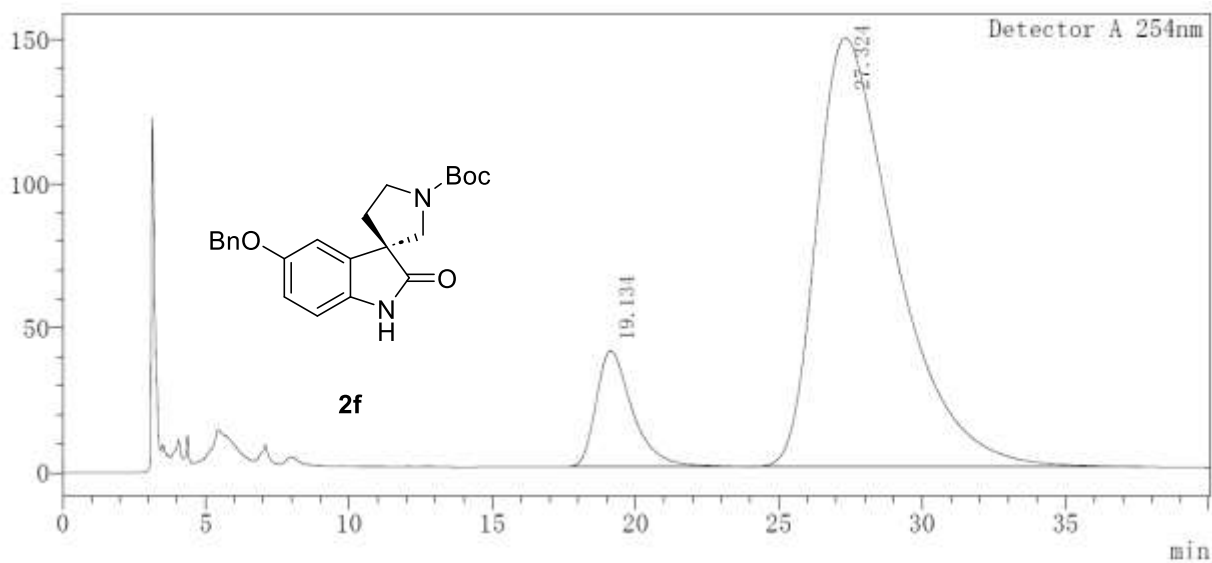
Peak	RetTime	Area	Height	Area%			
1	12.304	1483920	64368	8.816			
2	18.424	15349008	401544	91.184			
Total		16832928	465912				



<Peak Table>

Detector A 254nm

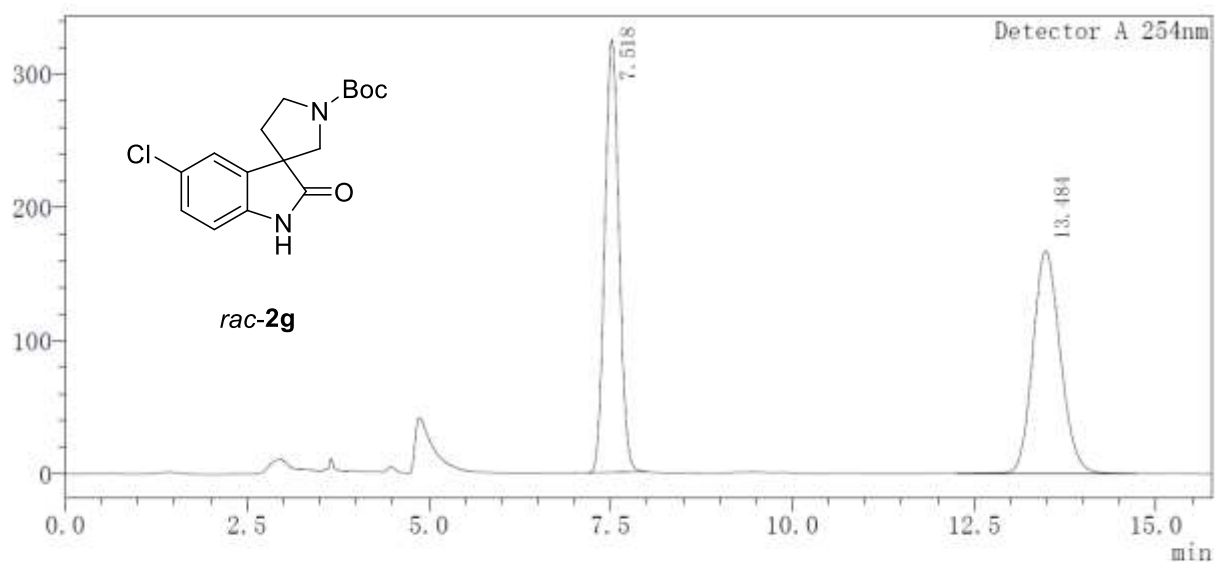
Peak	RetTime	Area	Height	Area%
1	19.253	21327774	227211	50.088
2	28.082	21252896	100731	49.912
Total		42580670	327942	



<Peak Table>

Detector A 254nm

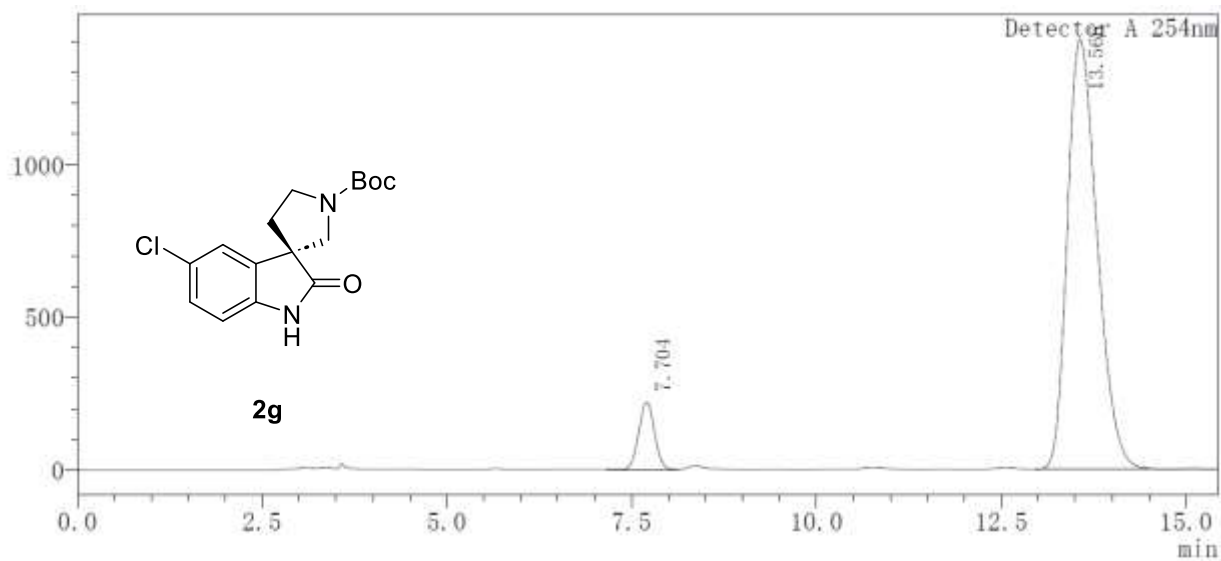
Peak	RetTime	Area	Height	Area%
1	19.134	3532845	40120	11.009
2	27.324	28558464	148491	88.991
Total		32091310	188611	



<Peak Table>

Detector A 254nm

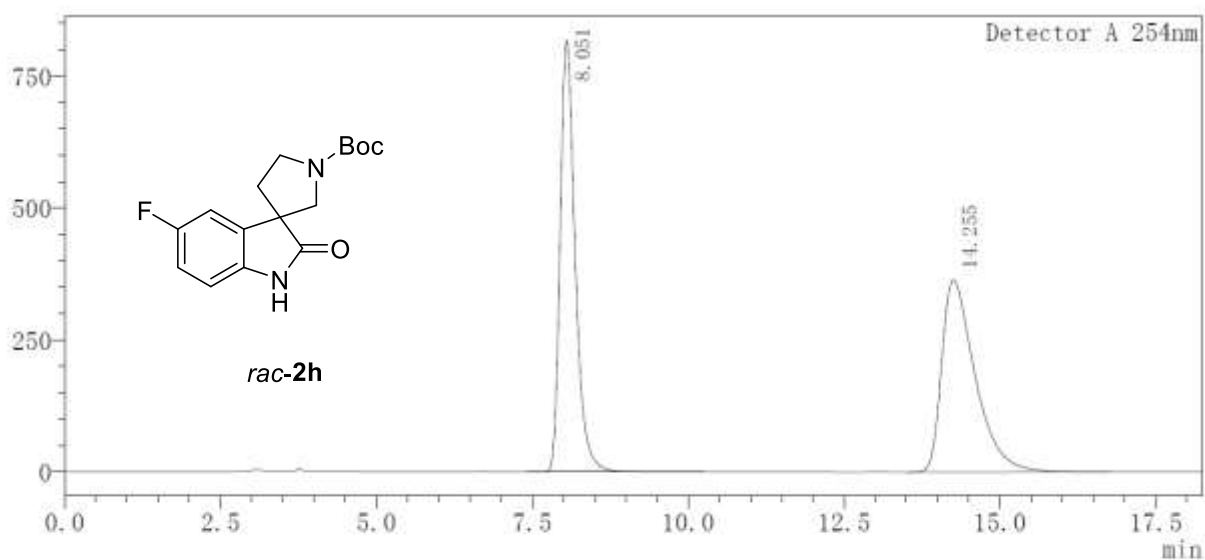
Peak	RetTime	Area	Height	Area%
1	7.518	4431375	324567	50.024
2	13.484	4427184	167394	49.976
Total		8858558	491962	



<Peak Table>

Detector A 254nm

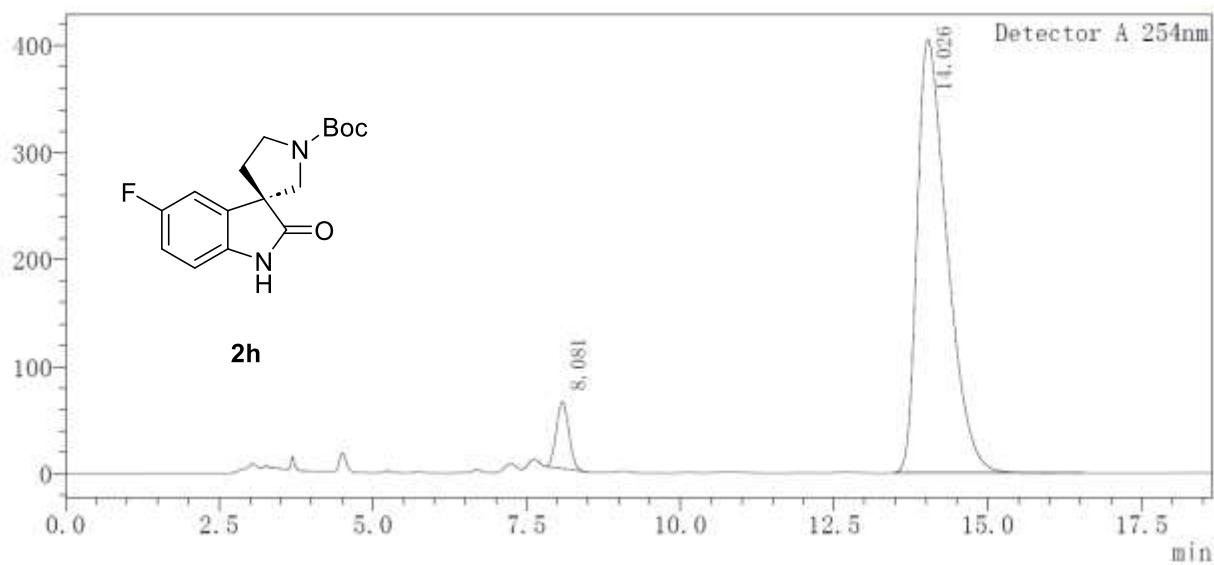
Peak	RetTime	Area	Height	Area%
1	7.704	3299672	219949	7.677
2	13.566	39682509	1409888	92.323
Total		42982182	1629837	



<Peak Table>

Detector A 254nm

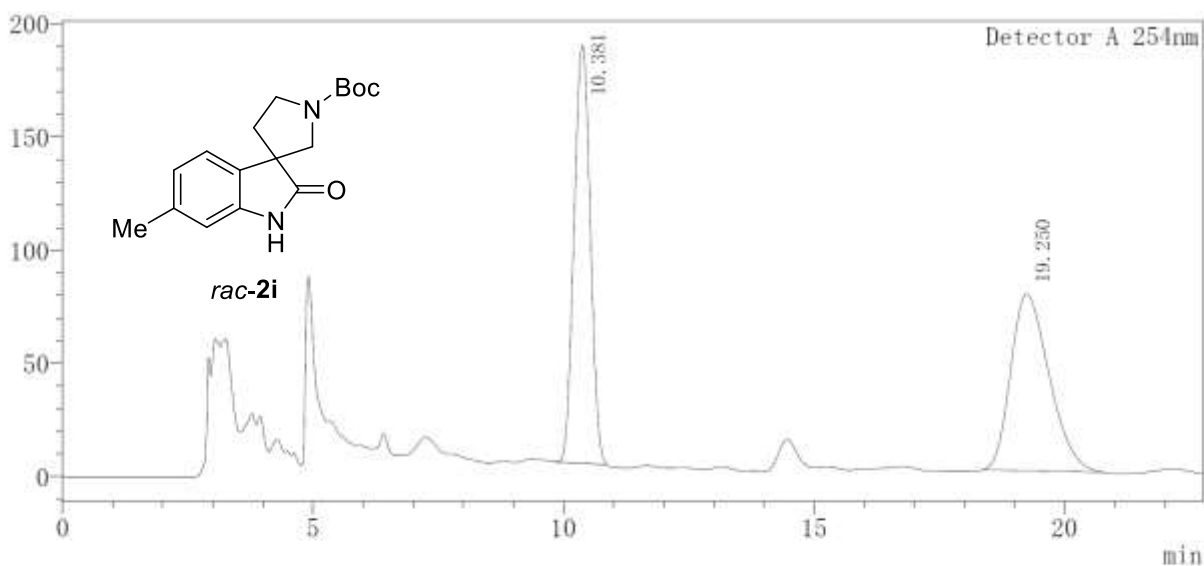
Peak	RetTime	Area	Height	Area%			
1	8.051	13703006	816734	50.260			
2	14.255	13561087	364282	49.740			
Total		27264093	1181016				



<Peak Table>

Detector A 254nm

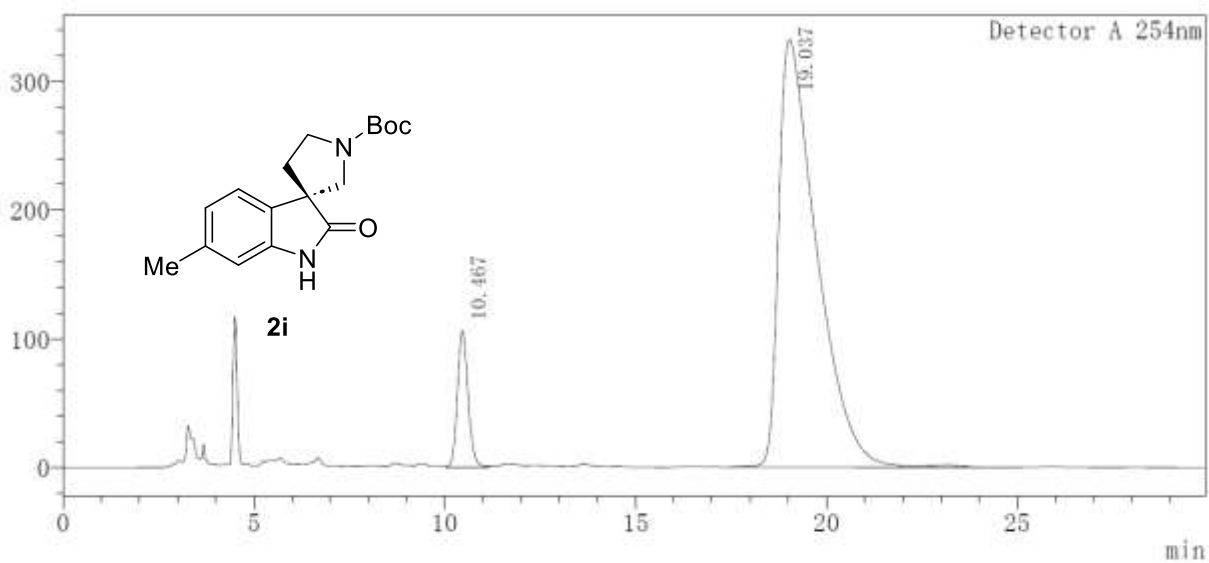
Peak	RetTime	Area	Height	Area%			
1	8.081	885917	62366	6.054			
2	14.026	13747623	405031	93.946			
Total		14633539	467397				



<Peak Table>

Detector A 254nm

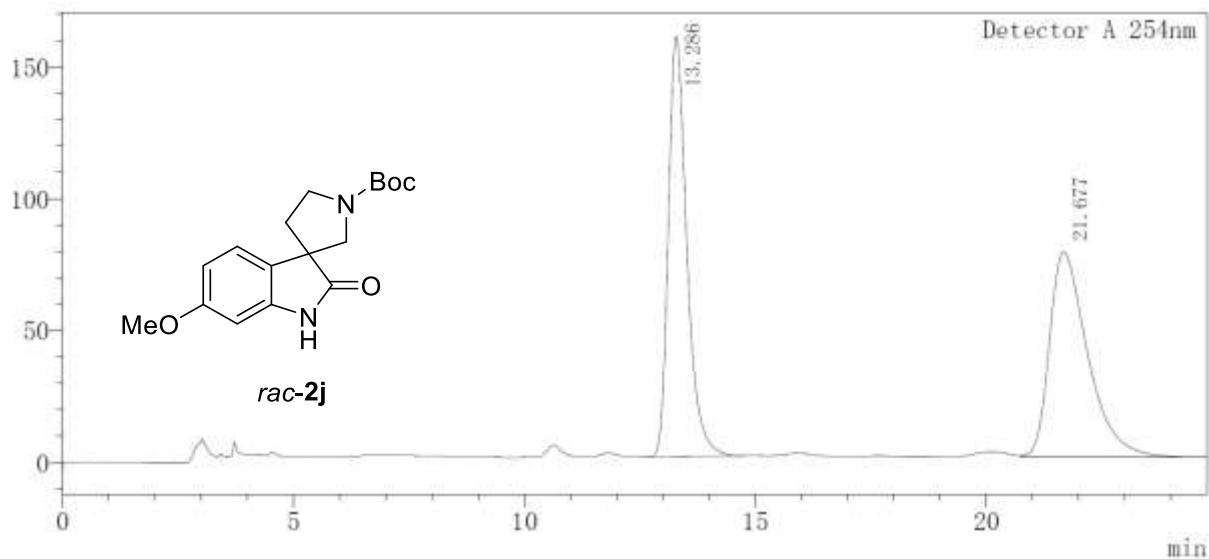
Peak	RetTime	Area	Height	Area%
1	10.381	4015975	181874	49.092
2	19.250	4164456	77831	50.908
Total		8180431	259706	



<Peak Table>

Detector A 254nm

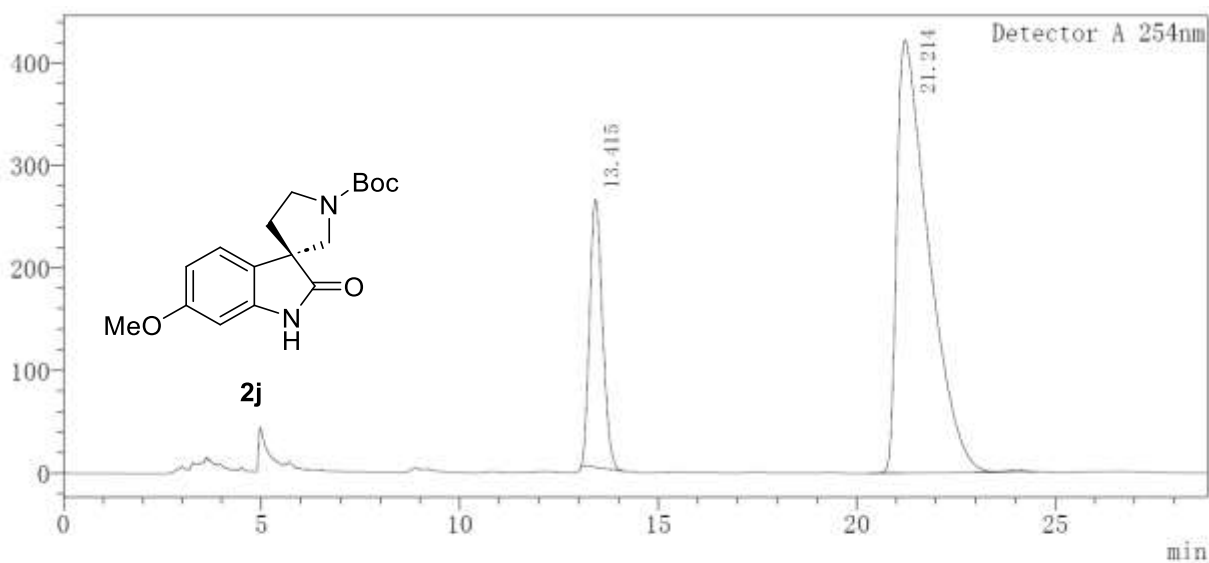
Peak	RetTime	Area	Height	Area%
1	10.467	2053899	106244	8.209
2	19.037	22966388	331992	91.791
Total		25020287	438236	



<Peak Table>

Detector A 254nm

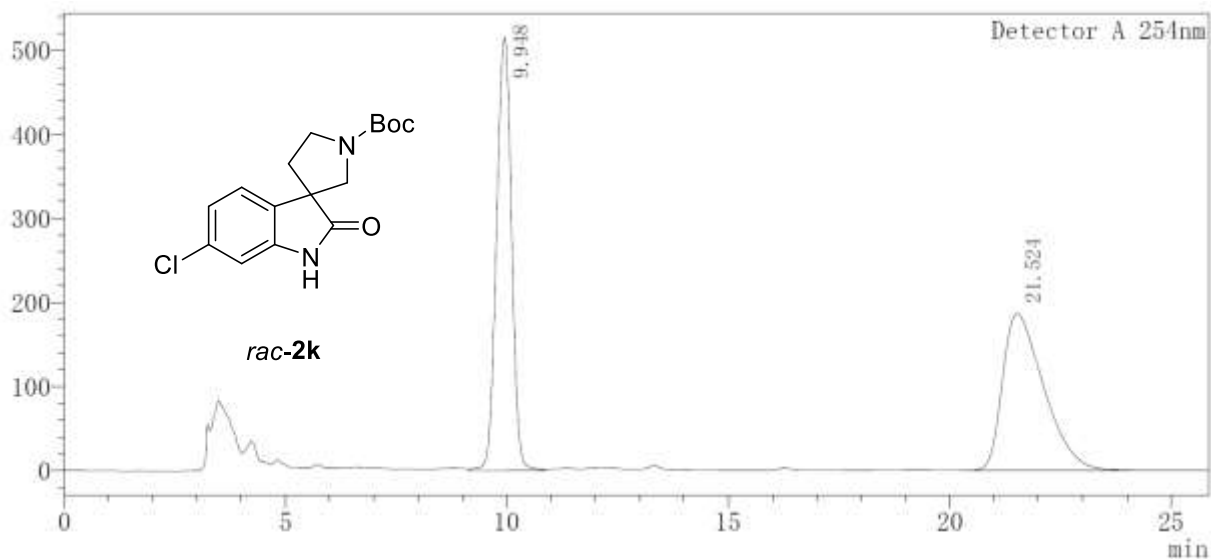
Peak	RetTime	Area	Height	Area%
1	13.286	4427975	159413	49.594
2	21.677	4500505	78029	50.406
Total		8928480	237442	



<Peak Table>

Detector A 254nm

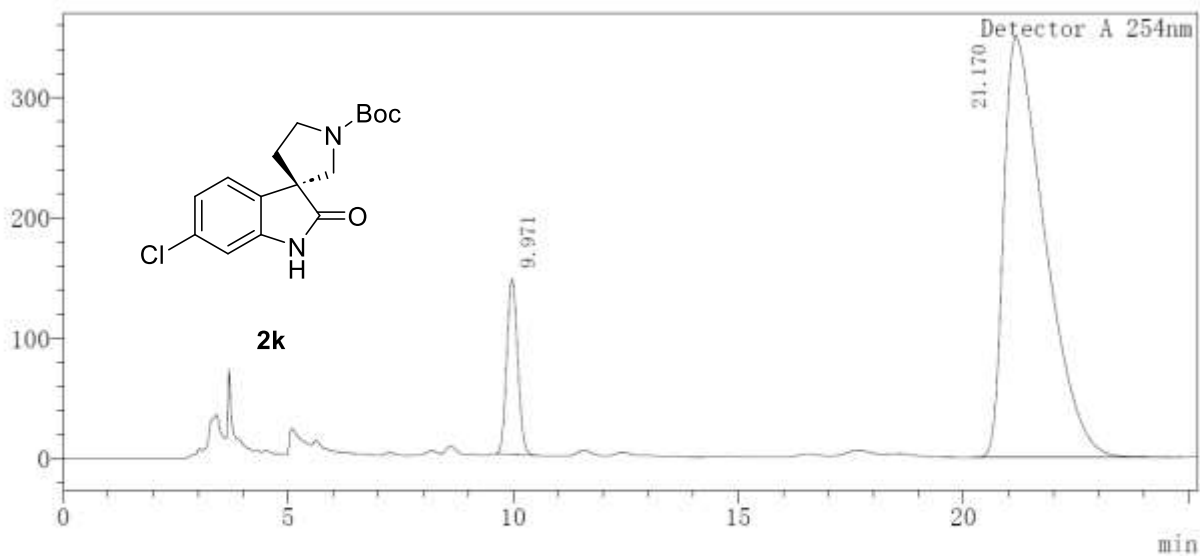
Peak	RetTime	Area	Height	Area%
1	13.415	5875472	261088	19.985
2	21.214	23524298	422048	80.015
Total		29399770	683136	



<Peak Table>

Detector A 254nm

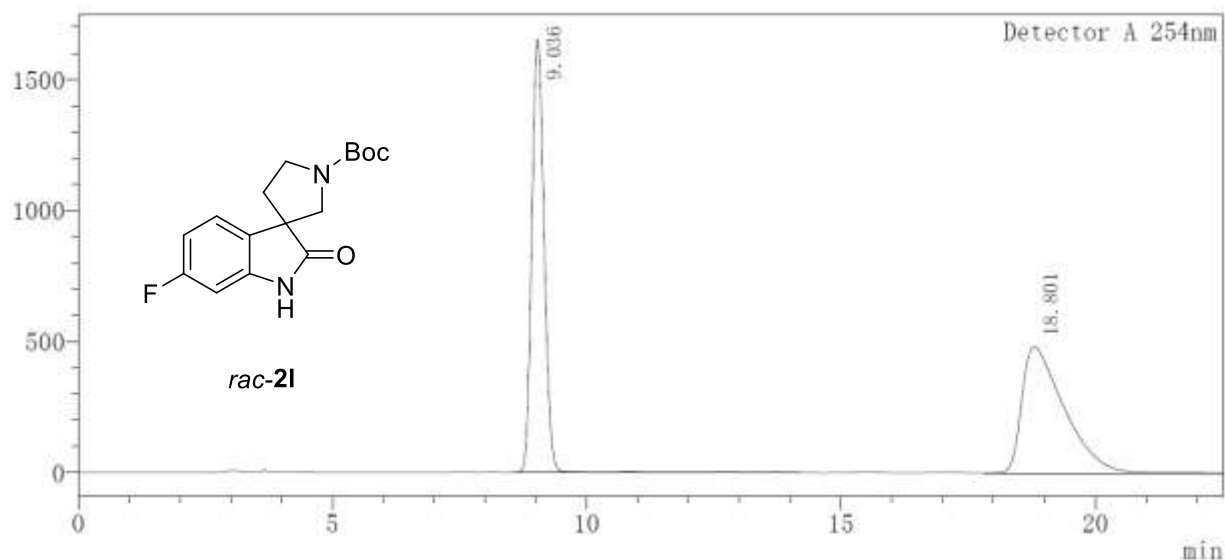
Peak	RetTime	Area	Height	Area%
1	9.948	12209329	515595	50.167
2	21.524	12127968	186824	49.833
Total		24337297	702420	



<Peak Table>

Detector A 254nm

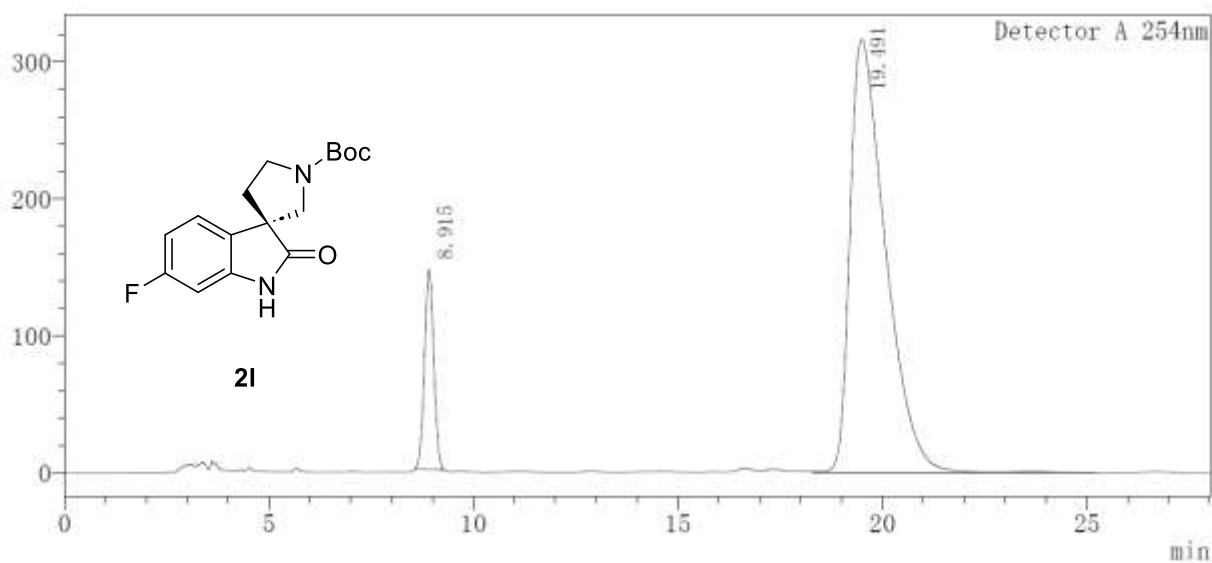
Peak	RetTime	Area	Height	Area%
1	9.971	2466042	145637	9.999
2	21.170	22197522	349211	90.001
Total		24663564	494847	



<Peak Table>

Detector A 254nm

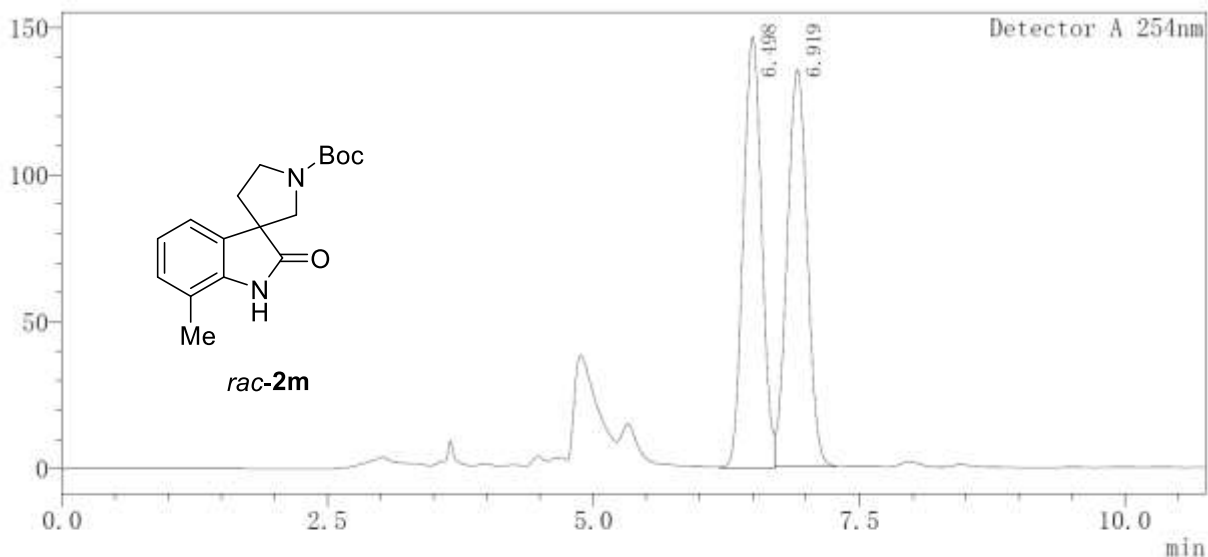
Peak	RetTime	Area	Height	Area%			
1	9.036	28086687	1654251	50.501			
2	18.801	27529883	479876	49.499			
Total		55616570	2134127				



<Peak Table>

Detector A 254nm

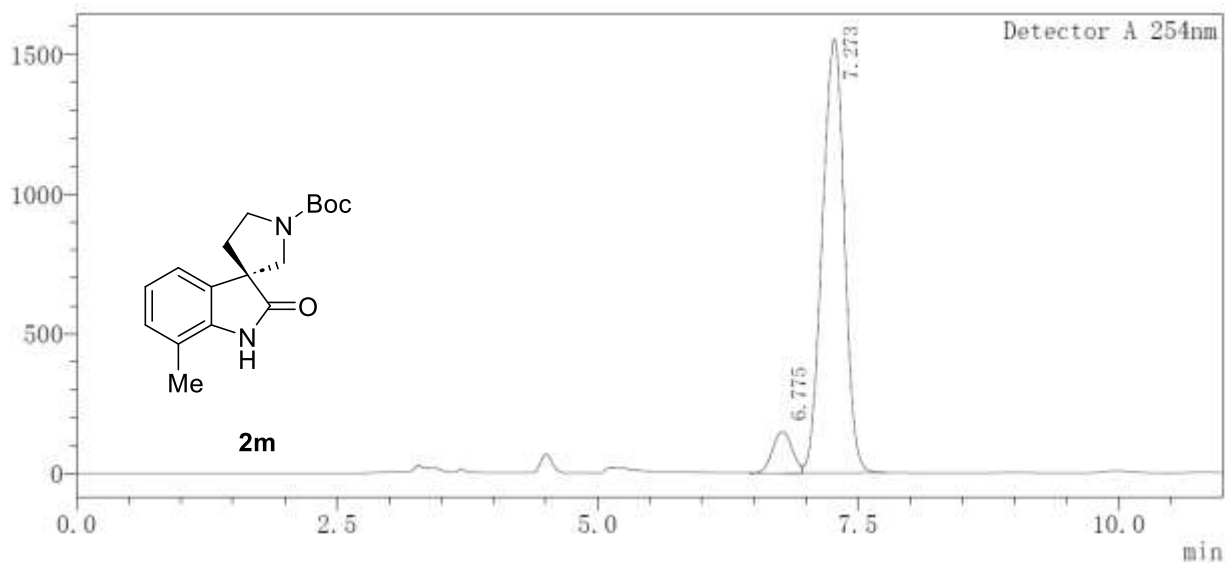
Peak	RetTime	Area	Height	Area%			
1	8.915	2316918	145684	10.791			
2	19.491	19154571	316667	89.209			
Total		21471489	462351				



<Peak Table>

Detector A 254nm

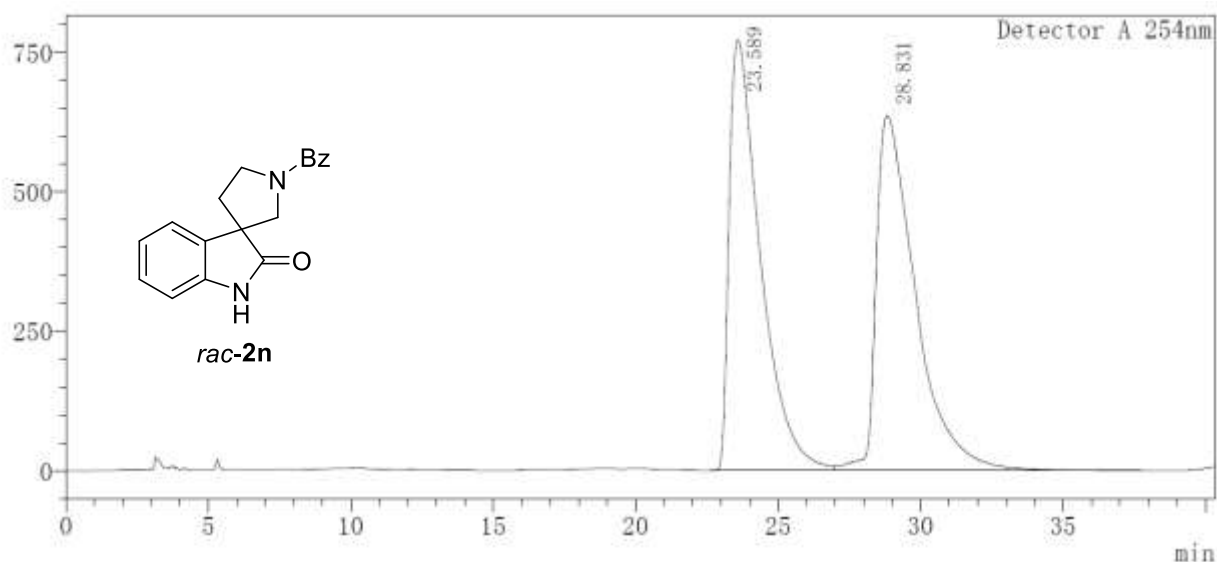
Peak	RetTime	Area	Height	Area%
1	6.498	1769138	146932	49.950
2	6.919	1772704	135082	50.050
Total		3541842	282014	



<Peak Table>

Detector A 254nm

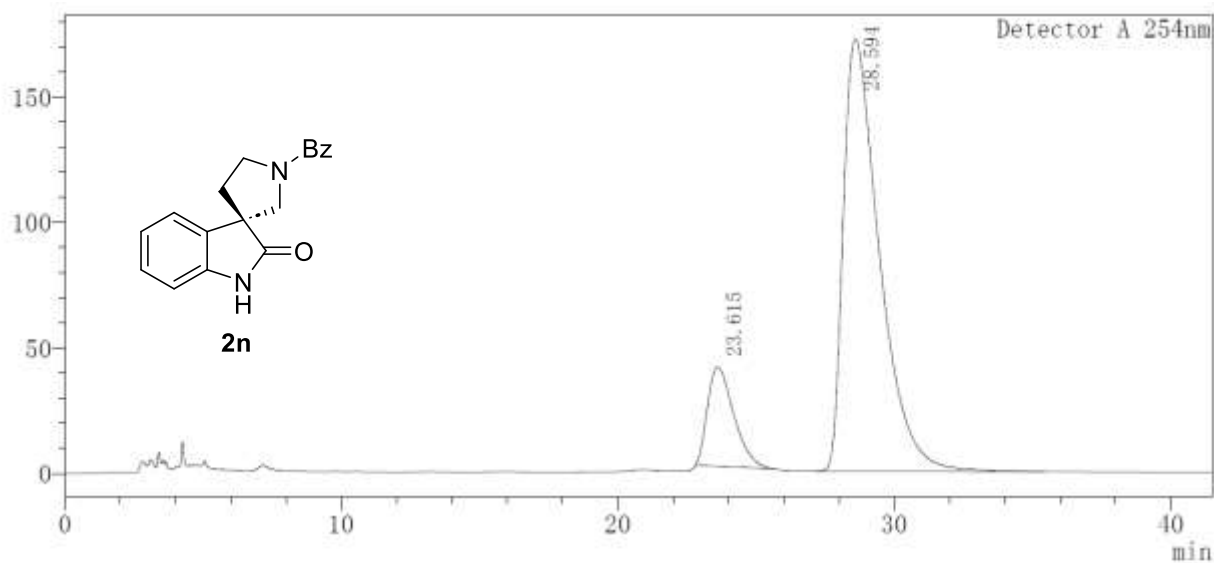
Peak	RetTime	Area	Height	Area%
1	6.775	1981836	149620	7.946
2	7.273	22959517	1553784	92.054
Total		24941353	1703404	



<Peak Table>

Detector A 254nm

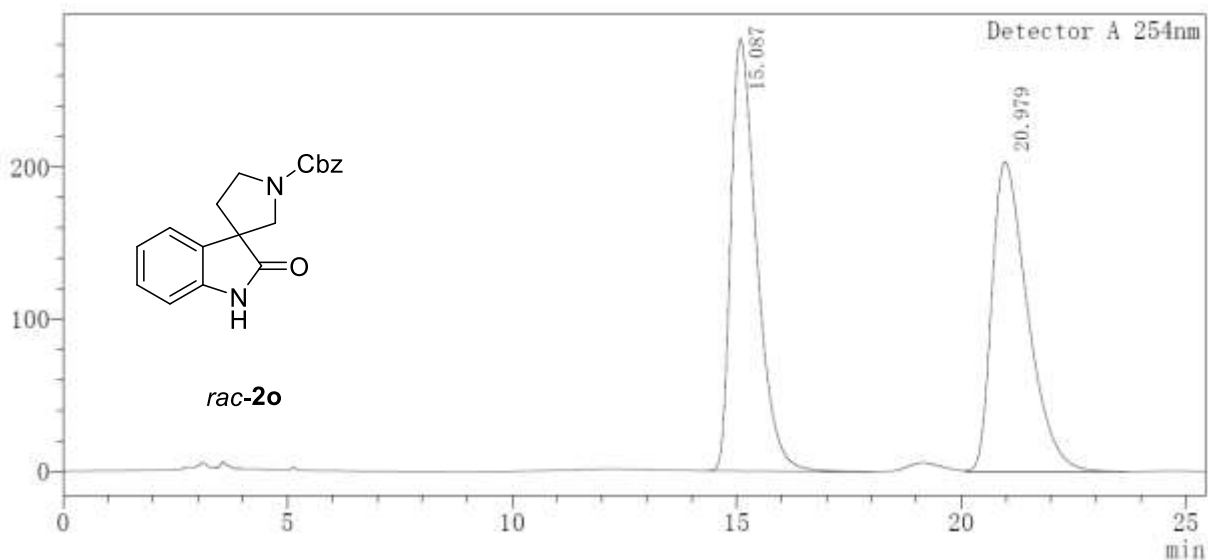
Peak	RetTime	Area	Height	Area%			
1	23.589	58921304	771817	49.042			
2	28.831	61222259	635952	50.958			
Total		120143563	1407769				



<Peak Table>

Detector A 254nm

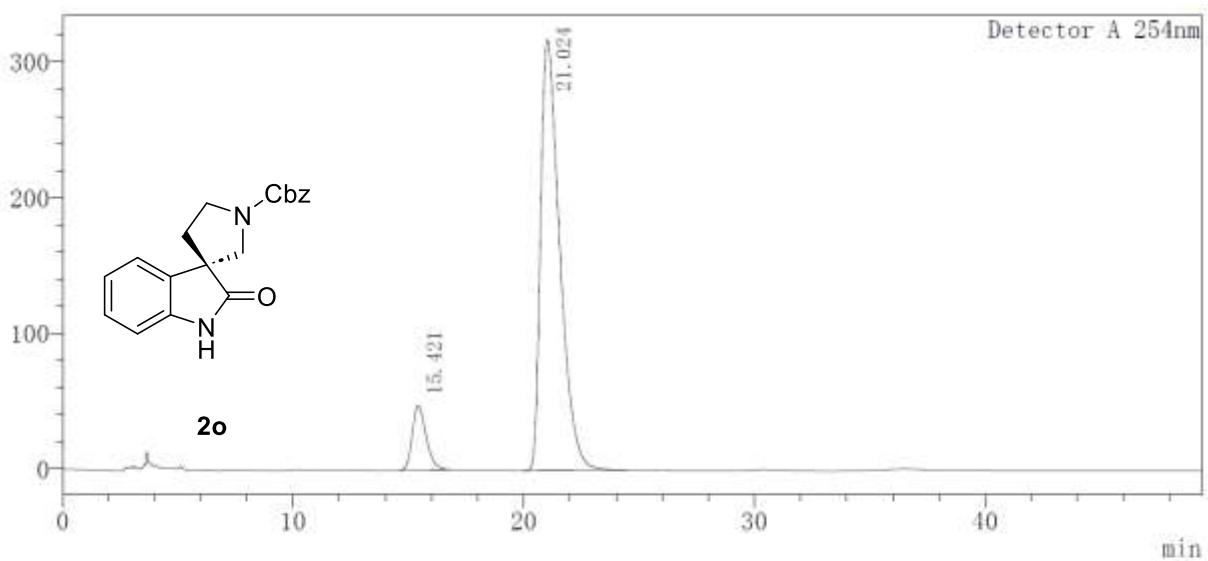
Peak	RetTime	Area	Height	Area%			
1	23.615	2611405	39473	14.296			
2	28.594	15655421	172184	85.704			
Total		18266826	211657				



<Peak Table>

Detector A 254nm

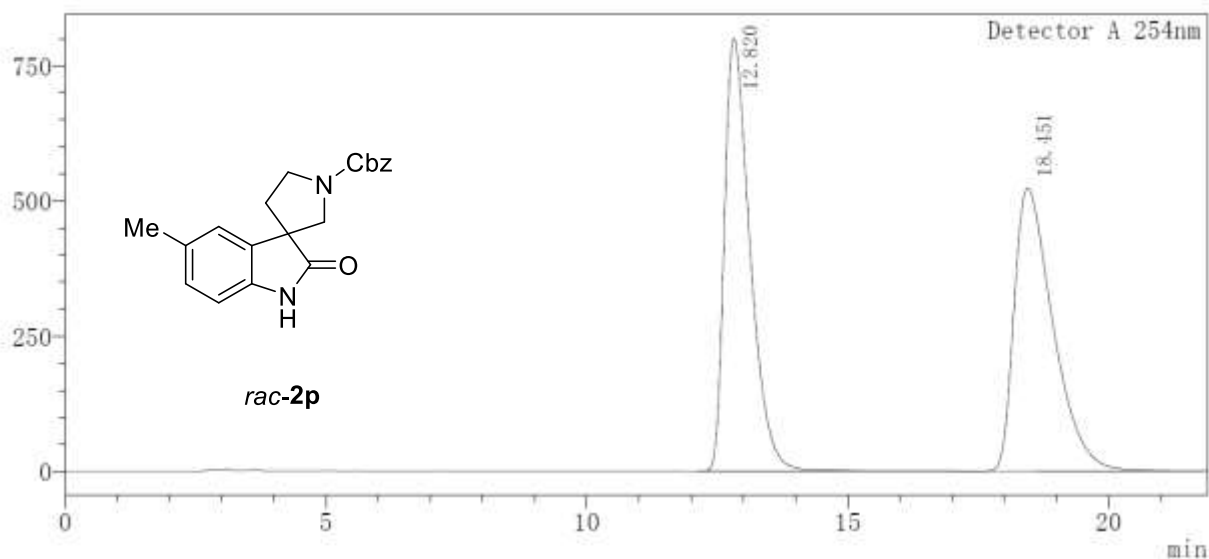
Peak	RetTime	Area	Height	Area%			
1	15.087	11349076	283672	50.179			
2	20.979	11268052	203272	49.821			
Total		22617128	486944				



<Peak Table>

Detector A 254nm

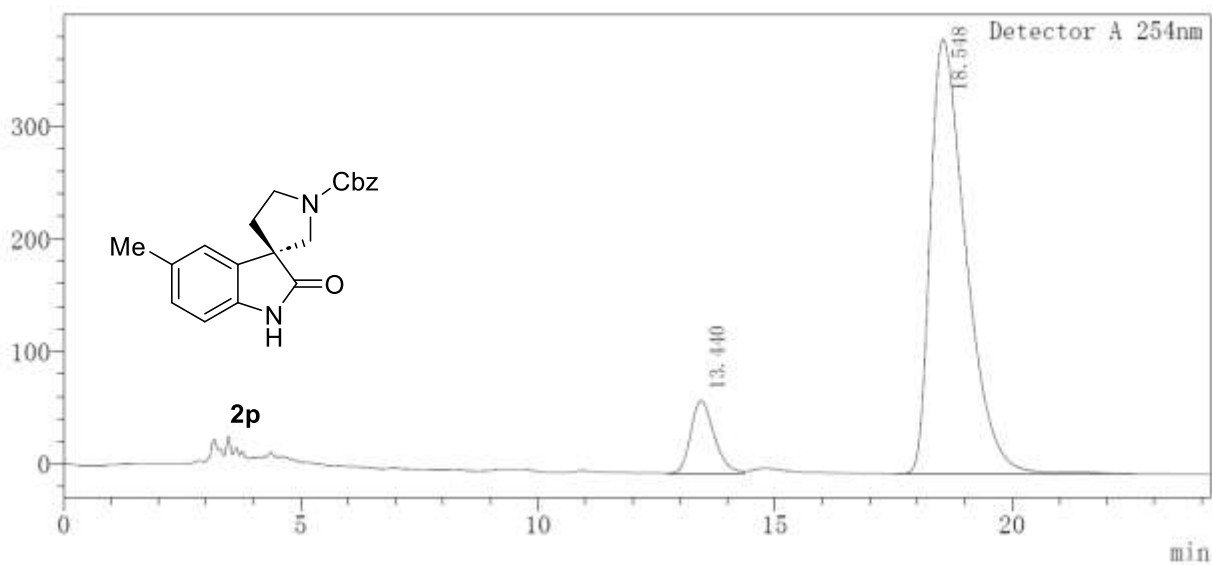
Peak	RetTime	Area	Height	Area%			
1	15.421	1964967	47483	9.686			
2	21.024	18321184	317359	90.314			
Total		20286151	364842				



<Peak Table>

Detector A 254nm

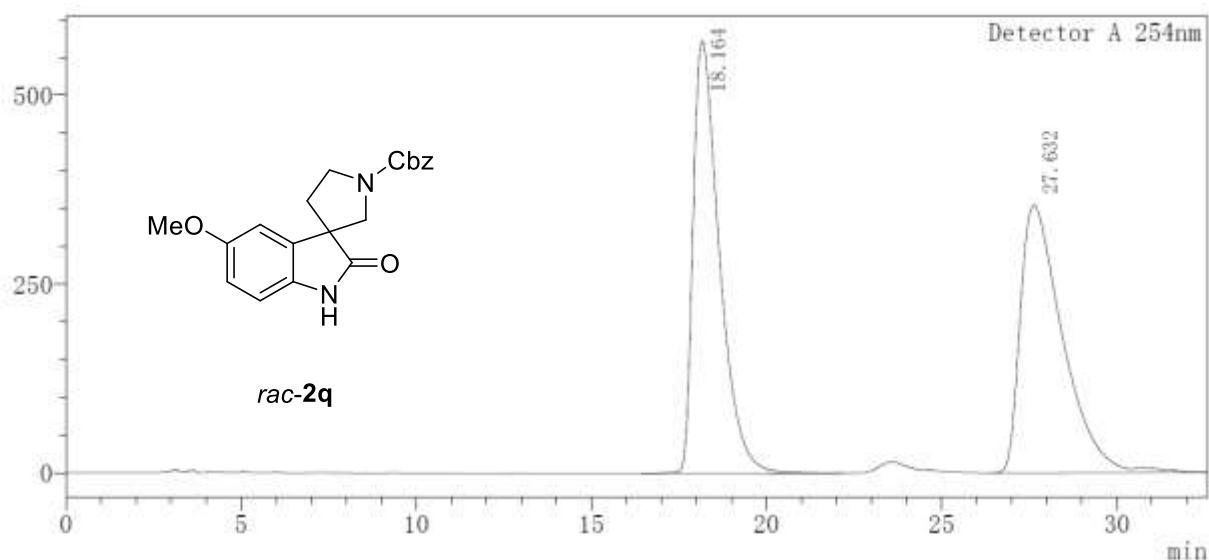
Peak	RetTime	Area	Height	Area%
1	12.820	27445816	799769	50.264
2	18.451	27158037	523334	49.736
Total		54603853	1323103	



<Peak Table>

Detector A 254nm

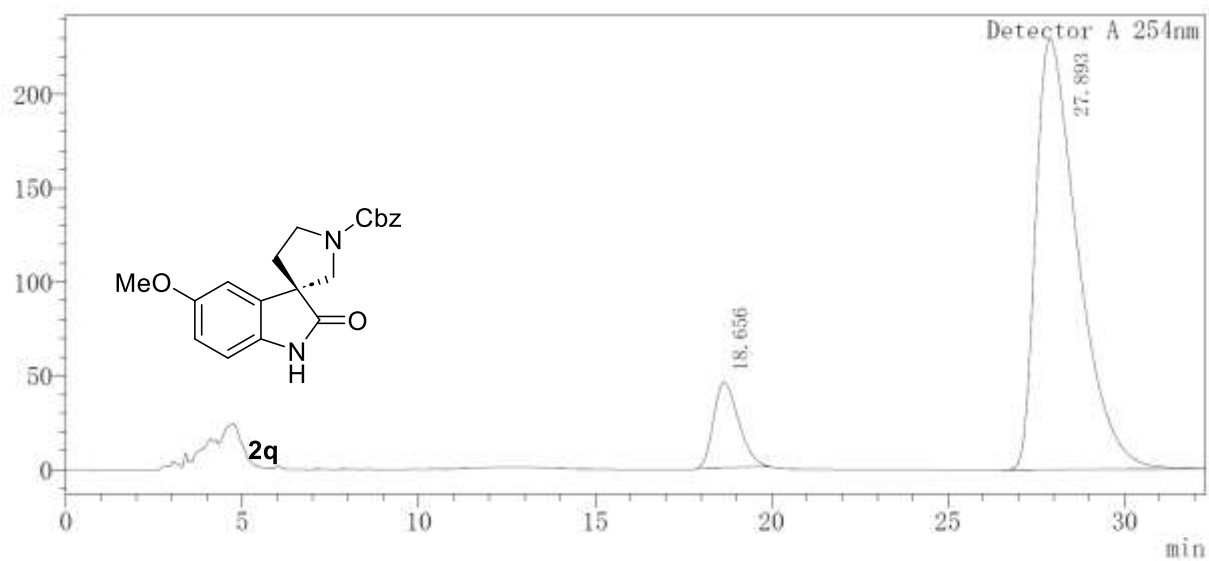
Peak	RetTime	Area	Height	Area%
1	13.440	2210102	64604	9.853
2	18.548	20220688	386387	90.147
Total		22430790	450992	



<Peak Table>

Detector A 254nm

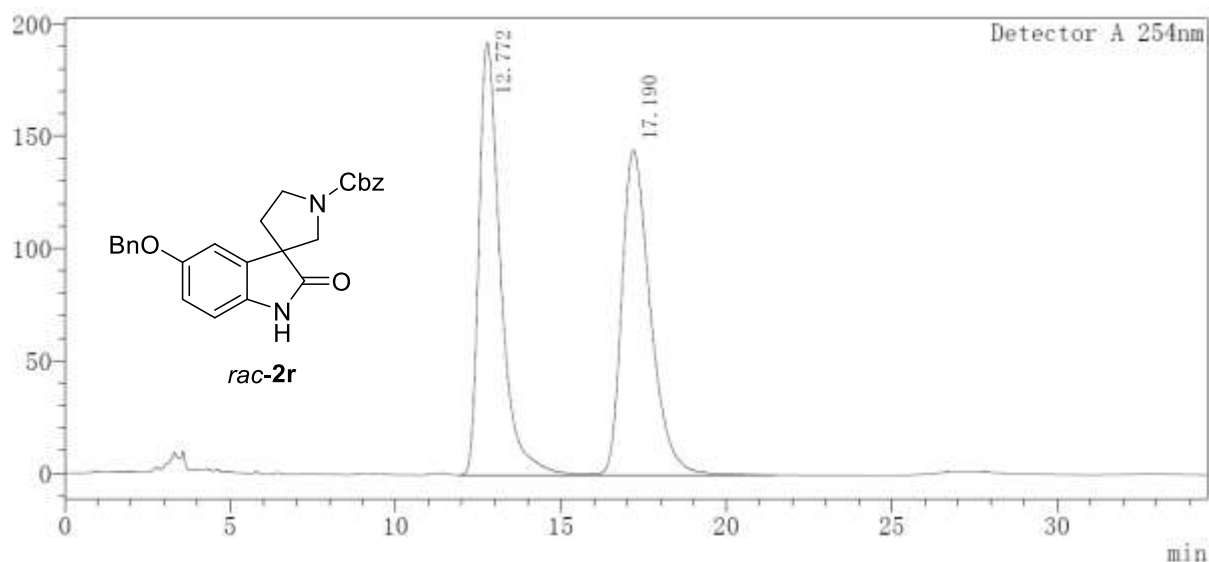
Peak	RetTime	Area	Height	Area%
1	18.164	29966465	573000	50.060
2	27.632	29894969	354577	49.940
Total		59861434	927577	



<Peak Table>

Detector A 254nm

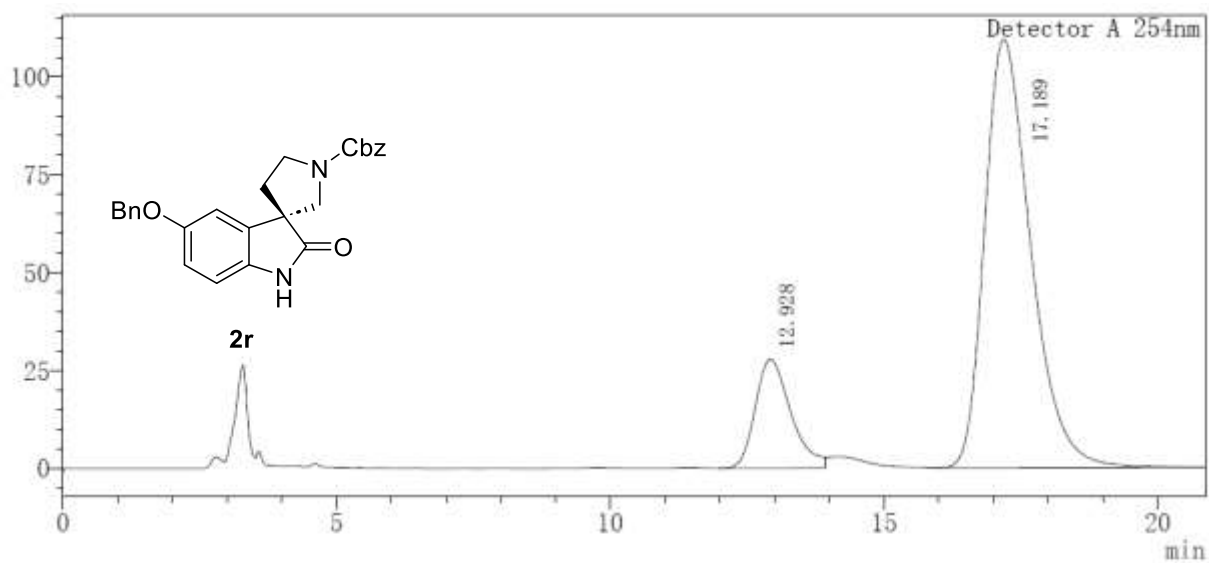
Peak	RetTime	Area	Height	Area%
1	18.656	2235573	45268	10.554
2	27.893	18947141	229351	89.446
Total		21182714	274618	



<Peak Table>

Detector A 254nm

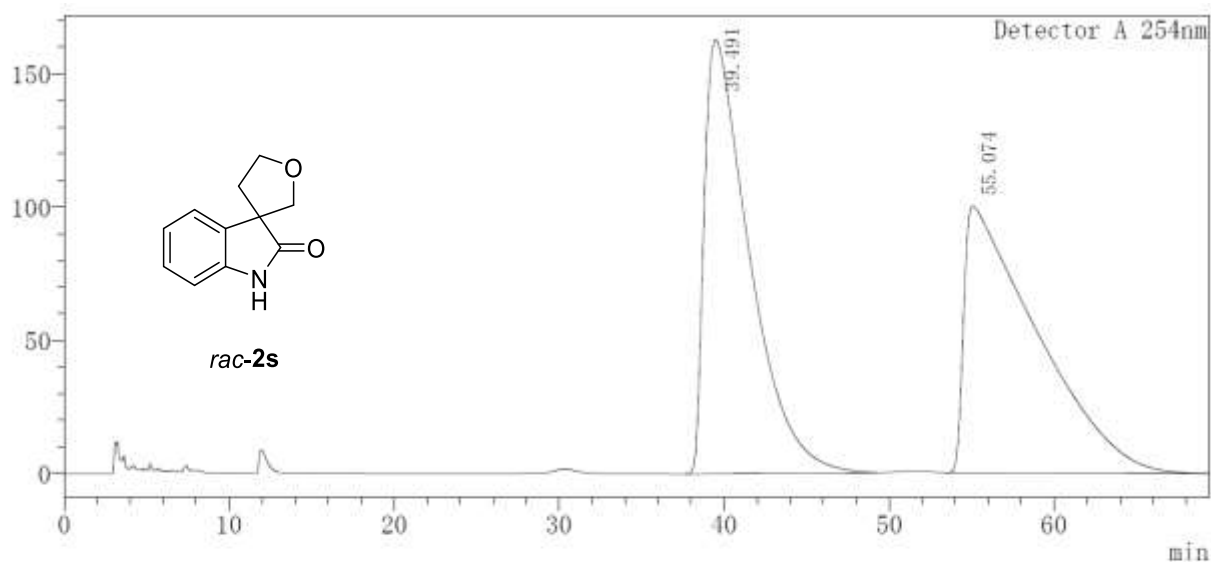
Peak	RetTime	Area	Height	Area%
1	12.772	8748129	192375	50.157
2	17.190	8693320	144616	49.843
Total		17441449	336991	



<Peak Table>

Detector A 254nm

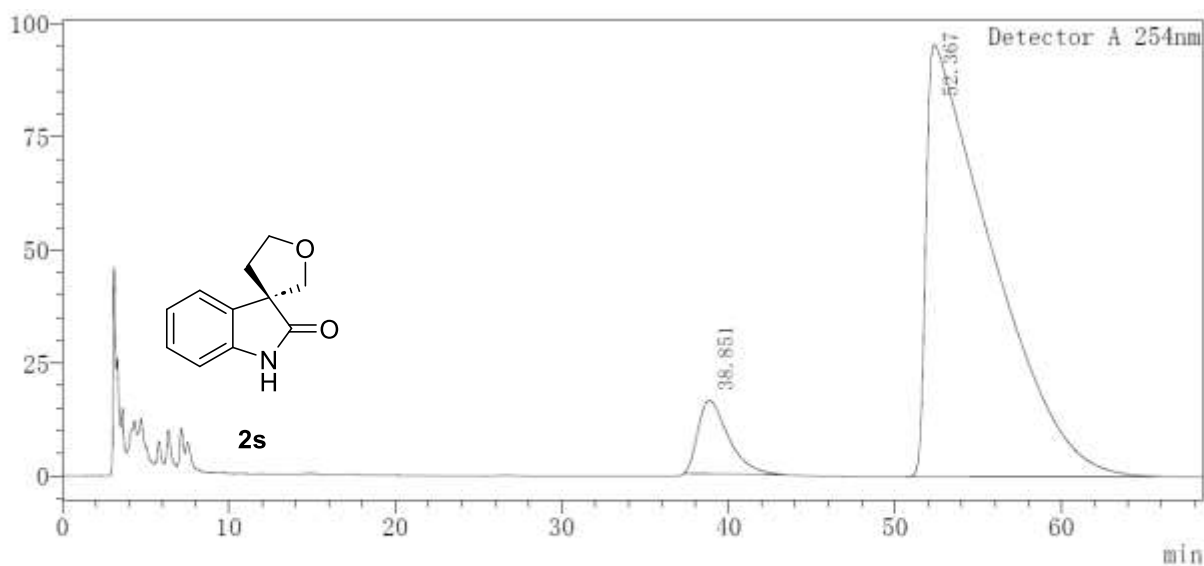
Peak	RetTime	Area	Height	Area%
1	12.928	1257853	27862	16.377
2	17.189	6422840	109475	83.623
Total		7680693	137338	



<Peak Table>

Detector A 254nm

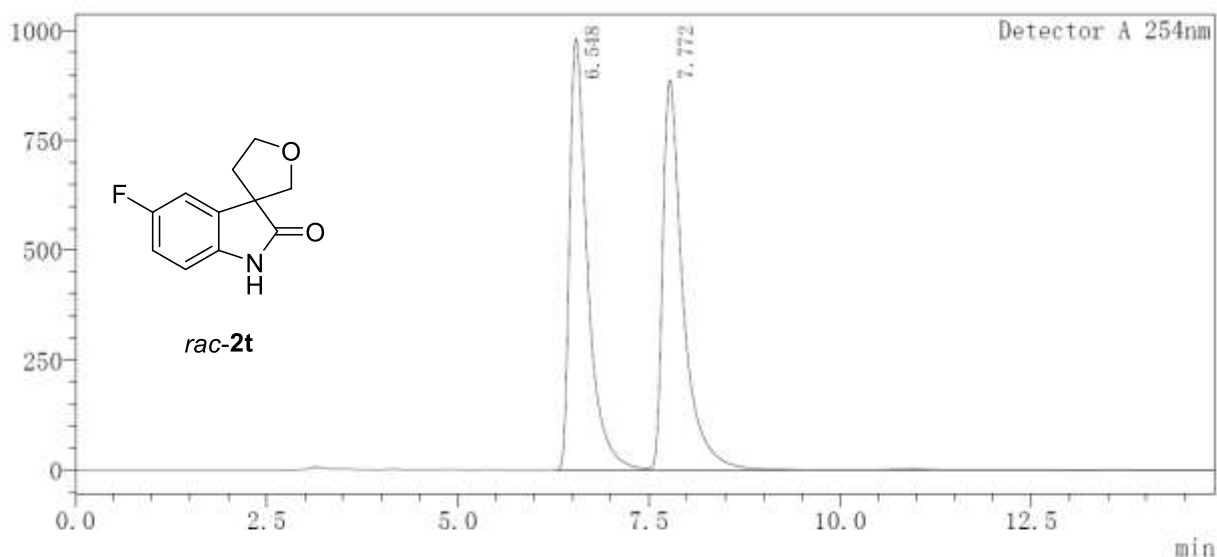
Peak	RetTime	Area	Height	Area%
1	39.491	30885888	162602	50.114
2	55.074	30744757	99868	49.886
Total		61630645	262470	



<Peak Table>

Detector A 254nm

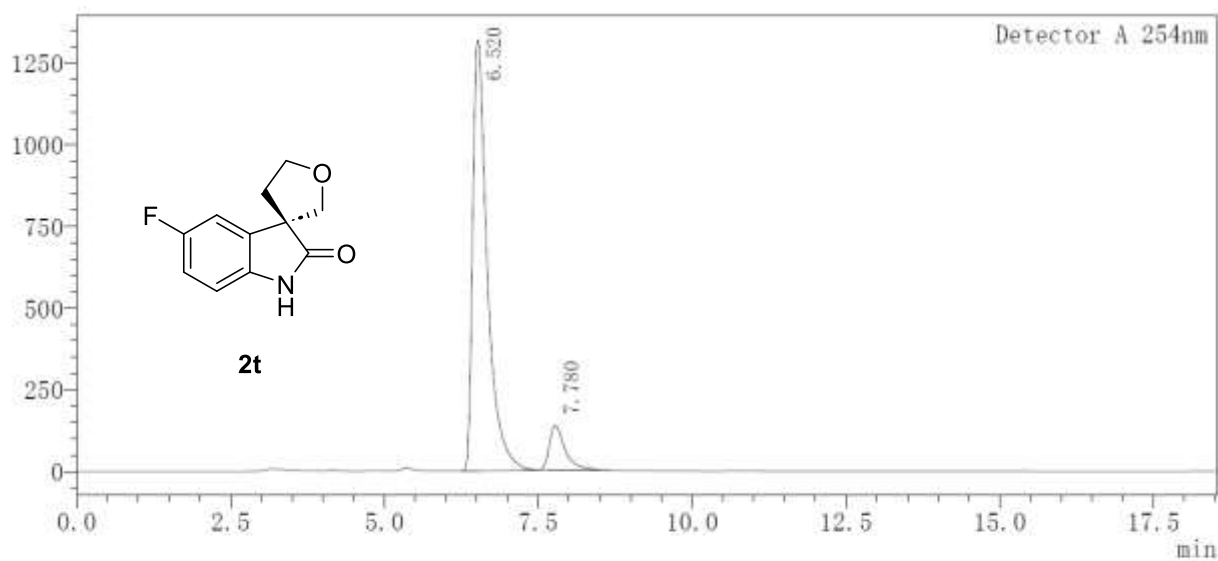
Peak	RetTime	Area	Height	Area%
1	38.851	2056003	16168	7.277
2	52.367	26195654	95511	92.723
Total		28251657	111679	



<Peak Table>

Detector A 254nm

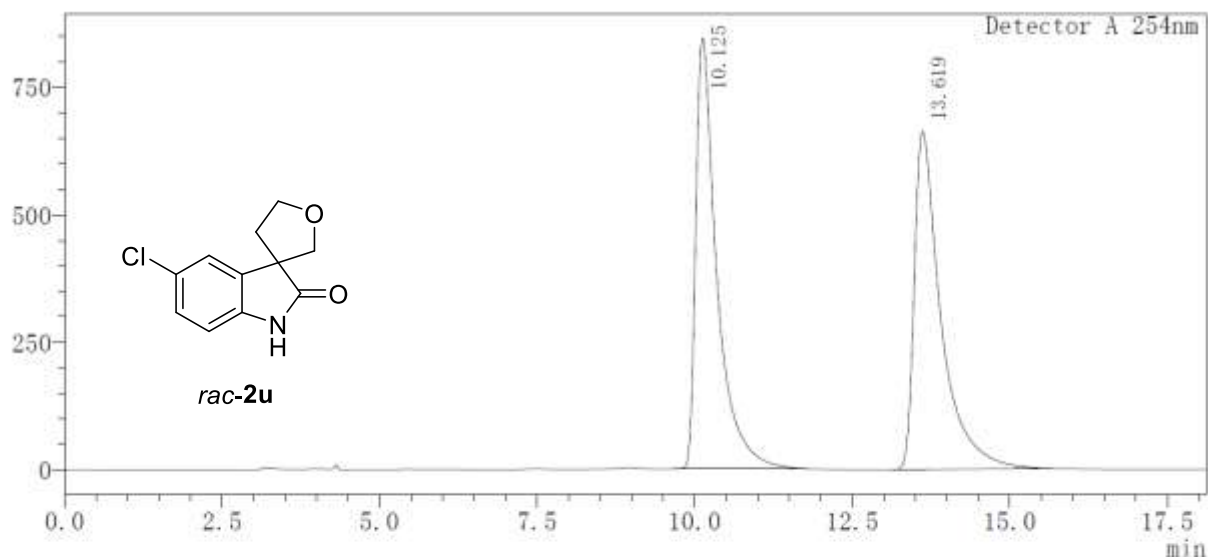
Peak	RetTime	Area	Height	Area%
1	6.548	16034306	981155	49.342
2	7.772	16462086	887183	50.658
Total		32496392	1868338	



<Peak Table>

Detector A 254nm

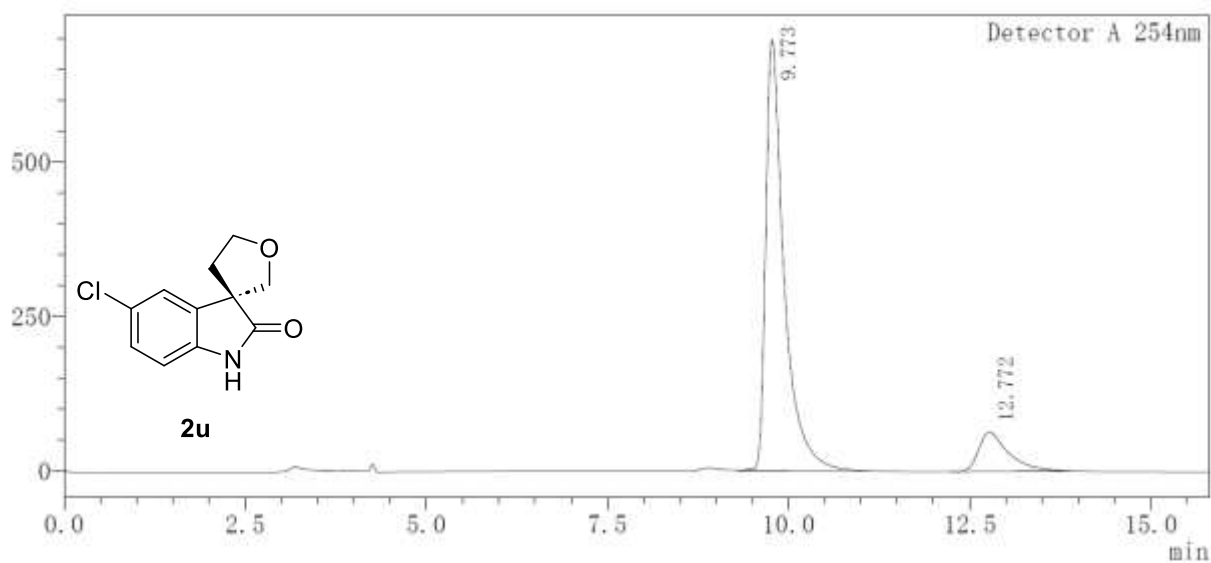
Peak	RetTime	Area	Height	Area%
1	6.520	21867056	1319331	90.021
2	7.780	2423877	136938	9.979
Total		24290933	1456269	



<Peak Table>

Detector A 254nm

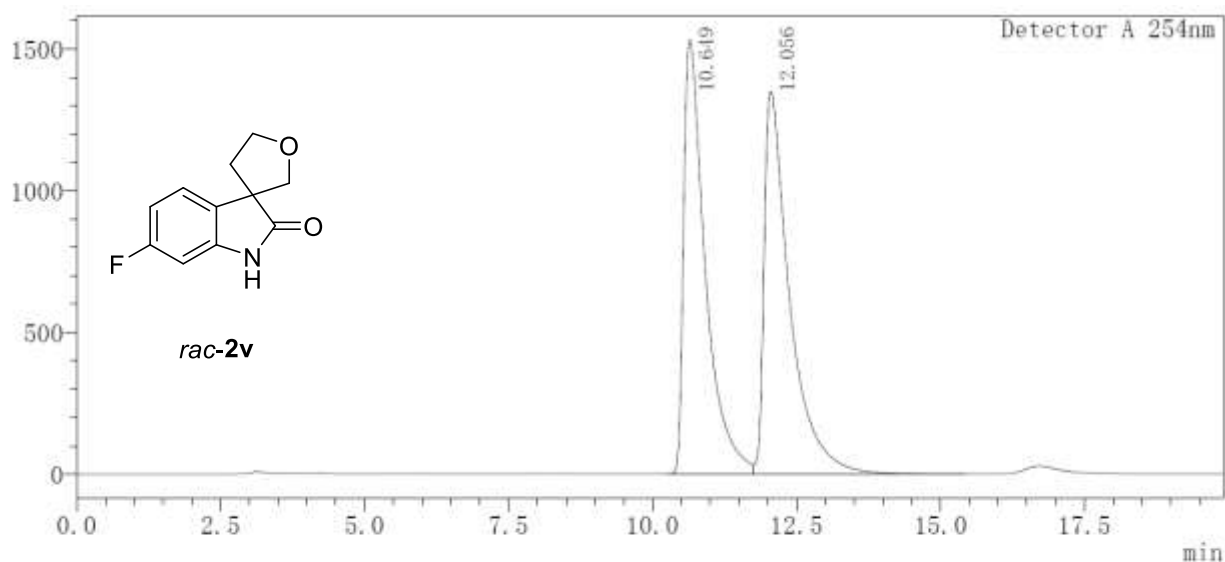
Peak	RetTime	Area	Height	Area%
1	10.125	19642796	843182	50.149
2	13.619	19526289	661816	49.851
Total		39169086	1504998	



<Peak Table>

Detector A 254nm

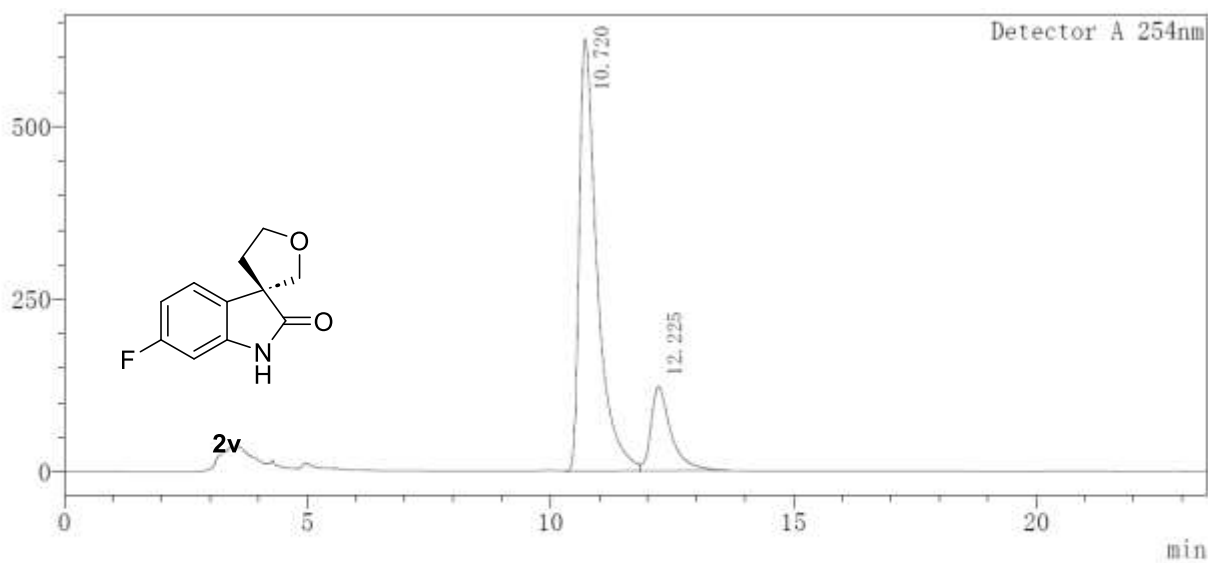
Peak	RetTime	Area	Height	Area%
1	9.773	12371237	698285	87.091
2	12.772	1833648	63610	12.909
Total		14204885	761894	



<Peak Table>

Detector A 254nm

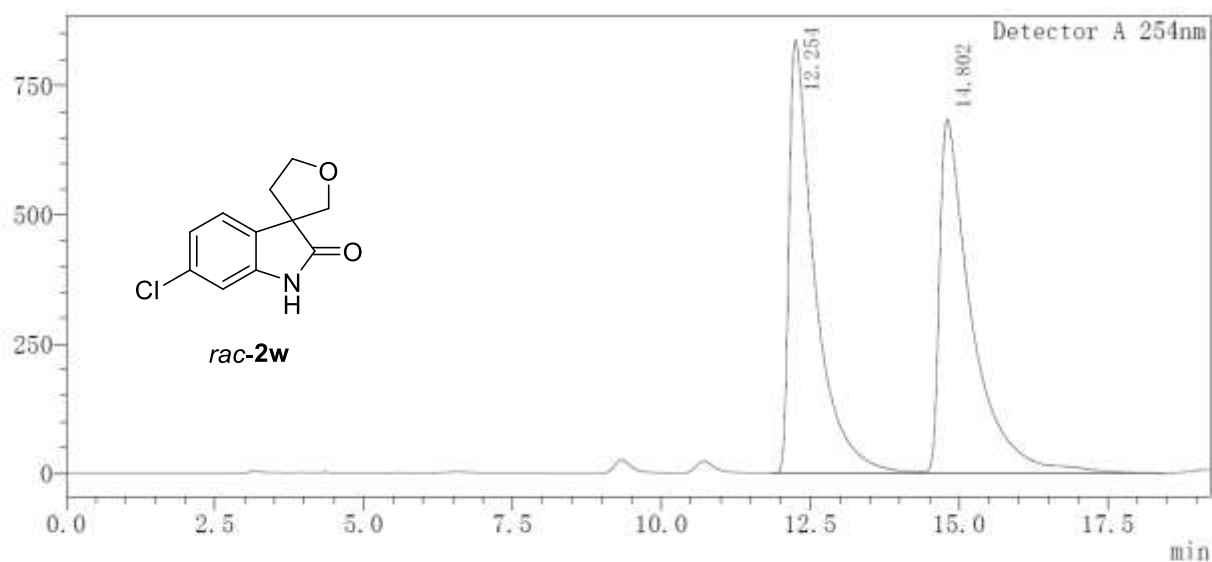
Peak	RetTime	Area	Height	Area%
1	10.649	40085219	1531116	49.040
2	12.056	41654035	1350983	50.960
Total		81739254	2882099	



<Peak Table>

Detector A 254nm

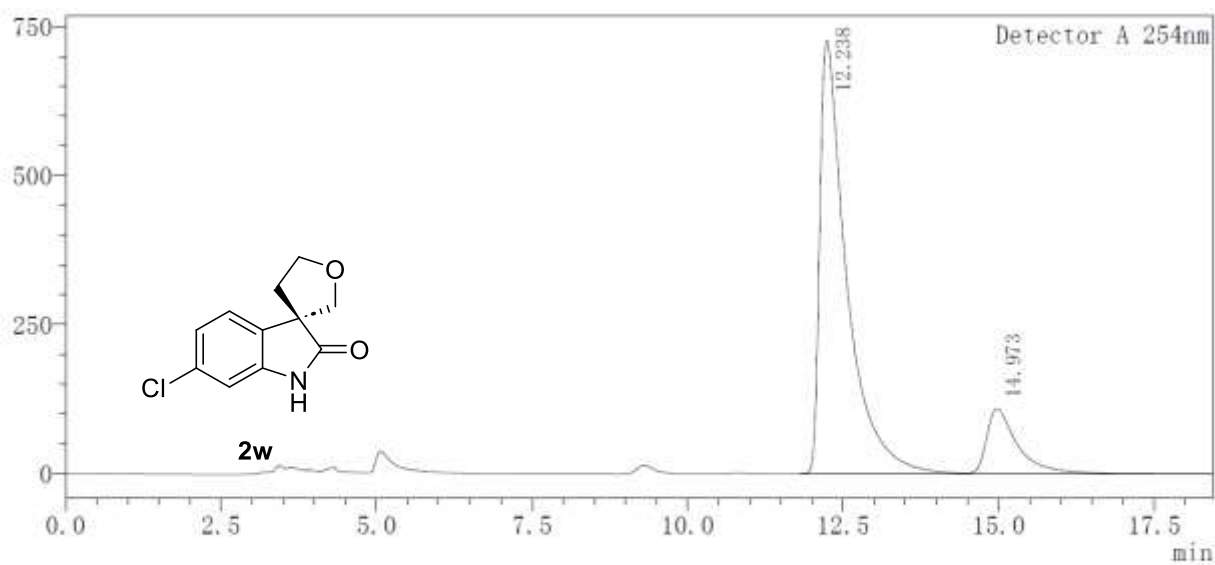
Peak	RetTime	Area	Height	Area%
1	10.720	16014361	626116	82.017
2	12.225	3511224	122174	17.983
Total		19525585	748290	



<Peak Table>

Detector A 254nm

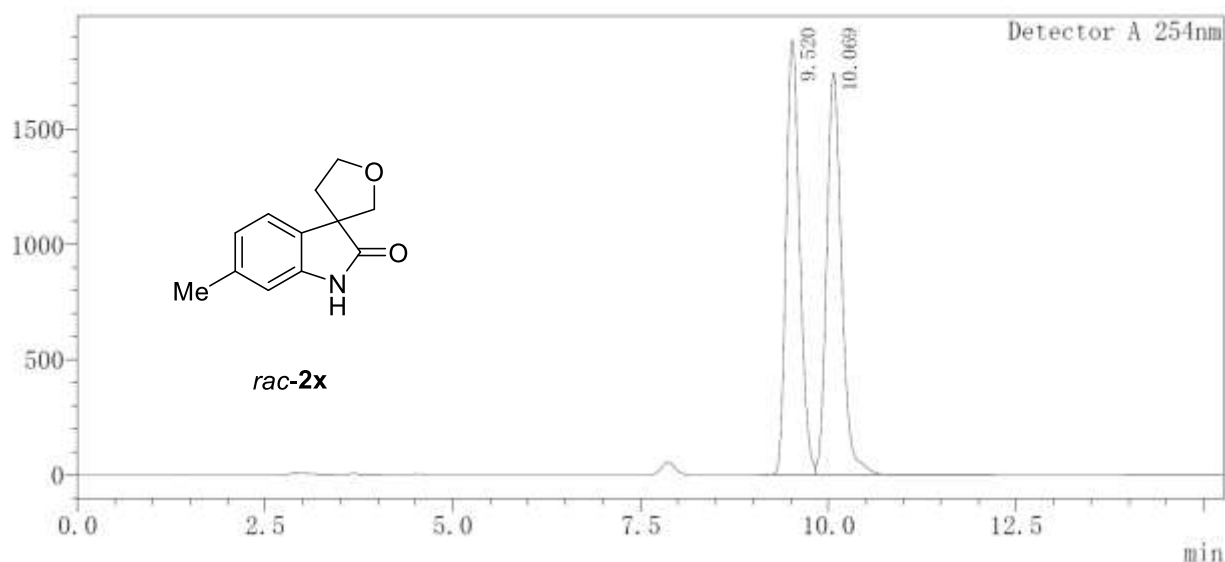
Peak	RetTime	Area	Height	Area%			
1	12.254	25102377	837638	49.448			
2	14.802	25663284	684648	50.552			
Total		50765661	1522287				



<Peak Table>

Detector A 254nm

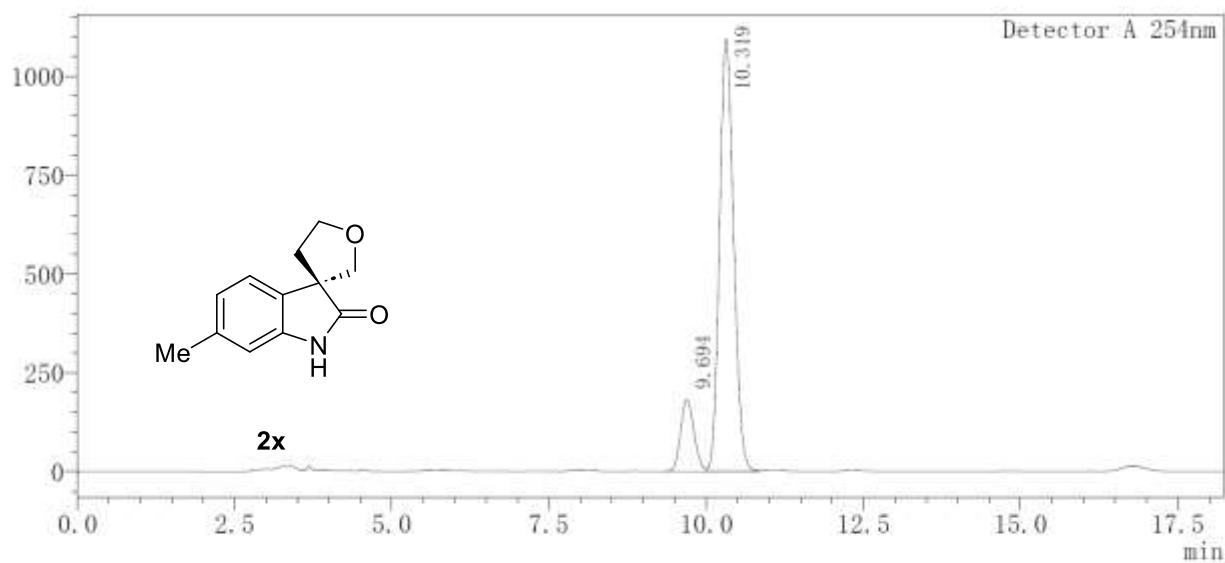
Peak	RetTime	Area	Height	Area%			
1	12.238	21983496	727769	85.020			
2	14.973	3873479	109742	14.980			
Total		25856975	837511				



<Peak Table>

Detector A 254nm

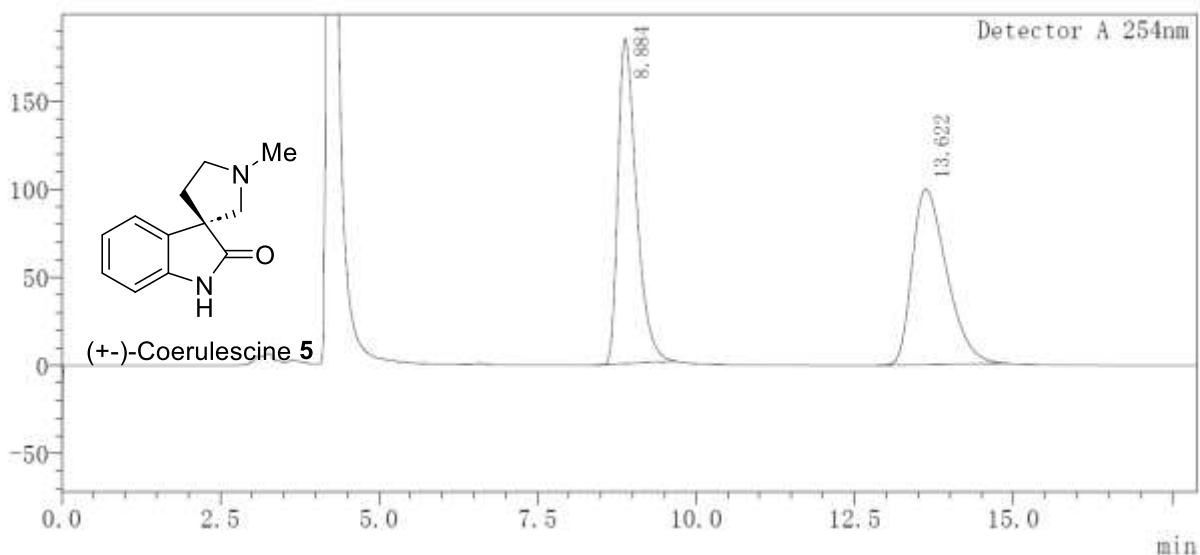
Peak	RetTime	Area	Height	Area%
1	9.520	23740196	1884112	49.505
2	10.069	24215246	1744193	50.495
Total		47955442	3628306	



<Peak Table>

Detector A 254nm

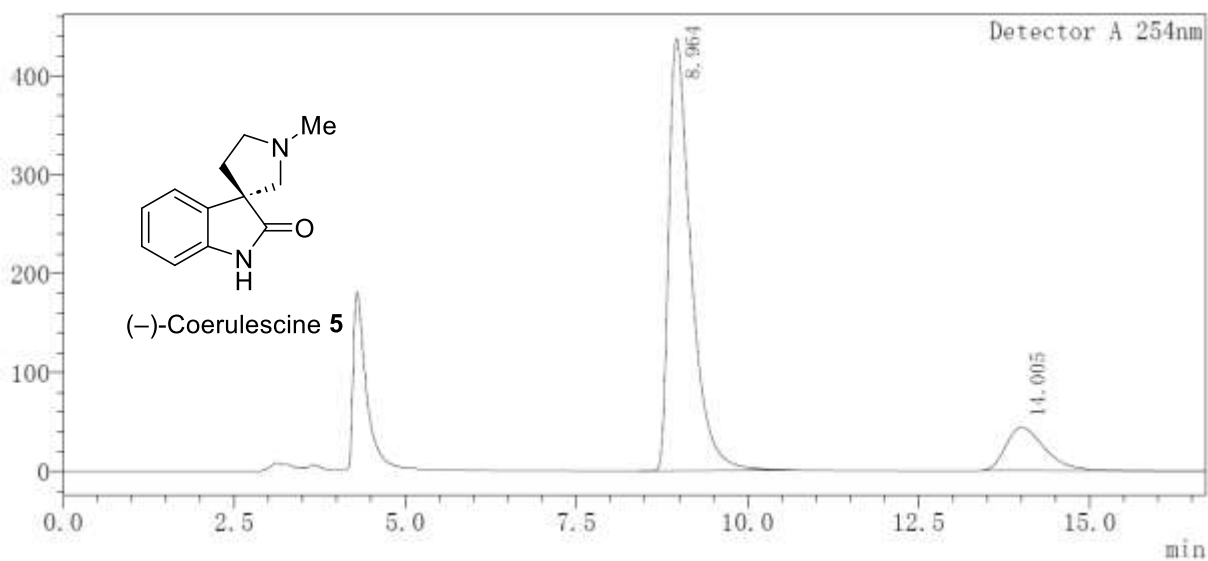
Peak	RetTime	Area	Height	Area%
1	9.694	2708156	182664	13.660
2	10.319	17118003	1095108	86.340
Total		19826159	1277772	



<Peak Table>

Detector A 254nm

Peak	RetTime	Area	Height	Area%
1	8.884	3743974	184690	49.791
2	13.622	3775353	99574	50.209
Total		7519327	284264	



<Peak Table>

Detector A 254nm

Peak	RetTime	Area	Height	Area%
1	8.964	9854734	437217	85.085
2	14.005	1727422	43586	14.915
Total		11582156	480803	