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Supporting Information

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Thermo-Responsive Water Purification: A Thermo-Switchable Molecular Brush for Precision Engineering of Antibacterial ZnO on Agro-Waste Filters

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16 **Text S1 Effect of PNAGA on the growth of ZnO NPs Crystal Facets**

17 To further corroborate the facet-selective growth, XRD analysis was performed.
18 The relative exposure of crystal facets in ZnO NPs is quantitatively reflected by the
19 intensity ratio (r) = I_{100}/I_{001} .¹ Values of $r > 1$ indicate predominant growth perpendicular
20 to the [001] direction (enhanced (100) facet exposure), characteristic of flower-like
21 needle-like morphologies, whereas $r < 1$ signifies preferential elongation along [001]
22 (greater (001) facet exposure), consistent with needle-like habits.

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24 **Text S2 Supplementary Experimental Methods**

25 2.1. Calculation of crystal size and crystallinity index

26 Crystalline phases within the samples were identified with an X'Pert-3 powder
27 diffractometer (PANalytical, Netherlands). Crystallite sizes (D) perpendicular to the
28 (002) lattice plane of cellulose I and the (101) plane of ZnO NPs were calculated using
29 Scherrer's equation (S1):²

$$30 \quad D = \frac{k\lambda}{\beta_{1/2}\cos\theta} \quad (S1)$$

31 where $k=0.943$, $\lambda=0.154056$ nm, and $\beta_{1/2}$ represents the full width at half-maximum
32 (4.21×10^{-2} rad for cellulose (002) and 7.80×10^{-3} rad for ZnO (101)).

33 The crystallinity index (CrI) of cellulose I was determined from peak
34 deconvolution according to Eq. (S2):³

$$35 \quad CrI(\%) = \frac{I_{002} - I_{am}}{I_{002}} \times 100\% \quad (S2)$$

36 where I_{002} is the intensity of the (002) peak at $2\theta=22.3^\circ$ and I_{am} is the minimum intensity
37 near $2\theta=18^\circ$. XRD patterns were analyzed using Jade 6.0 software without background
38 subtraction.

39 2.2. Computational details

40 Density functional theory (DFT) calculations were performed using the DMol³
41 module in Materials Studio to investigate the temperature-dependent interactions

42 between PNAGA segments and ZnO surfaces. The exchange–correlation interaction
43 was described using the Perdew–Burke–Ernzerhof functional within the generalized
44 gradient approximation (GGA-PBE). Ultrasoft pseudopotentials were used to describe
45 the core–electron interactions. The plane-wave cutoff energy was set to 400 eV, and
46 geometry optimization was performed until the residual force on each atom was less
47 than 0.03 eV Å⁻¹.⁴

48 The bulk hexagonal wurtzite ZnO structure was first optimized, and the optimized
49 lattice parameters were then used to construct ZnO surface slab models. The non-polar
50 ZnO-(100) and polar ZnO-(001) surfaces were modeled using periodic slab models. A
51 vacuum layer of 15 Å was introduced along the surface-normal direction to avoid
52 artificial interactions between neighboring periodic slabs.^{5,6} A Monkhorst-Pack k-point
53 mesh of 2 × 2 × 1 was used for the ZnO surface calculations.⁶ During geometry
54 optimization, the bottom layer of the ZnO slab was fixed to maintain the bulk-like
55 structure, whereas the upper surface atoms and PNAGA segments were fully relaxed.
56 For the polar ZnO-(001) surface, dipole correction was applied along the surface-
57 normal direction.

58 To represent the thermoresponsive states of PNAGA, two PNAGA segment
59 models were constructed. The low-temperature PNAGA model was built in a hydrogen-
60 bonded/collapsed conformation, in which part of the amide groups were involved in
61 intra/intermolecular hydrogen bonding. The high-temperature PNAGA model was built
62 in a hydrogen-bond-dissociated/extended conformation, exposing more amide carbonyl
63 and N–H groups toward the ZnO surface. Before adsorption calculations, the isolated
64 PNAGA segments and ZnO slab models were fully optimized using the same
65 calculation parameters. The adsorption energy was calculated according to:

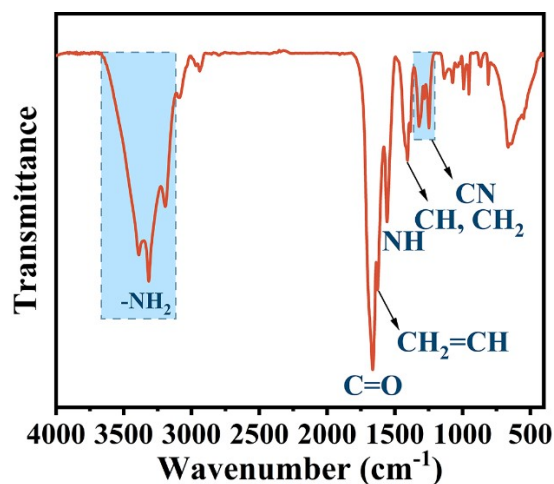
$$66 \quad E_{ads} = E_{\text{PNAGA/ZnO}} - E_{\text{PNAGA}} - E_{\text{ZnO}} \quad (\text{S2})$$

67 where $E_{\text{PNAGA/ZnO}}$, E_{PNAGA} , and E_{ZnO} are the total energies of the PNAGA/ZnO
68 adsorption complex, isolated PNAGA segment, and clean ZnO slab, respectively. A
69 more negative E_{ads} value indicates a stronger PNAGA-ZnO interaction.

70 Electrostatic potential (ESP) maps were generated from the optimized electronic
71 structures to visualize the polarity distribution of isolated PNAGA segments and the
72 interfacial electrostatic distribution of PNAGA/ZnO complexes. The dipole moments
73 of the isolated low-temperature and high-temperature PNAGA segments were
74 calculated from the optimized electronic structures. All ESP maps were plotted using
75 the same electrostatic potential scale for direct comparison.

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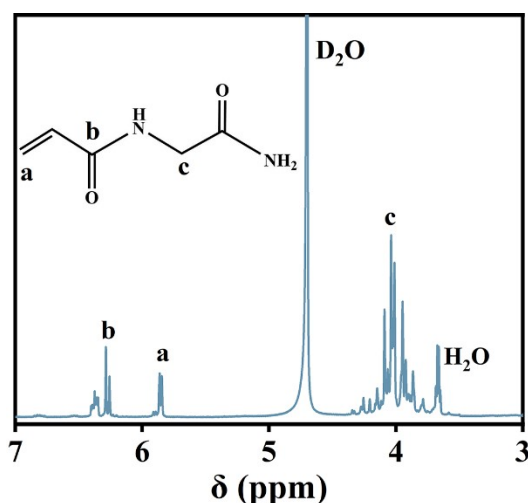
77 S3 Characterization of NAGA monomer



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79 **Figure S1** FT-IR spectra of NAGA. FT-IR spectroscopy confirmed successful synthesis of N-
80 acryloyl glycinamide, showing characteristic bands at 3390, 3317, and 3194 cm^{-1} (N-H stretching
81 with hydrogen bonding), 1662 cm^{-1} (amide I, C=O stretching), 1625 cm^{-1} (C=C stretching of vinyl
82 group), 1558 cm^{-1} (amide II), 1408 cm^{-1} (CH_2 bending), and 1258 cm^{-1} (C-N stretching).

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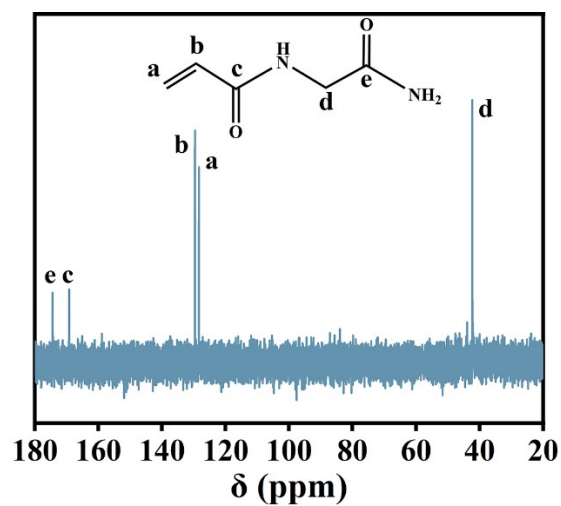


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85 **Figure S2** **Figure S1** ^1H -NMR spectra of NAGA. ^1H NMR spectroscopy confirmed the
86 successful synthesis of N-acryloyl glycinamide, displaying characteristic signals at δ 3.9 ppm (2H,
87 $\text{NH-CH}_2\text{-CONH}$), 5.8 ppm (2H, $\text{CH}=\text{CH}_2$), 6.2-6.3 ppm (1H, $\text{CH}=\text{CH}_2$), along with a residual
88 solvent peak at 4.7 ppm. The integrated peak areas were consistent with the expected molecular
89 structure.

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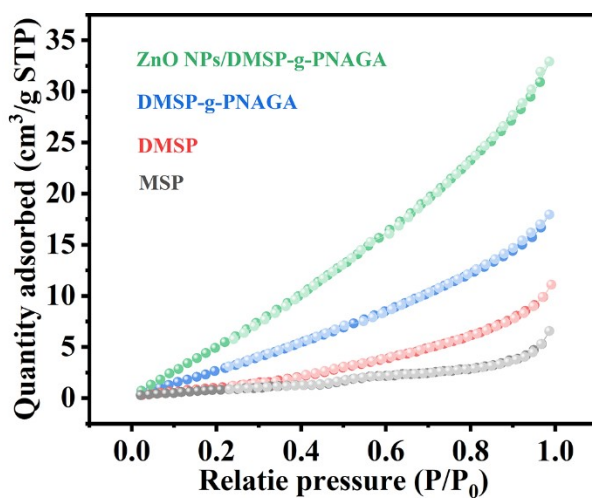
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93 **Figure S3** ¹³C-NMR spectra of NAGA. ¹³C NMR spectroscopy confirmed the structure of N-
 94 acryloyl glycinamide, with characteristic signals at δ 42.3 ppm (-NH-CH₂-CO-), 128.3 and 129.6
 95 ppm (CH=CH₂), 169.2 ppm (-CO-NH- acryloyl carbonyl), and 174.4 ppm (-CONH- glycinamide
 96 carbonyl), consistent with the expected molecular structure.

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99 S4 The microstructure of the composite column



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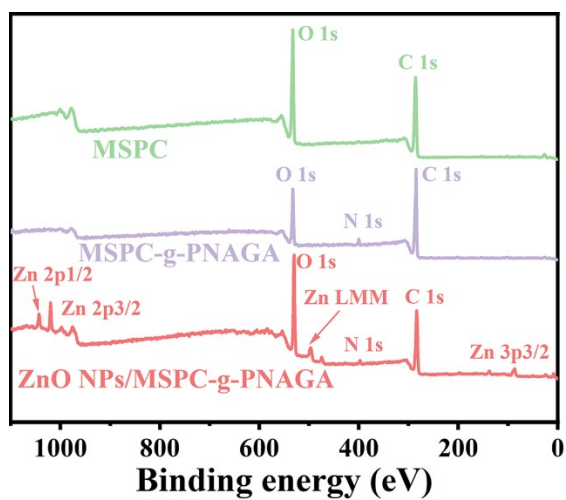
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Figure S4 Nitrogen adsorption isotherms of MSP, DMSP, DMSP-g-PNAGA and ZnO NPs/DMSP-g-PNAGA. N₂ adsorption-desorption isotherms revealed that both samples exhibited Type II isotherm behavior characteristic of macroporous structures, featuring reversible adsorption-desorption cycles at high relative pressures without a distinct plateau.

106 S5 XPS survey spectra of the composite column

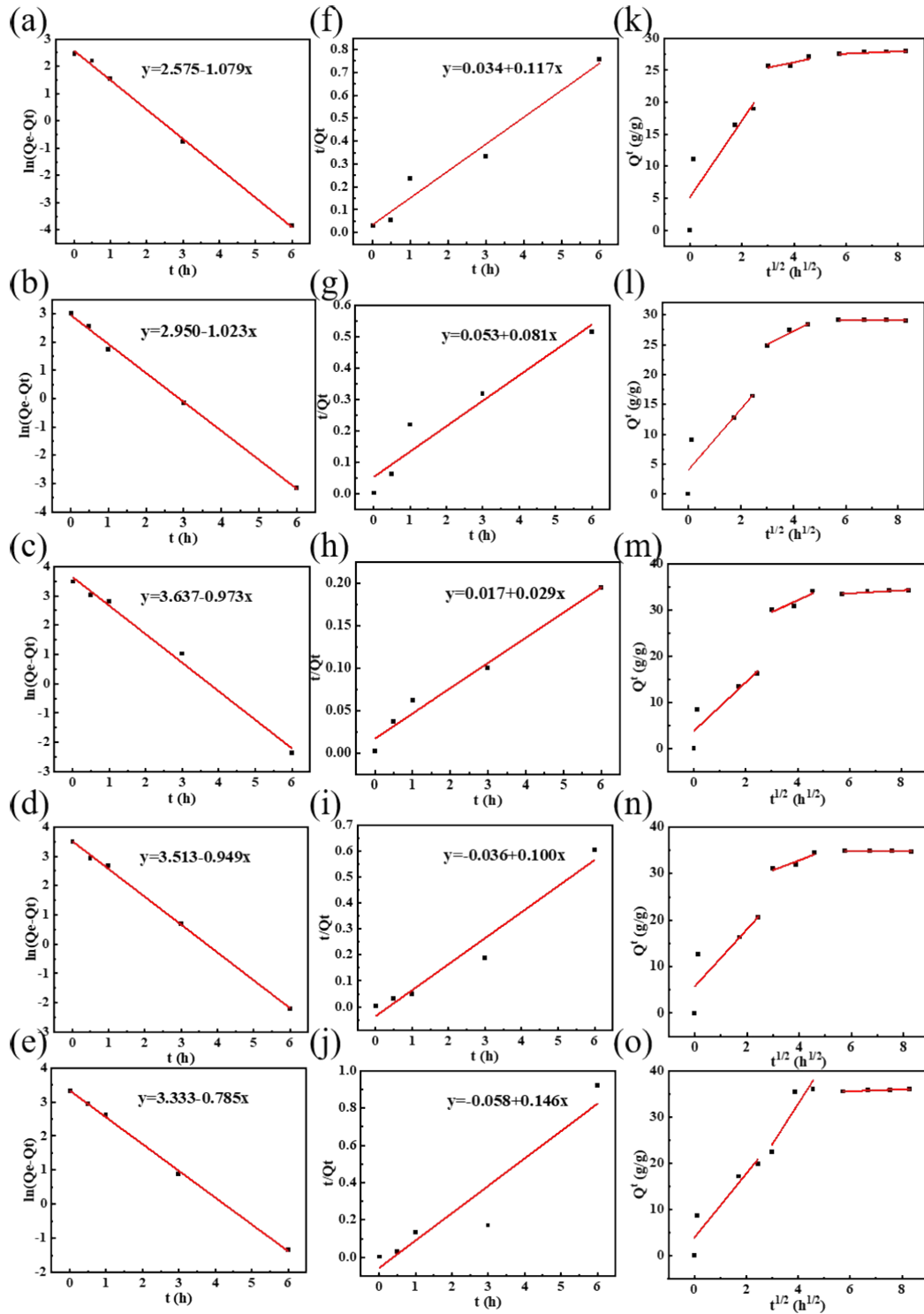


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108 **Figure S5** XPS survey spectra of DMSP, DMSP-g-PNAGA and ZnO NPs/DMSP-g-
109 PNAGA.

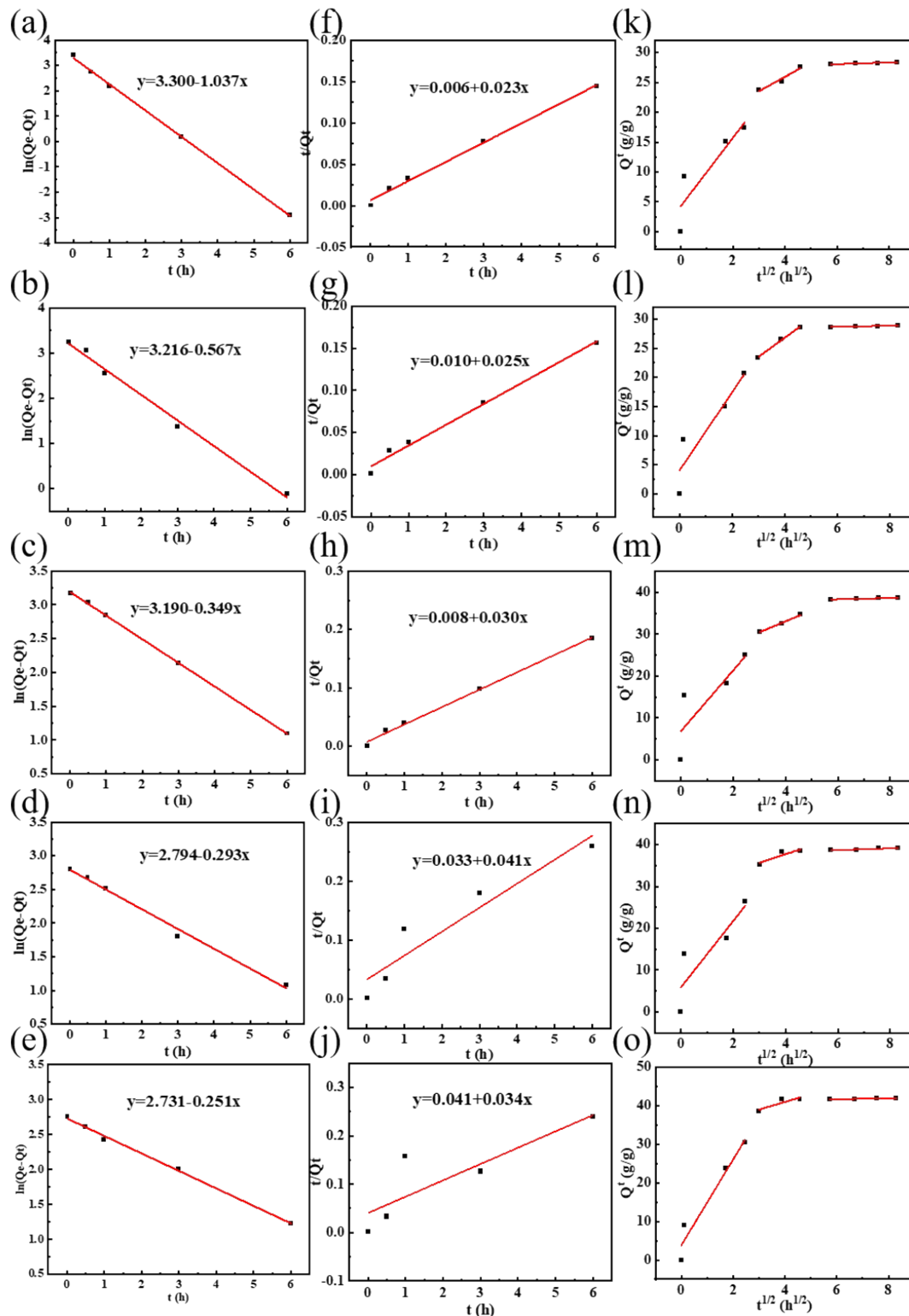
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111 S6 Pseudo-first-order, pseudo-second-order, and intraparticle diffusion models



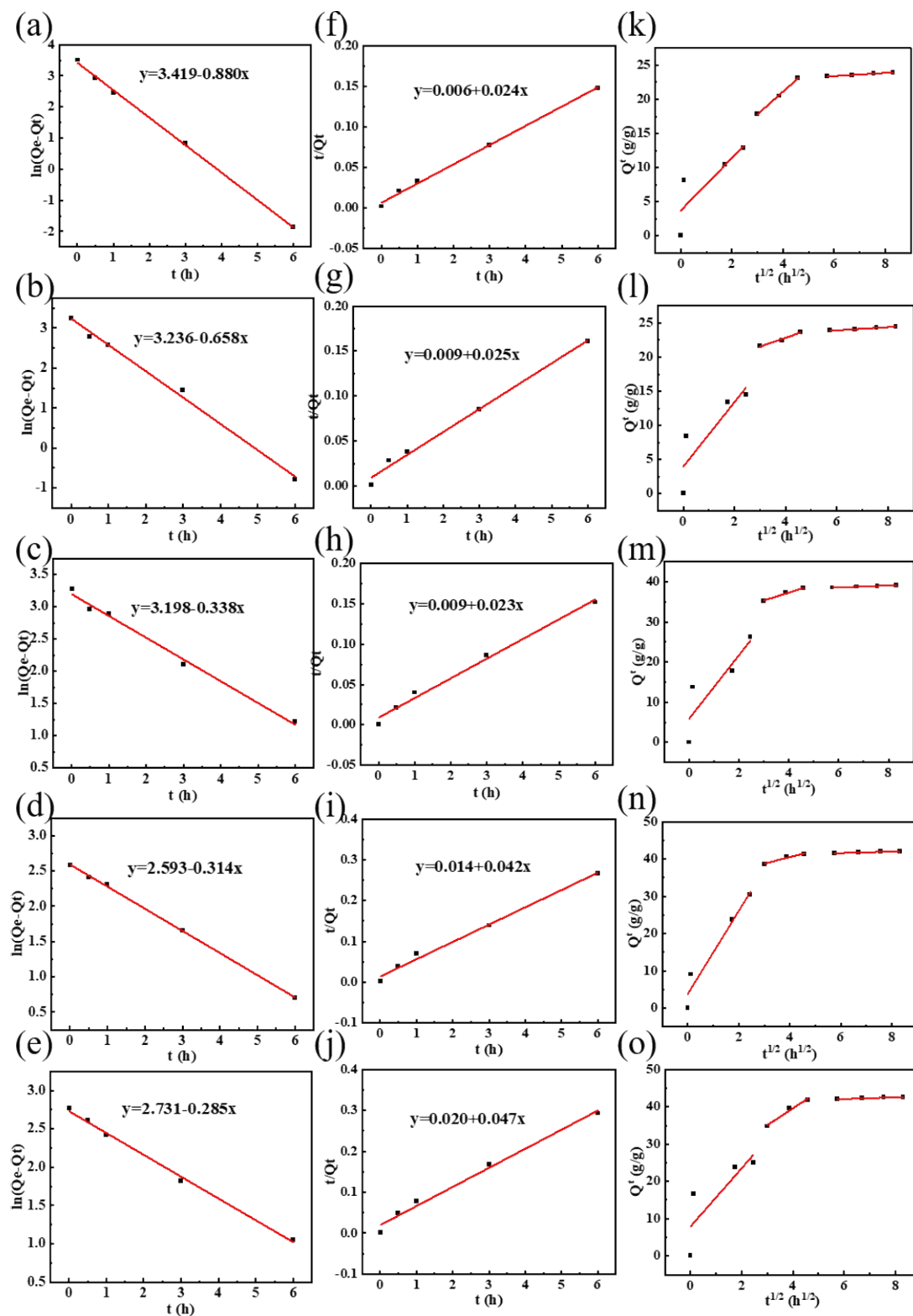
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Figure S6 Fitting curves for the pseudo-first-order, pseudo-second-order, and intra-particle diffusion models of DMSP-g-PNAGA1:1



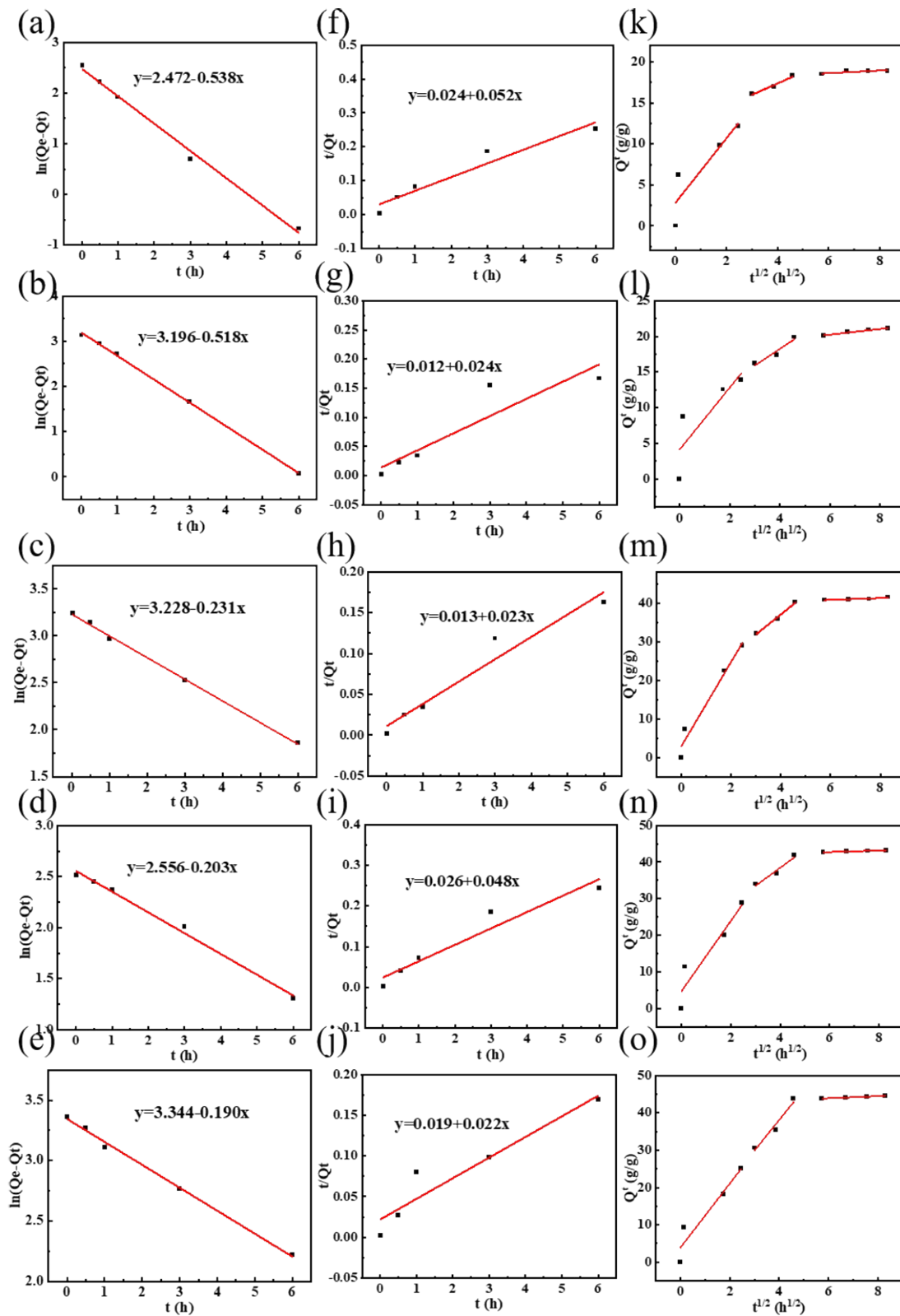
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Figure S7 Fitting curves for the pseudo-first-order, pseudo-second-order, and intra-particle diffusion models of DMSF-g-PNAGA100:1



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Figure S8 Fitting curves for the pseudo-first-order, pseudo-second-order, and intra-particle diffusion models of DMSP-g-PNAGA-T65



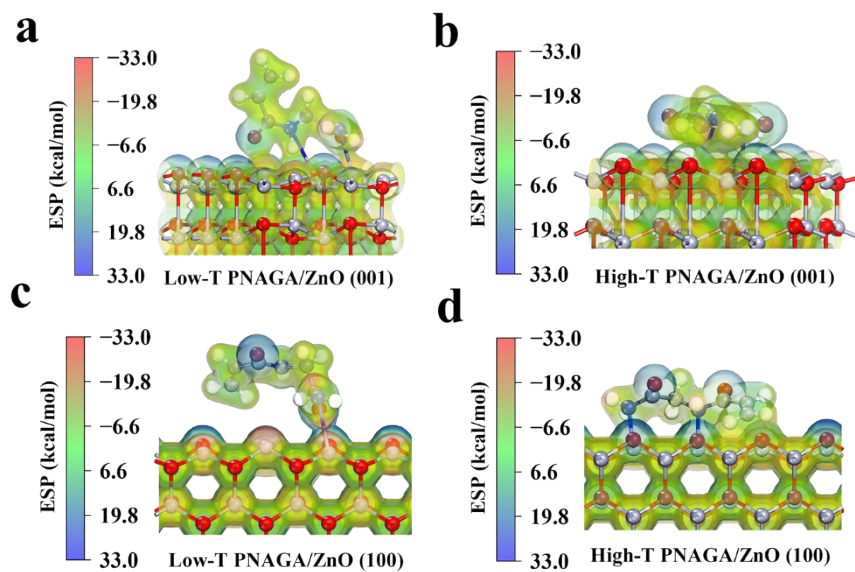
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Figure S9 Fitting curves for the pseudo-first-order, pseudo-second-order, and intra-particle diffusion models of DMSP-g-PNAGA-t21

135 S7 ESP maps of PNAGA/ZnO adsorption complexes



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137 **Figure S10** ESP maps of PNAGA/ZnO adsorption complexes: (a) low-T PNAGA/(100), (b)

138 high-T PNAGA/(100), (c) low-T PNAGA/(001), and (d) high-T PNAGA/(001). The high-T

139 PNAGA/(001) complex exhibits the most pronounced interfacial electrostatic complementarity,

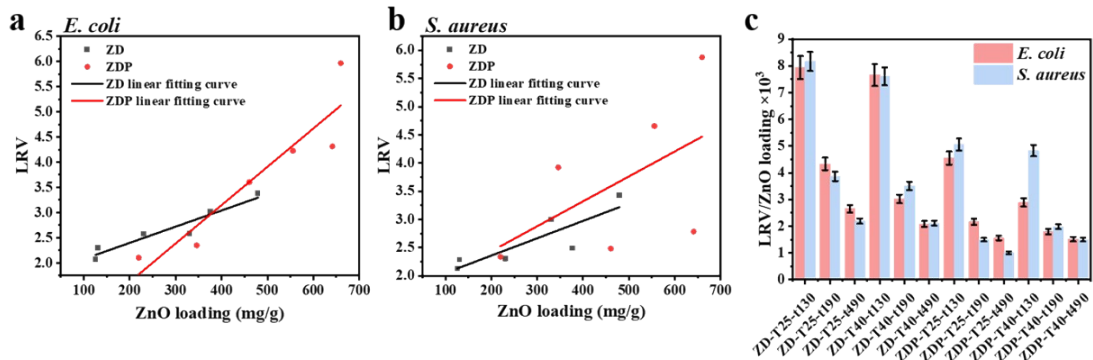
140 consistent with its strongest adsorption energy. All ESP maps were plotted using the same

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electrostatic potential scale.

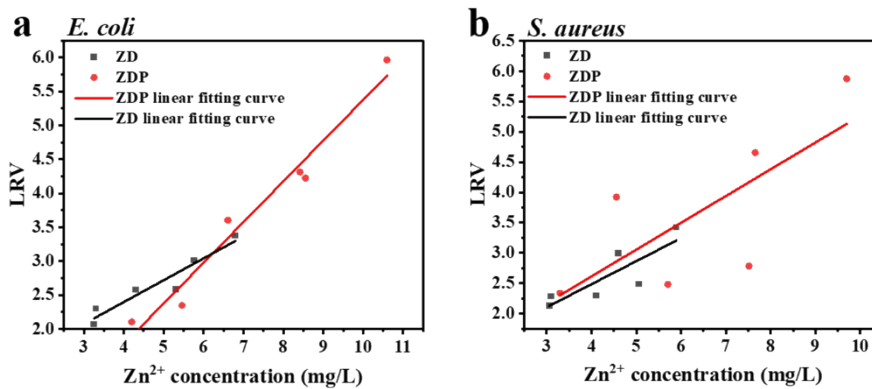
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143 **S8 Semi-quantitative analysis of the relationship between ZnO loading and**
 144 **antibacterial activity**



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 146 **Figure S11** (a, b) Scatter plots of LRV as a function of ZnO loading. (c) Apparent loading-
 147 normalized antibacterial index (LRV/ZnO loading) for *E. coli* and *S. aureus*, respectively. The
 148 fitting lines are presented solely to indicate trends and do not imply a strict linear correlation
 149 between ZnO loading and antibacterial performance.

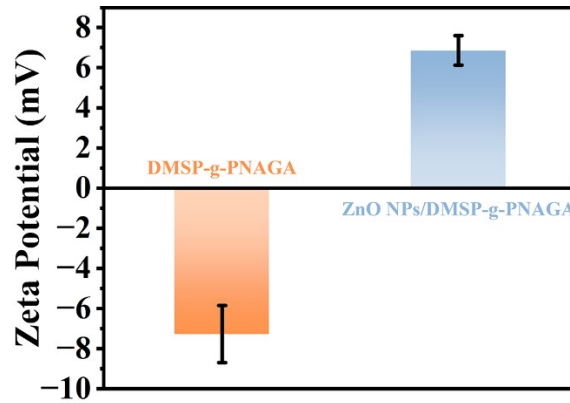
150 **S9 Correlation between Zn²⁺ release and antibacterial activity**



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 152 **Figure. S12 Correlation between Zn²⁺ release and antibacterial activity.** Scatter plots of LRV
 153 as a function of Zn²⁺ concentration at 30 min for (a) *E. coli* and (b) *S. aureus*. Each point
 154 represents one ZnO-loaded sample. Linear fitting was performed separately for the ZD and ZDP
 155 series. For *E. coli*, strong positive correlations were observed for both ZD and ZDP, with R²
 156 values of 0.9123 and 0.9653, respectively. For *S. aureus*, weaker correlations were observed, with
 157 R² values of 0.5380 and 0.7151 for ZD and ZDP, respectively. The fitting was used only for semi-
 158 quantitative trend analysis because LRV is logarithmic and Zn²⁺ release is not the sole
 159 antibacterial factor.

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161 **S10 Zeta potential of DMSP-g-PNAGA following in-situ ZnO NPs growth**



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163 **Figure S13** Zeta potentials of DMSP-g-PNAGA and ZnO NPs/DMSP-g-PNAGA. The zeta
 164 potential exhibited a substantial shift following the growth of ZnO NPs.

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166 **S11 Results of BET analysis for the isotherms used in the study**

167 **Table S1** Specific surface area and average pore size MSP, DMSP, DMSP-g-PNAGA and ZnO
 168 NPs/DMSP-g-PNAGA.

Sample	Pore size (nm)	Pore volume (cm ³ /g)	Surface area (m ² /g)
MSP	12.6 ± 0.04	0.030	3.73 ± 1.51
DMSP	13.3 ± 0.27	0.021	4.11 ± 1.51
DMSP-g-PNAGA	11.6 ± 0.06	0.012	5.26 ± 0.47
ZnO NPs/DMSP-g-PNAGA	14.1 ± 0.31	0.014	5.48 ± 0.33

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170 **S12 Pseudo-first-order, pseudo-second-order, and intraparticle diffusion models**

171 **fitting parameters**

172 **Table S2** Kinetic parameters for Water adsorption by DMSP-g-PNAGA1:1: linear pseudo-first-
 173 order and pseudo-second-order model

Model parameter	Linear fitting				
	20 °C	25 °C	30 °C	35 °C	40 °C
q_e (g/g)	28.1215	29.9491	35.0508	35.6125	37.1471
q_t (g/g)	27.9259	29.0734	34.3336	34.9146	35.0617
PFO k_1 (min ⁻¹)	1.0786	1.0227	0.9735	0.9494	0.7855
R^2	0.9980	0.9980	0.9927	0.9989	0.9990
RSS	0.0002	0.0004	0.0007	0.0002	0.0017
q_e (g/g)	29.0651	28.5593	25.1587	30.8897	24.0697
q_t (g/g)	27.9259	29.0734	34.3336	34.9146	35.0617
PSO k_2 (min ⁻¹)	0.1176	0.0808	0.0297	0.1002	0.1467
R^2	0.9664	0.9290	0.9754	0.9635	0.8962
RSS	0.0116	0.0120	0.0312	0.0451	0.1113

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176 **Table S3** Kinetic parameters for Water adsorption by DMSP-g-PNAGA1:1: intraparticle
 177 diffusion models

Parameter	Linear fitting				
	20 °C	25 °C	30 °C	35 °C	40 °C
q_e (g/g)	24.7612	24.7652	38.4325	42.0121	43.8563
q_t (g/g)	27.9259	29.0734	34.3336	34.9146	35.0617
k_{i1}	5.9947	5.0776	5.2493	6.1317	6.9258
k_{i2}	0.8453	2.2064	2.5384	2.1203	8.7645
k_{i3}	0.1717	0.0376	0.3255	0.0223	0.2110
R_{i1}^2	0.7407	0.7605	0.7963	0.7006	0.8683
R_{i2}^2	0.6877	0.9519	0.8537	0.8918	0.8327
R_{i3}^2	0.8747	0.4702	0.7342	0.9681	0.9401

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Table S4 Kinetic parameters for Water adsorption by DMSP-g-PNAGA100:1: linear

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pseudo-first-order and pseudo-second-order model

Model parameter	Linear fitting				
	20 °C	25 °C	30 °C	35 °C	40 °C
q_e (g/g)	28.7612	28.7652	38.4325	39.0121	41.8563
q_t (g/g)	28.4085	28.9051	38.6731	39.0982	41.8982
PFO k_1 (min ⁻¹)	1.0372	0.5673	0.3492	0.2931	0.2506
R^2	0.9990	0.9928	0.9998	0.9925	0.9969
RSS	0.0021	0.0101	0.0001	0.0114	0.0146
q_e (g/g)	27.7561	29.6151	38.3121	39.6451	30.1231
q_t (g/g)	28.4085	28.9051	38.6731	39.0982	41.8982
PSO k_2 (min ⁻¹)	0.0233	0.0248	0.0298	0.0407	0.0337
R^2	0.9948	0.9911	0.9962	0.9014	0.7466
RSS	0.0109	0.0134	0.0098	0.0437	0.0938

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Table S5 Kinetic parameters for Water adsorption by DMSP-g-PNAGA100:1: intraparticle

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diffusion models

Parameter	Linear fitting				
	20 °C	25 °C	30 °C	35 °C	40 °C
q_e (g/g)	28.7612	28.7652	38.4325	39.0121	41.8563
q_t (g/g)	28.4085	28.9051	38.6731	39.0982	41.8982
k_{i1}	5.7246	6.6913	7.2782	7.9234	11.1364
k_{i2}	2.4154	3.2817	2.6042	2.0528	1.9981
k_{i3}	0.1409	0.0914	0.1511	0.2031	0.0625
R_{i1}^2	0.7906	0.8414	0.6871	0.7604	0.9479
R_{i2}^2	0.9439	0.9964	0.9827	0.8288	0.8204
R_{i3}^2	0.9801	0.9698	0.9302	0.9468	0.9776

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Table S6 Kinetic parameters for Water adsorption by DMSP-g-PNAGA-T65: linear pseudo-first-order and pseudo-second-order model

Model parameter	Linear fitting				
	20 °C	25 °C	30 °C	35 °C	40 °C
q_e (g/g)	24.7612	24.7652	38.4325	42.0121	43.8563
q_t (g/g)	23.9464	24.4610	39.1627	42.0631	42.6428
PFO k_1 (min ⁻¹)	0.8803	0.6583	0.3380	0.3141	0.2849
R^2	0.9989	0.9946	0.9926	0.9993	0.9958
RSS	0.0015	0.0164	0.0066	0.0016	0.0082
q_e (g/g)	27.7561	29.6151	38.3121	39.6451	30.1231
q_t (g/g)	23.9464	24.4610	39.1627	42.0631	42.6428
PSO k_2 (min ⁻¹)	0.0237	0.0254	0.0243	0.0424	0.0466
R^2	0.9970	0.9922	0.9900	0.9923	0.9885
RSS	0.0034	0.0107	0.0141	0.0183	0.0439

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Table S7 Kinetic parameters for Water adsorption by DMSP-g-PNAGA-T65: intraparticle diffusion models

Parameter	Linear fitting				
	20 °C	25 °C	30 °C	35 °C	40 °C
q_e (g/g)	24.5681	24.6452	38.9984	42.1561	43.2481
q_t (g/g)	23.9464	24.4610	39.1627	42.0631	42.6428
k_{i1}	3.8300	4.7167	7.9235	11.1364	7.8191
k_{i2}	3.3237	1.2822	1.9749	1.7534	4.4941
k_{i3}	0.2330	0.2338	0.2341	0.2001	0.2338
R_{i1}^2	0.6889	0.8651	0.8720	0.9479	0.6724
R_{i2}^2	0.9959	0.9893	0.9867	0.9636	0.9765
R_{i3}^2	0.9971	0.9985	0.9971	0.9903	0.9971

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Table S8 Kinetic parameters for Water adsorption by DMSP-g-PNAGA-t21: linear pseudo-first-order and pseudo-second-order model

Model parameter	Linear fitting				
	20 °C	25 °C	30 °C	35 °C	40 °C
q_e (g/g)	18.9311	21.1142	41.5184	43.1857	44.6024
q_t (g/g)	18.9482	21.0661	41.5338	43.1578	44.5714
PFO k_1 (min ⁻¹)	0.5383	0.5184	0.2306	0.2033	0.1901
R^2	0.9992	0.9977	0.9985	0.9982	0.9950
RSS	0.0008	0.0022	0.0003	0.0003	0.0009
q_e (g/g)	20.4241	23.1422	36.4251	41.6242	40.1424
q_t (g/g)	18.9482	21.0661	41.5338	43.1578	44.5714
PSO k_2 (min ⁻¹)	0.0521	0.0244	0.0239	0.0479	0.0218
R^2	0.9384	0.9365	0.8911	0.9574	0.8822
RSS	0.8992	0.6056	1.8330	0.9560	2.6112

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Table S9 Kinetic parameters for Water adsorption by DMSP-g-PNAGA-t21: intraparticle diffusion models

Parameter	Linear fitting				
	20 °C	25 °C	30 °C	35 °C	40 °C
q_e (g/g)	24.7612	24.7652	38.4325	42.0121	43.8563
q_t (g/g)	18.9482	23.0661	41.5338	43.1578	44.5714
k_{i1}	3.9394	4.3563	10.9057	9.5850	8.5803
k_{i2}	1.3882	2.3227	5.1892	4.9029	8.1826
k_{i3}	0.1644	0.3998	0.2449	0.1954	0.2809
R_{i1}^2	0.8028	0.7039	0.9654	0.8811	0.9039
R_{i2}^2	0.9796	0.9406	0.9913	0.9521	0.9559
R_{i3}^2	0.7677	0.9426	0.8267	0.9499	0.9974

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202 **S13 Linear fitting was performed using ZnO loading as the independent variable**
 203 **and LRV as the dependent variable**

204 **Table S10** Linear fitting parameters for the relationship between ZnO loading and LRV.

Bacterial strain	Series	Fitting equation	Slope	Intercept	R²
<i>E. coli</i>	ZD	y=ax+b	0.0032	1.7555	0.9159
<i>E. coli</i>	ZDP	y=ax+b	0.0076	0.1004	0.8593
<i>S. aureus</i>	ZD	y=ax+b	0.0031	1.7447	0.7522
<i>S. aureus</i>	ZDP	y=ax+b	0.0044	1.5541	0.2973

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207 **S14 Linear fitting parameters for the relationship between Zn²⁺ release and LRV**

208 **Table S11** Linear fitting parameters for the relationship between Zn²⁺ release and LRV

Bacterial strain	Series	Fitting equation	Slope	Intercept	R²
<i>E. coli</i>	ZD	y=ax+b	0.3215	1.1126	0.9123
<i>E. coli</i>	ZDP	y=ax+b	0.5999	-0.6228	0.9653
<i>S. aureus</i>	ZD	y=ax+b	0.3803	0.9645	0.7151
<i>S. aureus</i>	ZDP	y=ax+b	0.4418	0.8476	0.5380

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211 **S15 Variation in r values for multiple DMSP and DMSP-g-PNAGA samples**

212 **Table S12** r values of DMSP and DMSP-g-PNAGA samples at reaction temperatures of 25 °C
213 and 40 °C and times of 130, 190, and 490 min

Sample	r
ZD-T25-t130	1.2
ZD-T25-t190	1.1
ZD-T25-t490	1.3
ZD-T40-t130	1.1
ZD-T40-t190	1.2
ZD-T40-t490	1.2
ZDP-T25-t130	0.8
ZDP-T25-t190	0.9
ZDP-T25-t490	0.9
ZDP-T40-t130	1.1
ZDP-T40-t190	1.3
ZDP-T40-t490	1.4

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216 **Supplementary References**

- 217 1T. Tian, S. Tu, A. Xu, S. Yin, A. L. Oechsle, T. Xiao, A. Vagias, J.
218 Eichhorn, J. Suo, Z. Yang, S. Bernstorff and P. Müller-Buschbaum, *Adv. Funct.*
219 *Mater.*, DOI:10.1002/adfm.202311793.
- 220 2X. Wu, Y. Liu, M. Li, B. Li, X. Mao, Q. Wang, X. Tang, H. Zhang, L.
221 Peng and X. Gao, *Chem. Eng. J*, DOI:10.1016/j.cej.2024.153757.
- 222 3G. E. Dawwam, M. T. Al-Shemy and A. S. El-Demerdash, *Sci. Rep.*,
223 2022, **12**, 1–18.
- 224 4G. Sun, Y. Chen, X. Xia, Y. Wang, J. Zeng, J. Zhu, L. Gao, L. Peng, L.
225 Wang, X. Kong and Y. Jin, DOI:10.1021/jacs.5c15840.
- 226 5S. Wang, Z. Zhao, L. Jia, X. Guo, R. Yang, Q. Deng and R. Sun,
227 DOI:10.1021/acssensors.4c01374.
- 228 6Y. Zhang, C. Wang, F. Gong, P. Wang, U. Guharoy, C. Yang, H. Zhang,
229 S. Fang and J. Liu, *J. Hazard. Mater.*, 2020, **388**, 122069.
- 230
231