Table S1. Extraction and fractionation yields of *Caryota mitis* seed extracts

Workflow	Fraction/extract	Starting material	Fraction mass (g)	Yield from current step (%)	Overall yield from dry material (%)
A1	A1-H	100 g dry seed powder	0.1441	0.1441	0.1441
	A1-EA	100 g dry seed powder	0.3670	0.3670	0.3670
	A1-M	100 g dry seed powder	8.1384	8.1384	8.1384
A2	A2-HEa	100 g dry seed powder	0.2834	0.2834	0.2834
	A2-EA	100 g dry seed powder	0.4642	0.4642	0.4642
	A2-EaM	100 g dry seed powder	5.7830	5.7830	5.7830
	A2-M	100 g dry seed powder	1.3968	1.3968	1.3968
EtOH	CMSE96	200 g dry seed powder	18.4246	9.2123	9.2123
B1	B1-H	9 g CME96	0.0741	0.8233	0.0758
	B1-EA	9 g CME96	0.2086	2.3178	0.2135
	B1-M	9 g CME96	7.0795	78.6611	7.2465
B2	B2-HEa	9 g CME96	0.1089	1.2100	0.1115
	B2-EA	9 g CME96	0.1566	1.7400	0.1603
	B2-EaM	9 g CME96	7.0964	78.8489	7.2638
	B2-M	9 g CME96	0.3550	3.9444	0.3634

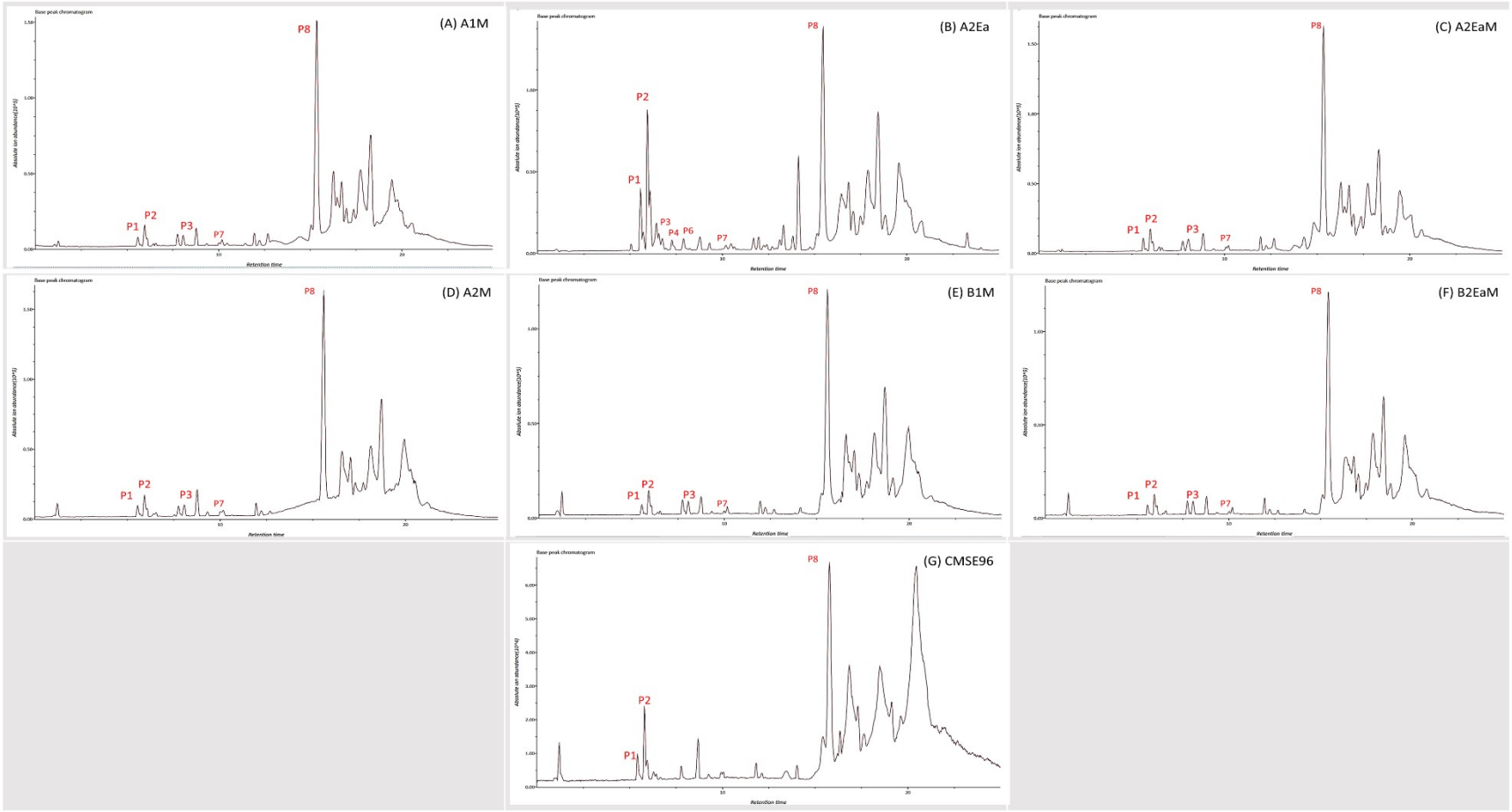

Figure S1A. Base peak chromatograms (BPCs) of the seven representative extracts analyzed by LC–MS/MS in negative ion mode. The annotated peaks were tentatively assigned as follows: P1, procyanidin C1; P2, catechin; P3, aromadendrin-O-glucoside; P4, isoquercitrin/quercetin-O-hexoside; P5, astragalin; P6, piceatannol; P7, naringenin; and P8, 6β,8β-dihydroxyeremophilenolide. All annotations are tentative and were based on accurate precursor ions and MS/MS fragmentation patterns.

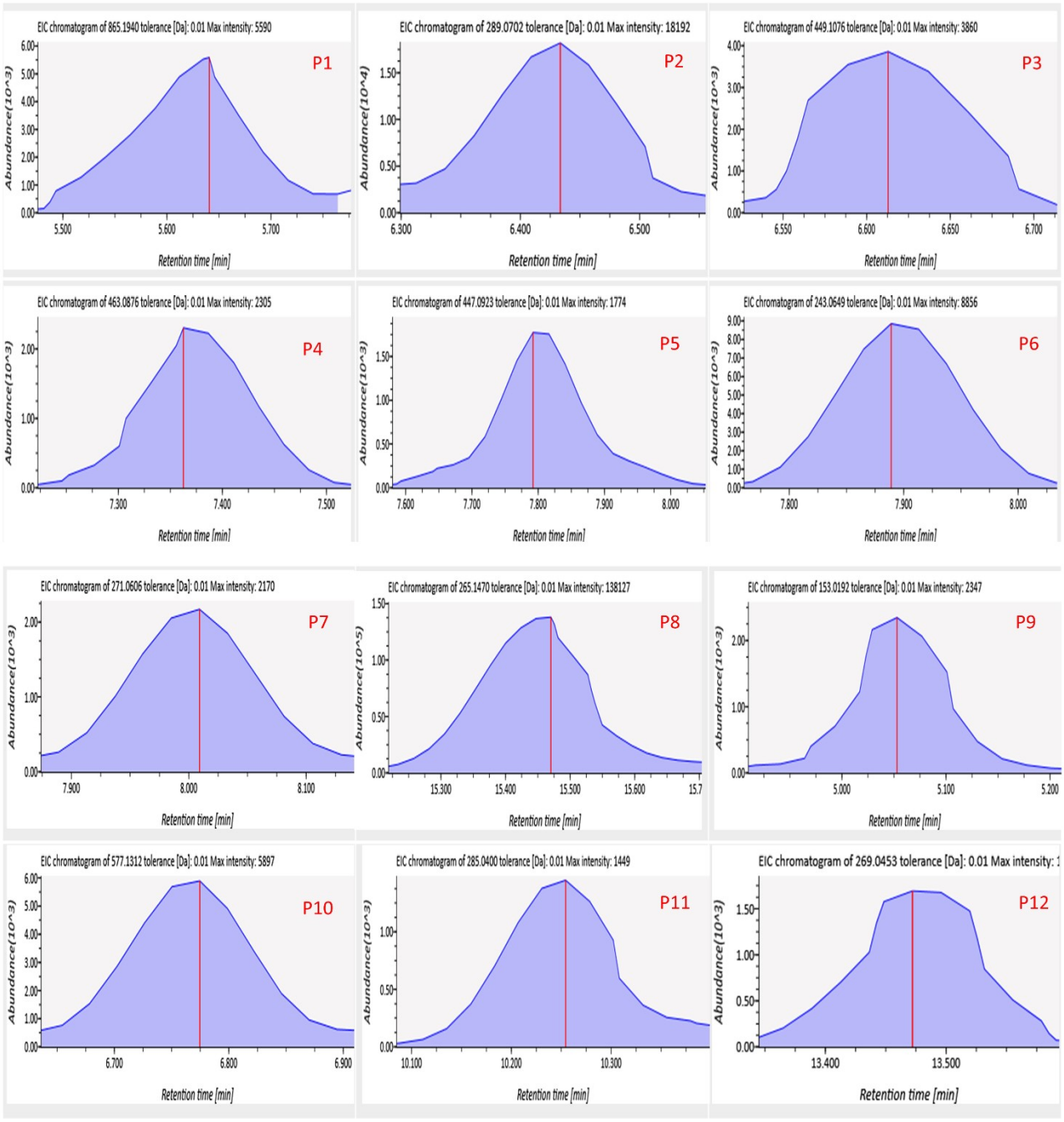

Figure S1B. Extracted ion chromatograms (EICs) of twelve representative compounds/features tentatively annotated by LC–MS/MS in negative ion mode. The peaks corresponding to those annotated in Figure S1A were labeled consistently as P1, procyanidin C1; P2, catechin; P3, aromadendrin-O-glucoside; P4, isoquercitrin/quercetin-O-hexoside; P5, astragalin; P6, piceatannol; P7, naringenin; and P8, 6β,8β-dihydroxyeremophilenolide. Additional representative EICs were labeled as P9, protocatechuic acid; P10, procyanidin B-type dimer isomer (B2-like); P11, kaempferol; and P12, apigenin. All annotations are tentative and were based on accurate precursor ions and MS/MS fragmentation patterns.

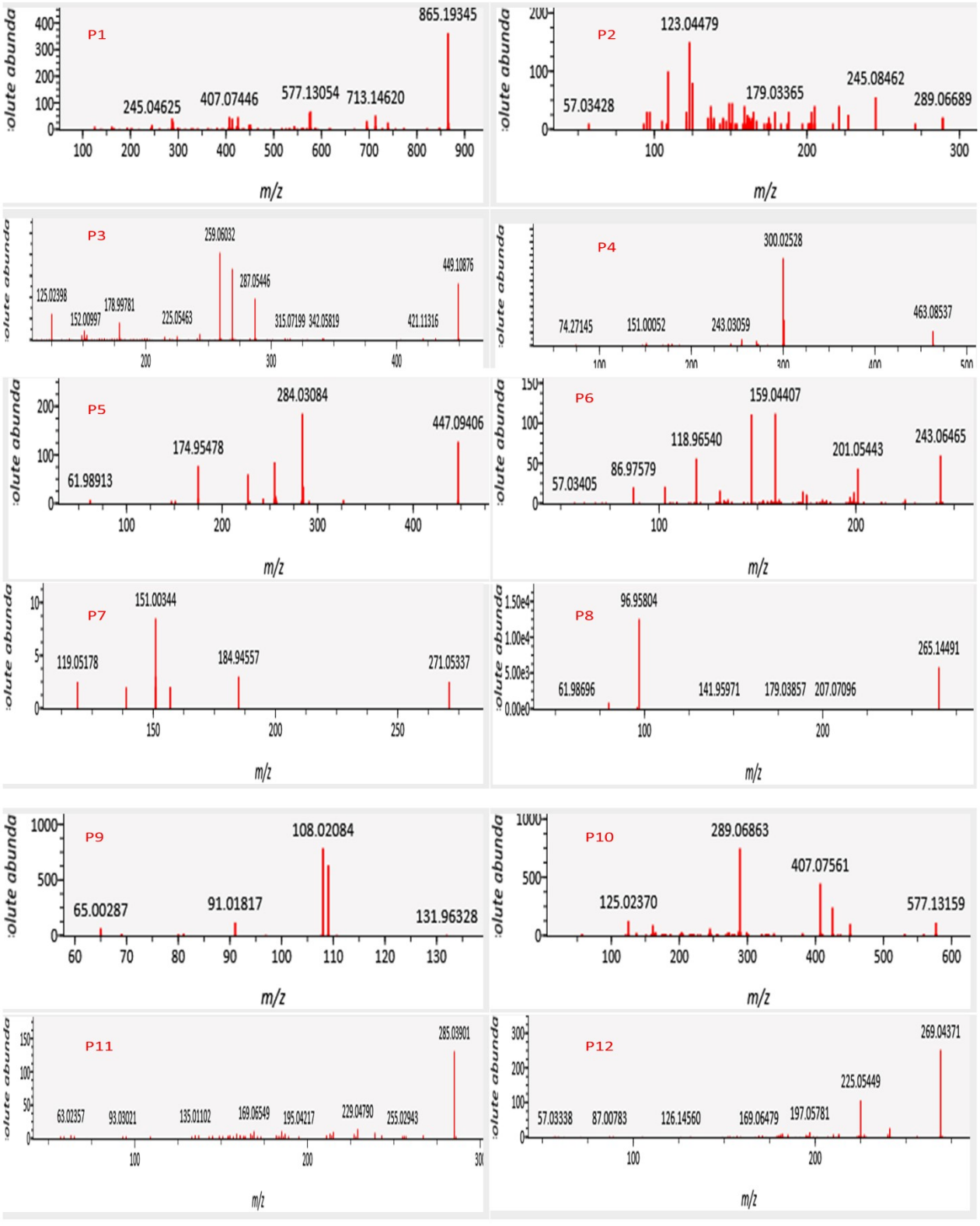
P1
865.19345
245.04625
407.07446
577.13054
713.14620
P2
123.04479
57.03428
179.03365
245.08462
289.06689
P3
259.06032
125.02398
152.00897
178.99781
225.05463
287.05446
315.07199
342.05819
421.11316
449.10876
P4
300.02528
74.27145
151.00052
243.03059
463.08537
P5
284.03084
174.95478
61.98913
447.09406
P6
159.04407
118.96540
86.97579
57.03405
201.05443
243.06465
P7
151.00344
119.05178
184.94557
271.05337
P8
96.95804
61.98696
141.95971
179.03857
207.07096
265.14491
P9
108.02084
65.00287
91.01817
131.96328
P10
289.06863
407.07561
125.02370
577.13159
P11
285.03901
63.02357
93.03021
135.01102
169.06549
195.04217
229.04790
255.02943
P12
269.04371
225.05449
57.03338
87.00783
126.14560
169.06479
197.05781
m/z

Figure S2. Representative MS/MS spectra of the compounds/features shown in Figure S1B. The spectra correspond to P1, procyanidin C1; P2, catechin; P3, aromadendrin-O-glucoside; P4, isoquercitrin/quercetin-O-hexoside; P5, astragalin; P6, piceatannol; P7, naringenin; P8, 6β,8β-dihydroxyeremophilenolide; P9, protocatechuic acid; P10, procyanidin B-type dimer isomer (B2-like); P11, kaempferol; and P12, apigenin. Diagnostic product ions were annotated to support the tentative LC–MS/MS assignments.