

Supporting Information

Iodine-Catalyzed Green and Efficient Synthesis of Secondary and *Tert*-Esters of *N*-Acetyl-Protected Amino Acids

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1 Material and instrumentation

Acetyl chloride, glycine, leucine, phenylalanine, methionine, tryptophan and proline were purchased from Sigma-Aldrich, Merck, and Fluka. All commercial solvents were distilled and dried prior to use. Thin layer chromatography (TLC) with silica gel-coated plates (0.5 mm thick, Merck) was used to monitor the reaction progress, and spots were visualized under a UV lamp and ninhydrine solution. Synthesized compounds were recrystallized using ethyl acetate and ethanol. Physical properties such as the melting point were determined using the Gallen Kamp melting point apparatus. An FT-IR 6300 SHIMADZU spectrophotometer was used to record the IR spectrum of synthesized compounds. A Bruker Advance nuclear magnetic resonance spectrometer resonating at 400 MHz was used for the structural elucidation of targeted compounds in CDCl₃ as the solvent and TMS as the internal standard.

1.1 Synthesis of Acetylated Amino Acids (3a-f)

5-7 mmol of L-amino acids (1a-f) were dissolved in water, and an equimolar amount of acetyl chloride was added, heating it for 2 hours at 70°C. *N*-acetyl amino acid formation was monitored by TLC and then by ninhydrin test, and the solvent was evaporated using a rotary evaporator. The crude product was dissolved in methanol and filtered to remove any unreacted amino acid. Methanol was removed till dryness under vacuum to get pure precipitates of *N*-acetyl amino acids.

Table 1: Physical and Chemical Parameters of Acetylation of Amino Acids

Compound	Yield %	m.p (°C)	[α] D ₂₀ (C=0.5g. 50mL Methanol)
3a	58	208-210	-
3b	94	190-195	-24.32
3c	96	176-178	+39.47
3d	93	103-105	-8.2
3e	96	182-188	+27.46
3f	91	117-120	-86

1.2 Synthesis of N-Protected Amino Esters:

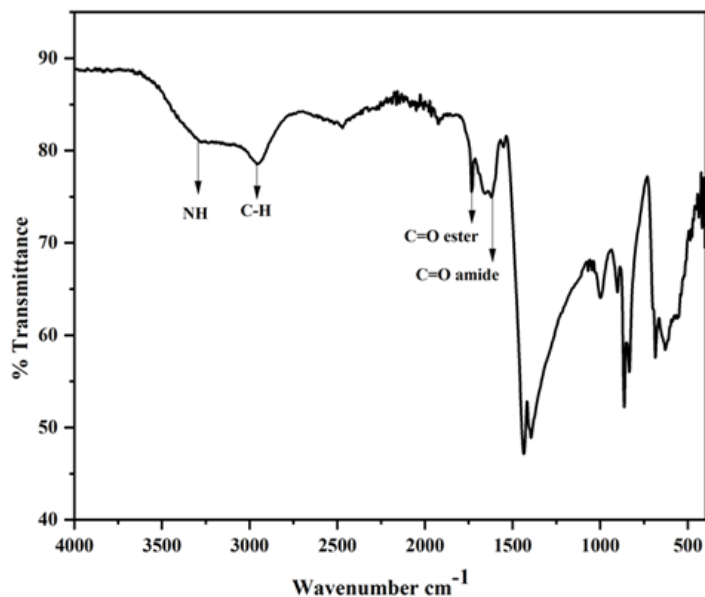
N-Protected amino acids (0.5g, 2.88 mmol) were dissolved in excess alcohol, followed by the addition of I₂ (25 mg, 0.01 equiv; doubled for *tert*-butyl esters) under stirring. The reaction mixture was refluxed for 6-8 hours and monitored by TLC. After completion, excess alcohol was removed under reduced pressure. The residue was extracted with diethyl ether or DCM, and the organic layer was washed successively with sodium thiosulfate, sodium bicarbonate, and water. It was then dried over anhydrous Na₂SO₄ or MgSO₄ and concentrated under vacuum. This general procedure was applied to all substrates for esterification. In case of DMAP, 1N HCl was used during work.

Table 2: Physical and chemical parameters of N-Acetylated Amino Esters

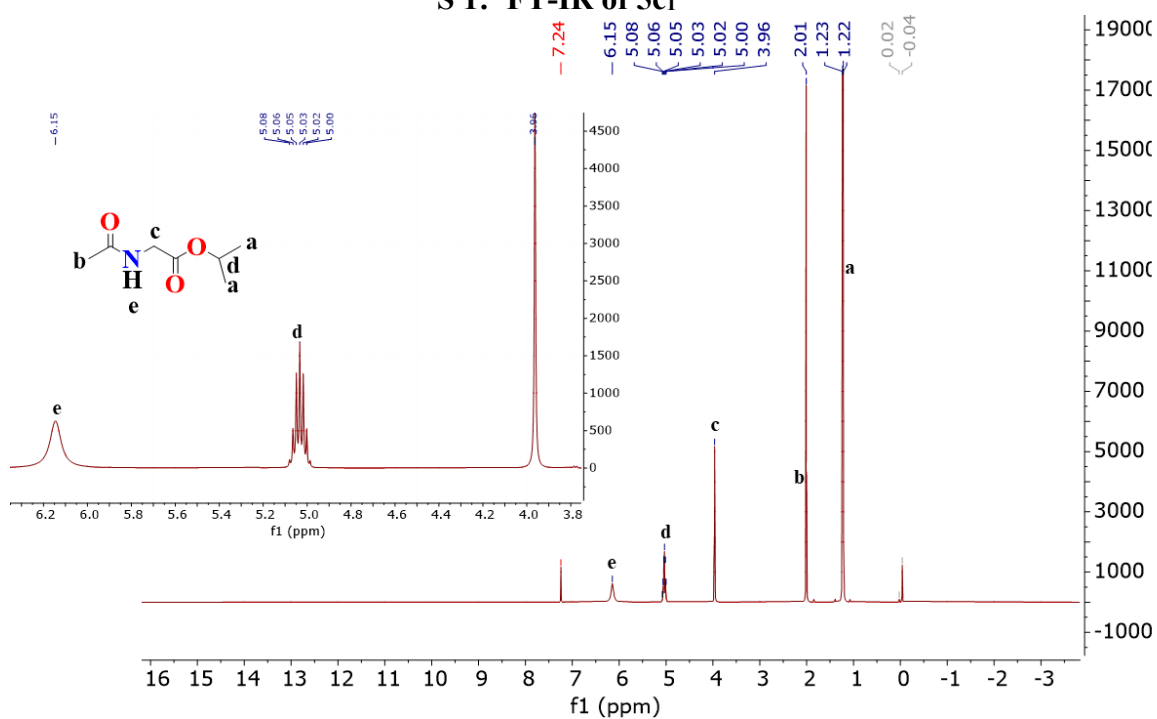
Compound	Melting Point °C	Yield %	[α] D ₂₀ (C=0.05g. 50mL Methanol)
5a ₁	58-60	75	-
5a ₂	47-50	80	-54
5a ₃	68-70	77	+40.1
5a ₄	80-82	80	-28.03
5a ₅	208-214	80	+18.98
5a ₆	72-73	70	-35.23
5b ₁	47-50	78	-
5b ₂	60-63	85	+22.34

5b₃	85-90	78	+13.2
5b₄	102-104	72	+25.15
5b₅	109-111	78	+22.40
5b₆	64-65	69	-9.3
5c₁	Oily liq.	76	-
5c₂	Semi-solid liq.	80	+9.27
5c₃	62-64	78	-0.20
5c₄	Oily liq.	80	-0.50
5c₅	150-152	74	-25.50
5c₆	Oily liq.	68	+11.62
5d₁	Oily liq.	70	-
5d₂	Oily liq.	75	+13.22
5d₃	Oily liq.	73	-1.23
5d₄	Oily liq.	75	+20.28
5d₅	Oily liq.	70	+9.33
5d₆	Oily liq.	60	-32.32

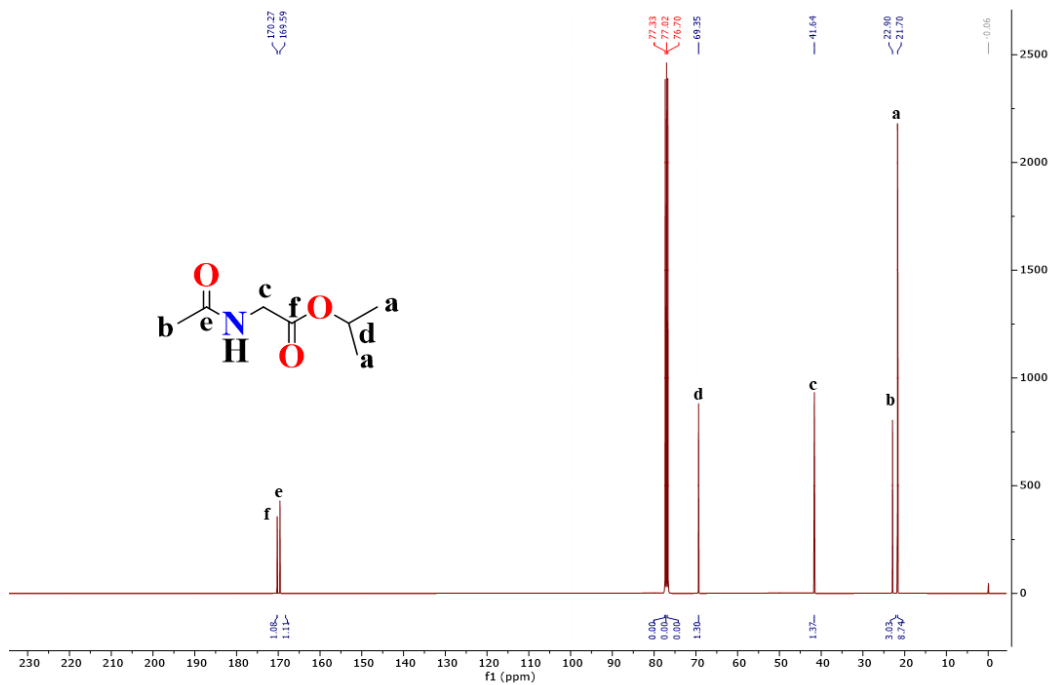
Spectroscopic Data of Secondary Esters of *N*-Acetylated Amino Acids



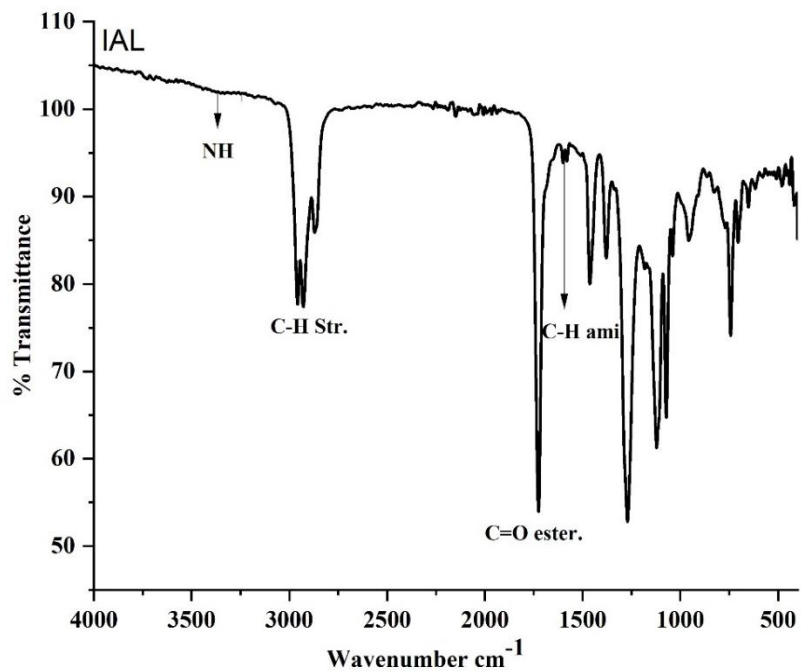
S 1: FT-IR of 5c1



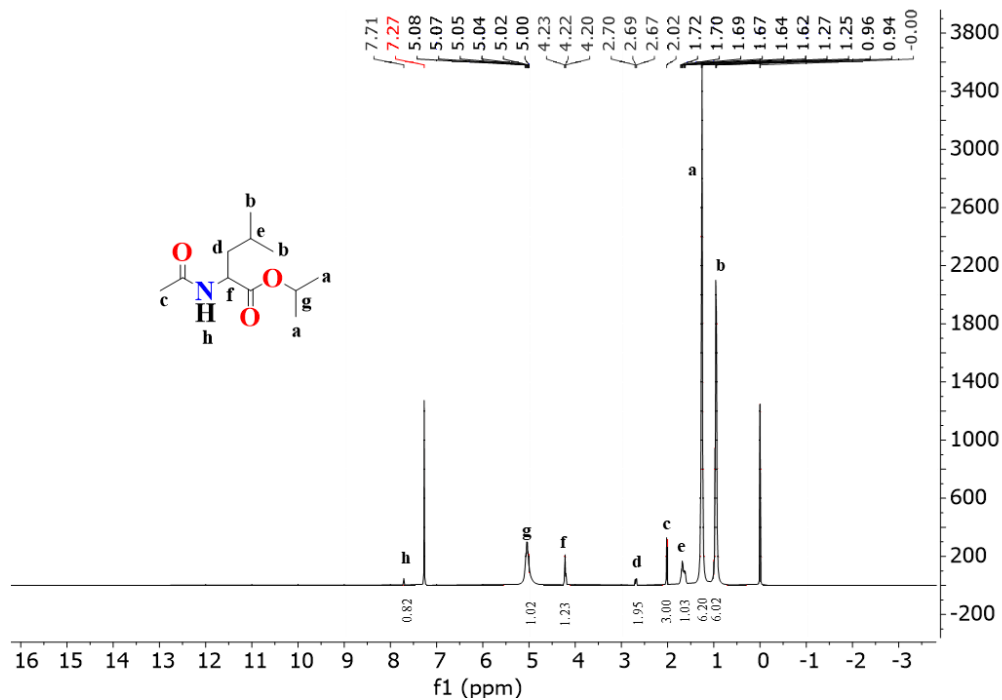
S1. $^1\text{H NMR}$ 5c1



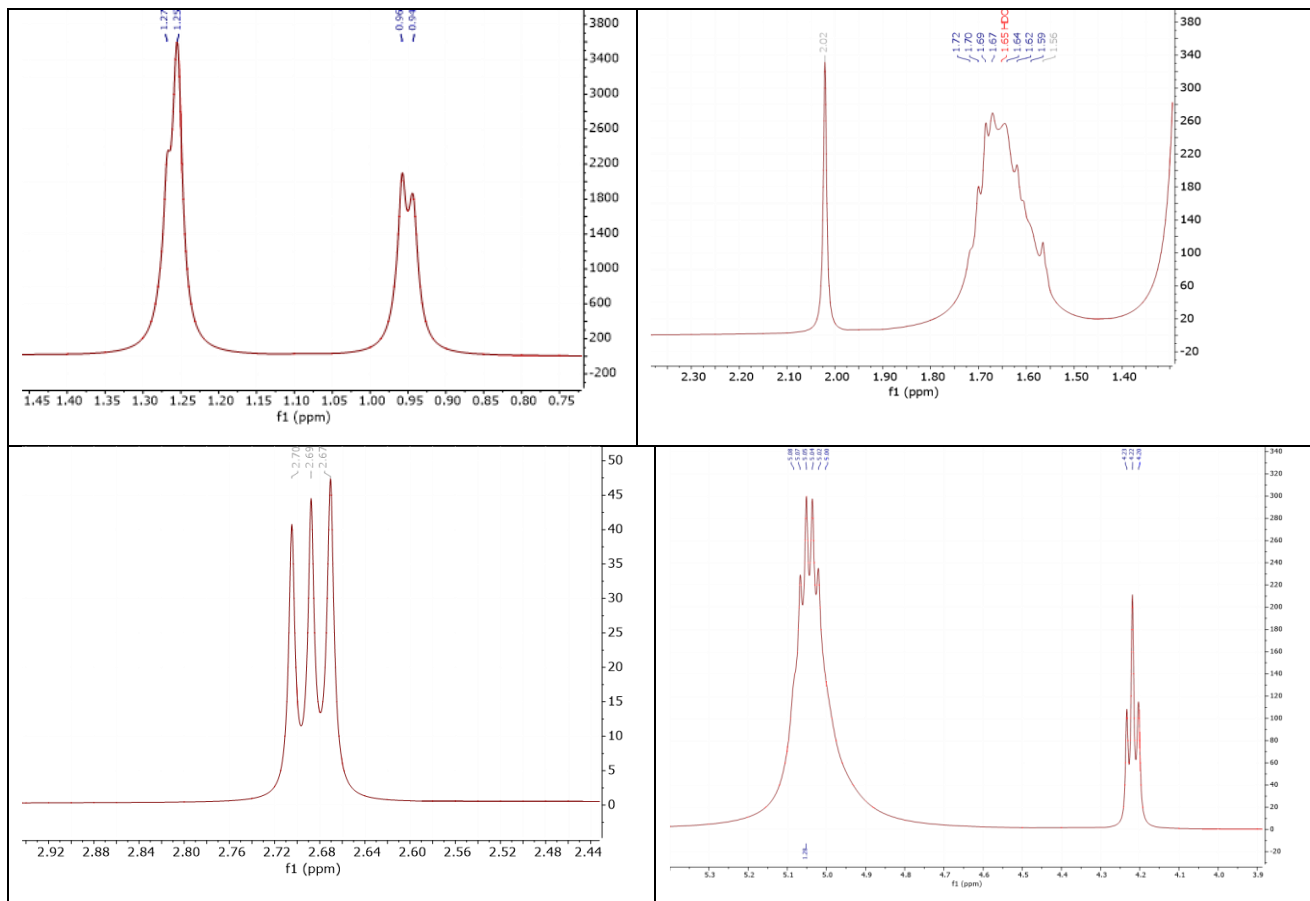
S 3: ^{13}C NMR5c1



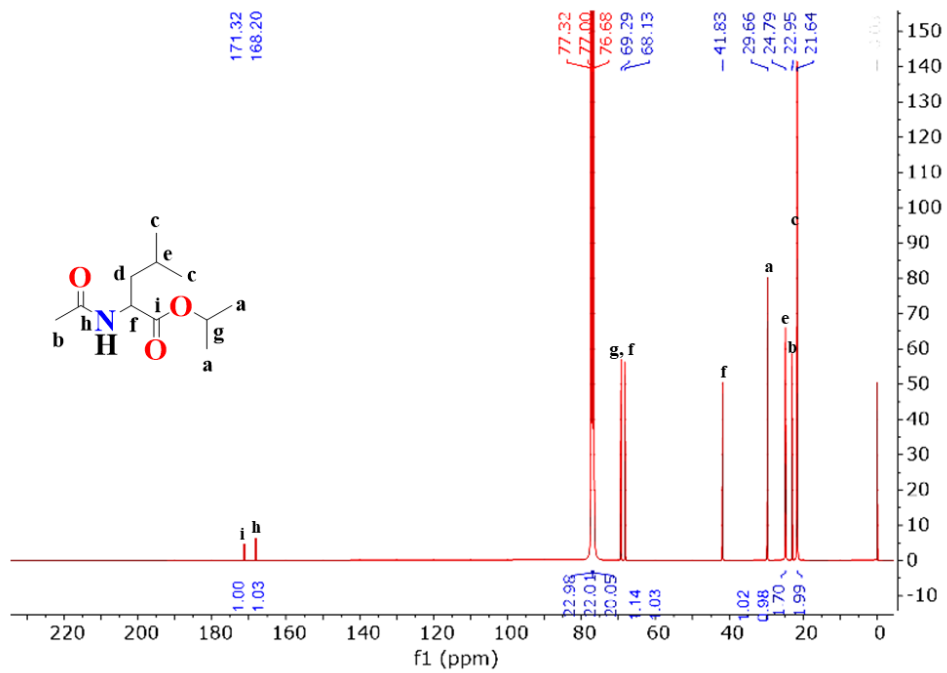
S 4: FT-IR of 5c2



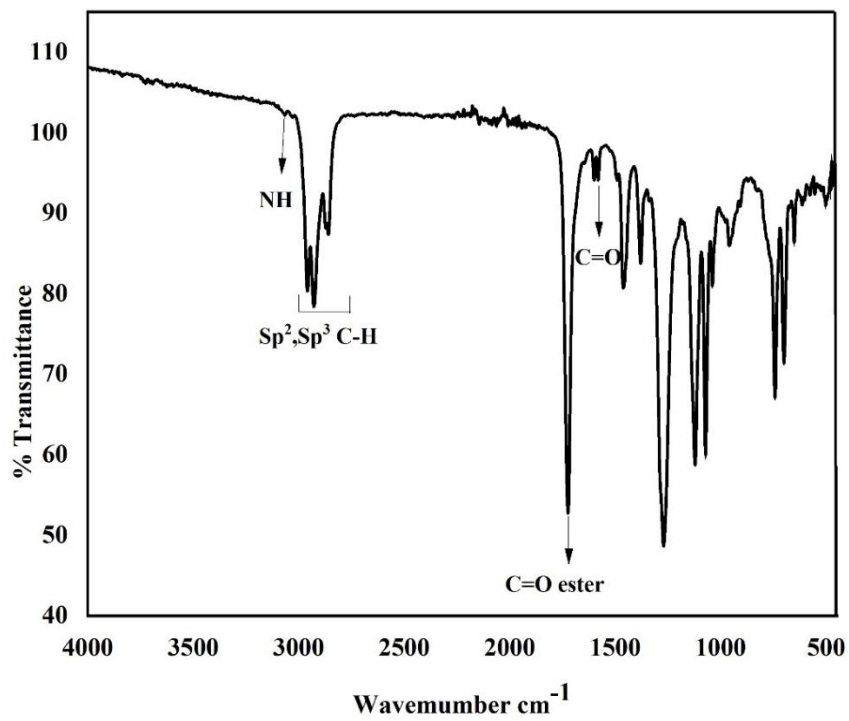
S 5: ¹H NMR 5c2



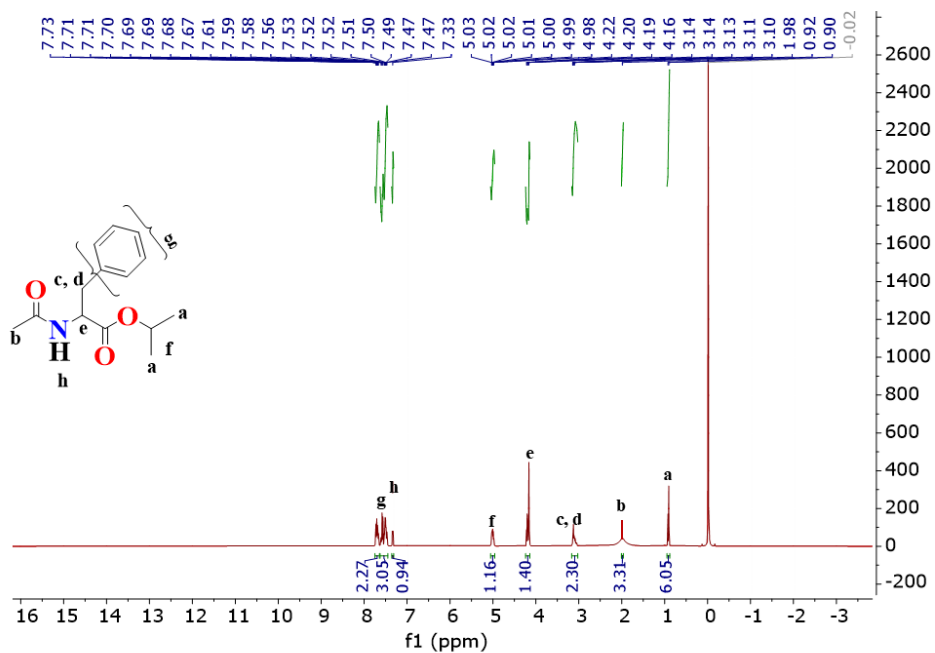
S 6: Extended ¹HNMR 5C₂



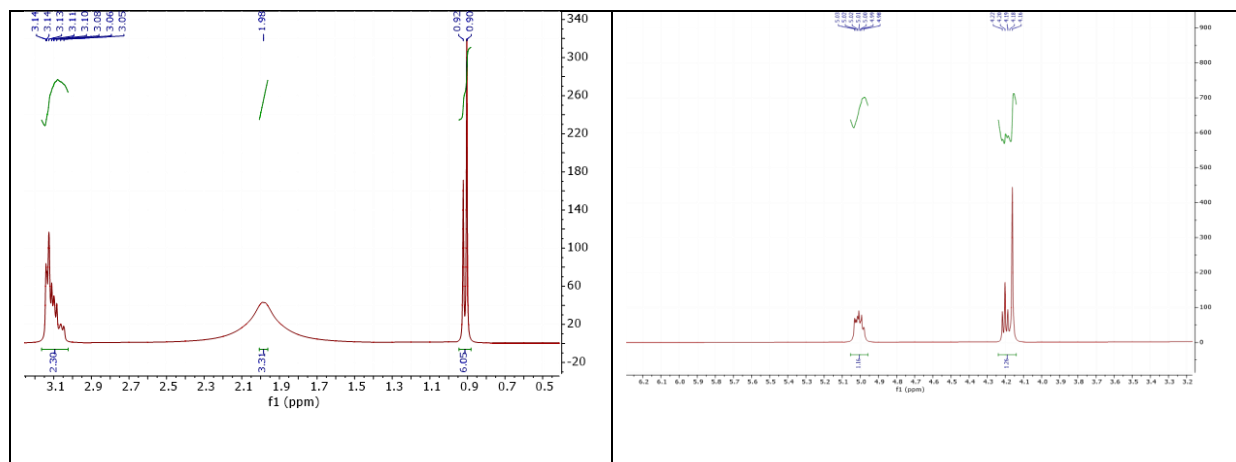
S 7: ¹³CNMR 5c₂

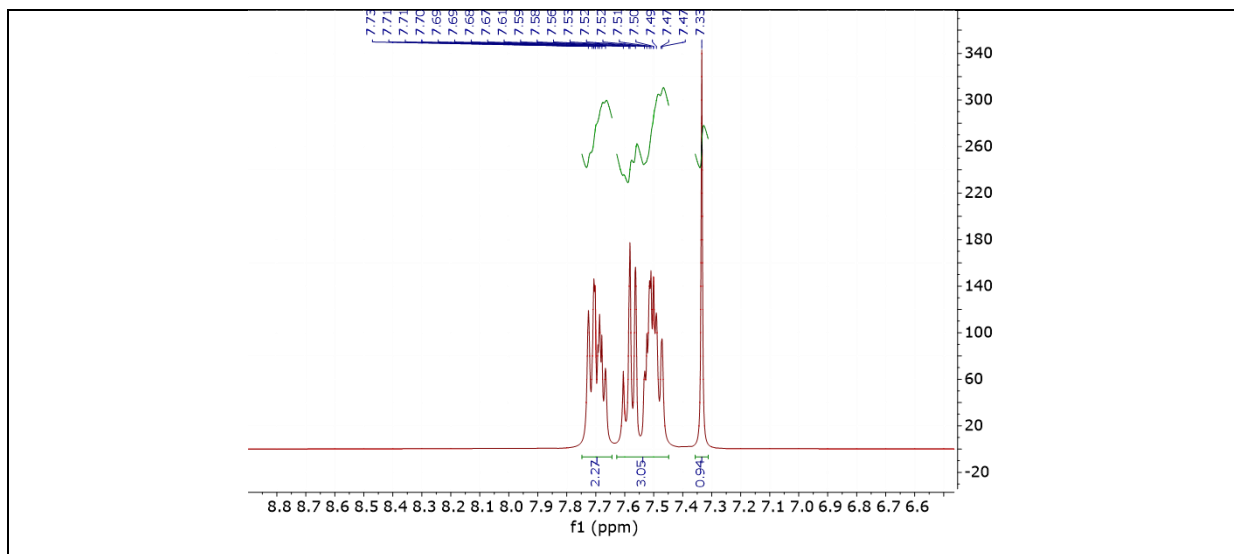


S 8: FT-IR of 5c₃

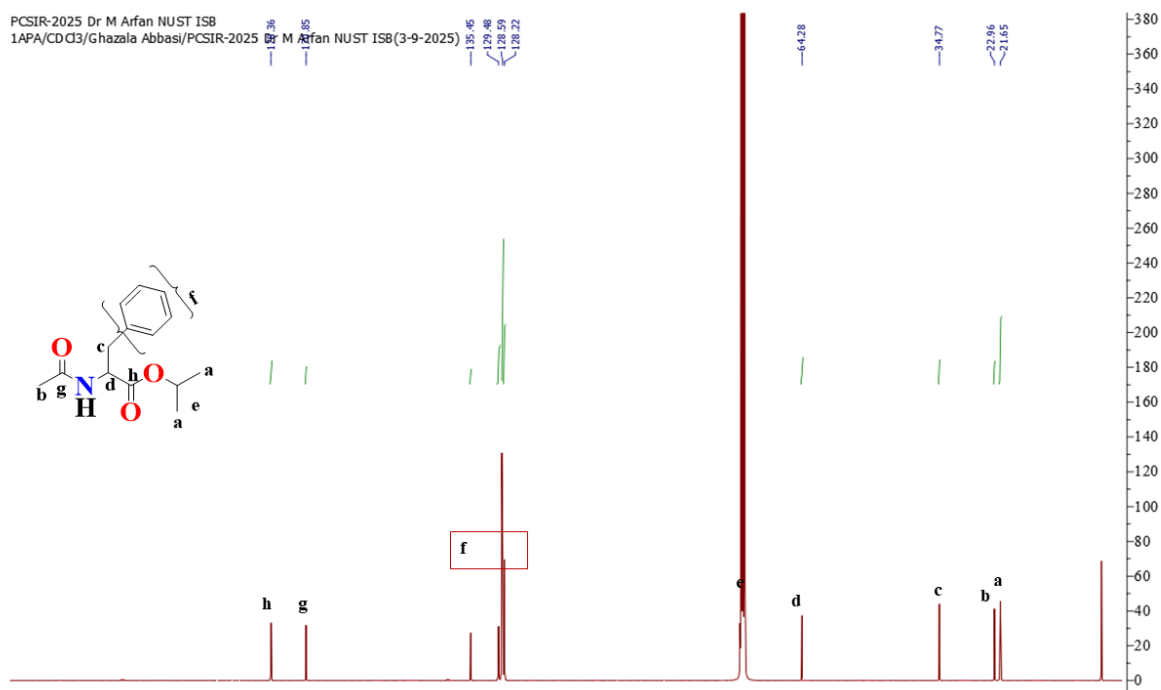


S 9: ¹H NMR 5c₃

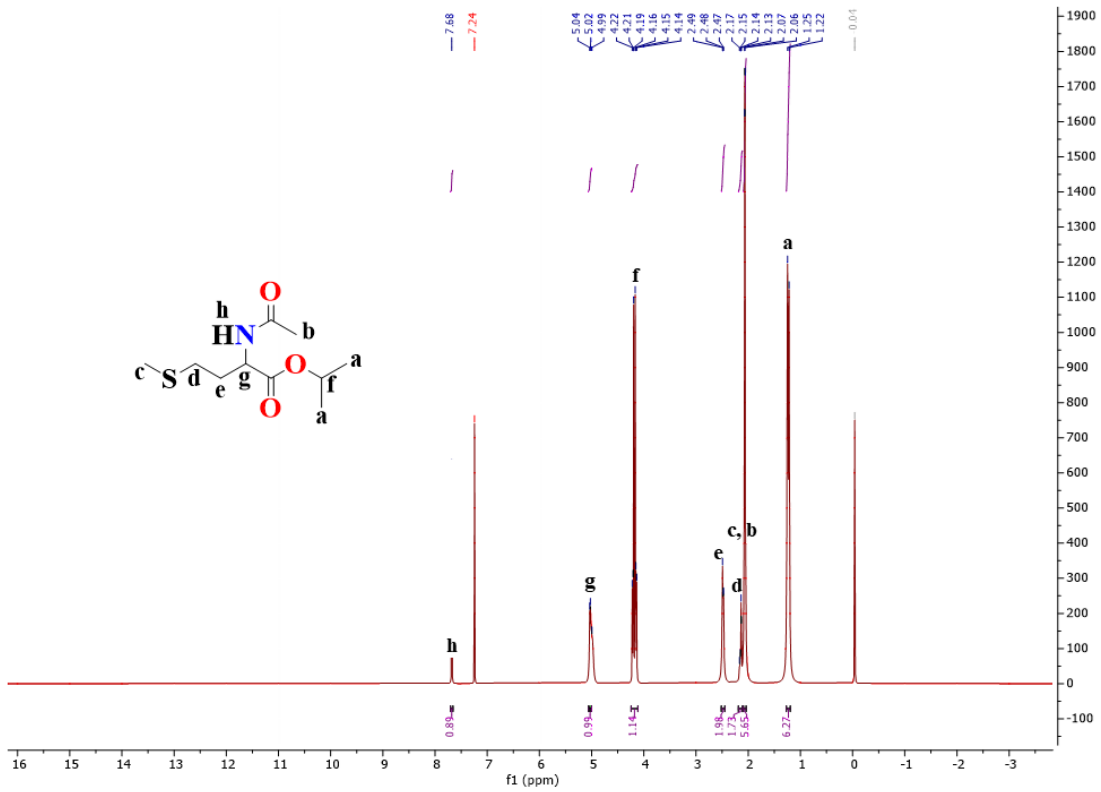




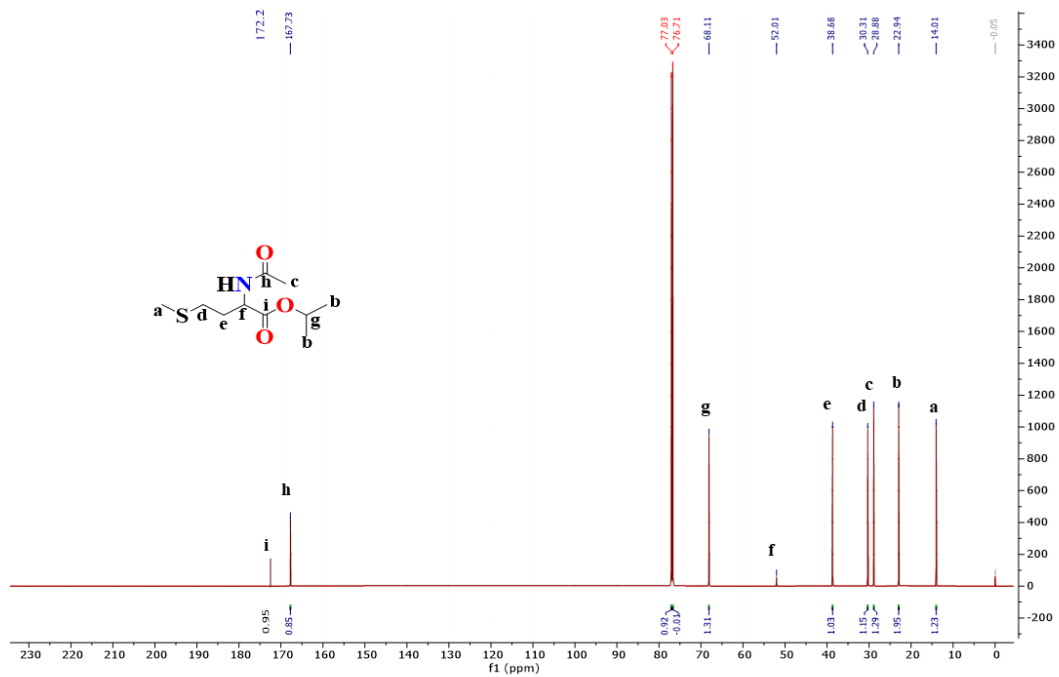
S 10: Extended ^1H NMR $5c_3$



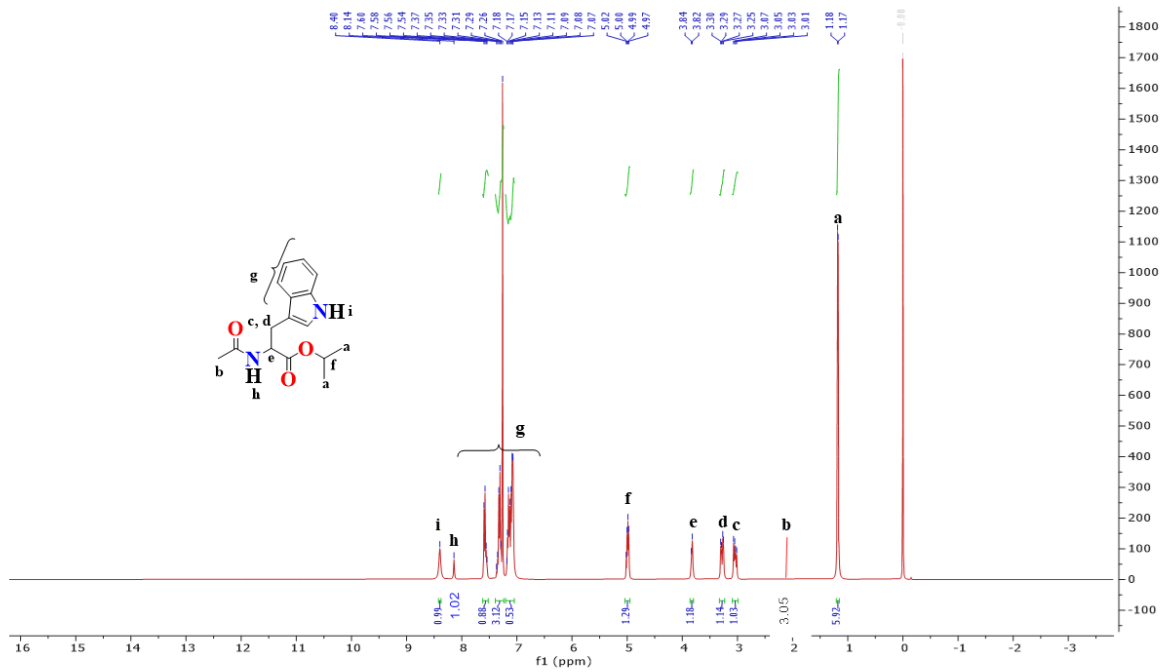
S 11: ^{13}C NMR $5c_3$



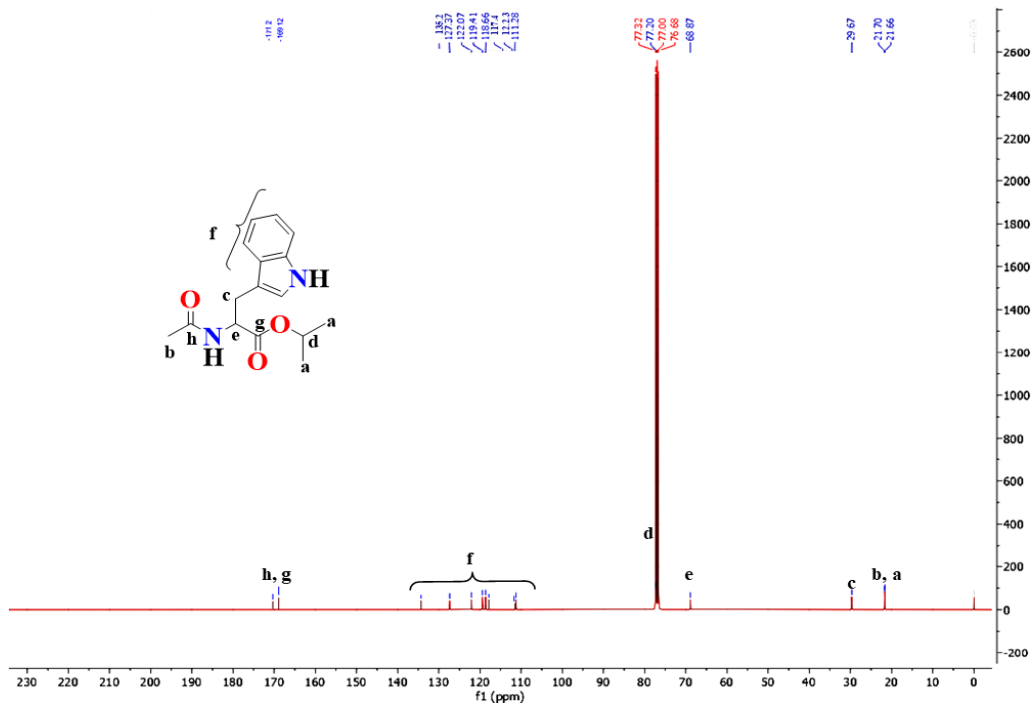
S 12: ¹H NMR 5c4



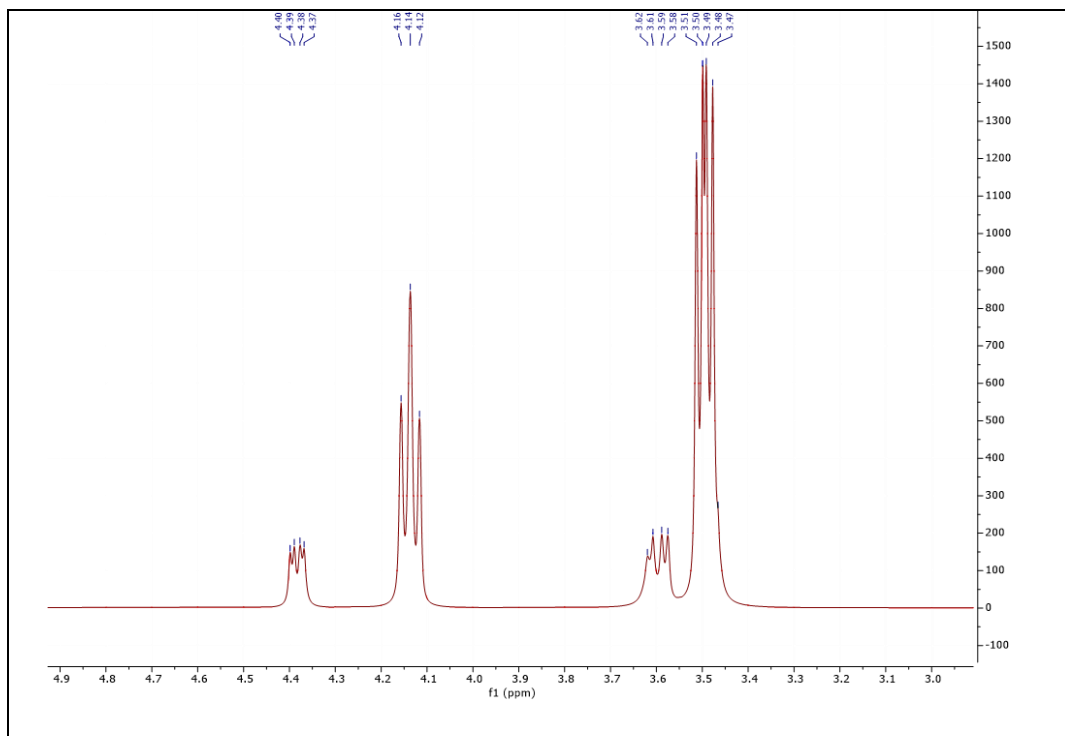
S 13: ¹³C NMR 5c4



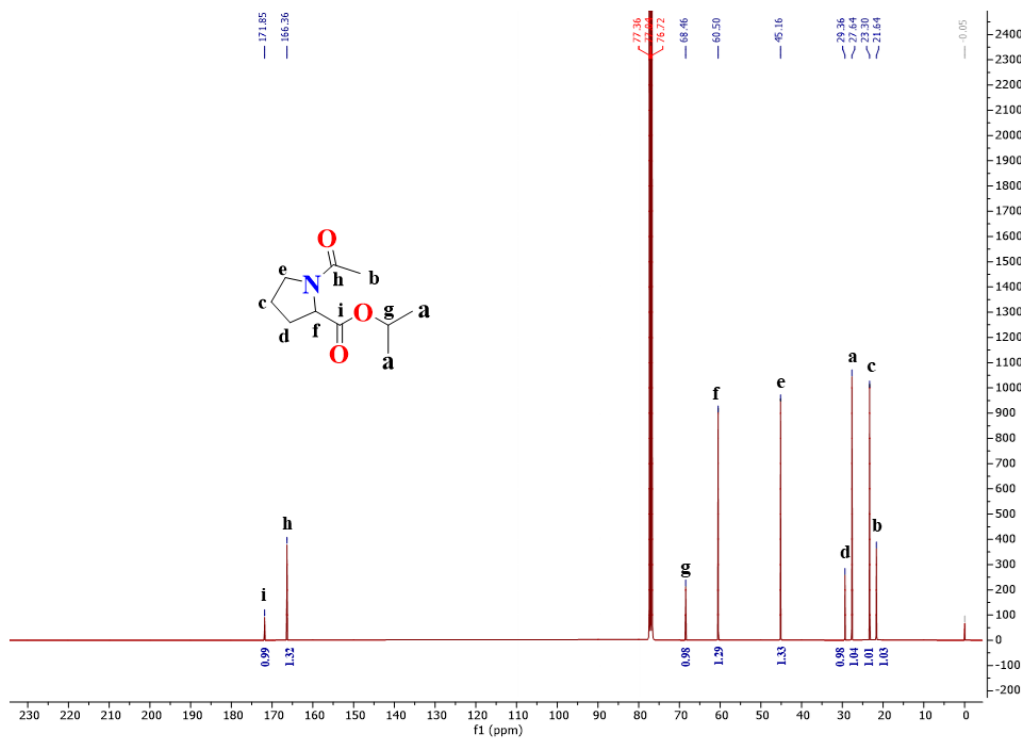
S 14: ¹H NMR 5cs



S 15: ¹³C NMR 5cs

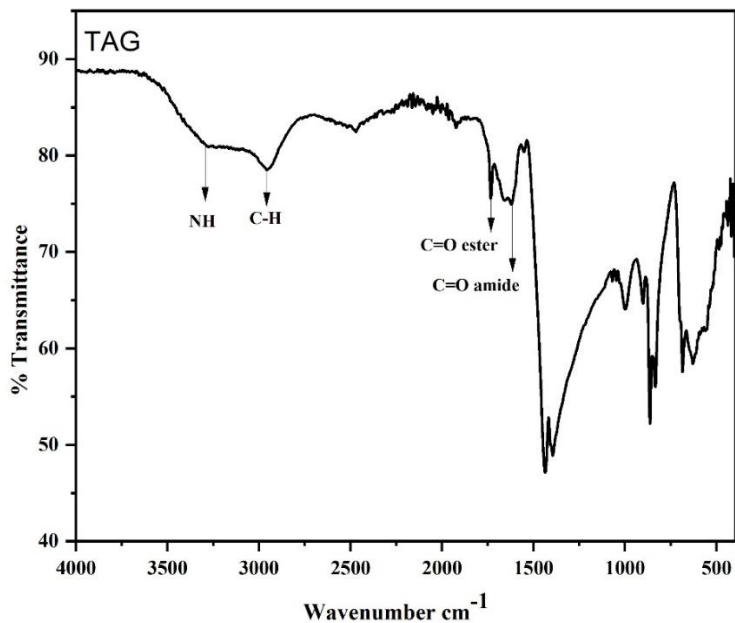


S 17: Extended ^1H NMR $5c_6$

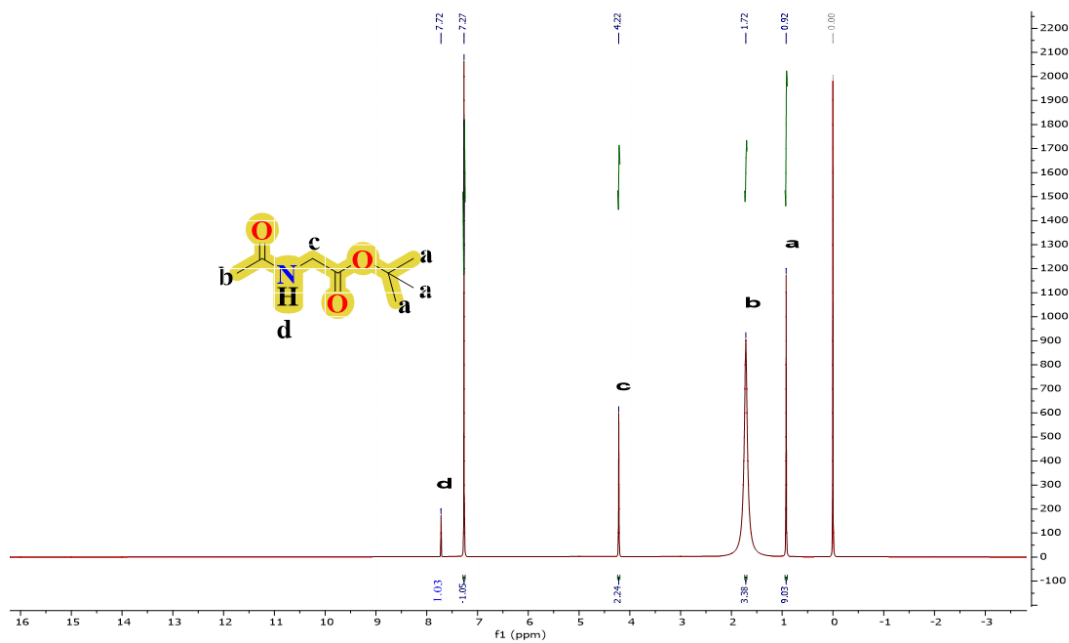


S 18: ^{13}C NMR $5c_6$

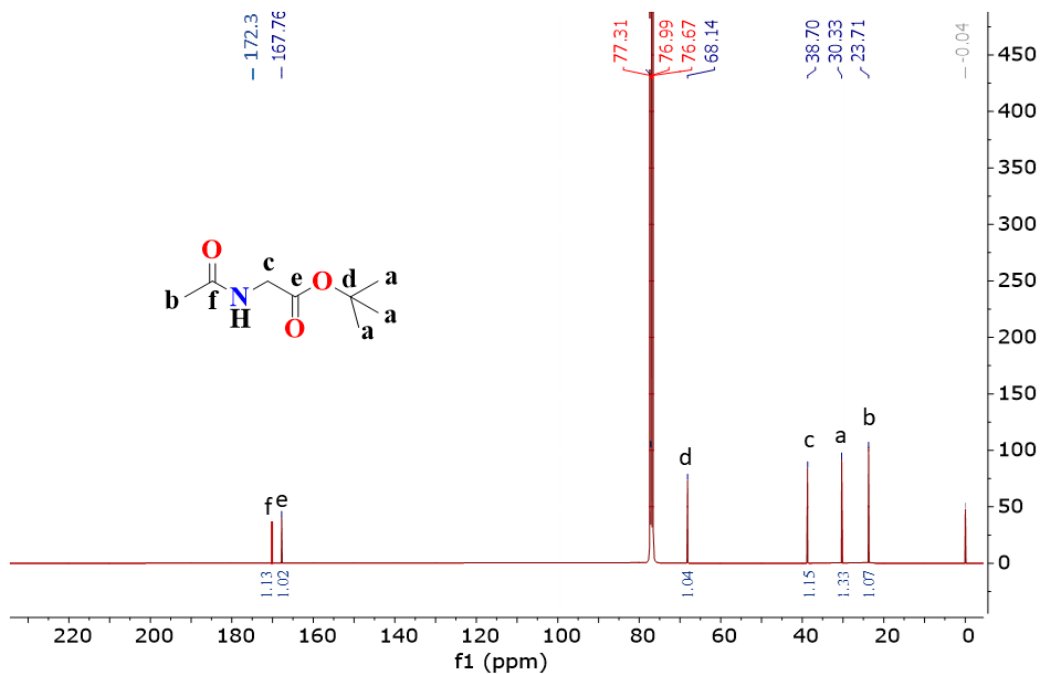
Spectroscopic Data of *Tert*-butyl Esters of *N*-Acetylated Amino Acids



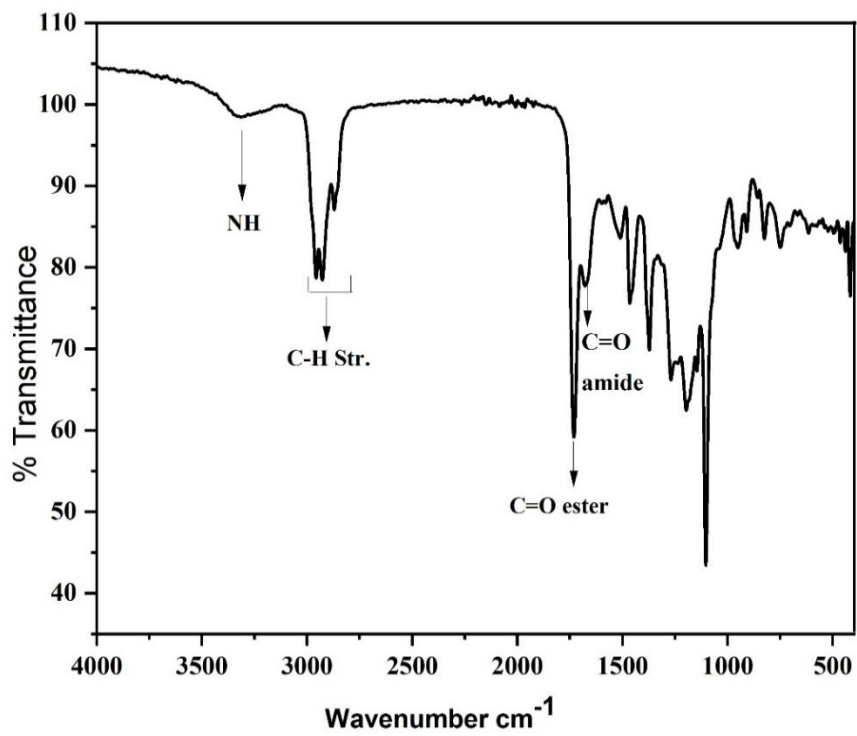
S 19: FT-IR of 5d₁



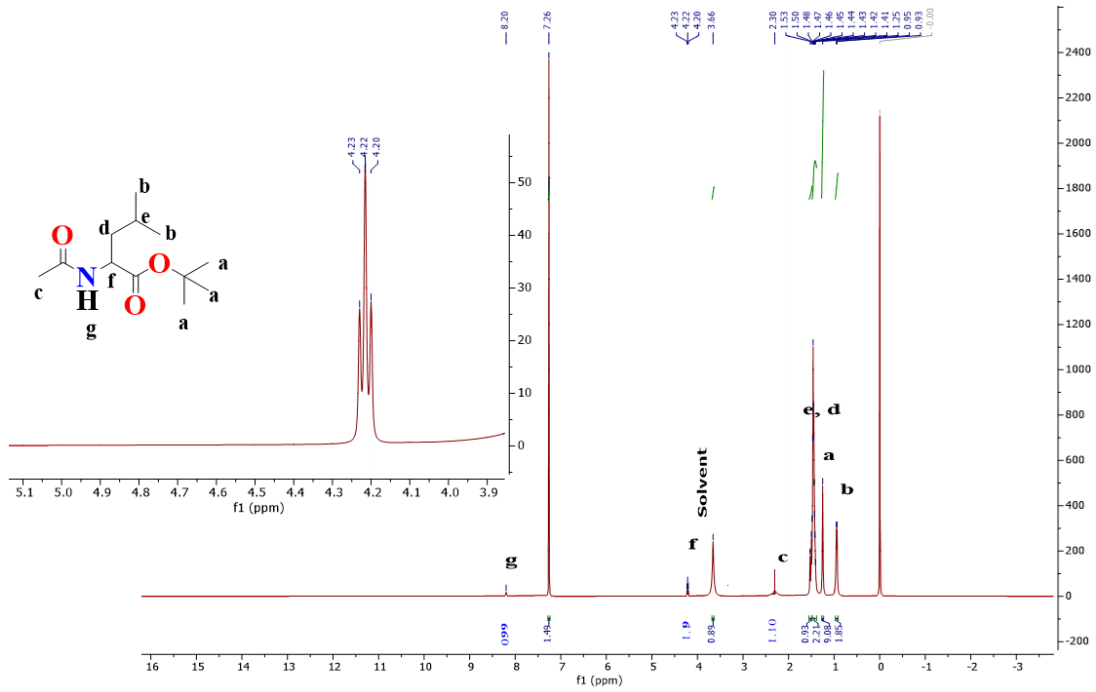
S 20: ¹H NMR of 5d₁



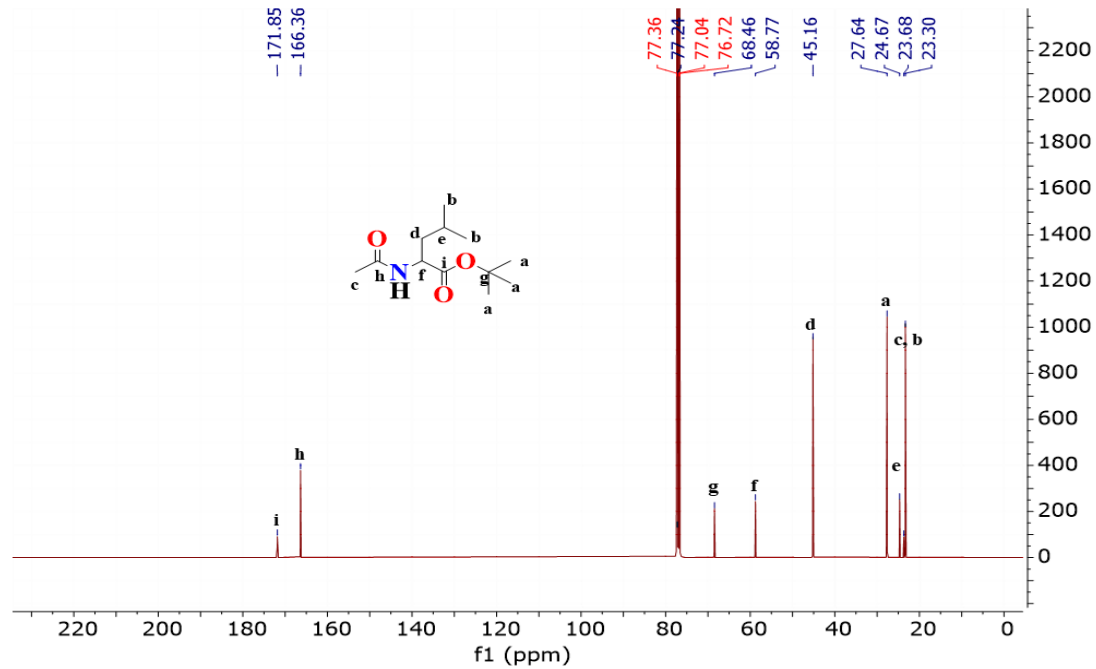
S 21: ¹³CNMR 5d₁



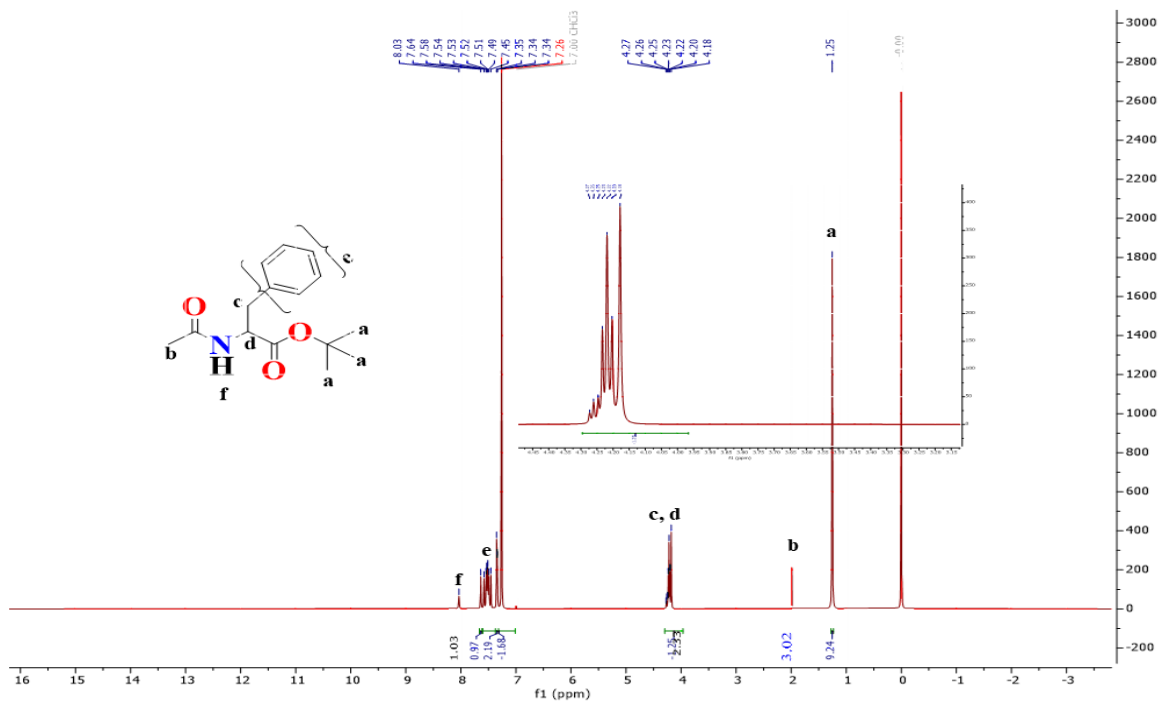
S 22: FT-IR of 5d₂



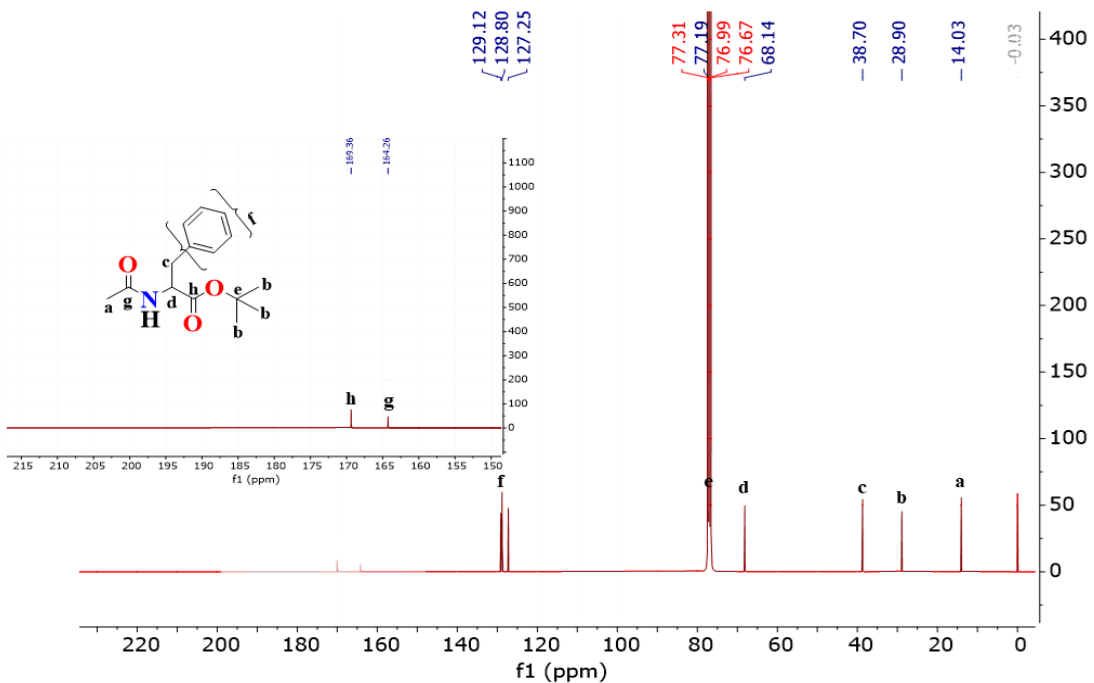
S 23: ¹H NMR of 5d₂



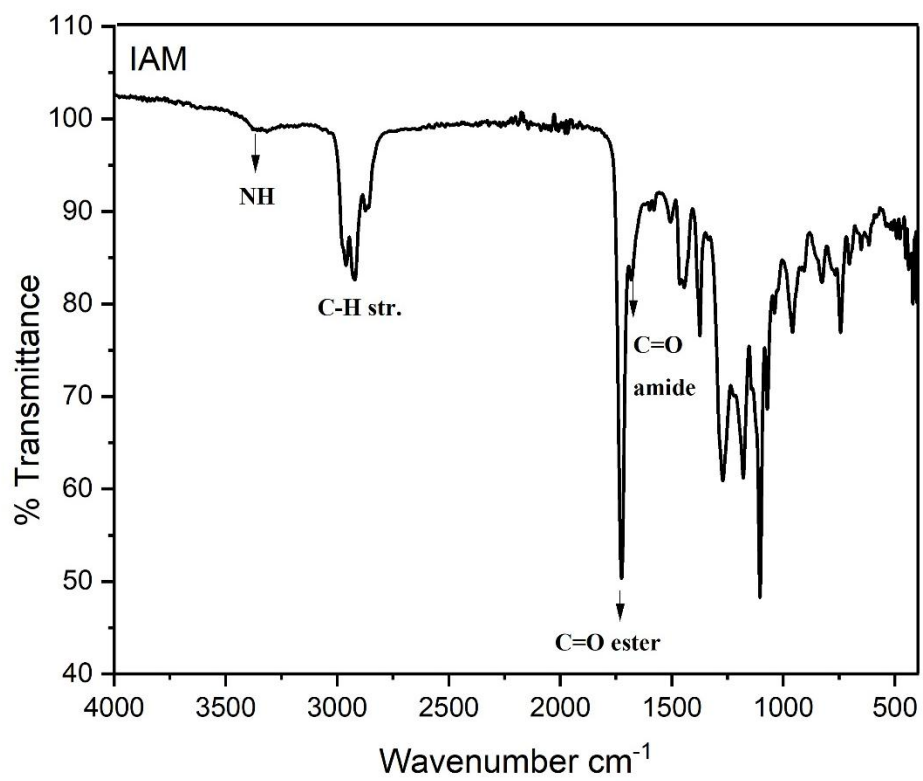
S 24: ¹³C NMR of 5d₂



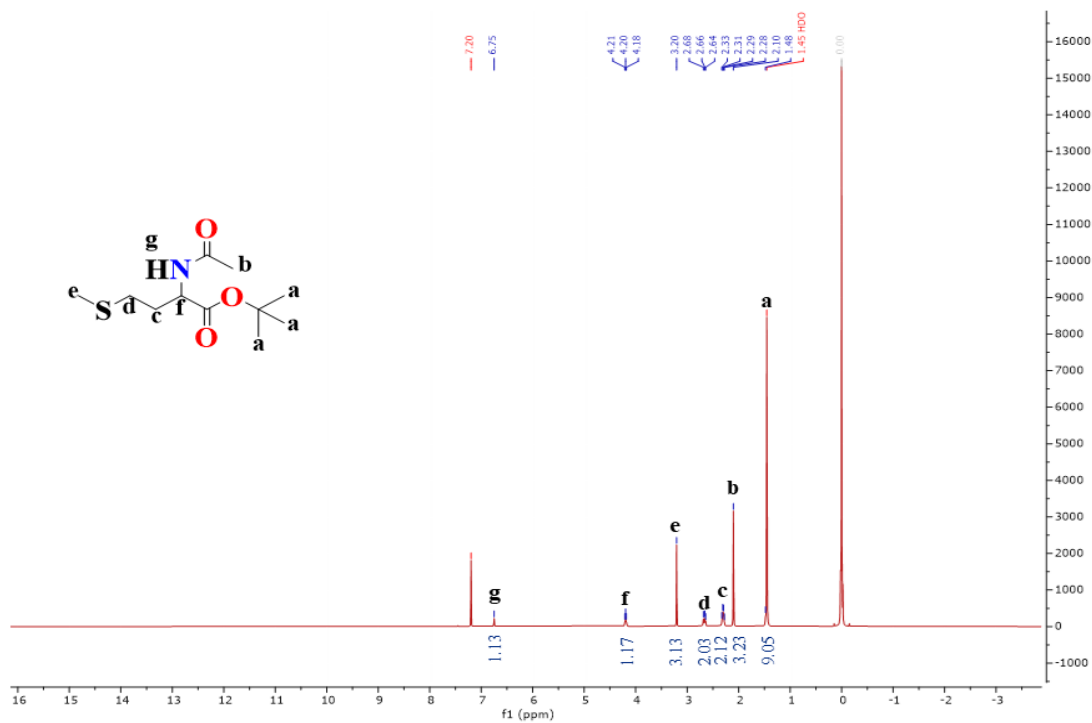
S 25: ¹H NMR of 5d₃



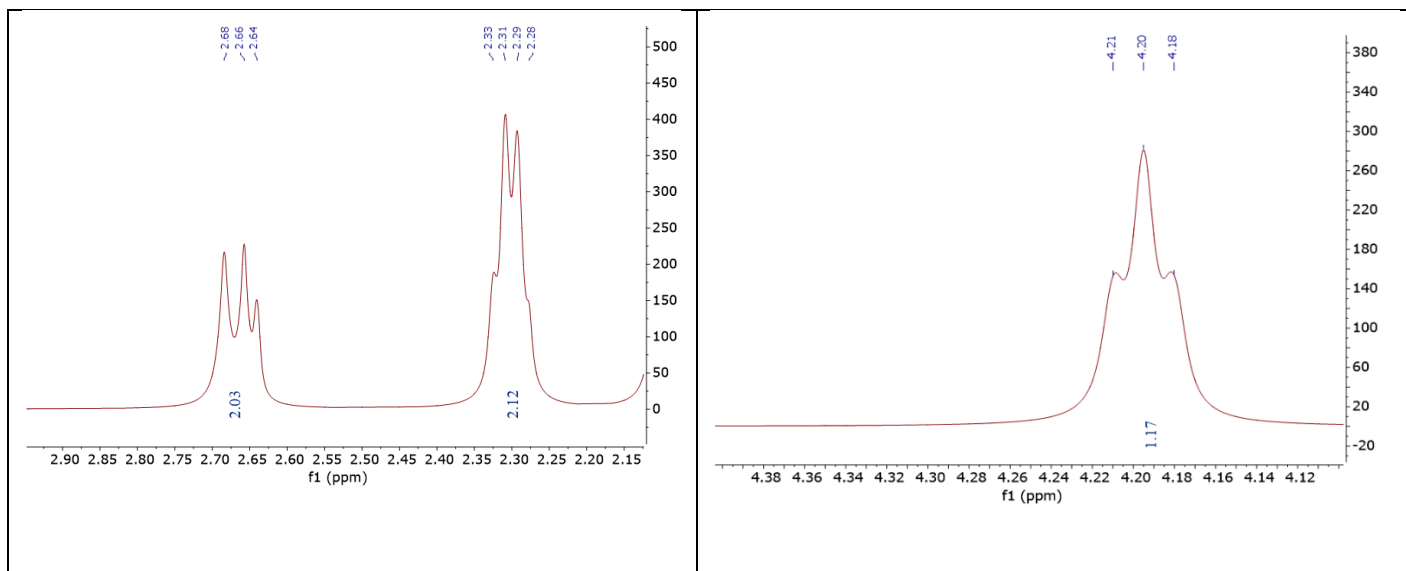
S 26: ¹³CNMR of 5d₃



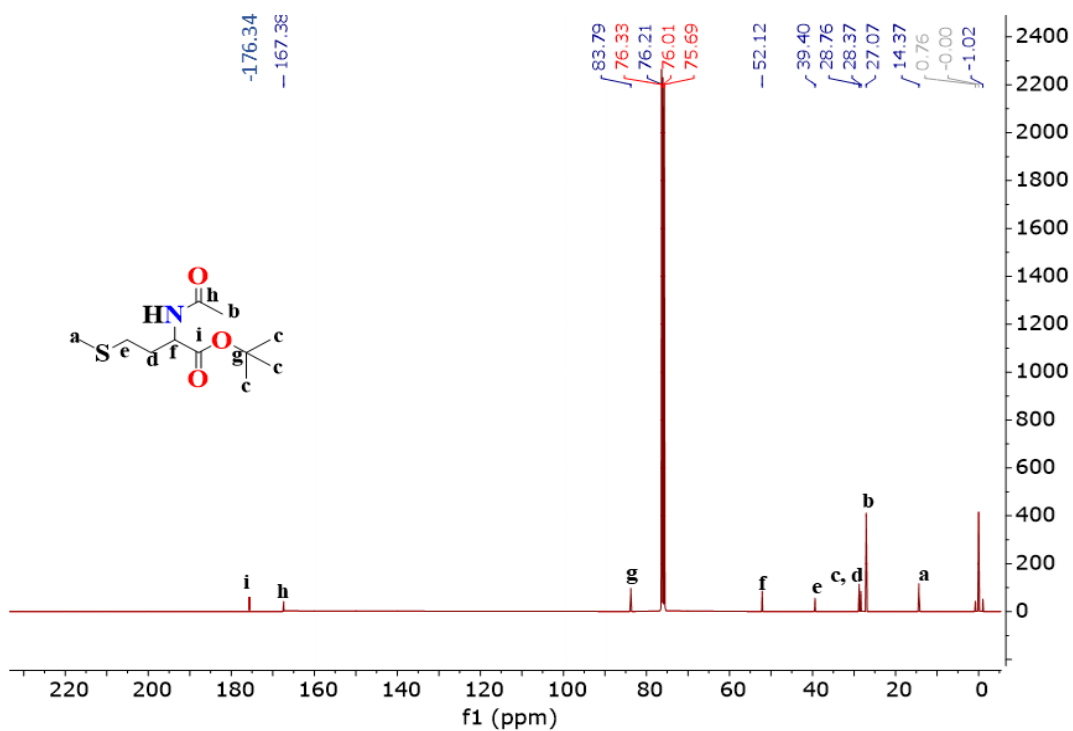
S 27: FT-IR of 5d₄



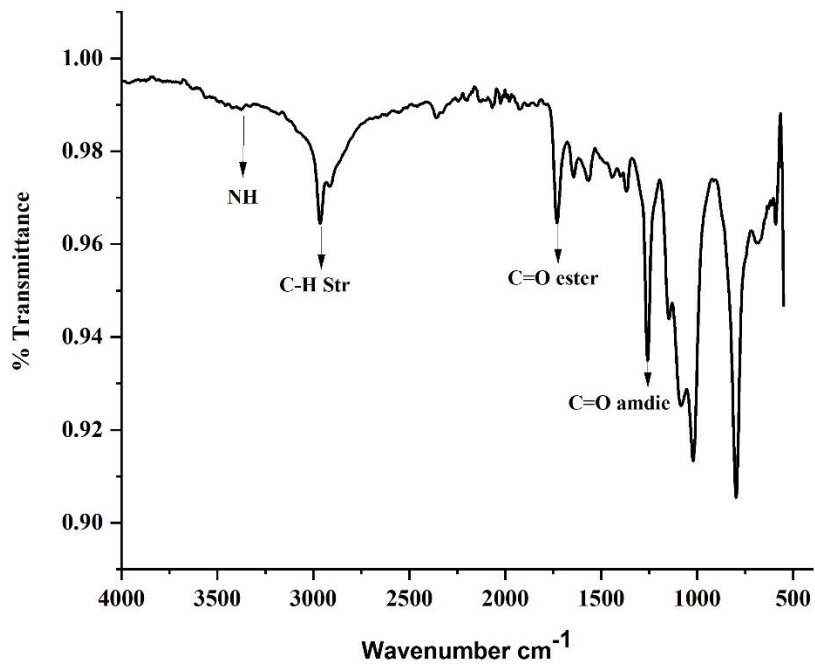
S 28: $^1\text{H NMR}$ $5d_4$



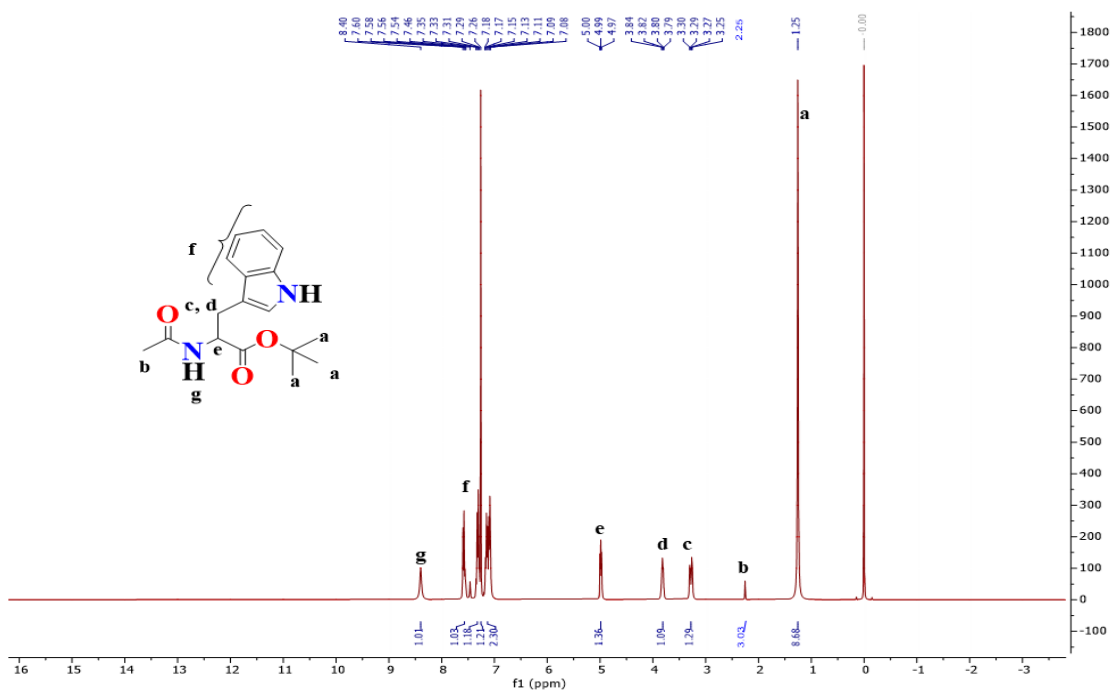
S 29: Extended ¹H NMR 5d₄



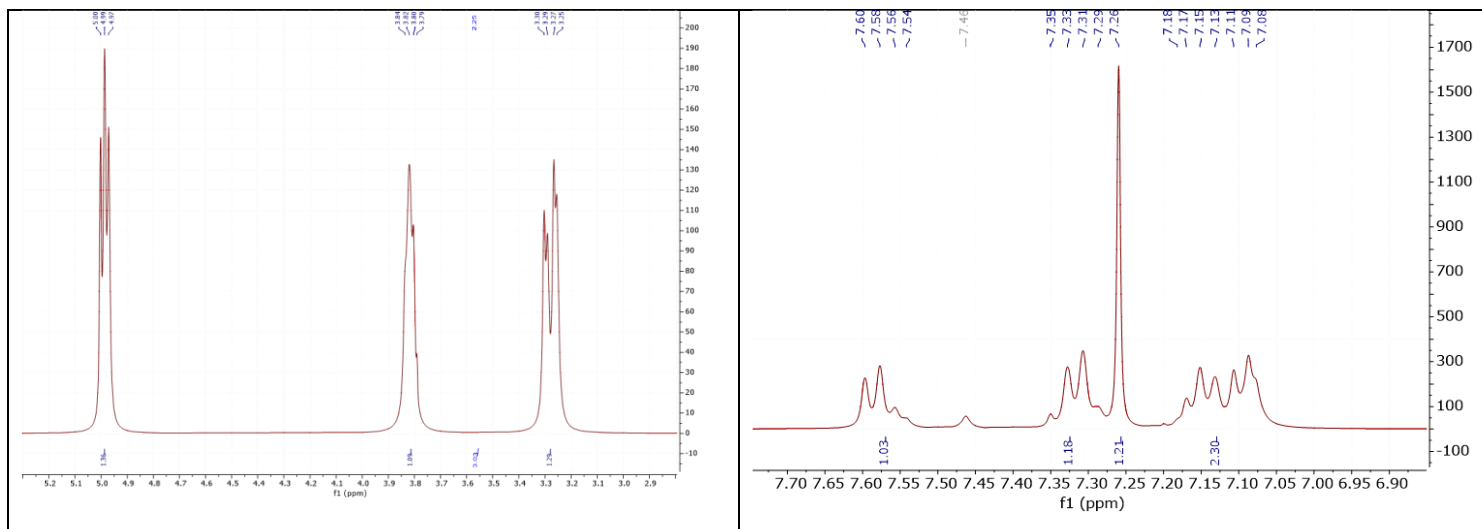
S 30: ¹³C NMR 5d₄



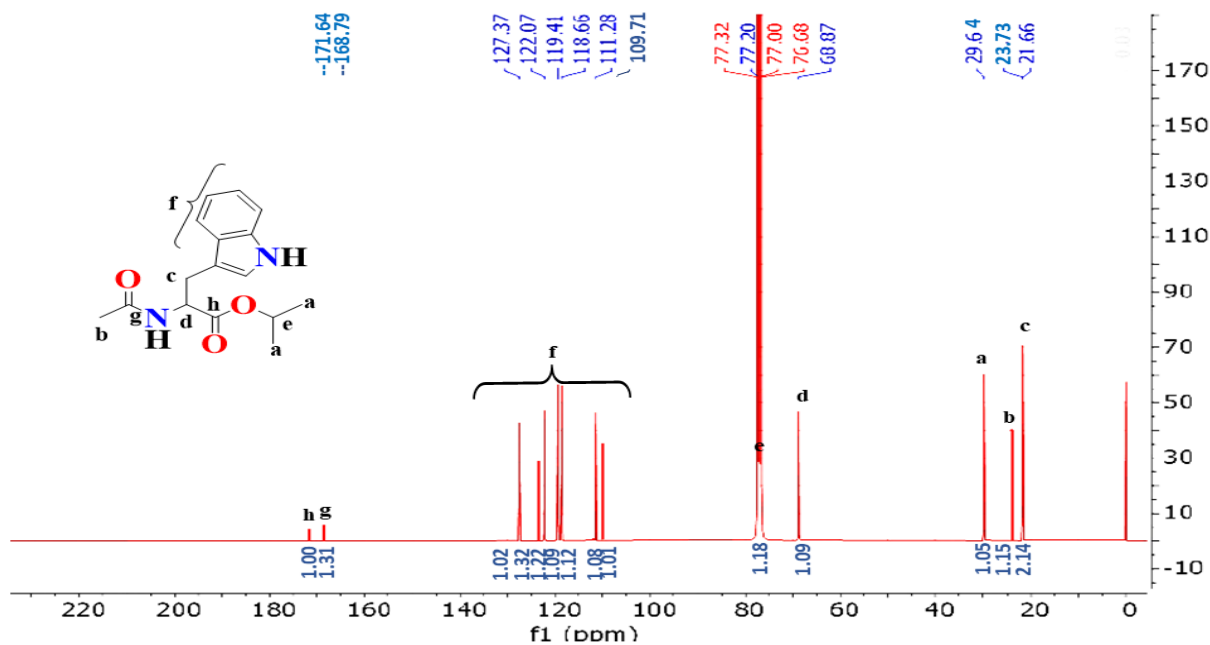
S 31: FT-IR of 5d₅



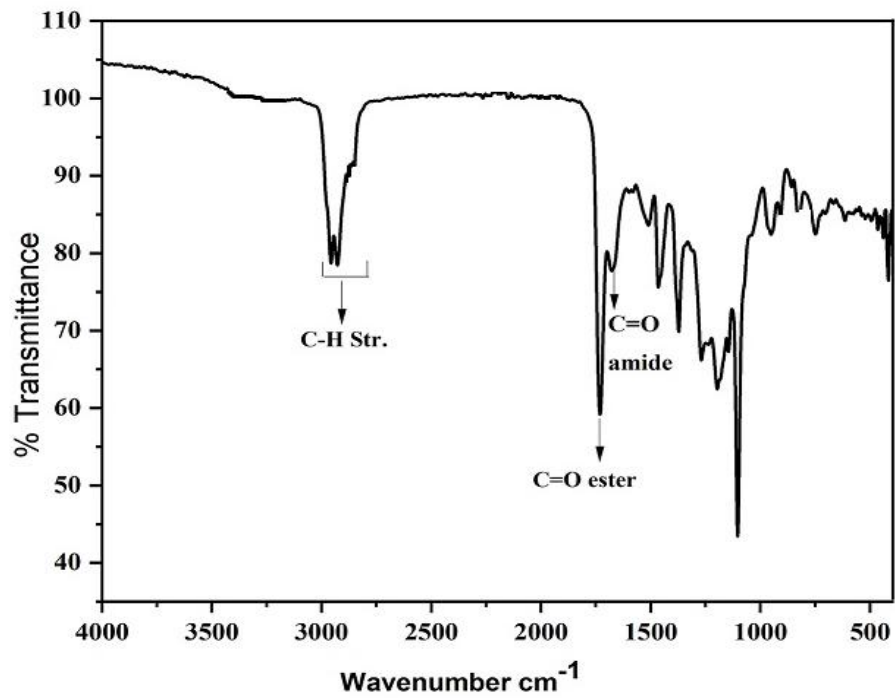
S 32: ¹H NMR 5d₅



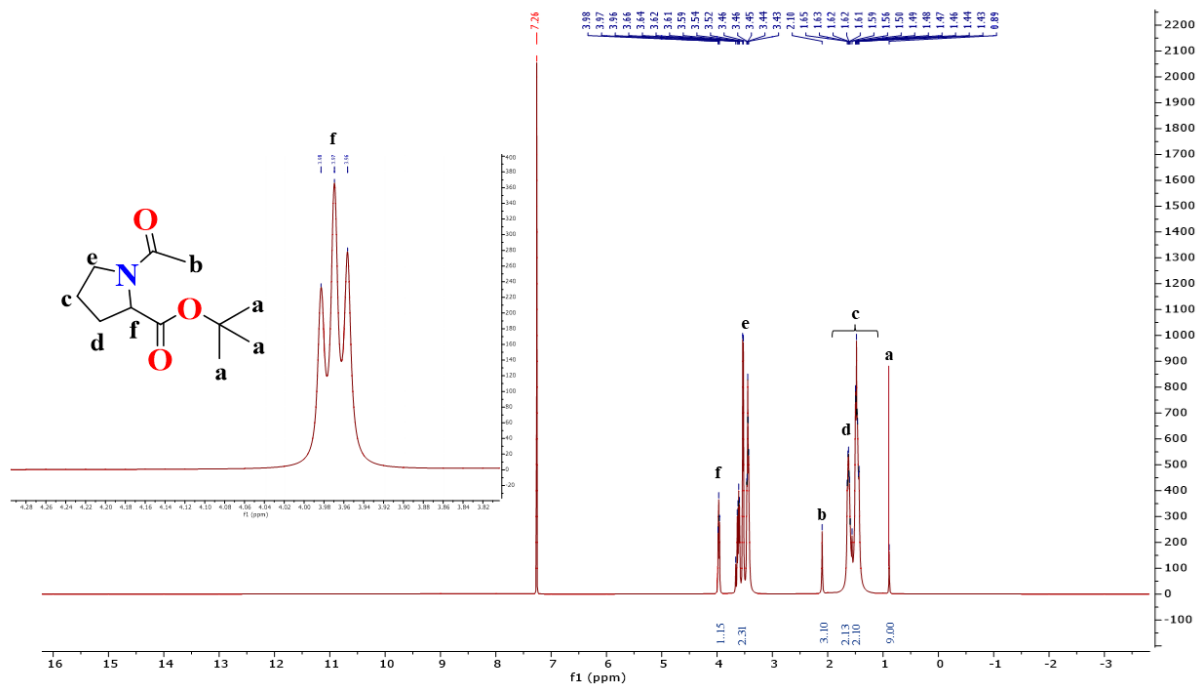
S 33: Extended ¹H NMR 5d₅



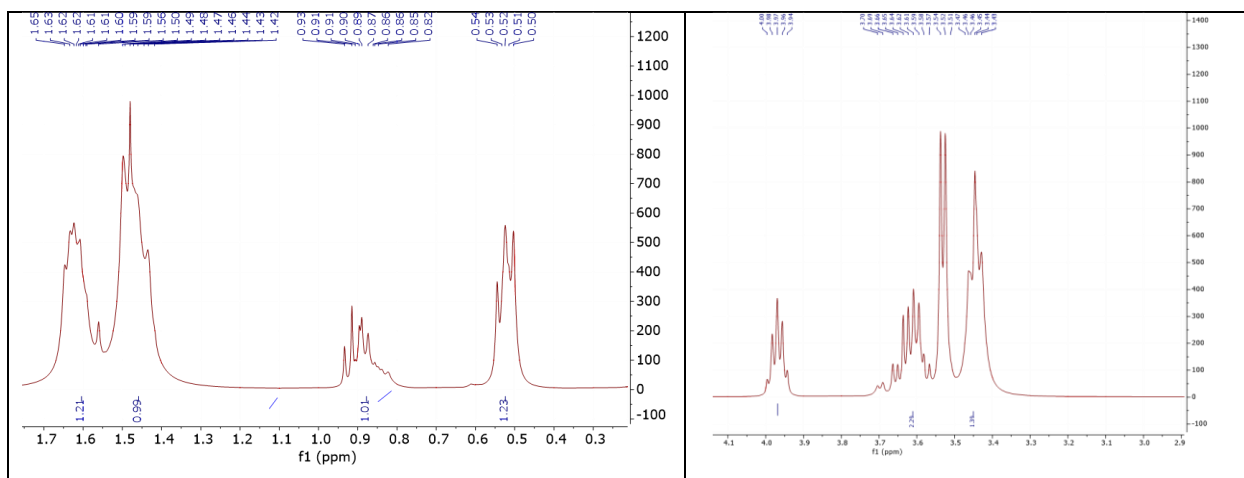
S 34: ¹³C NMR 5d₅



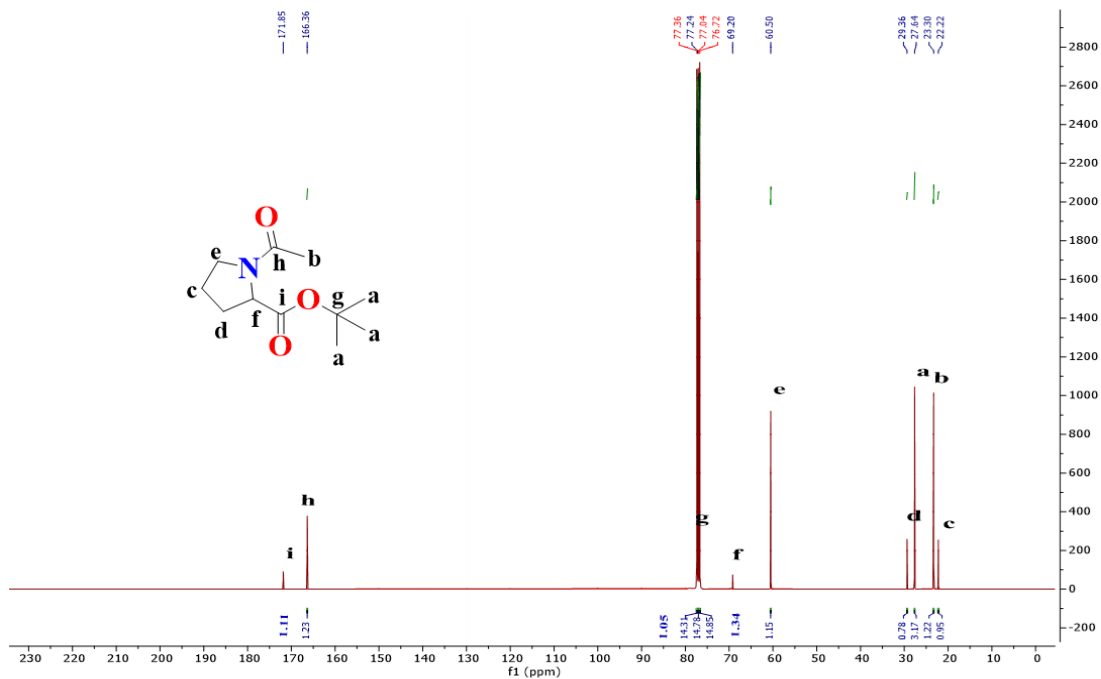
S 35: FT-IR of 5d₆



S 36: ¹H NMR 5d₆

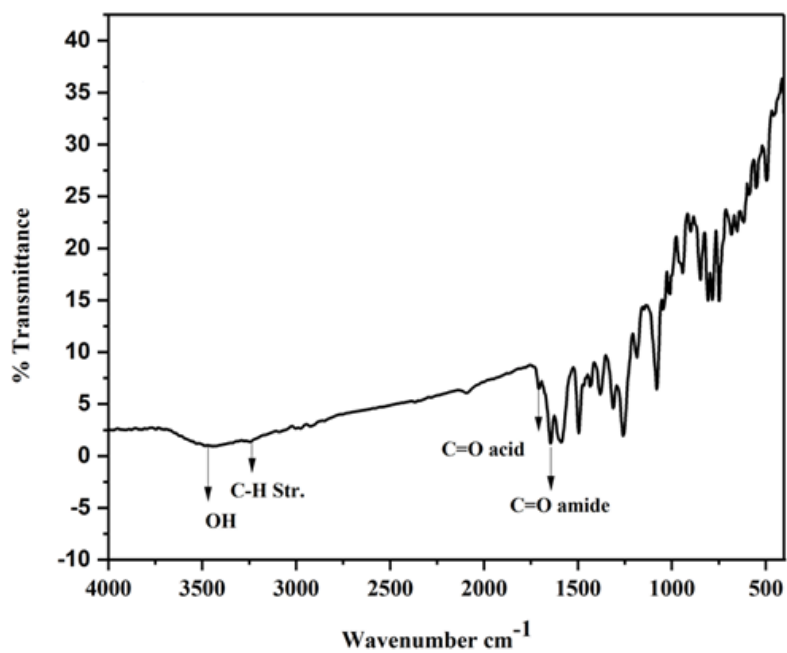


S 37: Extended $^1\text{H NMR}$ **5d6**



S 38: $^{13}\text{C NMR}$ **5d6**

FT-IR Spectroscopic Data of Acetylated Acids (De-esterified *tert*-butyl esters of *N*-acetylated amino acids)



S2. FT-IR of 6d1

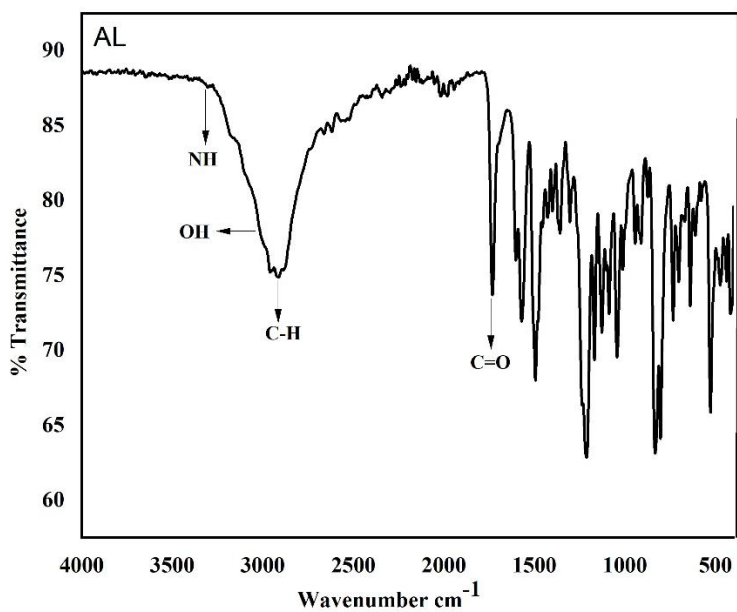
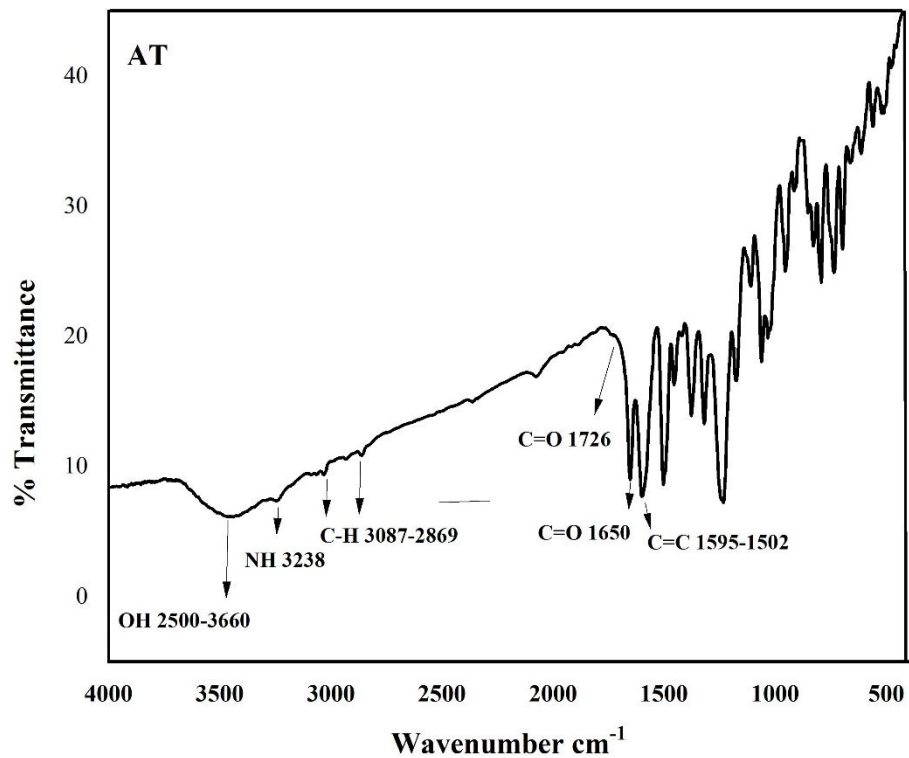
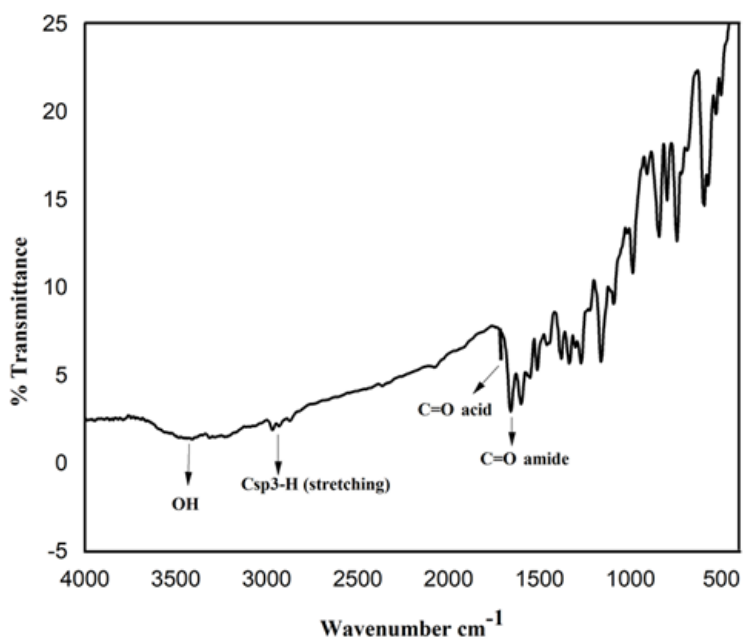


Fig. 3: FT-IR of 6d2



S4. FT-IR of 6d5



S5. FT-IR of 6d6