

Supplementary Information
for
Aerobic C–H Bond Activation at a Pd Center under
Aqueous Conditions

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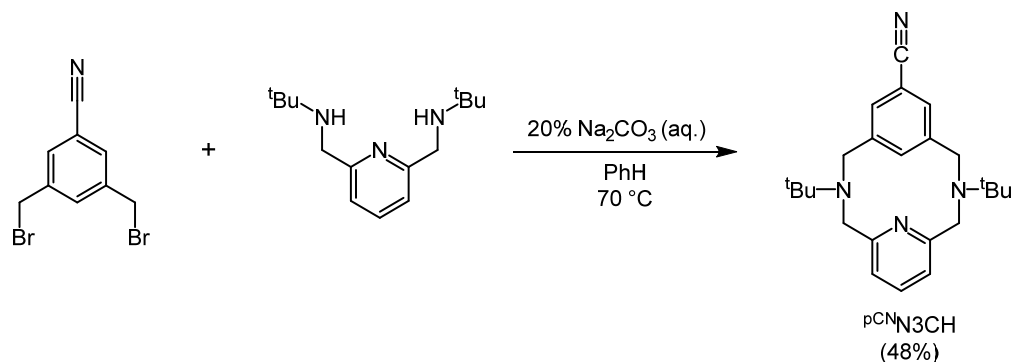
1. General Experimental Details

All operations were performed under a nitrogen atmosphere using standard Schlenk and glove box techniques if not indicated otherwise. All reagents for which the syntheses are not given were purchased from Sigma-Aldrich, Acros, STREM, or Pressure Chemical and were used as received without further purification. Solvents were purified prior to use by passing through a column of activated alumina using an MBRAUN SPS. The ligands and complex, $^{pMe}N_3CH$, $^{pH}N_3CH$, $^{pOMe}N_3CH^1$, N_3CBr^2 , and $[(^{pMe}N_3C)Pd^{III}(MeCN)_2](BF_4)_2^1$ were prepared following literature procedures.

NMR spectra were obtained on a Varian Mercury-300 spectrometer (300 MHz for 1H), a Varian Unity Inova-400 spectrometer (400 MHz for 1H), a Varian Unity Inova-500 spectrometer (500 MHz for 1H , 125 MHz for ^{13}C), or B600 Bruker NEO NMR spectrometer (600 MHz for 1H , 151 MHz for ^{13}C). Chemical shifts are reported in parts per million (ppm) with residual solvent resonance peaks as internal references. Abbreviations for the multiplicity of NMR signals are singlet (s), doublet (d), triplet (t), quartet (q), multiplet (m), broad resonance (br). EPR spectra were recorded on a Bruker 10" EMXPlus X-band Continuous Wave EPR spectrometer at 77 K. EPR spectra simulation and analysis were performed using Bruker WINEPR SimFonia program, version 1.25. UV-vis absorption spectra were recorded in 1 cm quartz cuvettes using a Varian Cary 50 Bio spectrophotometer or an Agilent Cary 60 spectrophotometer. Elemental Analysis was carried out by Intertek Pharmaceutical Services or the Microanalysis Laboratory at University of Illinois at Urbana-Champaign (UIUC) using an Exeter Analytical-Model CE440 CHN Analyzer. Cyclic voltammetry (CV) experiments were performed with a BASi EC Epsilon electrochemical workstation or a CHI 660D Electrochemical Analyzer. Electrochemical grade Bu_4NPF_6 from Fluka was used as the supported electrolyte. The electrochemical measurements were performed using a three-electrode system in a nitrogen-filled glove box. A 3 mm diameter glassy carbon disk electrode (GCE), Pt wire, and Ag wire were used as the working electrode, counter electrode, and pseudoreference electrode, respectively. The reference electrodes were calibrated against ferrocene at the end of each CV measurement. GC-MS data was collected using an Agilent 7890B GC Series System and an Agilent 5977B Mass Selective Detector. Electrospray ionization mass spectrometry (ESI-MS) was recorded on a Water Q-TOF Ultima ESI instrument by the Mass Spectrometry Laboratory at UIUC.

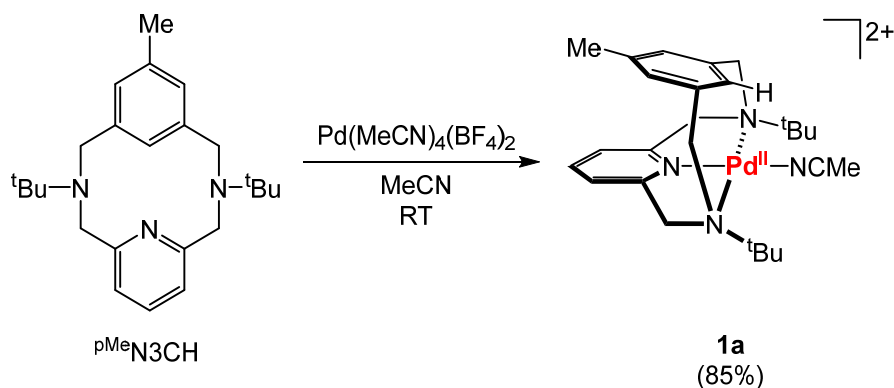
2. Synthesis of Ligands and Pd Complexes

Preparation of ${}^p\text{CN}{}^{\text{N}}\text{N}3\text{CH}$



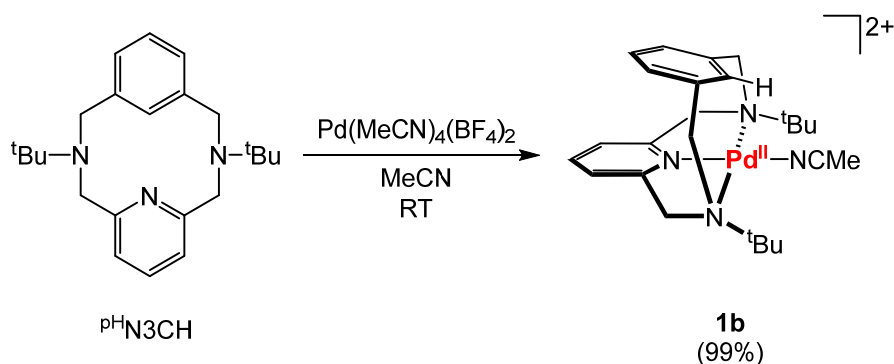
The ligand ${}^p\text{CN}{}^{\text{N}}\text{N}3\text{CH}$ was synthesized using a modified procedure from the literature.¹ A solution of bis[(*N*-*tert*-butylamino)methyl]pyridine (1.28 g, 5.14 mmol) in benzene (180 mL) and a 20% Na_2CO_3 aqueous solution (180 mL) was charged into a three-neck 1 L round-bottom flask equipped with an addition funnel and a magnetic stirring bar. The reaction mixture was stirred and heated to 70 °C, after which 3,5-bis(bromomethyl)benzonitrile (0.99 g, 3.43 mmol) in benzene (180 mL) was added dropwise to the reaction mixture over 1 h. The biphasic mixture was stirred at 70 °C for 2 days. Upon cooling to room temperature, the organic phase was separated. The organic layer was washed with concentrated K_2CO_3 (3 × 100 mL) and subsequently dried over MgSO_4 . The organic layer was then filtered and evaporated to give a sticky white-yellow solid. The solid was recrystallized in methanol at -20 °C overnight. The resulting white crystalline precipitated was filtered and dried under vacuum. Yield: 624 mg (48%). ${}^1\text{H}$ NMR (500 MHz, CDCl_3), δ : 1.31 (s, 18H, tBu-CH), 3.84 (d, J = 55.4 Hz, 8H, -CH₂-), 6.70 (d, J = 7.6 Hz, 2H), 6.98 (s, 2H), 7.13 (t, J = 7.6 Hz, 1H), 7.49 (s, 1H). ${}^{13}\text{C}$ NMR (151 MHz, CDCl_3) δ : 159.6, 140.8, 140.4, 135.9, 131.1, 122.6, 119.9, 110.4, 58.6, 56.0, 55.7, 27.8. Anal. Calcd. For $\text{C}_{24}\text{H}_{32}\text{N}_4$: C, 76.55; H, 8.57; N, 14.88; found: C, 76.92; H, 8.47; N, 14.66.

Preparation of [(p^{Me}N₃CH)Pd^{II}(MeCN)](BF₄)₂ (1a**)**



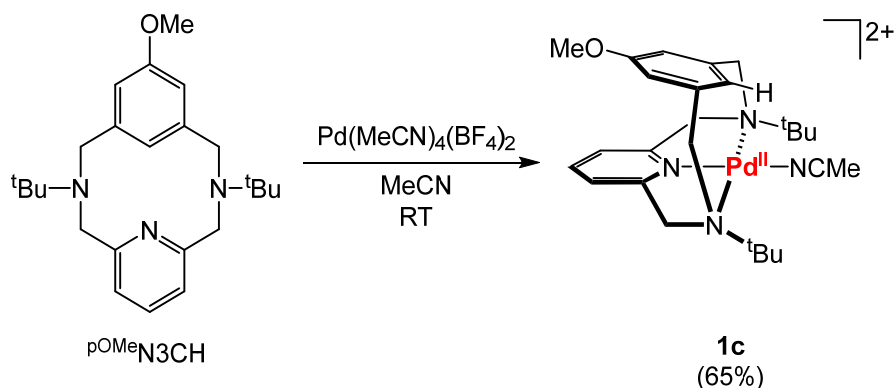
To a suspension of p^{Me}N₃CH (74.7 mg, 0.204 mmol) in MeCN (2 mL) was added a solution of Pd(MeCN)₄(BF₄)₂ (90.8 mg, 0.204 mmol) in MeCN (1 mL). The resulting yellow solution was stirred for 15 minutes. The reaction solution was then filtered through a pad of Celite. A large excess of diethyl ether was added to the filtrate and the mixture stored at -35 °C for 1.5 hours. The product crashed out of solution as a bright yellow solid. The mixture was decanted, and the isolated product was dried under vacuum. Yield: 119.3 mg (85%). ¹H NMR (300 MHz, CD₃CN), δ: 1.52 (s, 18H, tBu-CH), 1.96 (s, 3H, NC-Me) 1.98 (s, 3H, Ph-Me), 3.70-3.85 (br, 2H, -CH₂-), 3.97 (m, 2H, -CH₂-), 4.40-4.70 (br, 4H, -CH₂-), 6.81 (s, 2H, Ph-H_{meta}), 7.00 (d, *J* = 7.3 Hz, 2H, Py-H_{meta}), 7.68 (t, *J* = 7.9 Hz, 1H, Py-H_{para}). ¹³C NMR (125 MHz, CD₃COCD₃), δ: 164.8 151.3 141.7 140.7 140.3 138.5 138.5 138.2 136.5 134.1 133.1 118.2 68.4 67.5 57.0 56.9 55.7 25.3 21.2 20.5 4.5. Anal. Found: C, 46.11; H, 5.71; N, 8.63; calc. for C₂₆H₃₈B₂F₈N₄Pd: C, 45.48; H, 5.58; N, 8.16.

Preparation of $[(\text{P}^{\text{H}}\text{N}_3\text{CH})\text{Pd}^{\text{II}}(\text{MeCN})](\text{BF}_4)_2$ (1b**)**



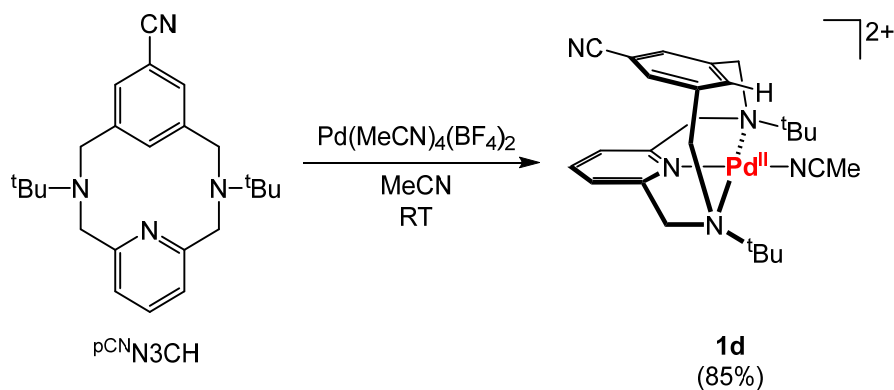
To a suspension of $\text{P}^{\text{H}}\text{N}_3\text{CH}$ (39.4 mg, 0.112 mmol) in MeCN (1 mL) was added a solution of $\text{Pd}(\text{MeCN})_4(\text{BF}_4)_2$ (49.8 mg, 0.112 mmol) in MeCN (0.75 mL). The resulting orange solution was stirred for 15 minutes. The reaction solution was then filtered through a cotton plug. A large excess of diethyl ether was added to the filtrate and the mixture stored at $-35\text{ }^\circ\text{C}$ for 1.5 hours. The product crashed out of solution as a yellow/orange solid. The mixture was decanted, and the isolated product was dried under vacuum. Yield: 75.0 mg (99%). ^1H NMR (300 MHz, CD_3CN), δ : 1.53 (s, 18H, tBu-CH), 1.97 (s, 3H), 3.80-3.95 (br, 2H, $-\text{CH}_2-$), 4.01 (m, 3H, $-\text{CH}_2-$), 4.35-4.60 (br, 3H, $-\text{CH}_2-$), 6.86 (t, $J = 7.5$ Hz, 1H, Ph- H_{para}), 6.97-7.04 (br, 4H, Ph- H_{meta} and Py- H_{meta}), 7.57 (t, $J = 7.8$ Hz, 1H, Py- H_{para}), 9.63 (br, 1H). ^{13}C NMR (125 MHz, CD_3COCD_3), δ : 164.7, 151.0, 141.3, 139.7, 138.9, 138.2, 136.2, 133.5, 133.2, 131.5, 130.0, 118.9, 68.5, 67.3, 57.0, 56.8, 55.7, 25.3, 15.4, 4.5. Anal. Found: C, 44.44; H, 5.58; N, 8.30; calc. for $\text{C}_{25}\text{H}_{36}\text{B}_2\text{F}_8\text{N}_4\text{Pd}$: C, 44.64; H, 5.40; N, 8.33.

Preparation of [(^pOMeN₃CH)Pd^{II}(MeCN)](BF₄)₂ (1c**)**



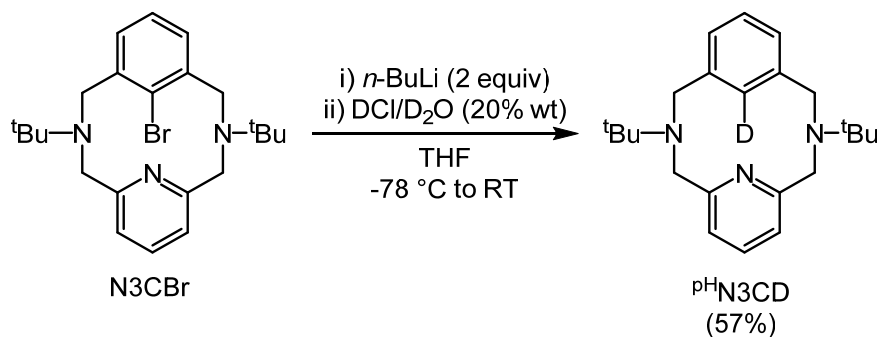
To a solution of ^pOMeN₃CH (50.0 mg, 0.131 mmol) in MeCN (2.5 mL) was added a solution of Pd(MeCN)₄(BF₄)₂ (58.2 mg, 0.131 mmol in MeCN (2.5 mL)). The resulting yellow solution was stirred for 15 minutes. The reaction solution was then filtered through a pad of Celite. A large excess of diethyl ether was added to the filtrate and the mixture stored at -35 °C for 1.5 hours. The product crashed out of solution as an off-yellow solid. The mixture was decanted, and the isolated product was dried under vacuum. Yield: 76.5 mg (83%). ¹H NMR (500 MHz, CD₃SOCD₃), δ: 1.46 (s, 18H, tBu-CH), 2.07 (s, 3H), 3.58 (s, 3H, OMe-H), 3.63-4.40 (m, 6H, -CH₂-), 3.99 (d, *J* = 16.6 Hz, 2H, -CH₂-), 6.50 (s, 2H, Ph-H_{meta}), 7.02 (d, *J* = 7.5 Hz, 2H, Py-H_{meta}), 7.61 (t, *J* = 7.8 Hz, 1H, Py-H_{para}), 9.16 (s, 1H). ¹³C NMR (151 MHz, CD₃SOCD₃), δ: 164.5, 160.0, 138.2, 136.6, 128.6, 128.6, 118.1, 117.8, 55.2, 55.2, 26.5, 24.5, 1.13. Anal. Found: C, 44.68; H, 5.64; N, 8.17; calc. for C₂₆H₃₈B₂F₈N₄OPd: C, 44.44; H, 5.45; N, 7.97.

Preparation of $[(p^{CN}N_3CH)Pd^{II}(MeCN)](BF_4)_2$ (**1d**)



To a solution of $p^{CN}N_3CH$ (50.0 mg, 0.133 mmol) in MeCN (1 mL) was added a solution of $Pd(MeCN)_4(BF_4)_2$ (59.0 mg, 0.133 mmol) in MeCN (1 mL). The resulting yellow solution was stirred for 15 minutes. The reaction solution was then filtered through a pad of Celite. A large excess of diethyl ether was added to the filtrate and the mixture was stored at $-35\text{ }^\circ\text{C}$ for 1.5 hours. The product crashed out of solution as an off-yellow solid. The mixture was decanted, and the isolated product was dried under vacuum. Yield: 79.0 mg (85%). ^1H NMR (500 MHz, CD_3CN), δ : 1.49 (br, 18H, tBu-CH), 1.96 (s, 3H), 3.82-4.54 (m, 8H), 7.10 (br, 2H), 7.36 (s, 2H), 7.68 (t, $J=7.8$ Hz, 1H), 9.61 (s, 1H). ^{13}C NMR (151 MHz, CD_3CN at $-20\text{ }^\circ\text{C}$), δ : 167.1, 163.5, 141.5, 140.9, 140.4, 138.1, 135.0, 128.5, 127.5, 121.4, 121.0, 119.3, 118.3, 114.1, 72.8, 65.6, 61.7, 57.5, 57.3, 54.8, 28.7, 27.4, 24.9, 23.8. Anal. Found: C, 44.28; H, 5.09; N, 9.98; calc. for $C_{26}H_{35}B_2F_8N_5Pd$: C, 44.76; H, 5.06; N, 10.04.

Preparation of $^p\text{H}^{\text{N}}\text{3CD}$



The synthesis of $^p\text{H}^{\text{N}}\text{3CD}$ ligand was modified from the literature.² A solution of N3CBr (100 mg, 0.232 mmol) in tetrahydrofuran (4 mL) was cooled to -78 °C. The two-equivalent amount of *n*-butyllithium (290.4 μL of 1.6 M in hexanes, 0.465 mmol, 2.0 equiv) was added dropwise to the mixture. The mixture was allowed to warm to room temperature while stirring. After reaching room temperature, the solution was stirred over 30 minutes. An excessive amount of DCl/D₂O (20 w/w%, 300 μL) was added to the mixture and the mixture stirred for an additional 10 minutes. The mixture was cooled using ice/water bath followed by the addition of saturated aqueous NaOH solution (1 mL). The organic layer was extracted with dichloromethane (3×1 mL). The organic layer was dried over MgSO₄. The organic layer was then filtered and evaporated to give white-yellow solid. The solid was washed with pentane (2×0.5 mL) then dried to afford white solid. The deuterium incorporation was greater than 99% which was determined by GC-MS isotope distribution or ¹H NMR (see Figure S1, S2, S13, and S14.). Yield: 47 mg (57%). ¹H NMR (400 MHz, C₆D₆), δ : 1.17(s, 18H), 3.77 (s, 4H), 3.95 (s, 4H), 6.65 (d, $J= 7.6$ Hz, 2H), 6.81-6.94 (m, 4H).

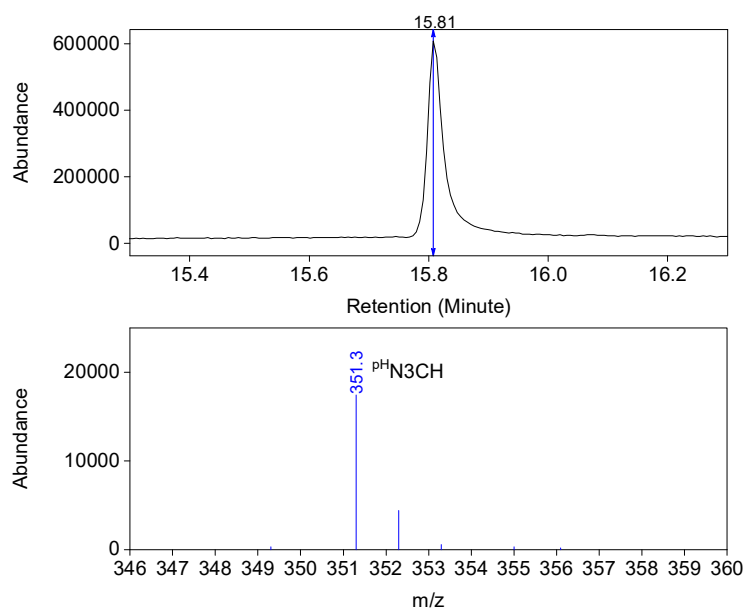


Figure S1. GC spectrum (top) and isotope distribution spectrum in MS (bottom) of $p^H N_3CH$.

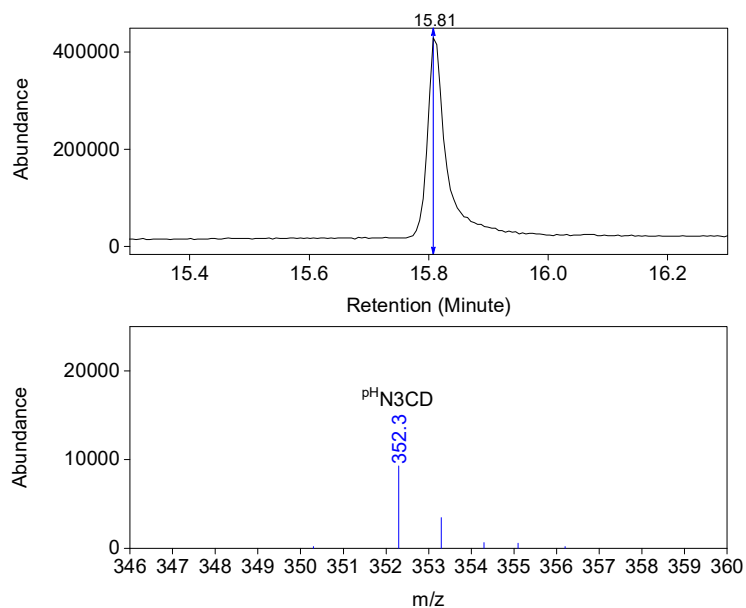


Figure S2. GC spectrum (top) and isotope distribution spectrum in MS (bottom) of $p^H N_3CD$.

3. NMR Spectral Data

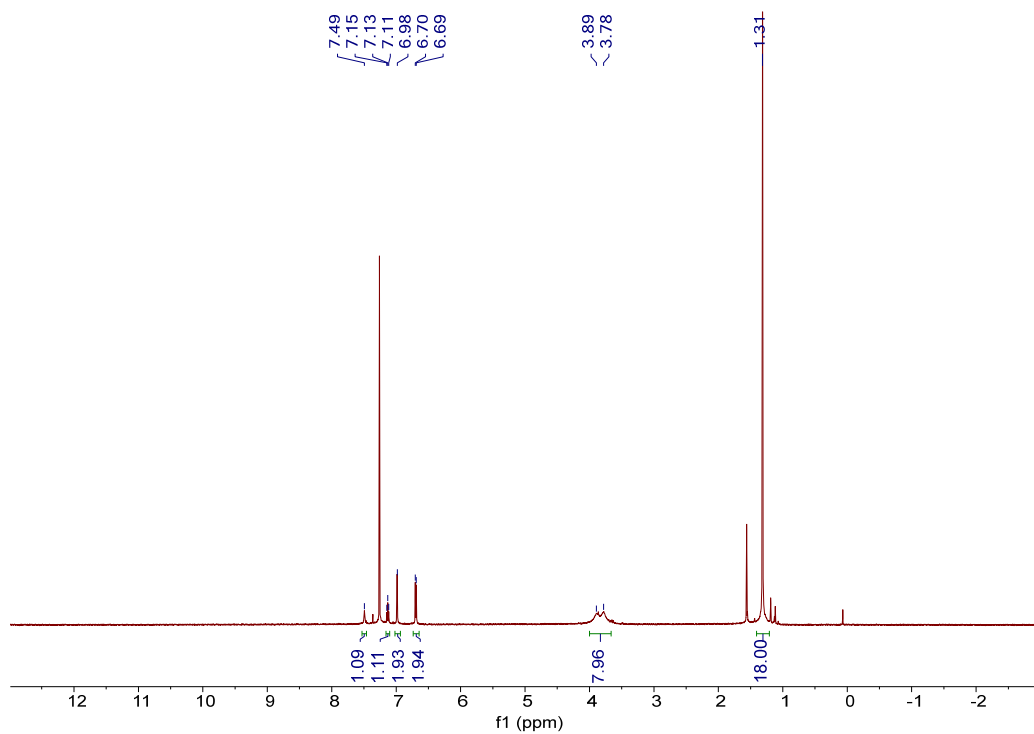


Figure S3. ¹H NMR spectrum of pCN-N3CH at 23 °C (CDCl₃).

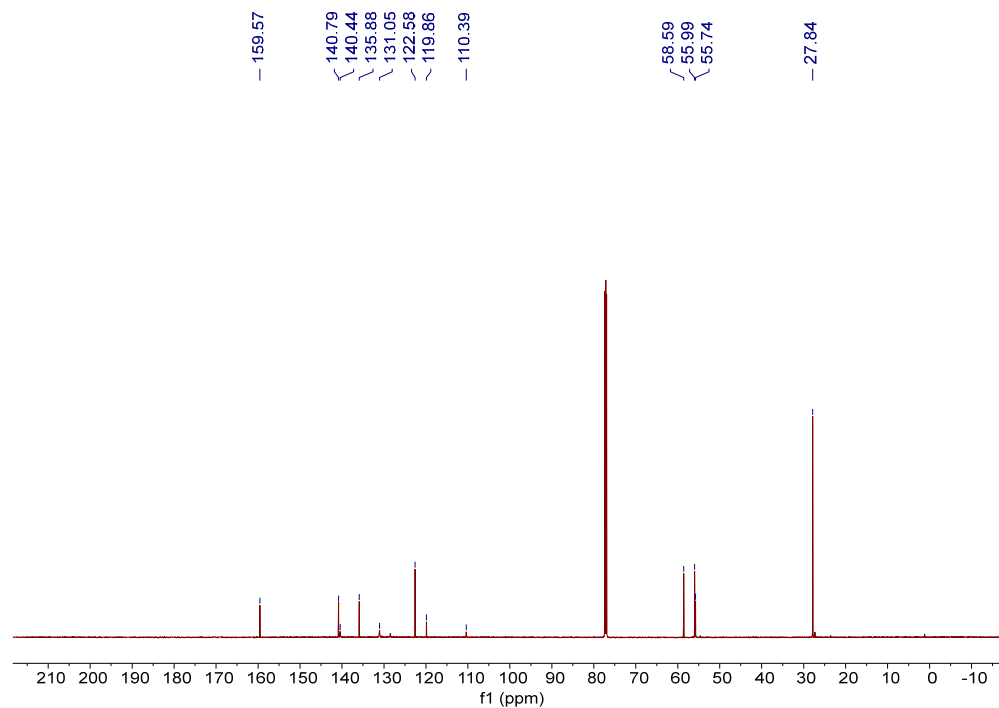


Figure S4. ¹³C NMR spectrum of pCN-N3CH at 23 °C (CDCl₃).

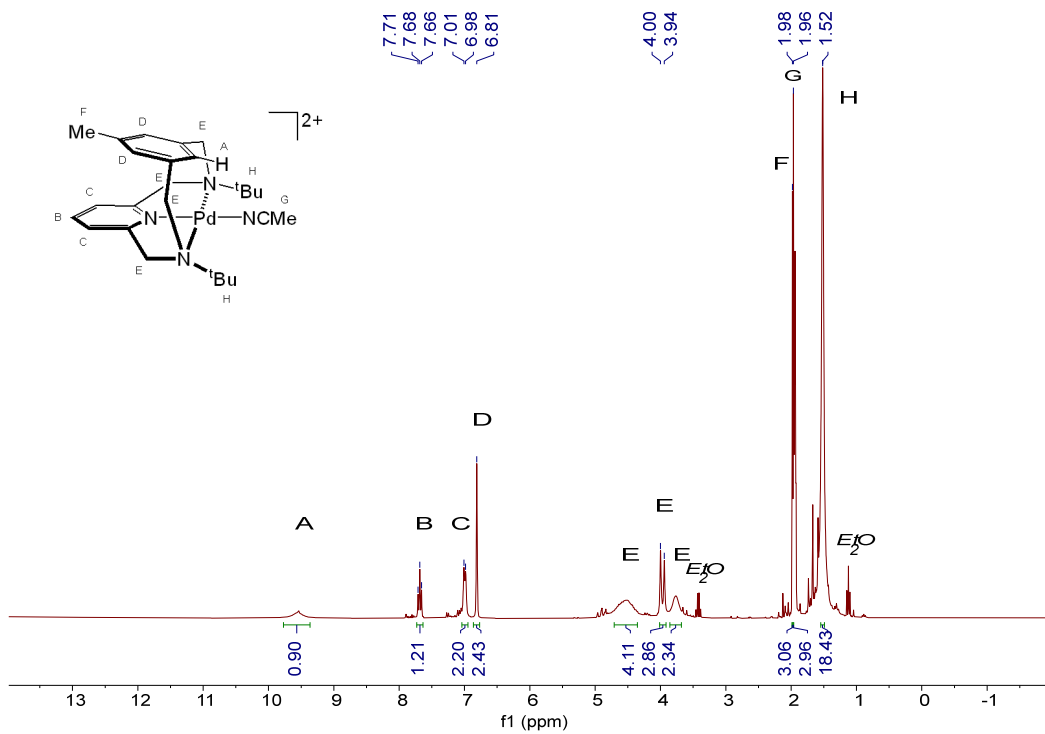


Figure S5. ^1H NMR spectrum of **1a** at 23 °C (CD_3CN).^a

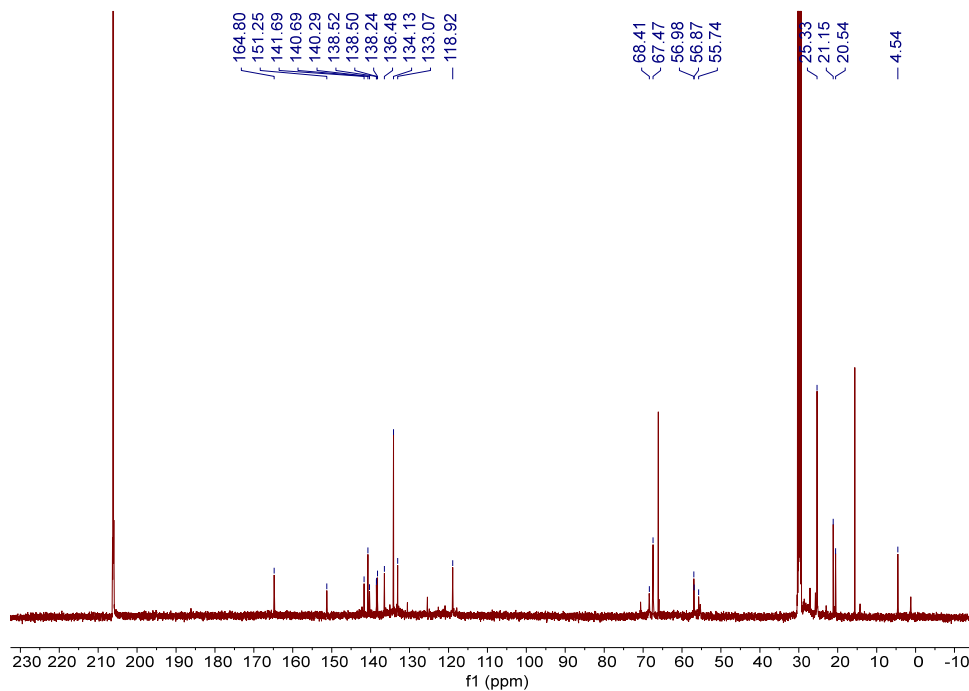


Figure S6. ^{13}C NMR spectrum of **1a** (CD_3COCD_3).

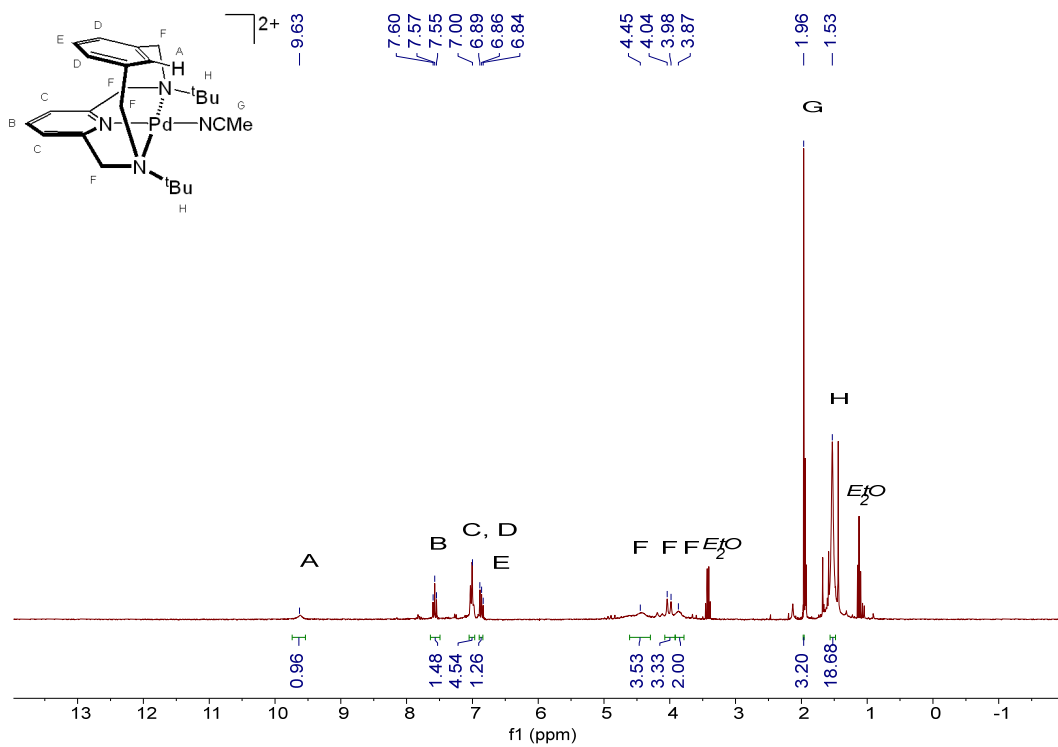


Figure S7. ¹H NMR spectrum of **1b** at 23 °C (CD₃CN).^a

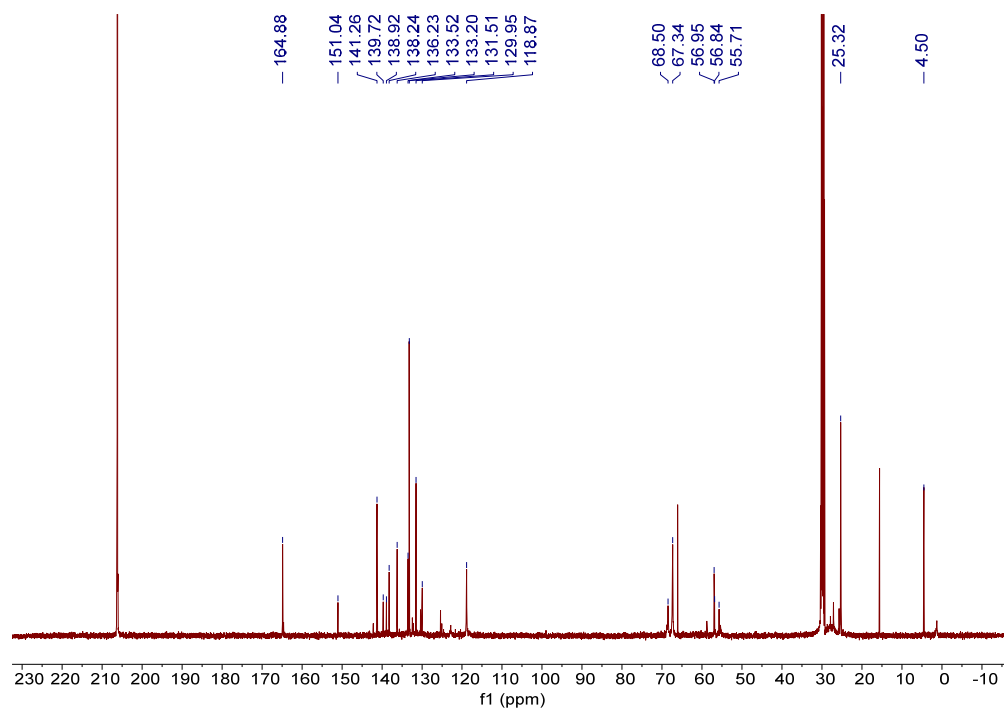


Figure S8. ¹³C NMR spectrum of **1b** (CD₃COCD₃).

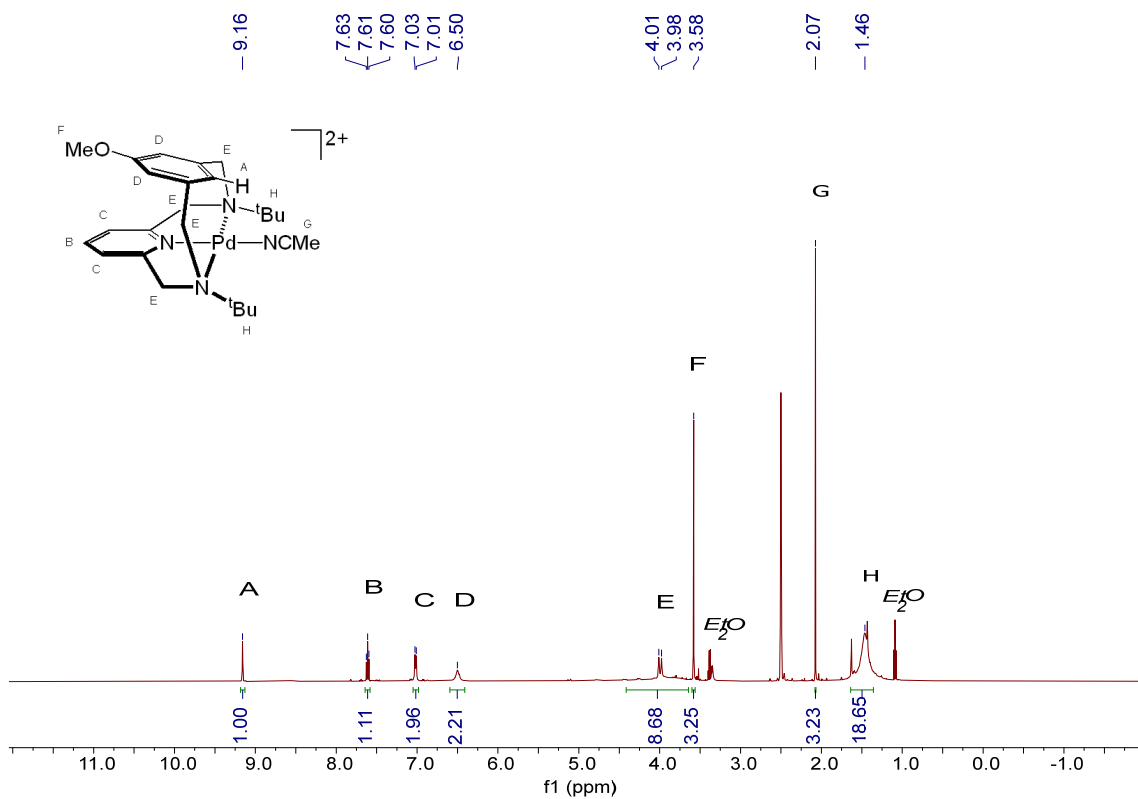


Figure S9. ¹H NMR spectrum of **1c** at 23 °C (CD₃SOCD₃).^a

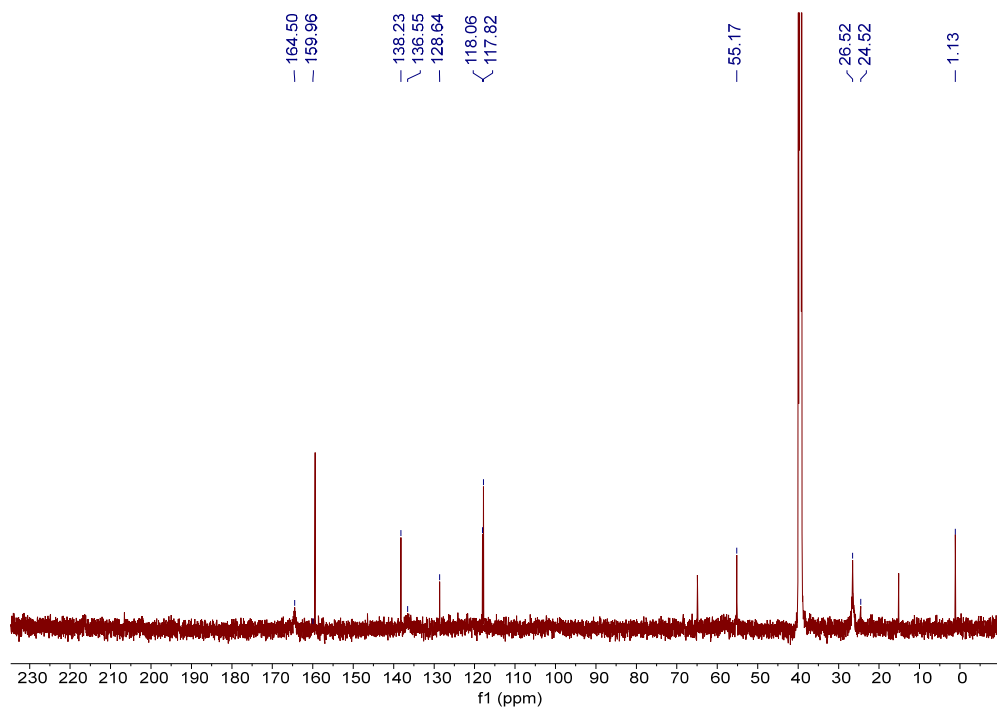


Figure S10. ¹³C NMR spectrum of **1c** at 23 °C (CD₃SOCD₃)

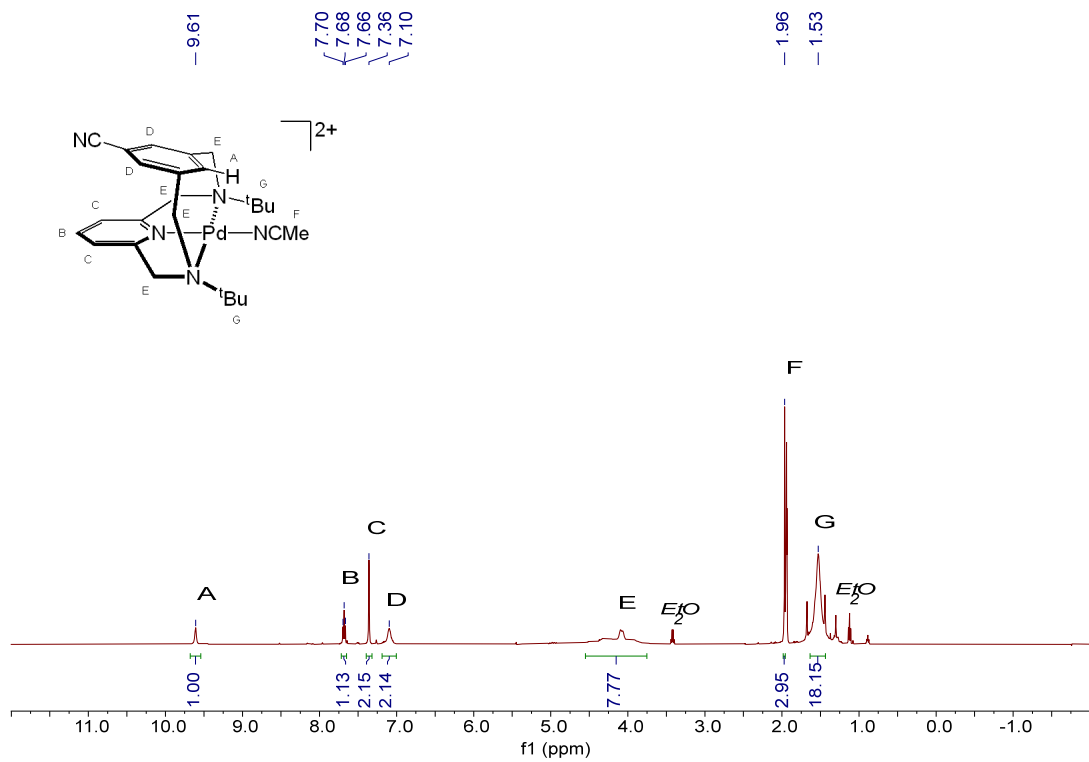


Figure S11. ^1H NMR spectrum of **1d** at 23 °C (CD₃CN).^a

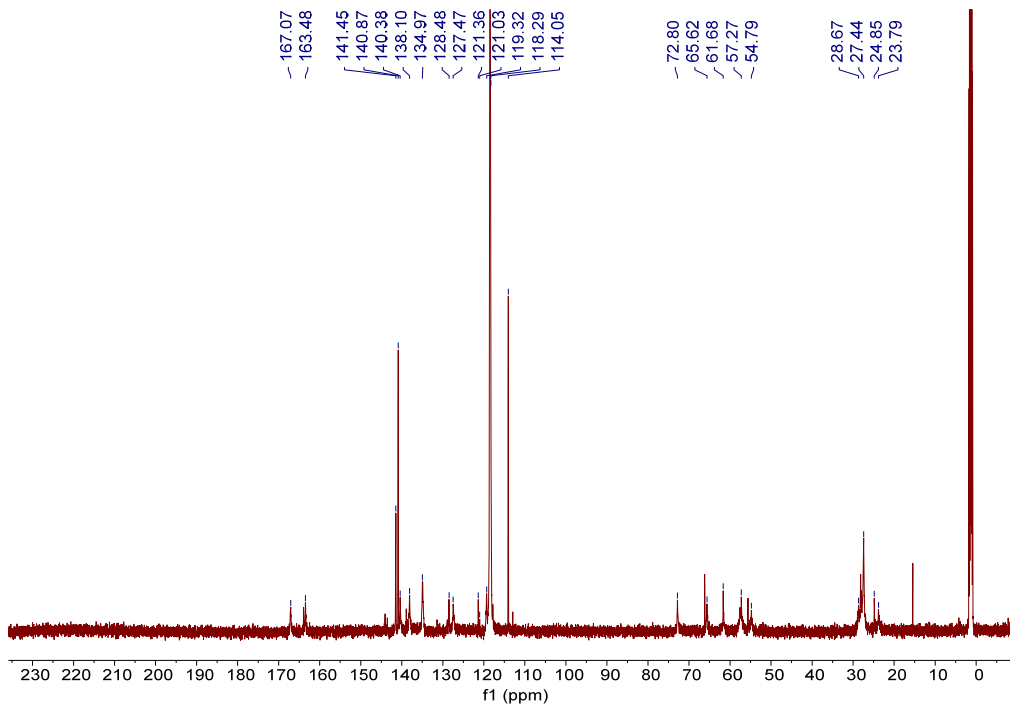


Figure S12. ^{13}C NMR spectrum of **1d** at -20 °C (CD₃CN).

^a The additional resonances and baseline features observed in the aryl and alkyl regions of the ¹H NMR spectra of **1a-d** are attributed to minor, Pd^{II} coordination species, such as **1ab**, that arise from MeCN coordination/ligand exchange and/or conformational flexibility of the N₃CH ligand framework. These weak features do not affect the assignment of the major diagnostic resonances of complexes **1a-d**.

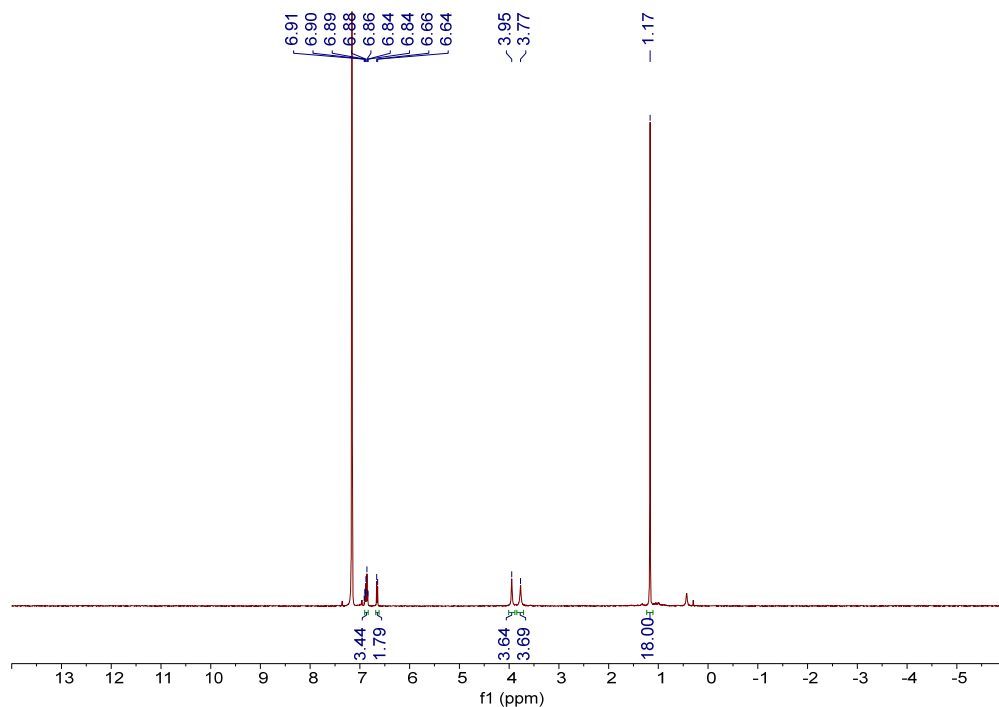


Figure S13. ¹H NMR spectrum of p^HN₃CD at 23 °C (C₆D₆).

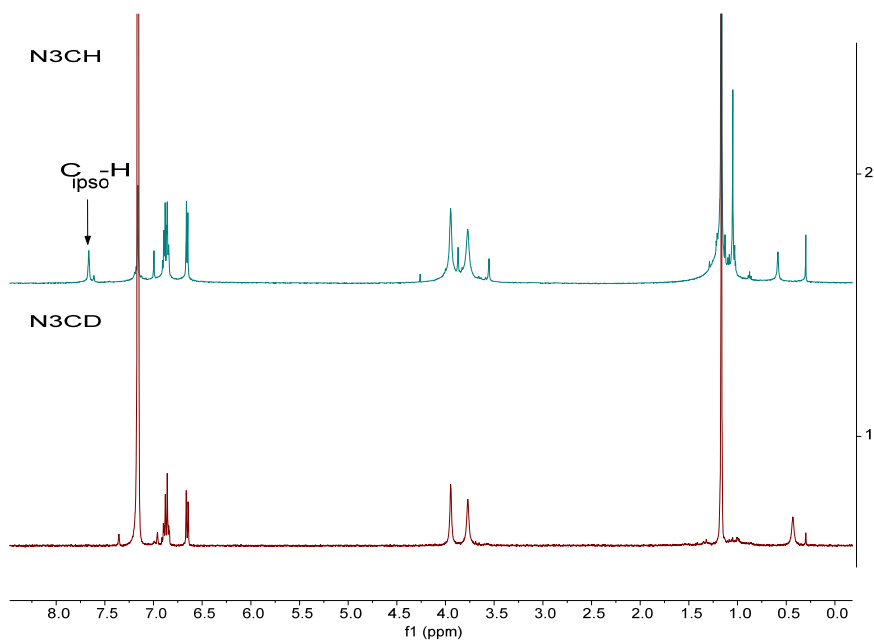


Figure S14. Comparison of ¹H NMR spectra of p^HN₃CH (top) and p^HN₃CD (bottom) at 23 °C (C₆D₆).

4. Electrochemical Characterization

Table S1. Summary of electrochemical data^a

Complex	Oxidative feature (V)	Reductive feature (V)
1a	$E_{pa,ox} = 1.094$	$E_{1/2,red} = -0.424$
2a	$E_{1/2,ox} = 0.818$	$E_{1/2,red} = -0.159$

^a Measured in 0.1 M *n*Bu₄NPF₆ acetonitrile solution with a scan rate of 0.1 V/s. All potentials are reported relative to Fc⁺⁰. The irreversible oxidative feature of **1a** is reported as the anodic peak potential, $E_{pa,ox}$. Reversible or quasi-reversible waves are reported as $E_{1/2}$ values.

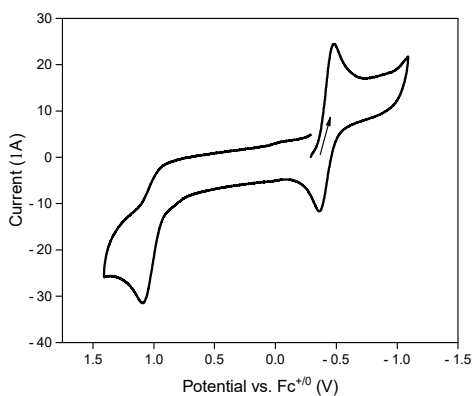


Figure S15. CV of **1a** in 0.1 M *n*Bu₄NPF₆/MeCN (scan rate= 100 mV/s).

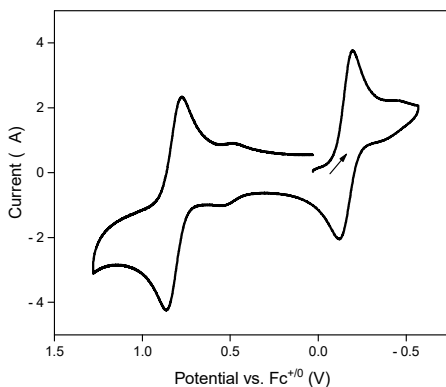


Figure S16. CV of **2a** in 0.1 M *n*Bu₄NPF₆/MeCN (scan rate= 100 mV/s).

5. UV-Vis and Kinetic Studies of the Aerobic Oxidation of **1**

a) In situ preparation of $[(^{pMe}N_3CH)Pd^{II}(MeCN)](BF_4)_2$ (**1a**) solution

Unless specified otherwise, all solutions for Pd^{II} reactant complexes were prepared according to the procedure outlined below. To a suspension of $^{pMe}N_3CH$ (4.6 mg, 0.0126 mmol) in MeCN (800 μ L) was added a solution of $Pd(MeCN)_4(BF_4)_2$ (5.6 mg, 0.0126 mmol) in MeCN (800 μ L). The resulting yellow solution was stirred at 880 rpm with a stir bar for 5 minutes to afford 7.86 mM stock solution **1a**. An aliquot (334 μ L) was extracted and diluted with 2166 μ L of MeCN to yield a 1.05 mM solution. Various concentrations of solutions were prepared by adjusting the volumes of aliquots from the stock solution and MeCN accordingly. For example, for 0.26 mM solution, 83 μ L of stock solution, 2417 μ L of MeCN; for 0.52 mM, 165 μ L of stock solution, 2335 μ L of MeCN; for 0.75 mM, 239 μ L of stock solution, 2261 μ L of MeCN; for 2.1 mM, 668 μ L of stock solution, 1832 μ L of MeCN were used. Due to slow decomposition into palladium black, only stock solutions prepared within 30 minutes were utilized for $[(^{pMe}N_3CH)Pd^{II}(MeCN)](BF_4)_2$. The same procedures were followed for other $[(^{pR}N_3CH)Pd^{II}(MeCN)](BF_4)_2$ (**1b-d**) solutions.

b) Aerobic oxidation of **1** and its UV-Vis monitoring

The solution of **1** (2 mL, 0.26-2.1 mM) in MeCN was placed into a quartz cuvette (10 mm path length) equipped with a septum-sealed cap and a magnetic stir bar. To control the O_2 concentration in MeCN, gases with different oxygen/nitrogen ratio were used. The ratio of O_2/N_2 used were 100/0, 75/25, 50/50, and 20/80 to make 8.1,³ 6.1, 4.1, and 2.4⁴ mM of oxygen in MeCN. Oxygen balloons and an oxygen/nitrogen gas mixer were used to match each ratio. The balloons underwent five rounds of oxygen purging, while the mixer was flushed with gas 30 seconds prior to utilization. The inlet needle (22 G) connecting to the balloon or the purging mixer and the outlet (23 G) needle were inserted orderly while the tip of inlet needle was placed in bottom of the solution and that of outlet needle was placed to the top of the headspace. Each gas was then bubbled through the solution for 45 seconds then needles were removed in the reverse order to make the cuvette a closed system, and the reaction mixture was stirred under closed gas system. The reaction progress was monitored by UV-Vis spectroscopy while stirring at 170 rpm with a stir bar. For the initial rate method, early time data points were obtained more than those of late and reactions that have progressed less than 5% was used for data to measure the initial rate.⁵ The concentration of $[(^{pR}N_3CPd^{III})(MeCN)_2](BF_4)_2$ during the reaction was calculated based on the extinction coefficient of the 550 nm absorption band (Figure S17, $\epsilon = 2128 \text{ M}^{-1}\text{cm}^{-1}$ in MeCN, $2148 \text{ M}^{-1}\text{cm}^{-1}$ in 9:1 $H_2O:MeCN$). The value of the extinction coefficient of the 550 nm absorption band has been updated from the previous literature¹ by

measuring thoroughly using highly purified single crystal of the complex **2a**. The same procedures were followed for other $[(P^R N_3 CH)Pd^{II}(MeCN)](BF_4)_2$ (**1b-d**) solutions.

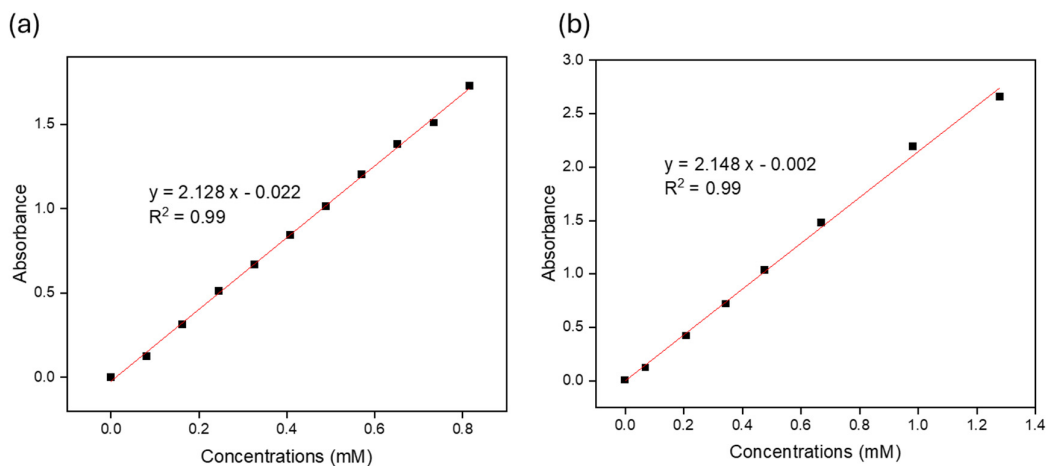


Figure S17. Beer-Lambert plots of the absorbance at 550 nm for complex **2a**. (a) MeCN ($\epsilon = 2,128 \text{ M}^{-1}\text{cm}^{-1}$); (b) 9:1 H₂O:MeCN ($\epsilon = 2,148 \text{ M}^{-1}\text{cm}^{-1}$).

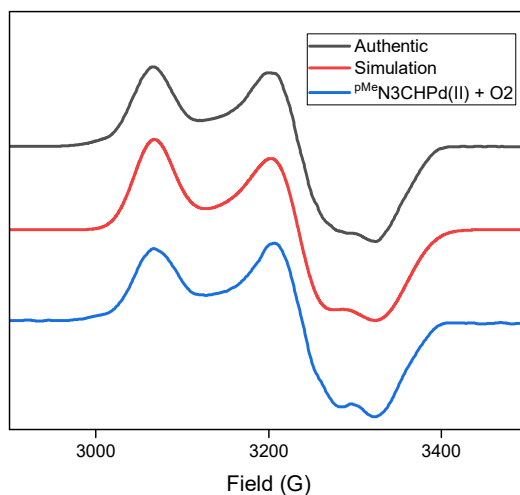


Figure S18. EPR spectrum of **2a** in 3:1 PrCN/MeCN at 77 K using the following parameters: $g_x = 2.193$, $g_y = 2.079$, $g_z = 2.018$ ($A_z(2N) = 15.0 \text{ G}$). Black: isolated **2a** from the reference.¹ Red: simulated spectrum of Pd^{III}. Blue: In situ generated **2a** by the reaction mixture of **1a** with O₂.

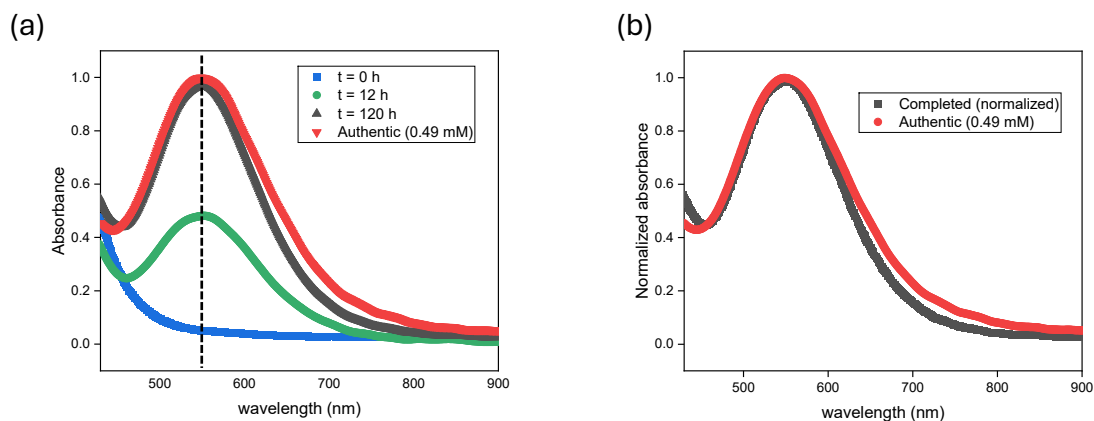


Figure S19. (a) Time-dependent UV-Vis spectra for the conversion of **1a** to **2a** in MeCN at 293 K, showing spectra at $t = 0$ (blue), an intermediate time point ($t = 12$ h, green), and at completion ($t = 120$ h, gray), overlaid with the spectrum of an authentic sample of **2a** (0.49 mM, red). The dashed line indicates the 550 nm wavelength. (b) Normalized overlay of the final reaction spectrum and authentic **2a** (normalized at 550 nm), highlighting the agreement in spectral shape at the monitoring wavelength.

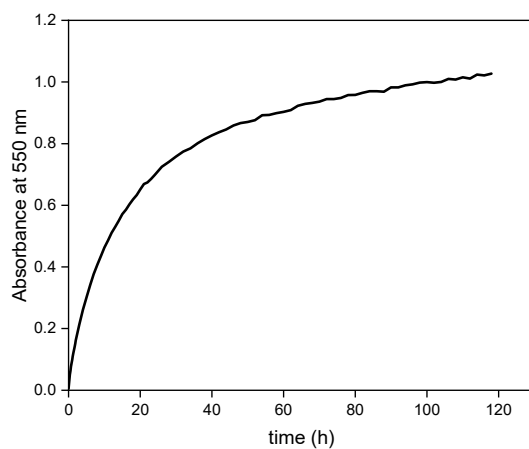


Figure S20. Absorbance (550 nm)-time graph for aerobic oxidation of **1a** at 1.05 mM O_2 -saturated (8.1 mM) acetonitrile at 293 K, monitored for the entire course of the reaction.

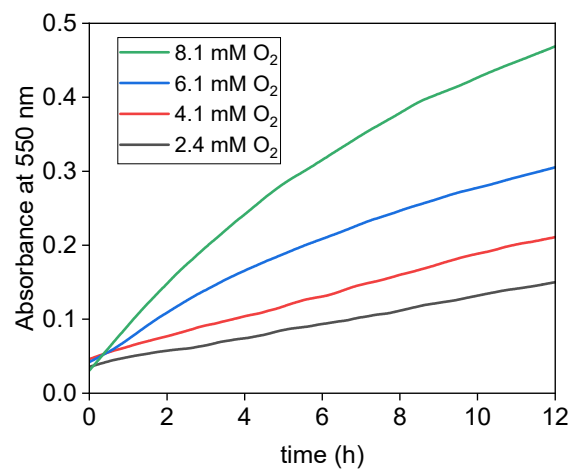


Figure S21. Absorbance (550 nm)-time graph for aerobic oxidation of **1a** (1.05 mM) with various oxygen concentrations in acetonitrile at 293 K.

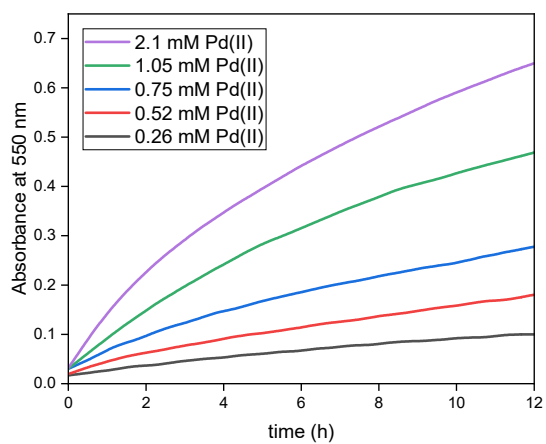


Figure S22. Absorbance (550 nm)-time graph for aerobic oxidation of **1a** at various concentrations in the O₂-saturated (8.1 mM) acetonitrile at 293 K.

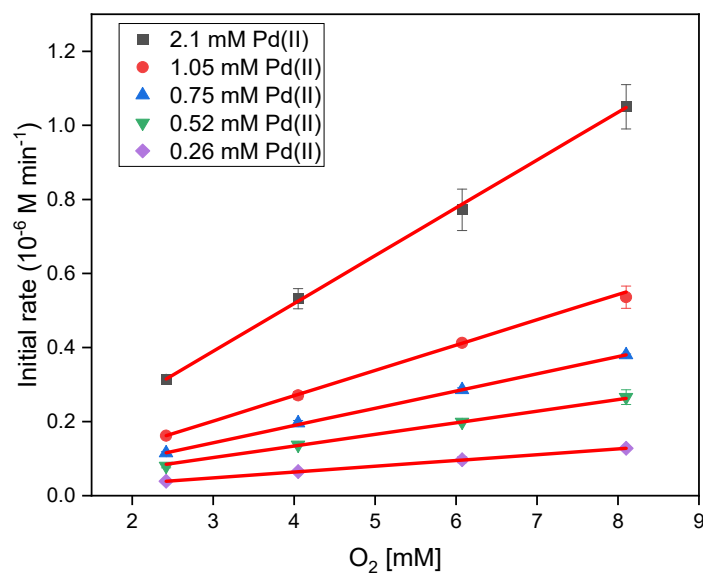


Figure S23. Plot of Initial rate vs. $[O_2]$ of the aerobic oxidation of **1a** at 293 K. $[Pd] = 0.26$ - 2.1 mM.

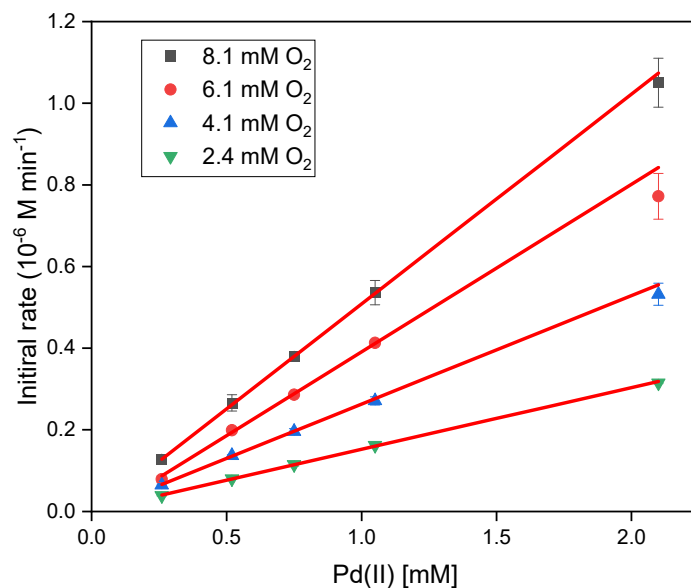


Figure S24. Plot of Initial rate vs. $[Pd^{II}]$ of the aerobic oxidation of **1a** at 293 K. $[O_2] = 2.4$ - 8.1 mM.

Table S2. Initial rates (unit: $10^{-9} M s^{-1}$) and rate constants for **1a** + O_2 at 293 K, $[Pd^{II}]$ varied, $[O_2] = 8.1$ mM.

1a (mM)	O_2 (mM)	Run 1	Run 2	Run 3	Average ($10^{-9} M s^{-1}$)	k_{obs} ($10^{-3} M^{-1} s^{-1}$)
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2.1	8.1	18.0	16.5	18.2	17.5 ± 0.9	1.03 ± 0.06
1.05		8.70	9.48	8.62	8.93 ± 0.48	1.05 ± 0.06
0.75		6.30	6.38	6.32	6.33 ± 0.04	1.04 ± 0.01
0.52		4.07	4.55	4.68	4.43 ± 0.32	1.05 ± 0.01
0.26		2.15	2.13	2.13	2.14 ± 0.01	1.02 ± 0.005

Table S3. Initial rates (unit: 10^{-9} M s^{-1}) and rate constants for **1a** + O₂ at 293 K, [Pd^{II}] varied, [O₂]= 6.08 mM.

1a (mM)	O ₂ (mM)	Run 1	Run 2	Run 3	Average (10^{-9} M s^{-1})	k _{obs} ($10^{-3} \text{ M}^{-1} \text{ s}^{-1}$)
2.1	6.08	11.8	13.7	13.1	12.9 ± 0.9	1.01 ± 0.07
1.05		6.77	6.95	6.92	6.88 ± 0.10	1.08 ± 0.02
0.75		4.82	4.68	4.83	4.78 ± 0.08	1.05 ± 0.02
0.52		3.37	3.38	3.23	3.33 ± 0.08	1.05 ± 0.03
0.26		1.77	1.53	1.53	1.61 ± 0.13	1.02 ± 0.09

Table S4. Initial rates (unit: 10^{-9} M s^{-1}) and rate constants for **1a** + O₂ at 293 K, [Pd^{II}] varied, [O₂]= 4.05 mM.

1a (mM)	O ₂ (mM)	Run 1	Run 2	Run 3	Average (10^{-9} M s^{-1})	k _{obs} ($10^{-3} \text{ M}^{-1} \text{ s}^{-1}$)
2.1	4.05	9.07	9.18	8.37	8.87 ± 0.44	1.04 ± 0.05
1.05		4.35	4.68	4.52	4.52 ± 0.17	1.06 ± 0.04
0.75		3.15	3.38	3.25	3.26 ± 0.12	1.07 ± 0.04
0.52		2.27	2.32	2.28	2.29 ± 0.03	1.09 ± 0.01
0.26		1.05	1.05	1.12	1.07 ± 0.04	1.02 ± 0.04

Table S5. Initial rates (unit: 10^{-9} M s^{-1}) and rate constants for **1a** + O₂ at 293 K, [Pd^{II}] varied, [O₂]= 2.42 mM.

1a (mM)	O ₂ (mM)	Run 1	Run 2	Run 3	Average (10^{-9} M s^{-1})	k _{obs} ($10^{-3} \text{ M}^{-1} \text{ s}^{-1}$)
2.1	2.42	5.22	5.30	5.23	5.25 ± 0.04	1.03 ± 0.01
1.05		2.70	2.70	2.70	2.70 ± 0.001	1.06 ± 0.001
0.75		1.87	1.98	1.90	1.92 ± 0.06	1.06 ± 0.03
0.52		1.32	1.33	1.35	1.33 ± 0.02	1.06 ± 0.01
0.26		0.63	0.65	0.65	0.64 ± 0.01	1.02 ± 0.02

Table S6. Initial rate comparison of [**1a**] and [O₂] variation at 293 K (unit: 10^{-9} M s^{-1}).

[O ₂], mM	[1a]=2.10 mM	[1a]=1.05 mM	[1a]=0.75 mM	[1a]=0.52 mM	[1a]=0.26 mM
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8.1	17.5 ± 0.9	8.93 ± 0.48	6.33 ± 0.04	4.43 ± 0.32	2.14 ± 0.01
6.08	12.9 ± 0.9	6.88 ± 0.10	4.78 ± 0.08	3.33 ± 0.08	1.61 ± 0.13
4.05	8.87 ± 0.44	4.52 ± 0.17	3.26 ± 0.12	2.29 ± 0.03	1.07 ± 0.04
2.42	5.25 ± 0.04	2.70 ± 0.001	1.92 ± 0.06	1.33 ± 0.02	0.64 ± 0.01

Table S7. Rate constant (k_{obs}) comparison of [**1a**] and [O_2] variation at 293 K (unit: $10^{-3} \text{ M}^{-1} \text{ s}^{-1}$).

[O_2], mM	[1a]=2.10 mM	[1a]=1.05 mM	[1a]=0.75 mM	[1a]=0.52 mM	[1a]=0.26 mM
8.1	1.03 ± 0.06	1.05 ± 0.06	1.04 ± 0.01	1.05 ± 0.01	1.02 ± 0.005
6.08	1.01 ± 0.07	1.08 ± 0.02	1.05 ± 0.02	1.05 ± 0.03	1.02 ± 0.09
4.05	1.04 ± 0.05	1.06 ± 0.04	1.07 ± 0.04	1.09 ± 0.01	1.02 ± 0.04
2.42	1.03 ± 0.01	1.06 ± 0.001	1.06 ± 0.03	1.06 ± 0.01	1.02 ± 0.02

c) Determination of Hammett ρ Value and the Kinetic Isotope Effect (KIE)

Table S8. Initial rates (unit: 10^{-9} M s^{-1}) and rate constants of reactions of $\text{Pd}^{\text{II}} + \text{O}_2$.

Pd complex	Pd (mM)	O_2 (mM)	Run 1	Run 2	Run 3	Average (10^{-9} M s^{-1})	k_{obs} ($10^{-3} \text{ M}^{-1} \text{ s}^{-1}$)
1a	1.05	8.1	8.70	9.48	8.62	8.93 ± 0.48	1.05 ± 0.06
1b			4.60	4.88	4.90	4.79 ± 0.17	0.56 ± 0.02
1c			14.2	14.8	14.5	14.5 ± 0.3	1.71 ± 0.03
1d			0.65	0.70	0.65	0.67 ± 0.03	0.08 ± 0.003
1b-D			1.87	2.02	1.83	1.95 ± 0.08	0.23 ± 0.01

Table S9. Initial rate and rate constant comparisons for $^{\text{pOMe}}\text{N}_3\text{CH}$, $^{\text{pMe}}\text{N}_3\text{CH}$, $^{\text{pH}}\text{N}_3\text{CH}$, and $^{\text{pCN}}\text{N}_3\text{CH}$ ligand systems ($[\text{Pd}]=1.05 \text{ mM}$, $[\text{O}_2]=8.1 \text{ mM}$ in all experiments) at 293 K.

Ligand system	Pd	σ_{para}	Initial rate (10^{-9} M s^{-1})	k_{obs} ($10^{-3} \text{ M}^{-1} \text{ s}^{-1}$)
$^{\text{pOMe}}\text{N}_3\text{CH}$	1c	-0.268	14.5 ± 0.3	1.71 ± 0.03
$^{\text{pMe}}\text{N}_3\text{CH}$	1a	-0.17	8.93 ± 0.48	1.05 ± 0.06
$^{\text{pH}}\text{N}_3\text{CH}$	1b	0.0	4.79 ± 0.17	0.56 ± 0.02
$^{\text{pCN}}\text{N}_3\text{CH}$	1d	0.66	0.67 ± 0.03	0.08 ± 0.003

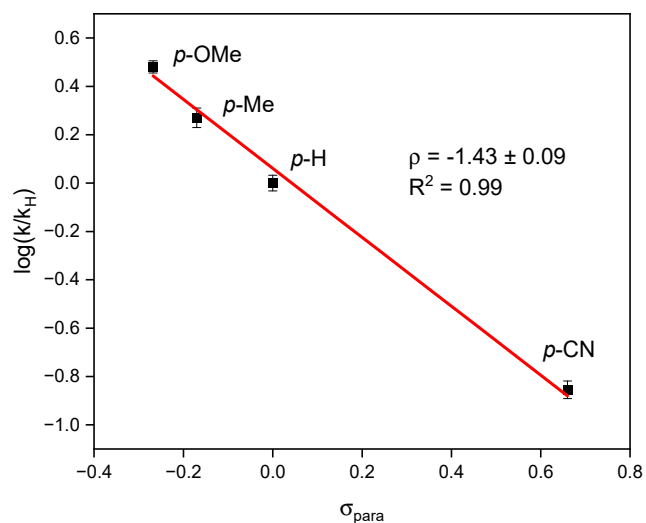


Figure S25. Hammett analysis of $\log(k/k_{\text{H}})$ vs. substituent constant, σ for *para*-substituted $[(^{\text{pR}}\text{N}_3\text{CH})\text{Pd}^{\text{II}}(\text{MeCN})](\text{BF}_4)_2$ in the aerobic C–H activation reactions at 293 K.

Table S10. Kinetic isotope effect experiments ($[\text{Pd}^{\text{II}}]=1.05 \text{ mM}$, $[\text{O}_2]=8.1 \text{ mM}$) at 293 K.

Pd	Initial rate (10^{-9} M s^{-1})	k_{obs} ($10^{-3} \text{ M}^{-1} \text{ s}^{-1}$)
1b	4.79 ± 0.17	0.56 ± 0.02
1b-D	1.95 ± 0.08	0.23 ± 0.01
KIE (kH/kD)	2.5 ± 0.1	2.5 ± 0.1

d) Derivation of the rate law for aerobic oxidation of **1**

$$\text{rate} = k_2[{}^3\text{Int1}] \quad (1)$$

$$\frac{d[{}^3\text{Int1}]}{dt} \approx 0 = k_1[\mathbf{1}][\text{O}_2] - (k_{-1} + k_2)[{}^3\text{Int1}] \quad (2)$$

$$[{}^3\text{Int1}] = \frac{k_1[\mathbf{1}][\text{O}_2]}{k_{-1} + k_2}$$

$$\text{rate} = \frac{k_1 k_2}{k_{-1} + k_2} [\mathbf{1}][\text{O}_2] \quad (3)$$

6. Aerobic Oxidation of **1** under Various Reaction Conditions

Aerobic oxidation of **1a** using H₂O:MeCN (v:v) solvent mixture with various ratio

a) Aerobic oxidation of **1a** with O₂ in 1:9 H₂O:MeCN conditions

Solution of **1a** (1.17 mM) in MeCN was prepared and 1.8 mL of the solution was placed into a 4 mL quartz cuvette equipped with a septum-sealed cap and a magnetic stir bar followed by the addition of 0.2 mL of degassed DI water to make the solution 1.05 mM. Oxygen balloon was prepared and the inlet needle (22 G) connecting to the balloon and the outlet (23 G) needle were inserted orderly while the tip of inlet needle was placed in bottom of the solution and that of outlet needle was placed to the top of the headspace. Oxygen was then bubbled through the solution for 45 seconds then needles were removed in the reverse order to make the cuvette a closed system, and the reaction mixture was stirred under closed gas system. The reaction progress was monitored by UV-Vis spectroscopy while stirring at 170 rpm with a stir bar.

b) Aerobic oxidation of **1a** with O₂ in 2:8 H₂O:MeCN conditions

Solution of **1a** (1.318 mM) in MeCN was prepared and 1.6 mL of solution **1a** was placed into a 4 mL quartz cuvette equipped with a septum-sealed cap and a magnetic stir bar followed by the addition of 0.4 mL of degassed DI water to make the solution 1.05 mM. Oxygen balloon was prepared and the inlet needle (22 G) connecting to the balloon and the outlet (23 G) needle were inserted orderly while the tip of inlet needle was placed in bottom of the solution and that of outlet needle was placed to the top of the headspace. Oxygen was then bubbled through the solution for 45 seconds then needles were removed in the reverse order to make the cuvette a closed system, and the reaction mixture was stirred under closed gas system. The reaction progress was monitored by UV-Vis spectroscopy while stirring at 170 rpm with a stir bar.

c) Aerobic oxidation of **1a** with O₂ in 5:5 H₂O:MeCN conditions

Solution of **1a** (2.1 mM) in MeCN was prepared and 1.0 mL of solution **1a** was placed into a 4 mL quartz cuvette equipped with a septum-sealed cap and a magnetic stir bar followed by the addition of 1.0 mL of degassed DI water to make the solution 1.05 mM. Oxygen balloon was prepared and the inlet needle (22 G) connecting to the balloon and the outlet (23 G) needle were inserted orderly while the tip of inlet needle was placed in bottom of the solution and that of outlet needle was placed to the top of the headspace. Oxygen was then bubbled through the solution for 45 seconds then needles were removed in the reverse order to make the cuvette a closed system, and the reaction mixture was stirred under closed gas system. The reaction progress was monitored by UV-Vis spectroscopy while stirring at 170 rpm with a stir bar.

d) Aerobic oxidation of **1a** with O₂ in 9:1 H₂O:MeCN conditions

To a suspension of ^pMeN₃CH (6.2 mg, 0.0169 mmol) in MeCN (800 μL) was added a solution of Pd(MeCN)₄(BF₄)₂ (7.5 mg, 0.0169 mmol) in MeCN (800 μL). The resulting yellow solution was stirred at 880 rpm with a stir bar for 5 minutes to afford 10.5 mM stock solution **1a**. An aliquot (200 μL) of the stock solution was placed into a 4 mL quartz cuvette equipped with a septum-sealed cap and a magnetic stir bar followed by the addition of 1.8 mL of degassed DI water to make the solution 1.05 mM. Oxygen balloon was prepared and the inlet needle (22 G) connecting to the balloon and the outlet (23 G) needle were inserted orderly while the tip of inlet needle was placed in bottom of the solution and that of outlet needle was placed to the top of the headspace. Oxygen was then bubbled through the solution for 45 seconds then needles were removed in the reverse order to make the cuvette a closed system, and the reaction mixture was stirred under closed gas system. The reaction progress was monitored by UV-Vis spectroscopy while stirring at 170 rpm with a stir bar.

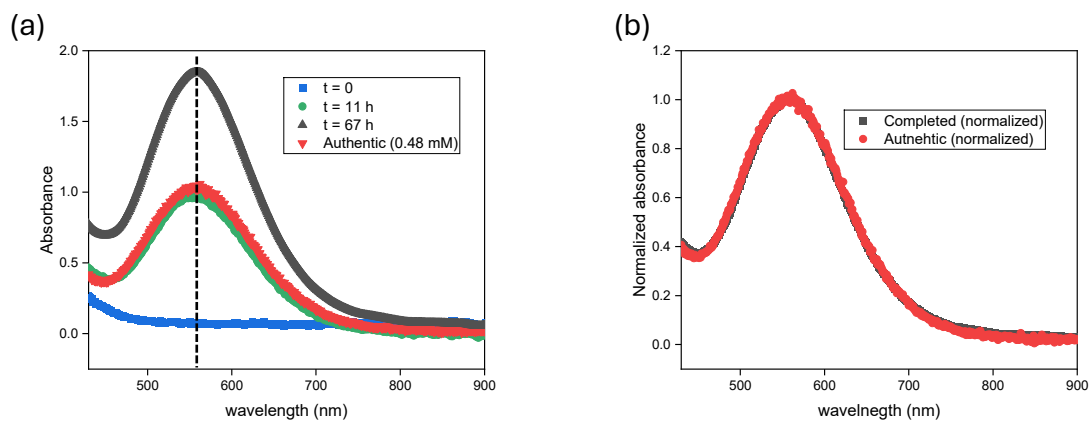


Figure S26. (a) Time-dependent UV-Vis spectra for the conversion of **1a** to **2a** in 9:1 H₂O:MeCN at 293 K, showing spectra at t = 0 (blue), an intermediate time point (t = 11 h, green), and at completion (t = 67 h, gray), overlaid with the spectrum of an authentic sample of **2a** (0.48 mM, red). The dashed line indicates the 550 nm wavelength. (b) Normalized overlay of the final reaction spectrum and authentic **2a** (normalized at 550 nm), highlighting the agreement in spectral shape at the monitoring wavelength.

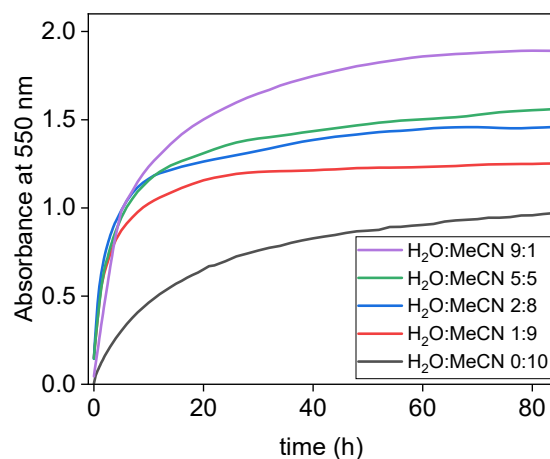


Figure S27. Absorbance (550 nm)-time graph of **1a** (1.05 mM) + O₂ in various H₂O:MeCN solvent mixture ratio.

Table S11. Yield of **2a** in various H₂O:MeCN ratio conditions. Yield was measured when the development of absorbance at 550 nm was fully completed.

H ₂ O:MeCN	Yield (%)
9:1	90
5:5	75
2:8	70
1:9	56
0:10	46

e) Aerobic oxidation of **1a** with O₂ in 9:1 H₂O:MeCN at different temperature

The procedure was identical to that described in part d), except that the reaction temperature was varied up to 70 °C. The concentration of O₂ in the 9:1 H₂O:MeCN solvent mixture at each temperature was estimated by a weighted average of the saturated O₂ concentrations in pure H₂O and MeCN, respectively. The solubilities of O₂ in each solvent at various temperatures were obtained from literature sources,^{3, 6-8} and the solubility of O₂ in MeCN does not vary to large extent in the temperature range of interest (305.51 K to 353.13 K, 9.52 mM to 10.9 mM).⁸

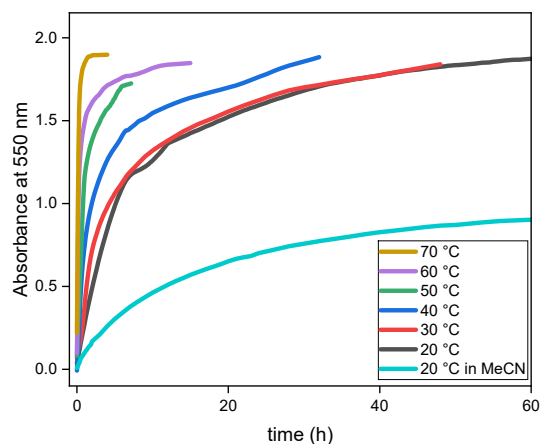


Figure S28. Time course of the 550 nm absorbance of **1a** (1.05 mM) + O₂ in 9:1 H₂O:MeCN mixture at different temperatures, and reference graph of **1a** (1.05 mM) + O₂ in MeCN (cyan).

Table S12. Yield of **2a** in 9:1 H₂O:MeCN at different temperature. Yield was measured when the development of absorbance at 550 nm was fully completed.

Temp (°C)	Reaction completion time (h)	Yield (%)
20	67	90
30	48	82
40	32	85
50	6	81
60	5	81
70	1.5	90

Table S13. Initial rate (unit: 10⁻⁹ M s⁻¹) and rate constant in the reaction of **1a** (1.05 mM) + O₂ in 9:1 H₂O:MeCN at different temperatures.

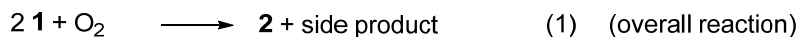
T (K)	1a (mM)	O ₂ (mM)	Run1	Run2	Run3	Average (10 ⁻⁹ M s ⁻¹)	k _{obs} (10 ⁻³ M ⁻¹ s ⁻¹)
293	1.05	1.91	32.5	35.7	31.1	33.1 ± 2.3	16.5 ± 1.2
303		2.04	56.3	47.9	48.5	50.9 ± 4.7	23.8 ± 2.2
313		1.91	119.9	115.0	109.8	114.9 ± 5.0	57.3 ± 2.5
323		1.87	206.2	214.3	227.9	216.1 ± 11.0	110 ± 5.6
333		1.82	659.3	641.1	713.4	671.3 ± 37.6	351 ± 20
343		1.81	1469.1	1393.1	1611.8	1491.3 ± 111.1	785 ± 58

Full Time-Course Kinetic Analysis and Comparison with Initial Rate Method

To carefully evaluate the validity of the initial rate treatment, the full time-course data were analyzed by nonlinear least-squares fitting to a second-order kinetic model. The rate constants derived from the initial rate method were then compared with those obtained from the global fitting analysis.

MeCN conditions

In MeCN conditions, the reaction proceeds through the following stepwise processes:



To enable global fitting of the full time-course data, we derived the integrated rate expression for a second-order kinetic model, which was used to simulate the product concentration vs time profile. Mass balance relationships were incorporated to ensure accurate kinetic fitting.

$$\text{Let } A = [\mathbf{1}], B = [\text{O}_2], \text{ and } P = [\mathbf{2}]. \quad ([\mathbf{1}] \neq [\text{O}_2])$$

From the overall reaction (eq. 1), the observed rate the corresponding second-order rate constant (k_{obs}) can be expressed as follows using the steady-state approximation:

$$\text{Observed rate} = \frac{dP}{dt} = \frac{k_1 k_2}{k_{-1} + k_2} [\mathbf{1}][\text{O}_2] = k_{obs} [\mathbf{1}][\text{O}_2] = k_{obs} AB \quad (5)$$

$A = A_0 - 2P$, $B = B_0 - P$, where A_0 , B_0 are initial concentrations of $\mathbf{1}$ and O_2 respectively.

$$\frac{dP}{dt} = k_{obs} [A_0 - 2P][B_0 - P] \quad (6)$$

$$\ln \left(\frac{(A_0 - 2P)B_0}{(B_0 - P)A_0} \right) = (A_0 - 2B_0)k_{obs}t \quad (7)$$

$$\text{Let } E = \exp((A_0 - 2B_0)k_{obs}t) \quad (8)$$

$$\frac{(A_0 - 2P)B_0}{(B_0 - P)A_0} = E(t) \quad (9)$$

$$P = \frac{A_0 B_0 (E - 1)}{E A_0 - 2B_0} \quad (10)$$

$$P = \frac{A_0 B_0 (\exp((A_0 - 2B_0)k_{obs}t) - 1)}{A_0 \exp((A_0 - 2B_0)k_{obs}t) - 2B_0} \quad (11)$$

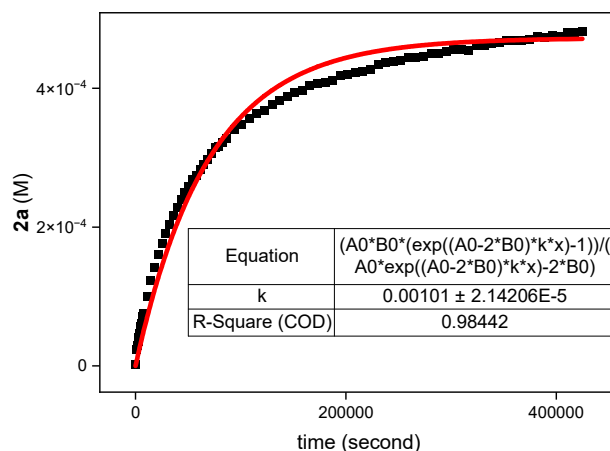


Figure S29. Nonlinear least-squares global fit of eq. 11 (red curve) to the experimental [2a] versus time data (black dotted) for the reaction of **1a** (1.05 mM) and O₂ (8.1 mM) in acetonitrile at 293 K. The red curve represents the simulated concentration profile obtained from the second-order kinetic model.

The integrated rate expression described above (eq. 11) was fitted to the experimental [2] versus time data obtained from UV-Vis spectroscopy. Nonlinear least-squares fitting afforded a global rate constant of

$$k_{\text{global}} = (1.01 \pm 0.02) \times 10^{-3} \text{ M}^{-1}\text{s}^{-1}.$$

This value is in good agreement with the rate constant obtained from the initial rate method,

$$k_{\text{initial}} = (1.05 \pm 0.06) \times 10^{-3} \text{ M}^{-1}\text{s}^{-1},$$

Supporting the validity of the initial rate approximation under the present experimental conditions.

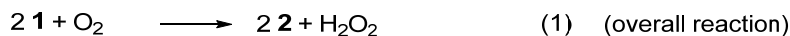
Under MeCN conditions, the overall reaction (eq. 1) produces one equivalent of Pd^{III} product (**2**) per equivalent of O₂ and half equivalents of Pd^{II} (**1**) consumed. The stoichiometry-normalized rate and its rate constant in MeCN (k_{MeCN}) are expressed as follows:

$$\frac{dP}{dt} = -\frac{1}{2} \frac{dA}{dt} = -\frac{dB}{dt} = k_{\text{MeCN}}[\mathbf{1}][\text{O}_2] = k_{\text{MeCN}}AB \quad (12)$$

From eq. 5 and eq. 12, the stoichiometry-normalized rate constant (k_{MeCN}) is identical to the observed rate constant under these MeCN conditions.

9:1 H₂O:MeCN mixture conditions

Under 9:1 H₂O:MeCN conditions, the reaction proceeds according to the following overall process:



To enable global fitting of the full time-course data, we derived the integrated rate expression for a second-order kinetic model (eq. 7), which was used to simulate the product concentration versus time profile. Mass balance relationships were incorporated to ensure accurate kinetic fitting.

Let A = [1], B = [O₂], and P = [2]. ([1] ≠ [O₂])

$$A = A_0 - P, \quad B = B_0 - 0.5P$$

From the overall reaction (eq. 1), the observed rate and experimentally observed second-order rate constant (k_{obs}) can be expressed as follows using the steady-state approximation:

$$\text{Observed rate} = \frac{dP}{dt} = \frac{k_1 k_2}{k_{-1} + k_2} [\mathbf{1}][\text{O}_2] = k_{obs} [\mathbf{1}][\text{O}_2] = k_{obs} AB \quad (2)$$

$$= k(A_0 - P)(B_0 - \frac{1}{2}P) \quad (3)$$

$$\frac{1}{(A_0 - P)(B_0 - \frac{1}{2}P)} dp = k_{obs} dt \quad (4)$$

$$\ln \frac{(B_0 - \frac{1}{2}P)A_0}{(A_0 - P)B_0} = k_{obs} \left(B_0 - \frac{A_0}{2} \right) t \quad (5)$$

$$\frac{(B_0 - \frac{1}{2}P)A_0}{(A_0 - P)B_0} = \exp \left(k_{obs} \left(B_0 - \frac{A_0}{2} \right) t \right) = E(t) \quad (6)$$

$$P = \frac{2A_0B_0(E - 1)}{2B_0E - A_0} \quad (7)$$

The integrated rate expression (eq. 7) was fitted to the experimental [2] versus time data obtained from UV-Vis spectroscopy using nonlinear least-squares analysis. The rate constants obtained from global fitting (k_{global}) were compared with those determined independently from the initial rate method ($k_{initial}$) at each reaction temperature (Table S14). The two sets of values are in good agreement, supporting the validity of the initial rate approximation under the present experimental conditions.

Under aqueous conditions (9:1 H₂O:MeCN), the overall reaction (eq. 1) produces two equivalents of Pd^{III} (**2**) per one equivalent of O₂ and two equivalents of Pd^{II} (**1**) consumed by the proposed mechanism. The stoichiometry-normalized rate and its rate constant in 9:1 H₂O:MeCN mixture conditions (k_{H_2O}) are expressed as follows:

$$\frac{1}{2} \frac{dP}{dt} = -\frac{1}{2} \frac{dA}{dt} = -\frac{dB}{dt} = k_{H_2O}[\mathbf{1}][O_2] = k_{H_2O}AB \quad (8)$$

From eq. 2 and eq. 8, the stoichiometry-normalized reaction rate constant (k_{H_2O}) is half of the observed rate constant under these 9:1 H₂O:MeCN conditions

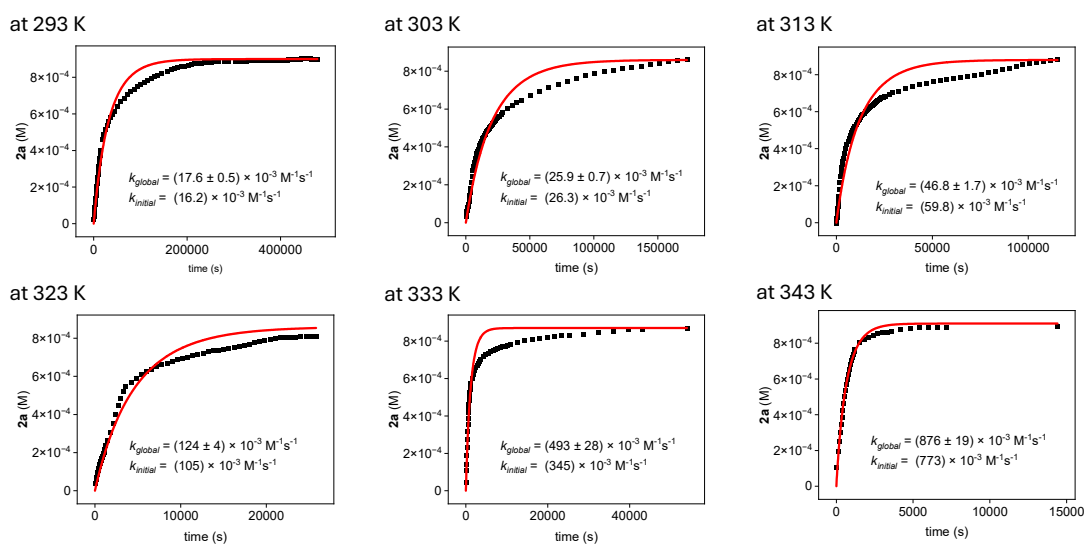


Figure S30. Global nonlinear fits of the full time-course data for formation of **2a** under 9:1 H₂O:MeCN conditions at 293–343 K (six temperatures). Experimental data (black dotted) are fit to eq. 7 (red solid). Insets list the corresponding k_{global} and k_{initial} values at each temperature.

Table S14. Comparison of observed rate constants obtained from initial rate method, second-order global kinetic fitting, and normalized rate constant (k_{H_2O}) at various reaction temperatures in the reaction of **1a** (1.05 mM) + O₂ under 9:1 H₂O:MeCN conditions.

Temp (K)	k_{initial} ($10^{-3} \text{ M}^{-1} \text{ s}^{-1}$)	k_{global} ($10^{-3} \text{ M}^{-1} \text{ s}^{-1}$)	k_{H_2O} ($10^{-3} \text{ M}^{-1} \text{ s}^{-1}$)
293	16.5 ± 1.2	17.6 ± 0.5	8.8 ± 0.3
303	23.8 ± 2.2	25.9 ± 0.7	13.0 ± 0.4
313	57.3 ± 2.5	46.8 ± 1.7	23.4 ± 0.9
323	110 ± 5.6	124 ± 4	62.0 ± 2
333	351 ± 20	493 ± 28	247 ± 14
343	773 ± 58	876 ± 19	438 ± 10

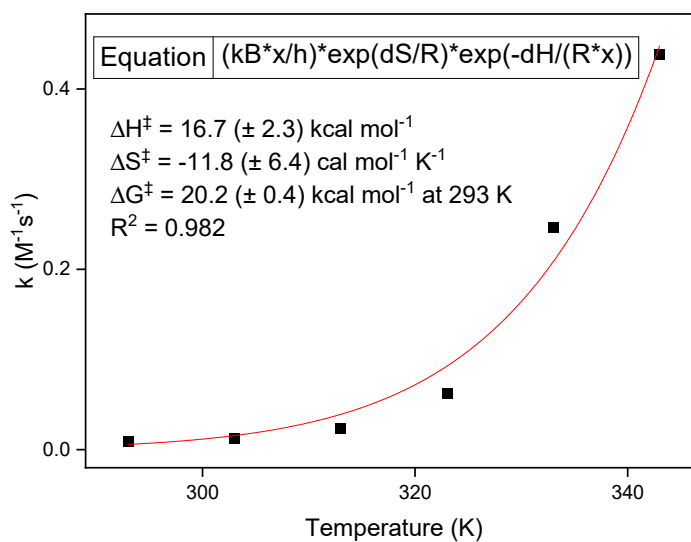


Figure S31. Nonlinear regression Eyring plot of rate constant versus temperature to determine activation parameters for the aerobic oxidation of **1a** in 9:1 H₂O:MeCN.

Stoichiometric O₂ study

Solution of **1a** (7.84 mM) in MeCN was prepared and 265 μL of solution **1a** was placed into a 4 mL quartz cuvette equipped with a septum-sealed cap and a magnetic stir bar. Additionally, 135 μL of acetonitrile and 3.6 mL of degassed deionized water to make 0.52 mM of 4.0 mL solution to reduce the headspace as much as possible. A gas tight syringe was prepared to inject the exact amount of molecular oxygen into the cuvette. The syringe was purged 5 times with molecular oxygen before measuring the amount of oxygen. Oxygen was measured exactly depending on each equivalent (for 1 equiv: 0.0125 mmol, 302 μL ; for 1/2 equiv: 0.00627 mmol, 151 μL ; for 1/4 equiv: 0.00314 mmol, 75 μL). The oxygen containing syringe was inserted into the cuvette and the tip of the needle placed bottom of the cuvette with care. The reaction progress was monitored by UV-Vis spectroscopy while stirring at 170 rpm with a stir bar.

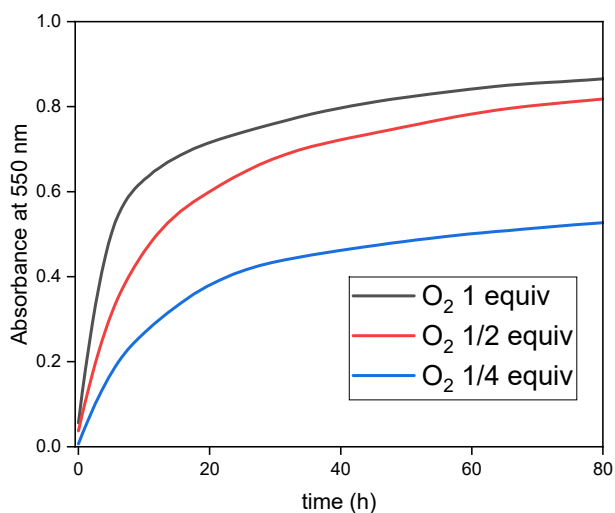


Figure S32. Absorbance (550 nm)-time graph of stoichiometric (1, 1/2, and 1/4 equiv) study of O₂ with **1a** (0.52 mM) in 9:1 H₂O:MeCN solvent mixture at 293 K.

Quantification of Hydrogen Peroxide

a) Calibration curve

The yield of the hydrogen peroxide was spectroscopically determined from the absorption at 407 nm by the Ti^{IV} oxysulfate method.⁹⁻¹¹. The calibration curve for the measurement of H₂O₂ was derived the following procedure. The 10 μ L of 30 wt% H₂O₂ solution was added to a 10 mL volumetric flask followed by H₂O addition to make 10 mL stock solution. The known amount of volume of the stock solution were added to each 10 mL volumetric flask followed by the addition of 0.1 mL of a 15 wt% Ti(O)(SO₄) solution in H₂SO₄ and H₂O to make 10 mL of 0 mM, 0.098 mM, 0.196 mM, 0.489 mM, 0.98 mM, and 1.96 mM solutions. After mixing each solution for 5 minutes, the absorptions at 407 nm of each solution were measured to make the calibration curve (Figure S33).

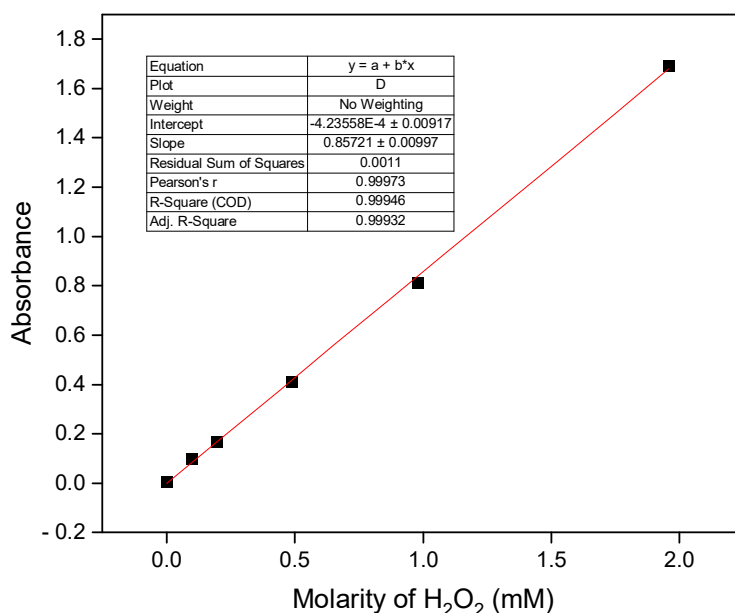


Figure S33. Calibration curve of the concentrations of H₂O₂ in aqueous solution with Ti(O)(SO₄).

b) Sample preparation

A reaction was run in the same way as the section 5 in Supporting Information. When the reaction reaches 90% of the yield of Pd^{III} complex **2a** by UV-Vis spectroscopy, a 1 mL of the solution was taken. Then, to the 1 mL of the solution, 0.1 mL of a 15 wt% Ti(O)(SO₄) solution in H₂SO₄ and H₂O was introduced to make the total volume of the solution to 10 mL. The absorption at 407 nm of the solution was measured and subtracted the absorbance effect from (diluted) **2a** to quantify H₂O₂ precisely, because product **2a** was highly soluble in aqueous phase which makes organic solvent extraction unable to separate H₂O₂ from **2a**. For example,

a 1.05 mM reaction solution **A** that reaches 90% yield of **2a** showed 1.05 absorbance value at 407 nm. The 1 mL of the solution **A** was taken and 0.1 mL of a 15 wt% Ti(O)(SO₄) solution and H₂O was introduced to **A** to make the 10 mL of the solution **B**. The absorption at 407 nm of solution **B** was measured as 0.145. Since the reaction solution **A** has diluted ten times in solution **B**, so the absorption without effect of **2a** will be $(0.145 - 1.05/10 =) 0.04$, which corresponds to 0.0467 mM of TiO₂ in solution **B**. The TiO₂ concentration in **B** can be considered as $(0.0467 \times 10 =) 0.467$ mM of H₂O₂ in solution **A** and the yield of H₂O₂ in solution **A** is $(0.467 \text{ mM} / 1.05 \text{ mM} \times 100 =) 44\%$. We also prepared the mock sample to investigate whether the complex in the reaction solution hinders the titanium oxysulfate method to quantify H₂O₂. The average yield of H₂O₂ in 3 trials was $42 \pm 3\%$.

A solution of independently synthesized **2a** (0.95 mM) in 9:1 H₂O:MeCN solvent mixture and TiO₂ (0.45 mM) solution was also prepared, separately. The 1 mL of each solution and 90 μ L of 15 wt% Ti(O)(SO₄) solution were added to a 10 mL volumetric flask followed by the addition of water to make 10 mL mock solution. The mock solution was measured and compared with the solution **B**. There was little difference in UV-Vis spectrum between solution **B** and the mock solution, so we conducted this method to quantify H₂O₂.

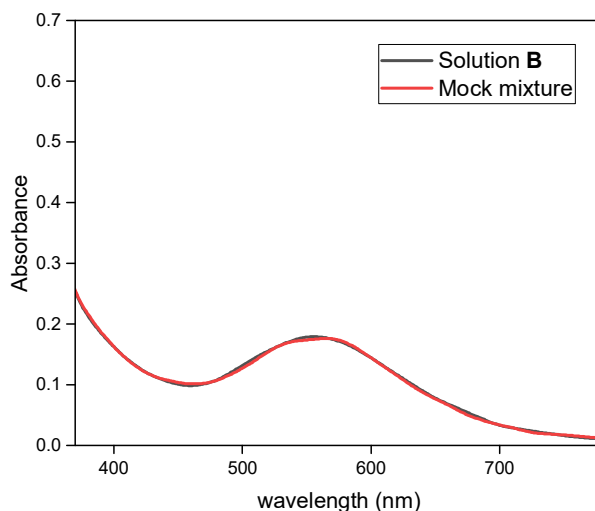


Figure S34. UV-Vis spectrum comparison of the solution **B** and mock solution: a mixture of **2a** (0.095 mM), TiO₂ (0.045 mM), and 90 μ L of 15 wt% Ti(O)(SO₄) solution.

Reaction with *p*-toluenesulfonic acid

The addition of 1 equiv *p*-toluenesulfonic (tosic) acid did not result in the development of the absorbance band at 550 nm, indicating no progress to afford complex **2a** (Figure S35). An attempt to add 0.5 equiv of tosic acid after the complete development of the absorbance band at 550 nm under standard conditions was conducted, but no further growth of the band was observed (Figure S36). Additionally, when tosic acid was introduced to independently synthesized **2a**, it led to a decrease in absorbance, suggesting the decomposition of **2a** (Figure S37). These results indicate that the presence of a strong acid suppresses the reaction and does not facilitate a higher yield of **2a**.

a) Aerobic oxidation of **1a** with O₂ in the presence of *p*-toluenesulfonic acid

4.05 mL solution of **1a** (1.08 mM) in MeCN was placed into a 20 mL vial. *p*-toluenesulfonic acid monohydrate (8.3 mg, 0.0437 mmol) was weighed into a 4 mL vial then 1 mL of MeCN was added to make a solution. 100 μ L of *p*-toluenesulfonic acid solution was poured into solution of **1a** to make 1.05 mM of solution **1a** in the presence of 1 equivalent of the acid. 2 mL of mixed solution was transferred into a quartz cuvette (10 mm path length) equipped with a septum-sealed cap and a magnetic stir bar. An oxygen balloon was prepared with an inlet needle, oxygen was then bubbled through the solution for 45 seconds, and the reaction mixture was stirred under closed gas system. The reaction progress was monitored by UV-Vis spectroscopy while stirring at 170 rpm.

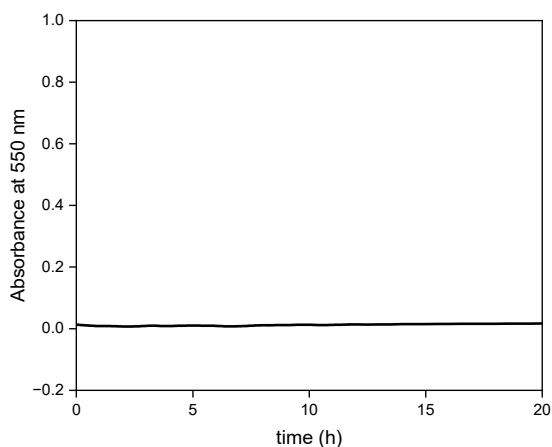


Figure S35. Absorbance (550 nm)-time graph for aerobic oxidation of **1a** with 1 equivalent of tosic acid in the O₂-saturated (8.1 mM) acetonitrile.

b) Aerobic oxidation of **1a** with O₂ then addition of *p*-toluenesulfonic acid after the complete development of the absorbance band at 550 nm under standard reaction conditions

2 mL solution of **1a** (1.05 mM) in MeCN was placed into a quartz cuvette (10 mm path length) equipped with a septum-sealed cap and a magnetic stir bar. Oxygen balloon was prepared and the inlet needle (22 G) connecting to the balloon and the outlet (23 G) needle were inserted orderly while the tip of inlet needle was placed in bottom of the solution and that of outlet needle was placed to the top of the headspace. The oxygen was then bubbled through the solution for 45 seconds then needles were removed in the reverse order to make the cuvette a closed system, and the reaction mixture was stirred under closed gas system. The reaction progress was monitored by UV-Vis spectroscopy while stirring at 170 rpm with a stir bar. The development of 550 nm band was almost completed at 72 hours, 1 mL of 0.105 M tosic acid solution in MeCN was prepared. 10 μ L of the acidic solution (0.5 equiv to **1a**, 0.00105 mmol) was inserted into the cuvette solution through a syringe.

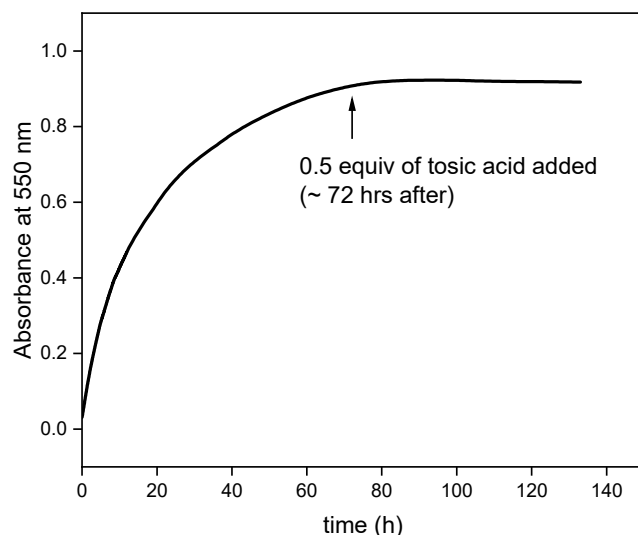


Figure S36. Absorbance (550 nm)-time graph for aerobic oxidation of **1a** in the O₂ saturated (8.1 mM) acetonitrile followed by the addition of 0.5 equiv of tosic acid when the development of absorbance at 550 nm was fully completed.

c) Stability test of **2a** in the presence of *p*-toluenesulfonic acid

The solution of independently synthesized complex **2a** in acetonitrile (2 mL, 0.178 mM) was prepared and was placed into a quartz cuvette (10 mm path length) equipped with a septum-sealed cap and a magnetic stir bar. *p*-toluenesulfonic acid monohydrate (6.7 mg, 0.0355 mmol) was weighed then dissolved by 1 mL of MeCN to obtain a solution. 10 μ L of the acid solution was injected into the cuvette and the reaction progress was monitored by UV-Vis spectroscopy while stirring at 170 rpm with a stir bar.

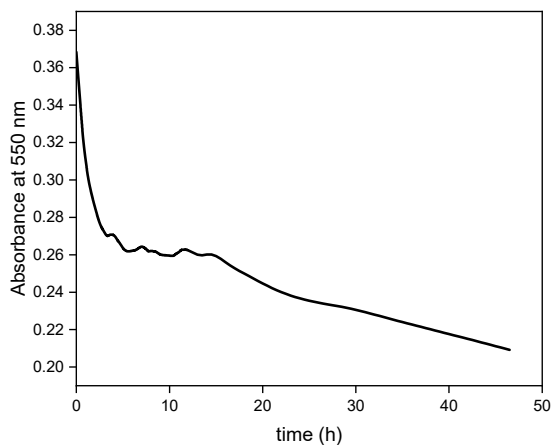


Figure S37. Absorbance (550 nm)-time graph of stability test for **2a** (0.177 mM) in the presence of *p*-toluenesulfonic acid (1 equivalent) in MeCN.

Reaction in phosphate buffered solution

We investigated the influence of a phosphate buffer¹² on reaction kinetics, expecting that the yield would increase. Surprisingly, this addition did not lead to any observable progress in the reaction (Figure S38). This suggests a potential interaction between the phosphate species and the metal complex that may impede the C–H bond activation process.

Solution of **1a** (1.17 mM) in MeCN was prepared and 1.8 mL of the solution was placed into a quartz cuvette equipped with a septum-sealed cap and a magnetic stir bar, followed by the addition of 200 μ L 0.1 M phosphate buffer solution (pH 7.4). Oxygen balloon was prepared and the inlet needle (22 G) connecting to the balloon and the outlet (23 G) needle were inserted orderly while the tip of inlet needle was placed in bottom of the solution and that of outlet needle was placed to the top of the headspace. Oxygen was then bubbled through the solution for 45 seconds then needles were removed in the reverse order to make the cuvette a closed system, and the reaction mixture was stirred under closed gas system. The reaction progress was monitored by UV-Vis spectroscopy while stirring at 170 rpm with a stir bar.

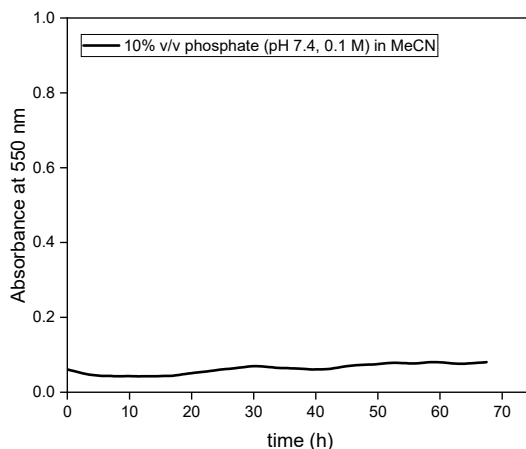


Figure S38. Absorbance (550 nm)-time graph of **1a** (1.05 mM) + O₂ in 10% v/v phosphate (pH 7.4, 0.1 M) in MeCN.

7. ESI-MS Analysis of Aerobic Oxidation of **1**

Electrospray ionization mass spectrometry (ESI-MS) was employed as a qualitative probe to compare the solution-state speciation of Pd complexes generated during aerobic oxidation of **1** under MeCN or H₂O:MeCN solvent conditions. In MeCN, a signal assigned to a Pd–OH containing species was detected (Figures S40, S43, S45). In contrast, no corresponding Pd–OH signal was observed when the reaction was conducted in a 9:1 H₂O:MeCN mixture (Figure S42). The appearance of a Pd–OH species under neat MeCN conditions may arise from decomposition of an oxygenated Pd intermediate generated following the initial C–H bond activation step. One possible interpretation is that, in the absence of water, this oxygenated intermediate does not undergo subsequent C–H bond activation and instead undergoes decomposition prior to detection by ESI-MS. Notably, the absence of Pd–OH signals under aqueous conditions is consistent with the enhanced formation of the C–H activated Pd^{III} product and hydrogen peroxide, as observed experimentally in H₂O-containing solvent mixtures.

a) Aerobic oxidation of **1a** in MeCN

A solution of **1a** (1.05 mM) in MeCN (2.0 mL) was placed in a 4 mL quartz cuvette equipped with a septum-sealed cap and a magnetic stir bar. An oxygen balloon was prepared and the inlet needle (22 G) connecting to the balloon and the outlet (23 G) needle were inserted orderly while the tip of inlet needle was placed in bottom of the solution and that of outlet needle was placed to the top of the headspace. Oxygen was then bubbled through the solution for 45 seconds then needles were removed in the reverse order to make the cuvette a closed system, and the reaction mixture was stirred under closed gas system. The reaction progress was monitored by UV-Vis spectroscopy while stirring at 170 rpm with a stir bar. Several aliquots were taken during the reaction proceeds to analyze ESI-MS. At early reaction times (0–2 h), no signal corresponding to a Pd–OH containing species ($m/z = 570.2419$) was detected. At longer reaction times (≥ 26 h), a peak at $m/z 570.2419$ (calcd for $[(^{\text{pMe}}\text{N}_3\text{CH})\text{Pd}(\text{OH})(\text{MeCN})_2]^+$, 570.2424) gradually emerged and increased in intensity, becoming clearly observable after 50 h. This signal disappeared following the addition of degassed DI water (2.0 mL) to the reaction mixture, without a concomitant increase in the yield of **2a**. The appearance of the Pd–OH assigned signal under neat MeCN conditions is consistent with formation and subsequent decomposition of an oxygenated Pd species generated during the reaction. In addition, ESI-MS spectra exhibited peaks at $m/z 366.2898$, 310.2274, and 254.1637, corresponding to ligand-derived fragment ions, as well as peaks at 470.1788, 414.1151, and 358.0498 assignable to

$[(^{pMe}N_3C)Pd]^+$ and its related fragment ions. A peak at m/z 235.0900 was assigned to the double charged charged species $[(^{pMe}N_3C)Pd]^{2+}$.

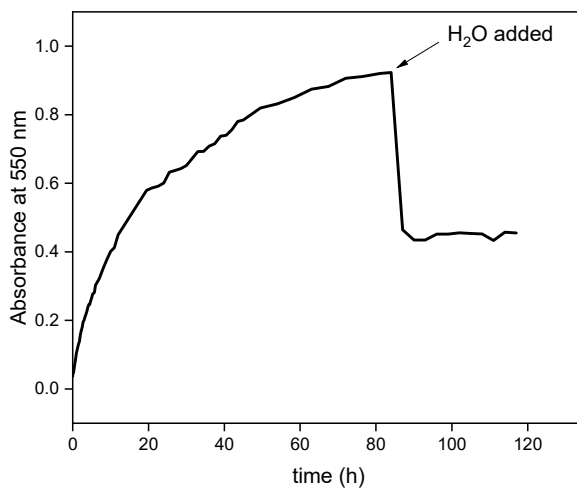


Figure S39. Time course for the 550 nm absorbance during aerobic oxidation of **1a** in O_2 -saturated MeCN (8.1 mM), followed by the addition of 2 mL of water when the development of absorbance at 550 nm was fully completed. The total concentration of palladium complexes was halved (1.05 mM \rightarrow 0.52 mM) upon addition of water to 2 mL MeCN.

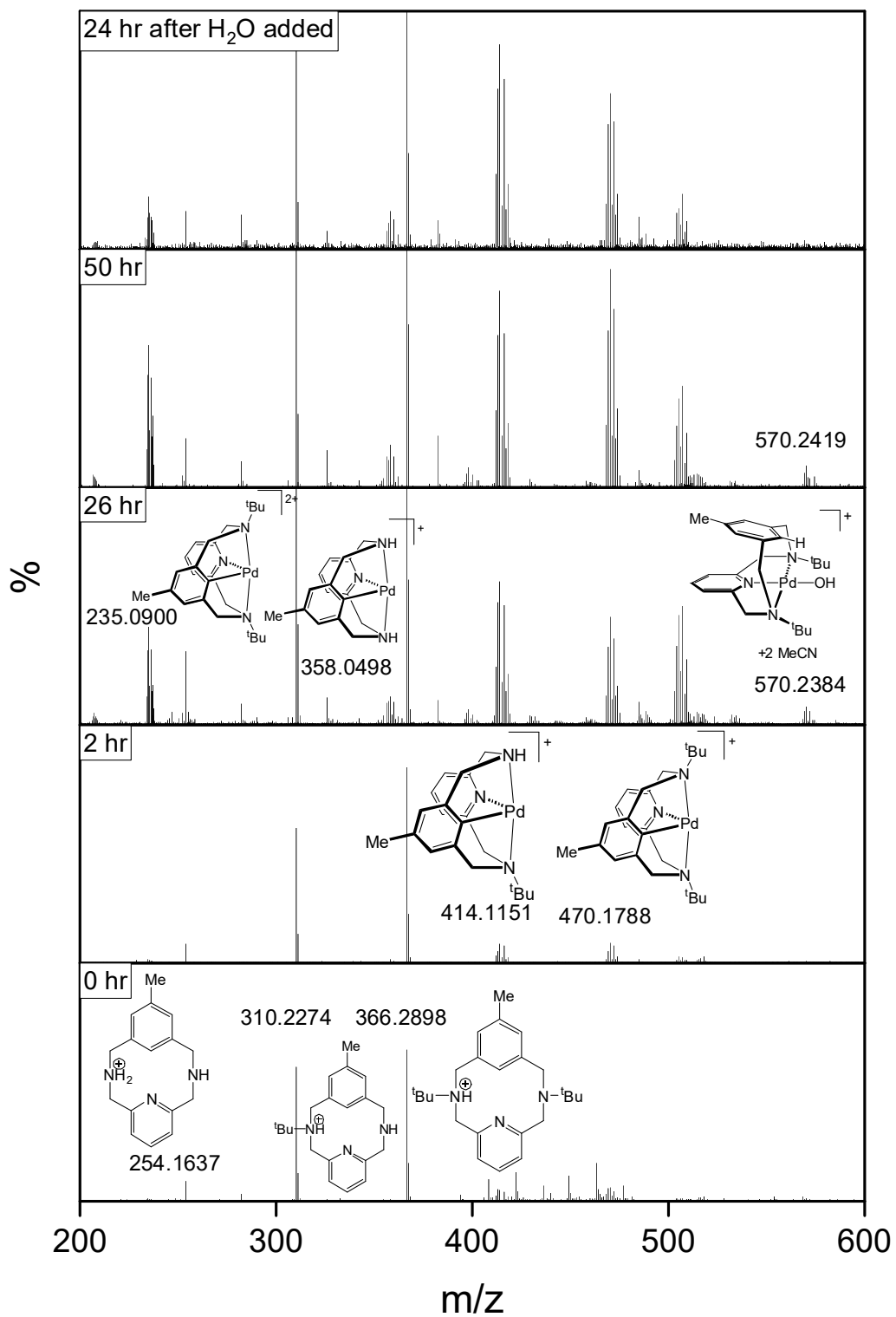


Figure S40. ESI-MS spectra for the aerobic oxidation of **1a** in MeCN according to time.

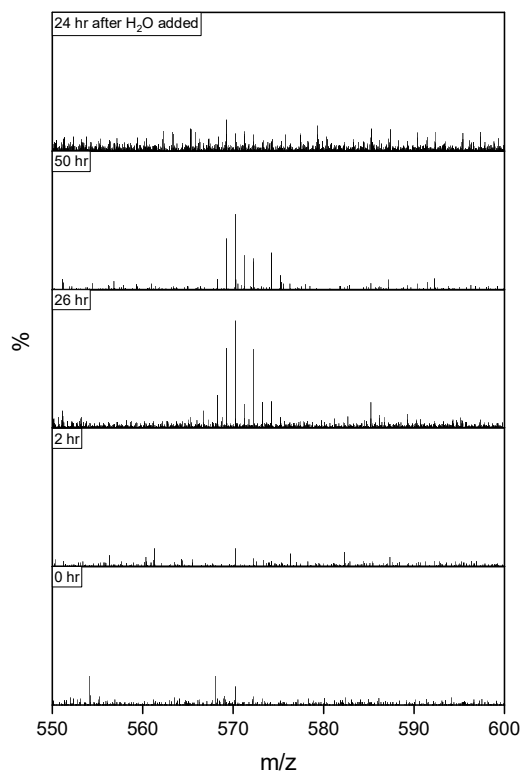


Figure S41. ESI-MS spectra for the aerobic oxidation of **1a** in MeCN in the magnified region (m/z 550–600).

b) Aerobic oxidation of **1a** in 2:8 H₂O:MeCN solvent mixture

Solution of **1a** (1.318 mM) in MeCN was prepared and 1.6 mL of solution **1a** was placed into a 4 mL quartz cuvette equipped with a septum-sealed cap and a magnetic stir bar followed by the addition of 0.4 mL of degassed DI water to make the solution 1.05 mM. Oxygen balloon was prepared and the inlet needle (22 G) connecting to the balloon and the outlet (23 G) needle were inserted orderly while the tip of inlet needle was placed in bottom of the solution and that of outlet needle was placed to the top of the headspace. Oxygen was then bubbled through the solution for 45 seconds then needles were removed in the reverse order to make the cuvette a closed system, and the reaction mixture was stirred under closed gas system. Reaction progress was monitored by UV-Vis spectroscopy, and aliquots were withdrawn periodically for ESI-MS analysis. Throughout the reaction period, no signal corresponding to a Pd–OH containing species ($m/z = 570.2419$) was observed by ESI-MS. Additional experiments conducted using other H₂O:MeCN ratios (1:9 and 5:5) similarly showed no detectable Pd–OH assigned signal over the course of the reaction.

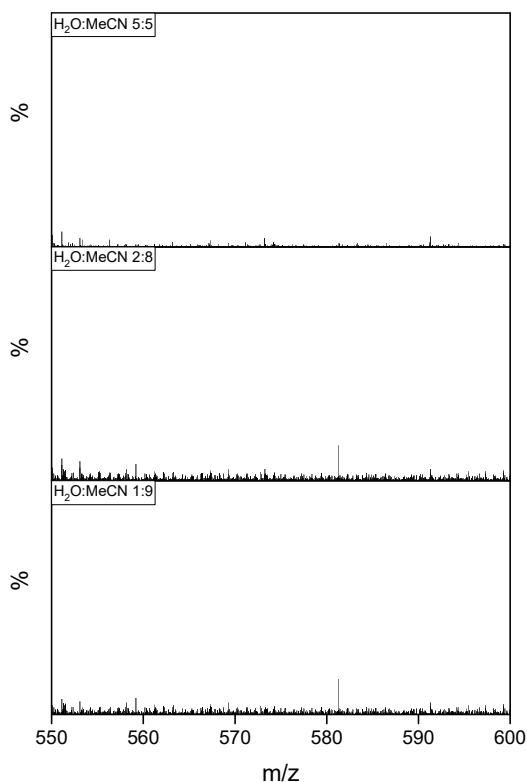


Figure S42. ESI-MS spectra for the aerobic oxidation of **1a** in H₂O:MeCN solvent mixture in the magnified region (m/z 550~600).

c) Aerobic oxidation of **1b** in MeCN

Aerobic oxidation of **1b** in MeCN was performed following the same general procedure described for **1a**. Aliquots were collected at various reaction times for ESI-MS analysis. At early reaction times (0–2h), no Pd–OH assigned signal was detected. At extended reaction times (≥ 26 h), a peak at m/z 556.2263 (calcd for $[(^{\text{P}}\text{H}^{\text{N}}\text{3CH})\text{Pd}(\text{OH})(\text{MeCN})_2]^+$, 556.2268) gradually emerged and increased in intensity, becoming clearly observable after 50 h. This signal disappeared following the addition of degassed DI water (200 μL) to the reaction mixture. Additional peaks corresponding to ligand-derived fragment ions were observed at m/z 352.2749, 296.2124, and 240.1493. Signals at m/z 456.1639, 400.0995, and 344.0332 were assigned to $[(^{\text{P}}\text{H}^{\text{N}}\text{3C})\text{Pd}]^+$ and related fragment ions, while a peak at m/z 228.0819 was assigned to the doubly charged species $[(^{\text{P}}\text{H}^{\text{N}}\text{3C})\text{Pd}]^{2+}$.

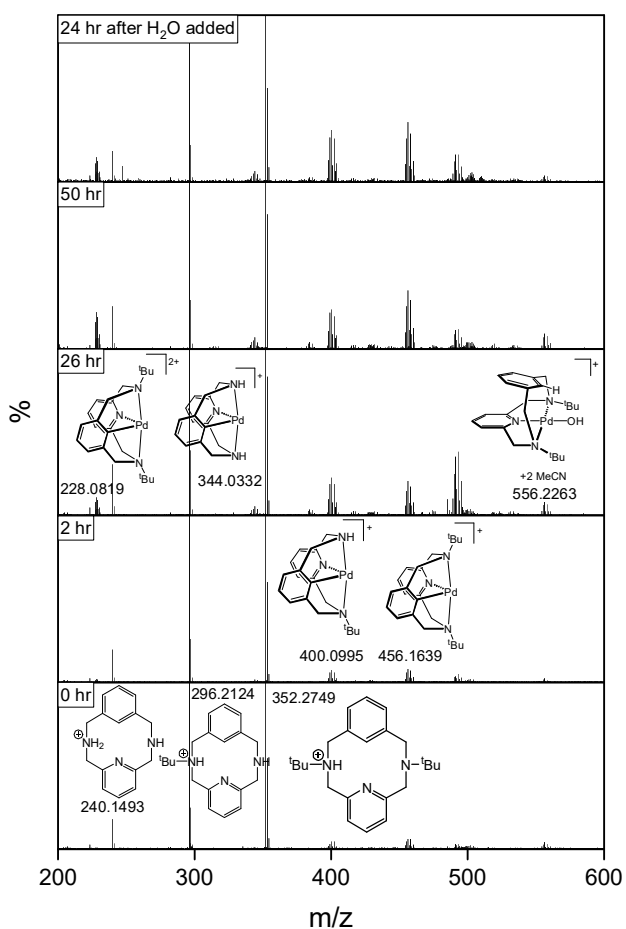


Figure S43. ESI-MS spectra for the aerobic oxidation of **1b** in MeCN according to time.

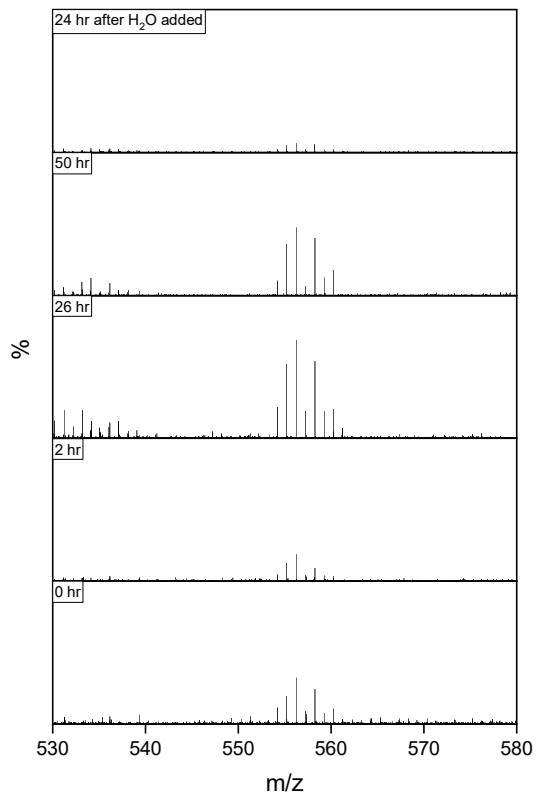


Figure S44. ESI-MS spectra for the aerobic oxidation of **1b** in MeCN in the magnified region (m/z 530~580).

d) Aerobic oxidation of **1b-D** in MeCN

Aerobic oxidation of the deuterated analogue **1b-D** in MeCN was conducted under the same conditions used for **1b**. Reaction aliquots were analyzed by ESI-MS at various time points. No Pd-OH assigned signal was observed at early reaction times. At longer reaction times (≥ 26 h), a peak at m/z 557.2325 (calcd for $[(^p\text{H}^3\text{N}^3\text{C}^3\text{D})\text{Pd}(\text{OH})(\text{MeCN})_2]^+$, 557.2325) was detected and increased in intensity, becoming prominent after 50 h. As observed for **1b**, this signal disappeared upon addition of degassed water (200 μL). Fragment ions corresponding to ligand-derived species were observed at m/z 353.2809, 297.2182, and 241.1552. Peaks at m/z 456.1673, 400.1043, 344.0343 were assigned to $[(^p\text{Me}^3\text{N}^3\text{C})\text{Pd}]^+$ and related fragment ions, while a peak at m/z 228.0823 corresponded to $[(^p\text{Me}^3\text{N}^3\text{C})\text{Pd}]^{2+}$.

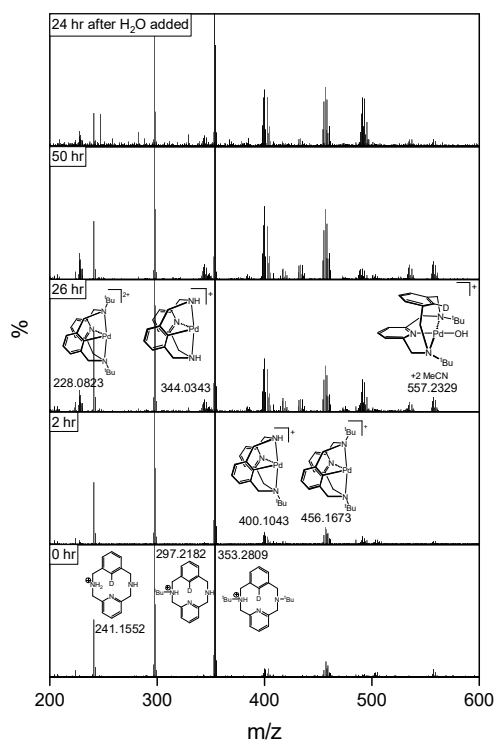


Figure S45. ESI-MS spectra for the aerobic oxidation of **1b-D** in MeCN according to time.

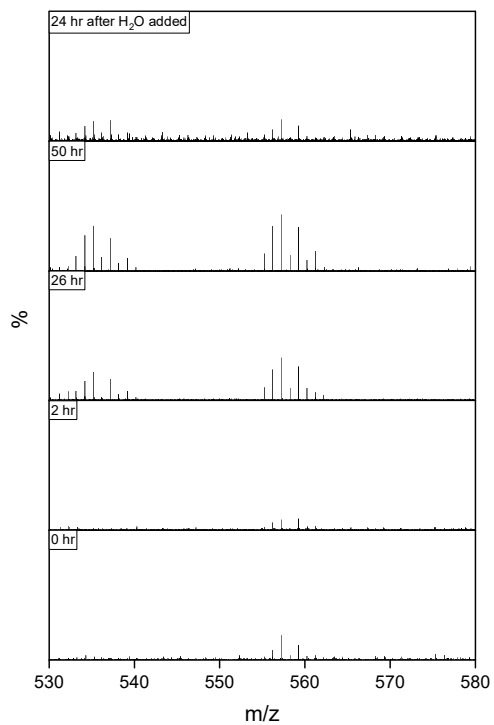


Figure S46. ESI-MS spectra for the aerobic oxidation of **1b-D** in MeCN in the magnified region (m/z 530~580).

8. X-ray Structure Characterization

General information.

Single crystals of **1a**, **1ab**, and **2a** were grown by vapor diffusion of diethyl ether into concentrated solution of acetonitrile at -35 °C. Crystals were mounted on a Bruker D8 Venture kappa diffractometer equipped with a Photon II CPAD detector. An I μ s microfocus Mo source ($\lambda = 0.71073$ Å) coupled with a multi-layer mirror monochromator provided the incident beam. The sample was mounted on a nylon loop with the minimal amount of Paratone-N oil. Data was collected as a series of φ and/or ω scans. Data were collected at 100 K using a cold stream of N₂(g). The collection, cell refinement, and integration of intensity data was carried out with the APEXIII software.¹³ A multiscan absorption correction was performed with SADABS or TWINABS-2012/1.¹⁴ The structure was phased with intrinsic methods using SHELXT and refined with the full-matrix least-squares program SHELXL.¹⁵ Hydrogen atoms were placed in calculated positions using the standard riding model and refined isotropically; all non-hydrogen atoms were refined anisotropically. Software and solutions: SAINT V8.38A (2016), SHELXT (Sheldrick, 2015), XL (Sheldrick, 2008), Olex2 (Dolomanov et al., 2009).¹⁵⁻¹⁷

The deposition numbers CCDC 2331324, 2331325, and 2331326 at the Cambridge Crystallographic Data Centre CCDC contain the supplementary crystallographic data. These data are provided free of charge by the Cambridge Crystallographic Data Centre. Crystallographic details are summarized in Tables S15-23.

X-ray structure determination of **1a**

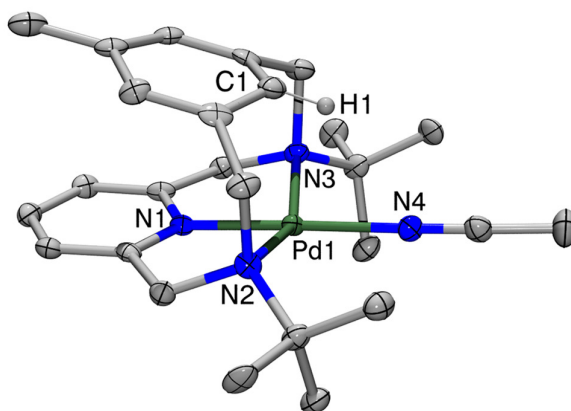


Figure S47. Solid-state structure of dication of **1a** with 50% probability ellipsoids.

Table S15. Crystal data and structure refinement for **1a**.

Empirical formula	C ₂₈ H ₄₁ B ₂ F ₈ N ₅ Pd
Formula weight	727.68
Temperature/K	100.05
Crystal system	monoclinic
Space group	P2 ₁ /c
a/Å	12.1854(10)
b/Å	13.2252(11)
c/Å	19.5898(17)
α /°	90
β /°	105.091(5)
γ /°	90
Volume/Å ³	3048.1(4)
Z	4
$\rho_{\text{calc}}/\text{cm}^3$	1.586
μ/mm^{-1}	0.686
F(000)	1488.0
Crystal size/mm ³	0.535 × 0.109 × 0.091
Radiation	MoK α (λ = 0.71073)
2 θ range for data collection/°	2.154 to 55.296

Index ranges	-15 ≤ h ≤ 15, -17 ≤ k ≤ 15, -25 ≤ l ≤ 25
Reflections collected	48647
Independent reflections	6936 [R _{int} = 0.0872, R _{sigma} = 0.0696]
Data/restraints/parameters	6936/1/411
Goodness-of-fit on F ²	1.063
Final R indexes [I ≥ 2σ (I)]	R ₁ = 0.0768, wR ₂ = 0.1967
Final R indexes [all data]	R ₁ = 0.1062, wR ₂ = 0.2208
Largest diff. peak/hole / e Å ⁻³	7.55/-1.22

Table S16. Fractional Atomic Coordinates ($\times 10^4$) and Equivalent Isotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for **1a**. U_{eq} is defined as 1/3 of the trace of the orthogonalised U_{ij} tensor.

Atom	x	y	z	U(eq)
Pd1	3143.1(4)	4199.2(3)	7233.9(2)	14.94(16)
N1	4745(4)	4569(4)	7479(3)	15.9(10)
N2	3501(4)	4038(4)	8394(3)	17.4(10)
N3	3442(4)	3952(4)	6186(3)	17.9(10)
N4	1474(4)	3871(4)	6963(3)	20.2(11)
C1	3791(5)	2350(5)	7378(3)	21.9(13)
C2	4344(5)	2426(5)	6843(3)	20.8(12)
C3	5528(5)	2365(4)	7018(3)	19.3(12)
C4	6158(5)	2311(5)	7726(3)	20.9(12)
C5	5556(5)	2420(5)	8244(3)	20.0(12)
C6	4389(6)	2477(5)	8074(3)	21.3(13)
C7	3732(6)	2905(5)	8557(3)	21.9(13)
C8	4585(5)	4610(5)	8671(3)	18.8(12)
C9	5307(5)	4661(4)	8164(3)	16.1(11)
C10	6462(5)	4861(5)	8353(3)	19.1(12)
C11	7029(5)	4945(5)	7826(3)	20.6(12)
C12	6441(5)	4811(5)	7124(3)	19.6(12)
C13	5282(5)	4611(5)	6963(3)	17.0(12)
C14	4548(5)	4507(5)	6230(3)	18.9(12)
C15	3639(6)	2799(5)	6137(3)	23.5(13)

C16	7404(6)	2262(5)	7907(4)	28.3(15)
C17	2603(5)	4442(5)	8761(3)	20.5(13)
C18	2202(5)	5467(5)	8441(3)	23.9(13)
C19	1597(5)	3729(5)	8651(3)	23.1(13)
C21	2561(5)	4328(5)	5508(3)	20.7(13)
C23	1547(5)	3599(5)	5293(3)	24.7(14)
C24	2135(5)	5357(6)	5668(3)	25.3(14)
C25	527(5)	3686(5)	6781(3)	23.5(13)
C26	-667(6)	3445(7)	6532(4)	36.6(18)
C20	3133(6)	4551(6)	9561(3)	25.0(14)
C22	3090(6)	4390(6)	4878(3)	27.8(15)
N1S	1009(8)	955(7)	5645(5)	60(2)
C1S	295(7)	1253(7)	5170(5)	48(2)
C2S	-607(7)	1639(9)	4613(5)	54(2)
B1	6287(6)	2612(7)	5248(4)	27.6(17)
F1	5610(3)	3337(3)	4814(2)	27.8(9)
F2	5574(3)	1843(3)	5379(2)	29.0(9)
F3	6867(4)	3067(4)	5873(2)	42.0(12)
F4	7039(4)	2185(4)	4905(3)	50.8(14)
B2	601(7)	3932(7)	2968(5)	33.2(18)
F5	182(5)	4148(4)	3533(3)	67.1(18)
F6	1750(4)	3714(4)	3205(3)	47.6(12)
F7	444(4)	4733(4)	2502(2)	41.7(11)
F8	35(5)	3079(5)	2639(4)	75(2)

Table S17. Bond lengths for **1a**.

Atom	Atom	Length/Å	Atom	Atom	Length/Å
Pd1	N1	1.947(5)	C8	C9	1.490(8)
Pd1	N2	2.210(5)	C9	C10	1.384(8)
Pd1	N3	2.201(5)	C10	C11	1.388(8)
Pd1	N4	2.011(5)	C11	C12	1.386(8)
Pd1	C1	2.563(6)	C12	C13	1.390(8)
N1	C9	1.343(7)	C13	C14	1.488(8)
N1	C13	1.340(7)	C17	C18	1.520(9)
N2	C7	1.542(8)	C17	C19	1.517(9)
N2	C8	1.496(8)	C17	C20	1.538(8)
N2	C17	1.553(8)	C21	C23	1.538(9)
N3	C14	1.518(8)	C21	C24	1.518(9)
N3	C15	1.551(8)	C21	C22	1.535(9)
N3	C21	1.555(8)	C25	C26	1.445(9)
N4	C25	1.142(8)	N1S	C1S	1.165(12)
C1	C2	1.390(9)	C1S	C2S	1.427(12)
C1	C6	1.378(9)	B1	F1	1.400(9)
C2	C3	1.395(9)	B1	F2	1.403(9)
C2	C15	1.509(9)	B1	F3	1.382(9)
C3	C4	1.402(9)	B1	F4	1.390(9)
C4	C5	1.406(8)	B2	F5	1.364(11)
C4	C16	1.468(10)	B2	F6	1.386(10)
C5	C6	1.376(9)	B2	F7	1.380(10)
C6	C7	1.500(9)	B2	F8	1.390(10)

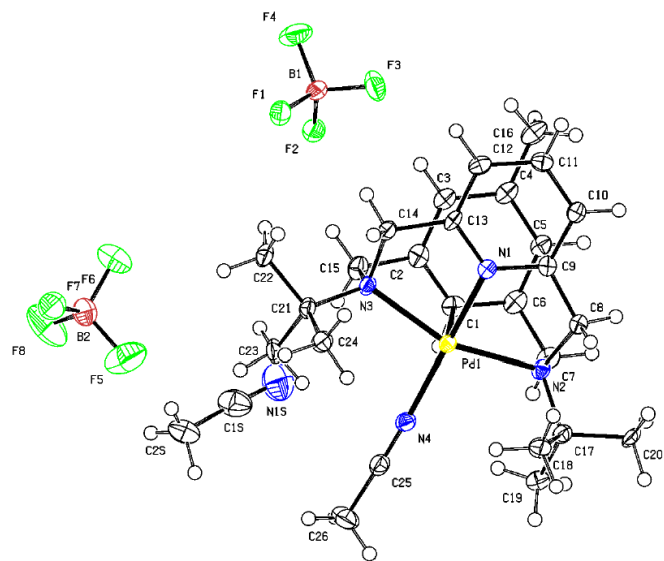


Figure S48. Projection view of **1a** with 50% probability ellipsoids.

X-ray structure determination of **1ab**.

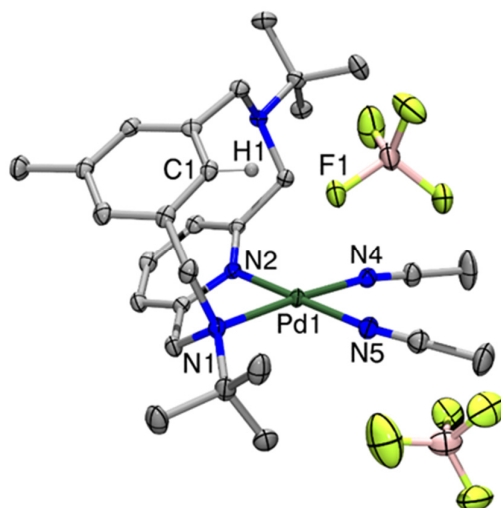


Figure S49. Solid-state structure of **1ab** with 50% probability ellipsoids. Selected distances (Å) and angles (°): H1⋯F1 2.473, C1–H1⋯F1 151.9.

Table S18. Crystal data and structure refinement for **1ab**.

Identification code	P1
Empirical formula	C ₃₀ H ₄₄ B ₂ F ₈ N ₆ Pd
Formula weight	768.73
Temperature/K	100.00
Crystal system	triclinic
Space group	P-1
a/Å	7.7290(3)
b/Å	9.6870(4)
c/Å	22.8572(9)
α/°	85.0080(10)
β/°	84.3020(10)
γ/°	86.3840(10)
Volume/Å ³	1693.86(12)
Z	2
ρ _{calc} /cm ³	1.507
μ/mm ⁻¹	0.623
F(000)	788.0

Crystal size/mm ³	0.115 × 0.108 × 0.023
Radiation	MoK α (λ = 0.71073)
2 θ range for data collection/ $^{\circ}$	4.228 to 52.88
Index ranges	-9 \leq h \leq 9, -12 \leq k \leq 12, -28 \leq l \leq 28
Reflections collected	72887
Independent reflections	6961 [R _{int} = 0.0491, R _{sigma} = 0.0222]
Data/restraints/parameters	6961/3/464
Goodness-of-fit on F ²	1.094
Final R indexes [I \geq 2 σ (I)]	R ₁ = 0.0273, wR ₂ = 0.0623
Final R indexes [all data]	R ₁ = 0.0309, wR ₂ = 0.0641
Largest diff. peak/hole / e \AA^{-3}	0.66/-0.55

Table S19. Fractional Atomic Coordinates ($\times 10^4$) and Equivalent Isotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for **1ab**. U_{eq} is defined as 1/3 of the trace of the orthogonalised U_{ij} tensor.

Atom	x	y	z	U(eq)
Pd1	4574.3(2)	6070.3(2)	7181.3(2)	15.67(5)
F1	7607.6(19)	8020.2(13)	7700.0(6)	31.9(3)
F4	6203(2)	9959.8(15)	7302.7(7)	44.9(4)
F3	7876(2)	10116.2(16)	8042.6(8)	48.3(4)
N2	3213(2)	4770.5(17)	7762.8(7)	15.9(3)
N1	6414(2)	4413.7(17)	7187.1(7)	19.1(4)
N3	1661(2)	6475.7(17)	9061.7(7)	16.4(3)
F2	5307(2)	9071.5(17)	8224.6(8)	56.2(5)
N4	2601(2)	7520.5(18)	7136.1(8)	21.1(4)
N5	5799(2)	7416.9(19)	6583.6(8)	22.7(4)
C2	6558(3)	4303(2)	8304.8(9)	18.3(4)
C15	1974(2)	5087(2)	8200.0(8)	15.5(4)
C14	972(3)	4045(2)	8480.5(9)	18.4(4)
C1	5697(3)	5454(2)	8546.7(9)	17.6(4)
C3	7533(3)	4461(2)	7700.3(9)	21.0(4)
C7	4584(3)	5302(2)	9060.6(9)	18.3(4)

C13	1285(3)	2687(2)	8336.3(9)	21.2(4)
C5	5372(3)	2827(2)	9141.5(9)	19.5(4)
C4	6417(3)	2999(2)	8611.3(9)	20.3(4)
C10	5290(3)	3179(2)	7274.5(9)	20.6(4)
C12	2698(3)	2359(2)	7942.9(9)	20.3(4)
C16	1779(3)	6545(2)	8410.0(8)	16.9(4)
C11	3661(3)	3431(2)	7669.6(8)	17.7(4)
C8	3423(3)	6514(2)	9265.2(9)	19.1(4)
C21	367(3)	7524(2)	9323.6(9)	19.2(4)
C25	1671(3)	8415(2)	7006.9(10)	24.8(5)
C6	4446(3)	3985(2)	9354.6(9)	19.6(4)
C27	6153(3)	8334(2)	6259.6(10)	23.9(5)
C9	5186(3)	1399(2)	9456.4(10)	26.2(5)
F5	2846(3)	6509(2)	5668.3(14)	96.4(8)
C17	7665(3)	4281(2)	6608.3(10)	25.8(5)
F6A	355(7)	7868(8)	5667(6)	49(2)
C22	-1444(3)	7214(2)	9172.3(9)	23.2(4)
C28	6628(4)	9502(3)	5843.0(12)	39.5(6)
C18	9092(3)	5314(2)	6571.1(11)	30.7(5)
C24	340(3)	7338(2)	9999.2(9)	26.5(5)
C26	487(4)	9579(3)	6846.0(14)	47.4(8)
C20	6599(3)	4516(3)	6073.9(10)	31.5(5)
C19	8540(3)	2810(2)	6607.3(11)	34.7(6)
C23	772(3)	9027(2)	9103.8(11)	27.4(5)
N6	3116(3)	11972(3)	6261.9(12)	54.9(7)
F8A	2924(5)	8908(4)	5504.4(16)	62.7(15)
C29	2974(4)	12525(3)	5811.3(13)	44.3(7)
F7A	1771(6)	7832(3)	4808.2(12)	75.2(14)
B1	6733(4)	9294(3)	7821.7(12)	30.1(6)
C30	2810(5)	13271(4)	5242.1(14)	61.8(9)
B2	2035(4)	7699(4)	5408.1(14)	42.4(7)

F8B	3364(7)	8122(8)	5095(4)	93(4)
F7B	993(7)	6880(8)	5111(3)	72(3)
F6B	900(30)	8209(18)	5812(4)	95(5)

Table S20. Bond lengths for **1ab**.

Atom	Atom	Length/Å	Atom	Atom	Length/Å
Pd1	N2	2.0143(16)	C15	C16	1.525(3)
Pd1	N1	2.0759(17)	C14	C13	1.385(3)
Pd1	N4	2.0115(18)	C1	C7	1.388(3)
Pd1	N5	2.0129(18)	C7	C8	1.510(3)
F7Aa	B2	1.400(4)	C7	C6	1.394(3)
F6Aa	B2	1.378(7)	C13	C12	1.383(3)
F8Aa	B2	1.438(4)	C5	C4	1.391(3)
F1	B1	1.405(3)	C5	C6	1.391(3)
F4	B1	1.386(3)	C5	C9	1.511(3)
F3	B1	1.385(3)	C10	C11	1.495(3)
N2	C15	1.355(3)	C12	C11	1.383(3)
N2	C11	1.352(3)	C21	C22	1.528(3)
N1	C3	1.531(3)	C21	C24	1.537(3)
N1	C10	1.510(3)	C21	C23	1.539(3)
N1	C17	1.569(3)	C25	C26	1.452(3)
N3	C16	1.479(2)	C27	C28	1.455(3)
N3	C8	1.485(3)	F5	B2	1.393(4)
N3	C21	1.500(3)	C17	C18	1.527(3)
F2	B1	1.379(3)	C17	C20	1.533(3)
N4	C25	1.128(3)	C17	C19	1.538(3)
N5	C27	1.135(3)	N6	C29	1.131(4)
C2	C1	1.390(3)	C29	C30	1.446(4)
C2	C3	1.507(3)	B2	F8B	1.262(6)
C2	C4	1.395(3)	B2	F7B	1.422(6)
C15	C14	1.384(3)	B2	F6B	1.315(7)

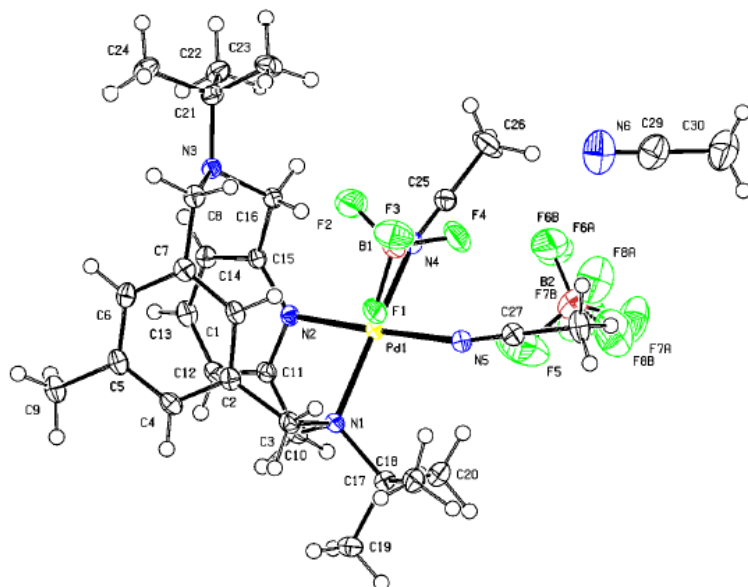


Figure S50. Projection view of **1ab** with 50% probability ellipsoids.

X-ray structure determination of 2a

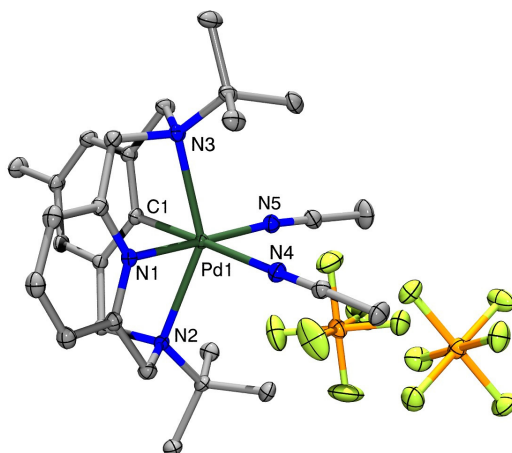


Figure S51. Solid-state structure of **2a** with 50% probability ellipsoids. Selected bond distances (Å) and angles (°): Pd1–C1 1.959(1), Pd1–N1 2.020(1), Pd1–N2 2.396(1), Pd1–N3 2.382(1), Pd1–N4 2.163(1), Pd1–N5 2.094(1), N2–Pd1–N3 146.97(4).

Table S21. Crystal data and structure refinement for **2a**.

Identification code	P21n_a
Empirical formula	C ₃₂ H ₄₆ F ₁₂ N ₇ P ₂ Pd
Formula weight	925.10
Temperature/K	100.00
Crystal system	monoclinic
Space group	P2 ₁ /n
a/Å	18.4894(4)
b/Å	10.4350(2)
c/Å	20.2727(4)
α /°	90
β /°	96.9980(10)
γ /°	90
Volume/Å ³	3882.21(14)
Z	4
$\rho_{\text{calc}}/\text{cm}^3$	1.583
μ/mm^{-1}	0.652

F(000)	1884.0
Crystal size/mm ³	0.28 × 0.151 × 0.081
Radiation	MoK α (λ = 0.71073)
2 θ range for data collection/ $^{\circ}$	4.398 to 61.132
Index ranges	-26 \leq h \leq 26, -14 \leq k \leq 14, -28 \leq l \leq 28
Reflections collected	153495
Independent reflections	11891 [R _{int} = 0.0442, R _{sigma} = 0.0185]
Data/restraints/parameters	11891/0/498
Goodness-of-fit on F ²	1.054
Final R indexes [$I \geq 2\sigma(I)$]	R ₁ = 0.0250, wR ₂ = 0.0595
Final R indexes [all data]	R ₁ = 0.0291, wR ₂ = 0.0617
Largest diff. peak/hole / e \AA^{-3}	0.80/-0.59

Table S22. Fractional Atomic Coordinates ($\times 10^4$) and Equivalent Isotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for **2a**. U_{eq} is defined as 1/3 of the trace of the orthogonalised U_{ij} tensor.

Atom	x	y	z	U(eq)
Pd1	7523.1(2)	2909.7(2)	5076.1(2)	11.39(3)
P2	4503.8(2)	1932.3(4)	3884.1(2)	18.91(7)
P1	6144.5(2)	3605.2(4)	1537.1(2)	23.50(8)
F7	5295.5(5)	1482.0(10)	4213.4(5)	26.42(19)
F9	4440.6(6)	686.7(9)	3419.8(5)	30.5(2)
F12	4561.5(5)	3187.1(10)	4348.0(5)	29.1(2)
F11	4144.1(5)	1160.6(10)	4444.3(5)	30.7(2)
F8	4862.4(6)	2699.8(10)	3324.2(5)	31.1(2)
F10	3707.6(5)	2386.3(11)	3558.0(5)	32.3(2)
F4	5491.3(6)	2646.6(11)	1619.6(6)	36.5(2)
F3	6093.4(8)	3244.1(11)	762.4(5)	43.9(3)
F2	6704.4(6)	2427.9(12)	1688.7(6)	42.9(3)
N1	7916.8(6)	3792.2(10)	5932.0(5)	13.19(19)
F1	6809.2(8)	4548.3(13)	1440.5(7)	53.6(3)
N5	7133.3(6)	1986.9(11)	4186.3(6)	16.6(2)
N4	6507.9(6)	2446.3(12)	5449.7(6)	17.2(2)
F5	5596.1(8)	4754.2(12)	1355.1(8)	60.4(4)

F6	6221.1(10)	3933.3(15)	2298.8(6)	64.7(4)
N2	7301.8(6)	5138.4(11)	4851.0(5)	13.23(19)
N3	8350.7(6)	1326.1(11)	5561.4(6)	14.4(2)
C1	8440.2(7)	3373.2(12)	4745.1(6)	13.2(2)
C5	9681.6(7)	3945.9(13)	4184.5(7)	16.8(2)
C2	8551.5(7)	4636.8(12)	4571.1(6)	13.5(2)
C11	8600.4(8)	3701.0(14)	6995.0(7)	18.8(3)
C3	8040.8(7)	5662.7(12)	4755.8(6)	14.0(2)
C10	8407.1(7)	3185.6(12)	6366.7(6)	14.6(2)
C7	8947.6(7)	2410.5(13)	4708.0(7)	15.0(2)
C4	9179.3(7)	4909.7(13)	4277.6(7)	16.1(2)
C15	7109.8(7)	5526.9(13)	5515.2(6)	15.8(2)
C8	8805.5(7)	1125.0(13)	5004.7(7)	16.6(2)
C23	6602.2(8)	6980.7(13)	4270.2(7)	19.2(3)
C14	7626.8(7)	4933.2(12)	6065.4(6)	14.5(2)
C6	9573.2(7)	2708.9(13)	4418.0(7)	17.1(2)
C25	5956.0(7)	2184.4(13)	5607.3(6)	15.6(2)
N7	3288.7(10)	7179.9(15)	2762.3(8)	36.5(3)
C13	7816.0(7)	5501.4(13)	6681.5(7)	17.8(2)
C19	7475.8(8)	-447.8(14)	5253.8(7)	20.9(3)
C16	8786.7(7)	2039.6(13)	6112.2(7)	16.1(2)
C17	8037.8(7)	83.1(12)	5800.1(7)	16.7(2)
C12	8298.8(8)	4864.1(14)	7150.0(7)	19.3(3)
C21	6731.0(7)	5522.7(13)	4280.8(7)	15.6(2)
C22	6015.3(7)	4827.7(14)	4347.0(7)	20.7(3)
C27	6829.8(8)	1492.8(13)	3735.0(7)	18.7(3)
C18	7654.8(8)	363.2(14)	6413.8(7)	19.7(3)
C9	10339.4(8)	4264.7(15)	3845.5(8)	22.9(3)
C20	8641.7(8)	-920.7(14)	5967.0(8)	22.7(3)
C26	5247.7(8)	1881.1(15)	5806.5(7)	21.7(3)
N6	5476.2(10)	7708(2)	2667.8(9)	48.2(5)
C24	6999.7(8)	5115.6(14)	3626.8(7)	19.9(3)
C29	5993.9(10)	7633.0(18)	2417.6(8)	31.7(4)

C30	6662.8(10)	7542.6(19)	2109.2(9)	32.5(4)
C31	3702.2(10)	6371.9(16)	2836.9(9)	31.6(4)
C28	6437.9(10)	876.9(18)	3152.6(8)	31.1(4)
C32	4223.3(13)	5325(2)	2944.9(12)	50.0(6)

Table S23. Bond lengths for **2a**.

Atom	Atom	Length/Å	Atom	Atom	Length/Å
Pd1	N1	2.0201(11)	C1	C2	1.3867(18)
Pd1	N5	2.0937(11)	C1	C7	1.3828(18)
Pd1	N4	2.1626(12)	C5	C4	1.3973(19)
Pd1	N2	2.3955(11)	C5	C6	1.3977(19)
Pd1	N3	2.3819(11)	C5	C9	1.5052(19)
Pd1	C1	1.9588(12)	C2	C3	1.5047(18)
P2	F7	1.6034(10)	C2	C4	1.3966(18)
P2	F9	1.6008(10)	C11	C10	1.3887(18)
P2	F12	1.6081(10)	C11	C12	1.388(2)
P2	F11	1.6016(10)	C10	C16	1.5090(19)
P2	F8	1.5977(10)	C7	C8	1.5062(18)
P2	F10	1.6099(10)	C7	C6	1.3951(18)
P1	F4	1.5927(11)	C15	C14	1.5105(18)
P1	F3	1.6066(11)	C23	C21	1.5397(19)
P1	F2	1.6118(12)	C14	C13	1.3883(18)
P1	F1	1.6050(12)	C25	C26	1.4516(19)
P1	F5	1.5845(13)	N7	C31	1.136(2)
P1	F6	1.5711(12)	C13	C12	1.391(2)
N1	C10	1.3433(16)	C19	C17	1.527(2)
N1	C14	1.3467(17)	C17	C18	1.5327(19)
N5	C27	1.1370(18)	C17	C20	1.5384(19)
N4	C25	1.1392(18)	C21	C22	1.5292(19)
N2	C3	1.5060(16)	C21	C24	1.5315(19)
N2	C15	1.4900(16)	C27	C28	1.457(2)
N2	C21	1.5209(16)	N6	C29	1.139(3)
N3	C8	1.5020(17)	C29	C30	1.455(3)

N3	C16	1.4938(17)	C31	C32	1.456(3)
N3	C17	1.5231(17)			

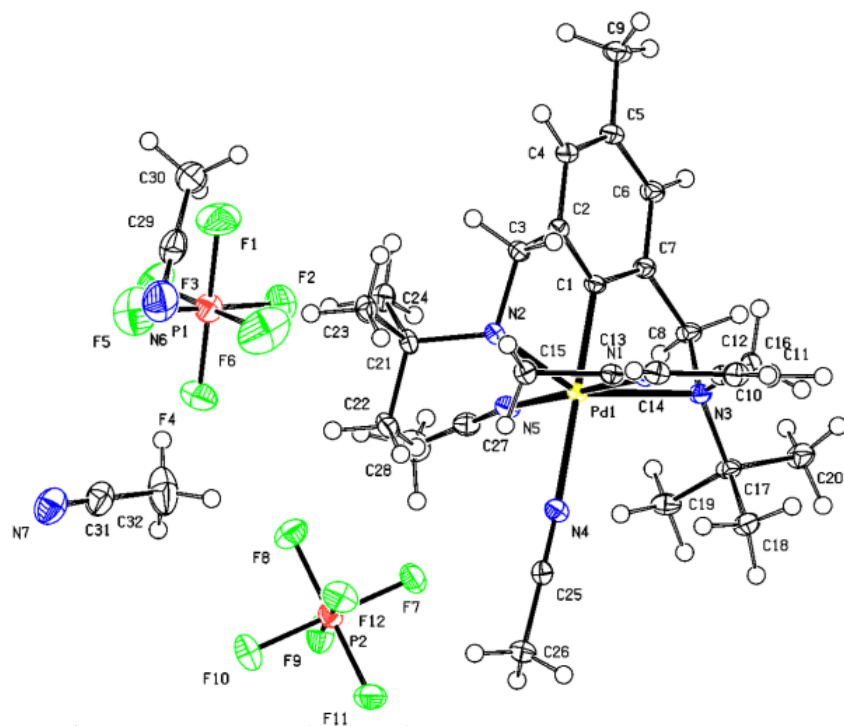


Figure S52. Projection view of **2a** with 50% probability ellipsoids.

9. Computational Details

Geometry optimizations of all compounds were carried out using the atomic coordinates derived from energy minimized or crystal structures. No constraints were forced on the geometry optimization. Geometry optimizations and frequency calculations were performed at the B3LYP level of theory utilizing the Stuttgart/Dresden ECP (SDD) basis set using the Gaussian 16 package.^{18, 19} Water or acetonitrile were used as the solvent, and solvation was modeled utilizing the SMD variant of the IEFPCM SCRF model.²⁰

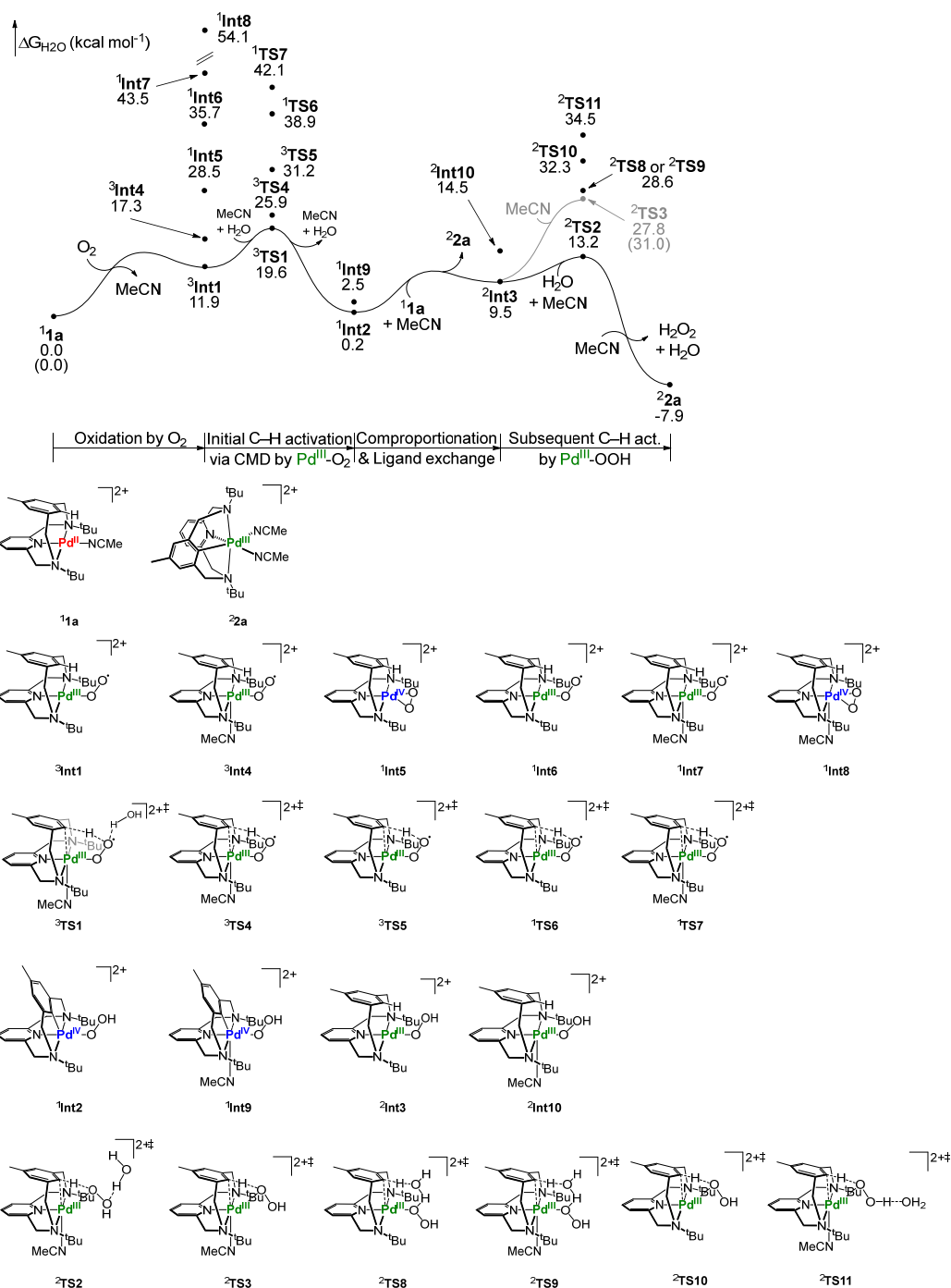


Figure S53. The overall energy profile for the proposed mechanism of the aerobic C-H activation of **1a** by molecular oxygen to afford complex **2a** with possible optimized structures in the reaction pathway. The solvation calculations were utilized using H₂O as solvent. Gibbs energies in the parenthesis on **1a** and **2TS3** stand for Gibbs energies in acetonitrile using solvent model density (SMD) method. Several binuclear Pd-O₂ species were also examined. In these calculations, we did not identify a binuclear structure competitive in energy with the mononuclear Pd^{III}-superoxo intermediate **3Int1**. However, because the absence of a low-energy optimized structure does not exclude all possible binuclear pathways, the present calculations

should be viewed as supporting a mononuclear pathway rather than definitively ruling out binuclear species.

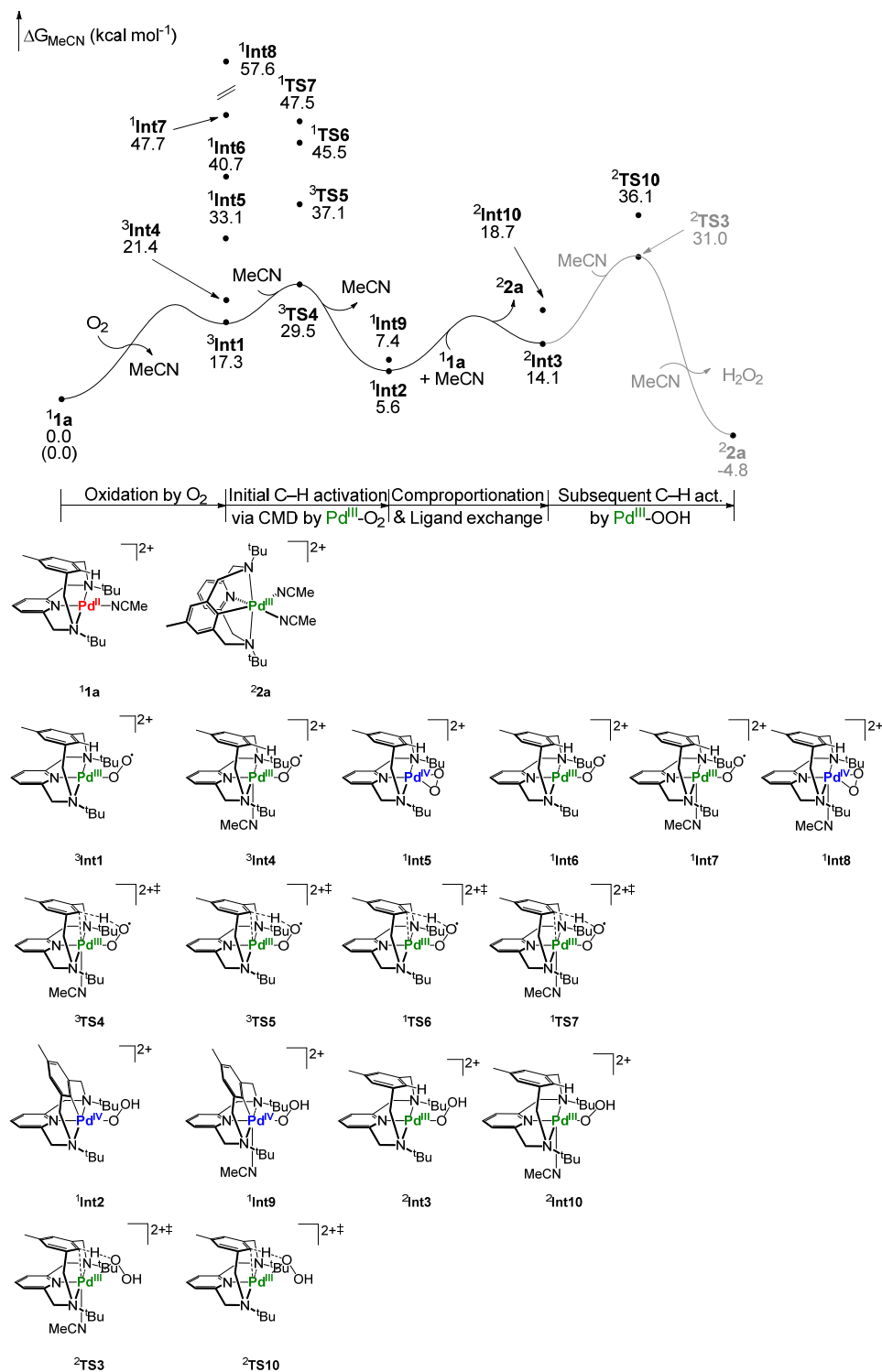


Figure S54. The overall energy profile for the proposed mechanism of the aerobic C-H activation of **1a** by molecular oxygen to afford complex **2a** with possible optimized structures in the reaction pathway. The solvation calculations were utilized using MeCN as solvent.

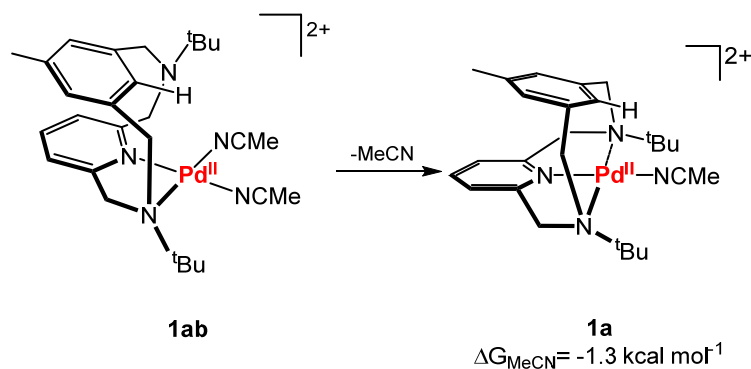
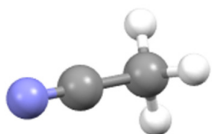


Figure S55. DFT calculation of a reaction from **1ab** to **1a**.

Table 24. Computed energy components for DFT-optimized structures (unit: Hartree)

	E^0	G^0	$E^0_{\text{H}_2\text{O}}$	$G^0_{\text{H}_2\text{O}}$	E^0_{MeCN}	G^0_{MeCN}
MeCN	-132.728543	-132.707055	-132.734784	-132.713296	-132.738698	-132.71721
O ₂	-150.315314	-150.332079	-150.316673	-150.333439	-150.319219	-150.335984
H ₂ O	-76.4143925	-76.411943	-76.4327383	-76.4302888	-76.42478	-76.4223305
H ₂ O ₂	-151.540687	-151.53823	-151.558742	-151.556285	-151.552451	-151.549994
1a	-1359.9134	-1359.36852	-1360.1349	-1359.59002	-1360.15503	-1359.61015
1ab	-1492.67044	-1492.08813	-1492.88364	-1492.30134	-1492.90758	-1492.32527
2a	-1492.06552	-1491.49579	-1492.26789	-1491.69816	-1492.29394	-1491.7242
Int1	-1377.44668	-1376.94425	-1377.69369	-1377.19126	-1377.70378	-1377.20135
Int2	-1377.46628	-1376.96181	-1377.71435	-1377.20987	-1377.72455	-1377.22007
Int3	-1378.07377	-1377.55877	-1378.31526	-1377.80025	-1378.32469	-1377.80969
Int4	-1510.21068	-1509.66882	-1510.43776	-1509.8959	-1510.45385	-1509.91199
Int5	-1377.42313	-1376.91907	-1377.66886	-1377.1648	-1377.68016	-1377.1761
Int6	-1377.41281	-1376.90806	-1377.65803	-1377.15328	-1377.66877	-1377.16402
Int7	-1377.42313	-1509.62685	-1510.39793	-1509.85422	-1510.41386	-1509.87015
Int8	-1510.15625	-1509.61071	-1510.3828	-1509.83726	-1510.39983	-1509.85429
Int9	-1510.24043	-1509.69356	-1510.46639	-1509.91953	-1510.4812	-1509.93433
Int10	-1510.83679	-1510.28253	-1511.05978	-1510.50552	-1511.07376	-1510.5195
TS1	-1586.632563	-1586.074244	-1586.853091	-1586.294772	-	-
TS2	-1587.24941	-1586.67897	-1587.46706	-1586.93781	-	-
TS3	-1510.81633	-1510.26759	-1511.03314	-1510.4844	-1511.04867	-1510.49993
TS4	-1510.19689	-1509.65859	-1510.42035	-1509.88204	-1510.43739	-1509.89909
TS5	-1377.41082	-1376.91309	-1377.65819	-1377.16046	-1377.66755	-1377.16982
TS6	-1377.39361	-1376.89218	-1377.64954	-1377.14811	-1377.65787	-1377.15644
TS7	-1510.16773	-1509.62316	-1510.40092	-1509.85634	-1510.41505	-1509.87047
TS8	-1454.49272	-1453.96194	-1454.73079	-1454.20002	-	-
TS9	-1587.26927	-1586.69582	-1587.48674	-1586.91329	-	-
TS10	-1378.03125	-1377.52364	-1378.27146	-1377.76385	-1378.28221	-1377.7746
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MeCN



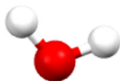
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O₂



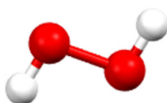
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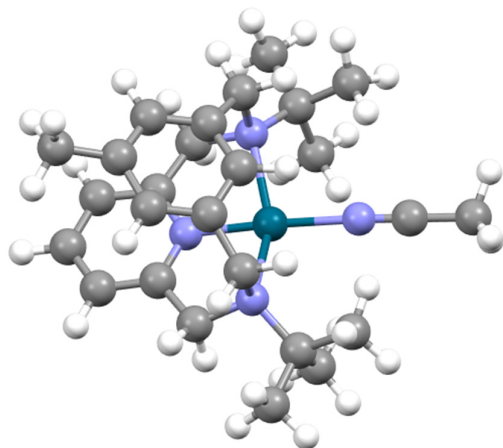
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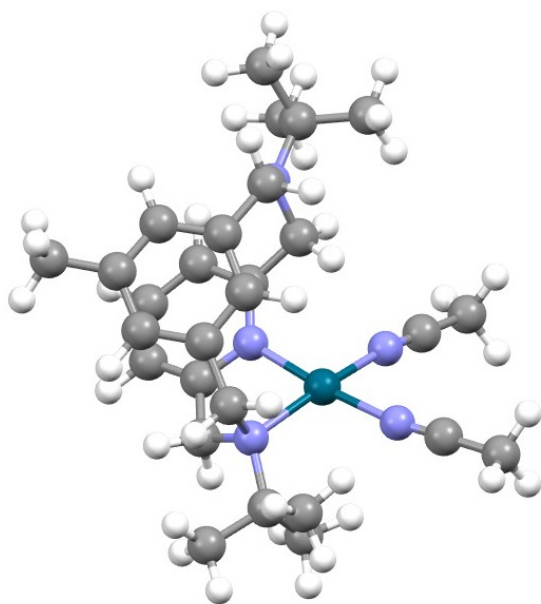
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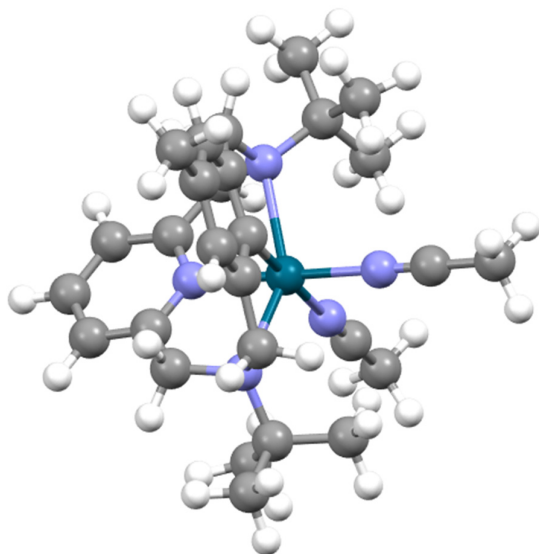
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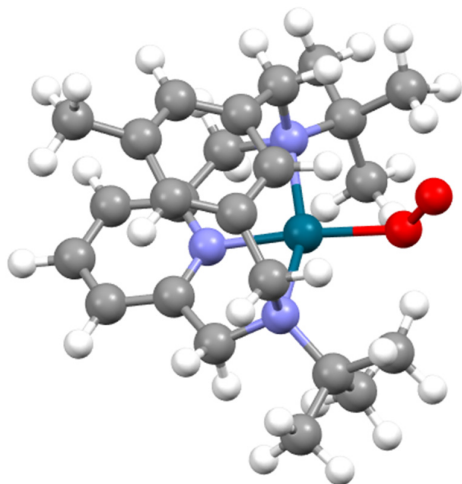
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2a



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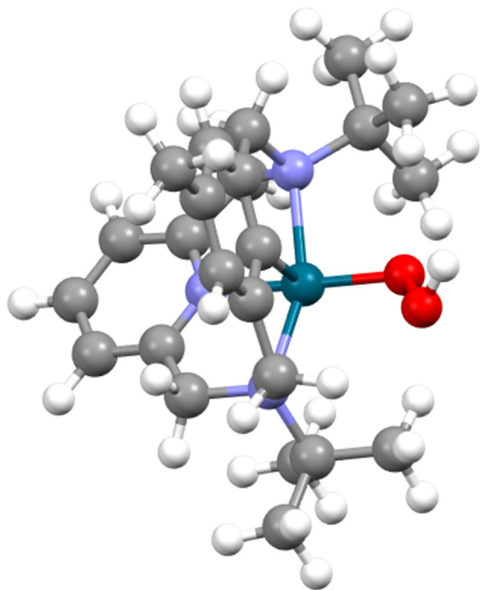
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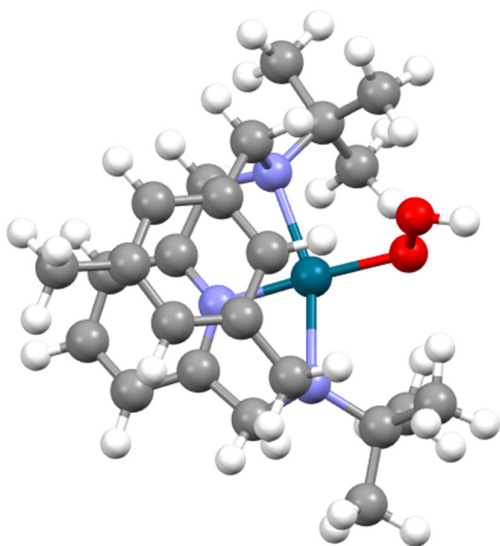
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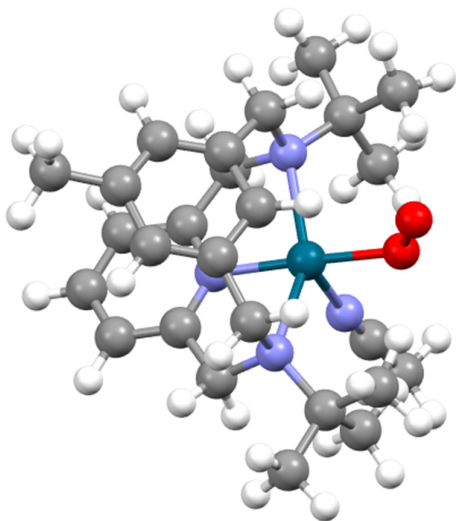
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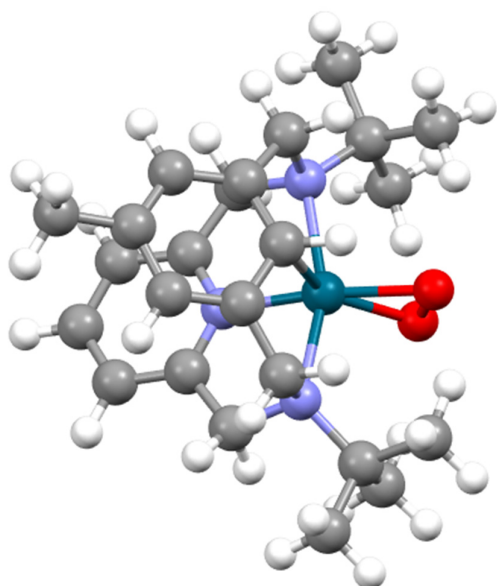
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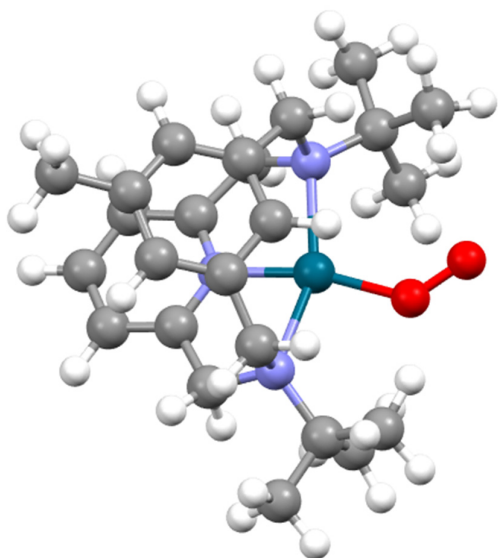
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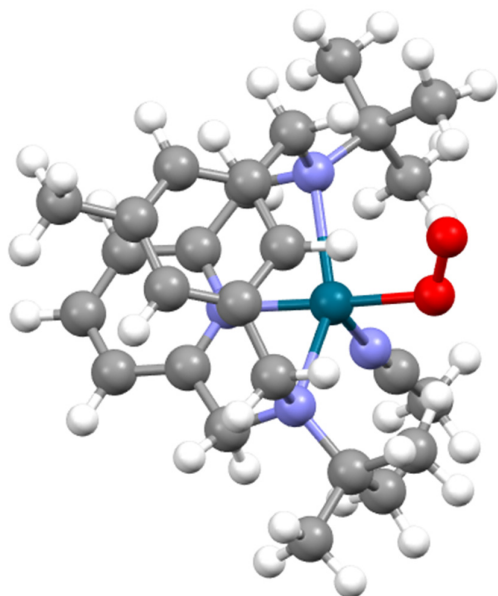
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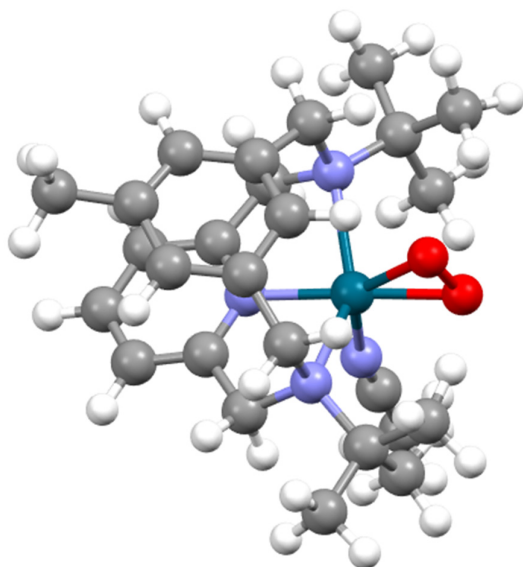
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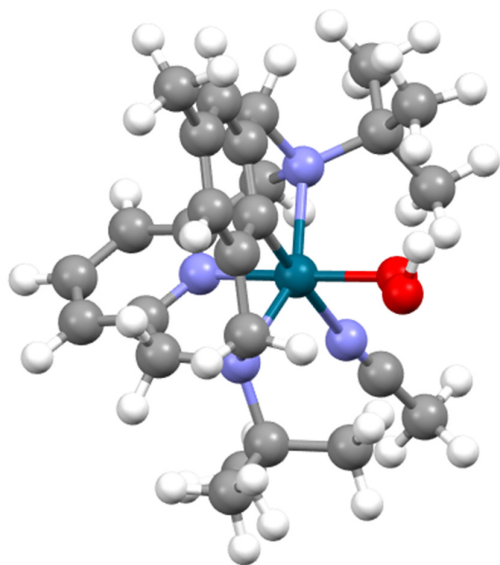
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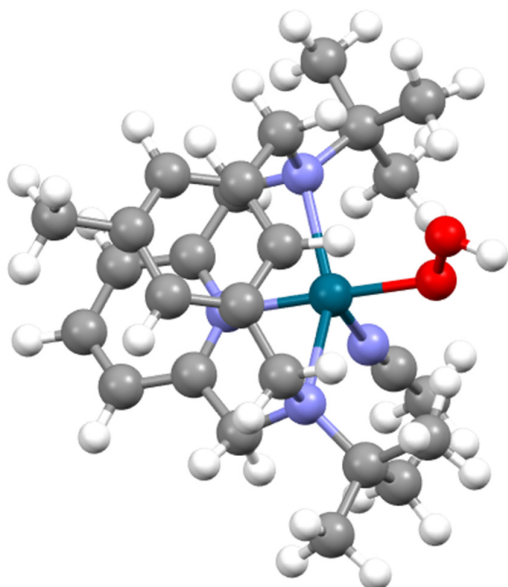
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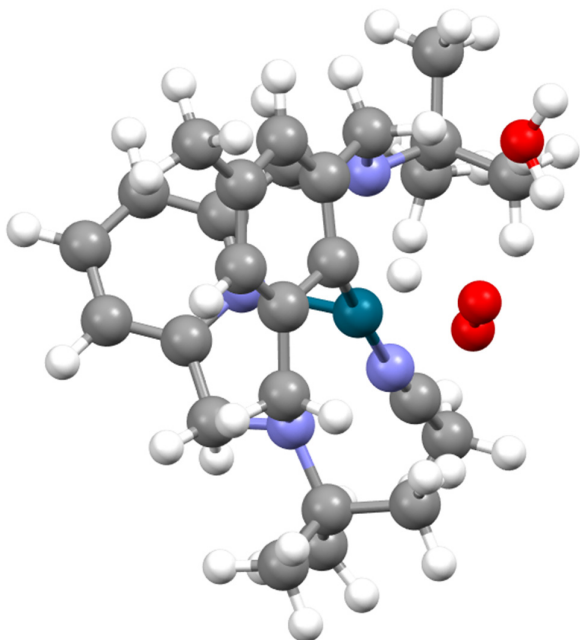
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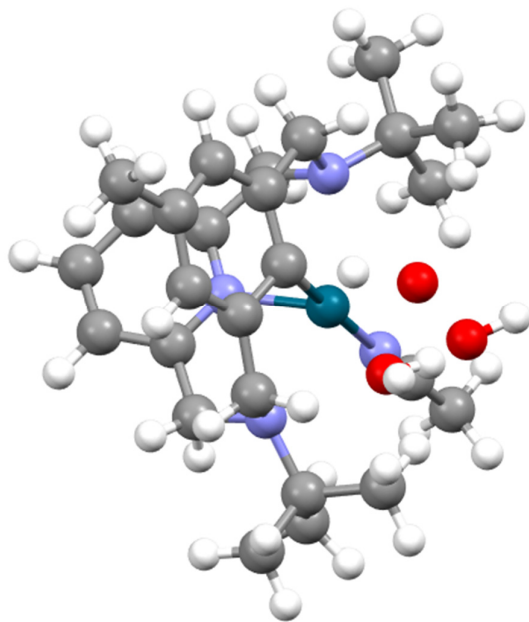
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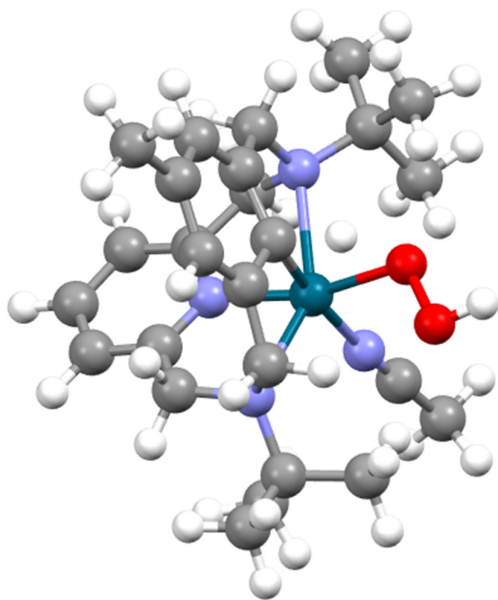
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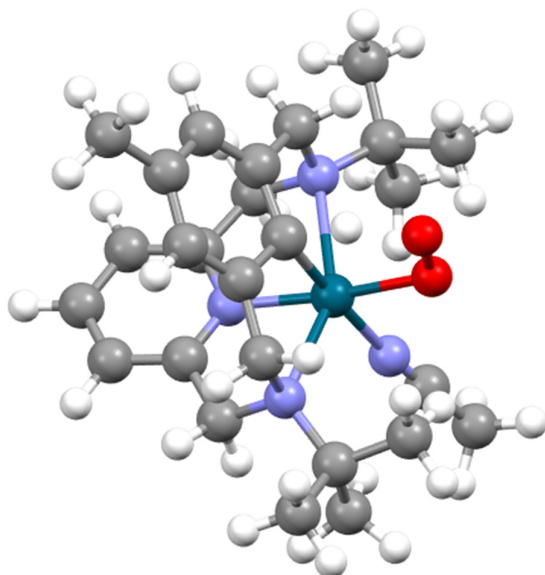
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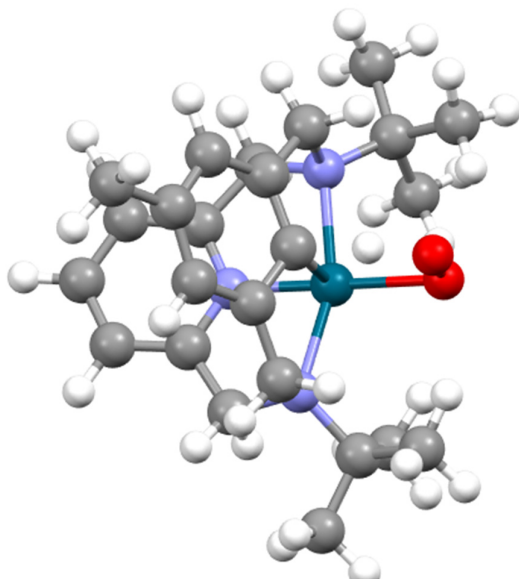
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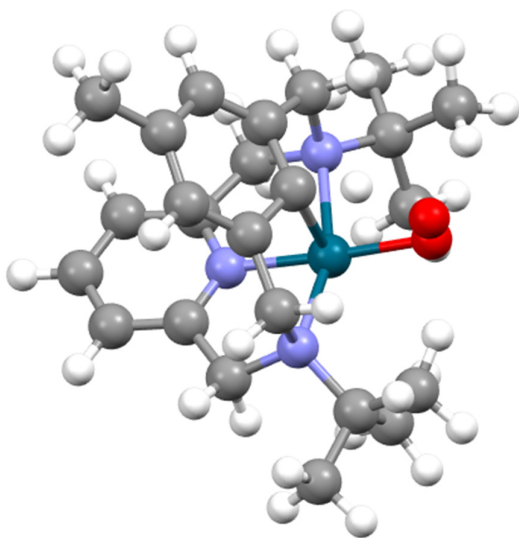
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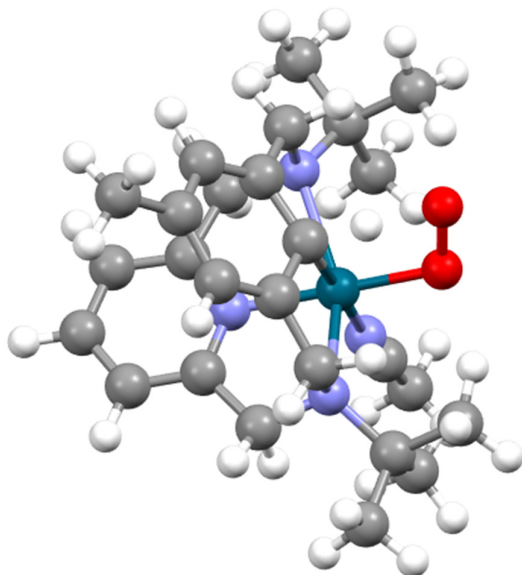
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C 3.5751677998 2.3808556928 14.6251280408
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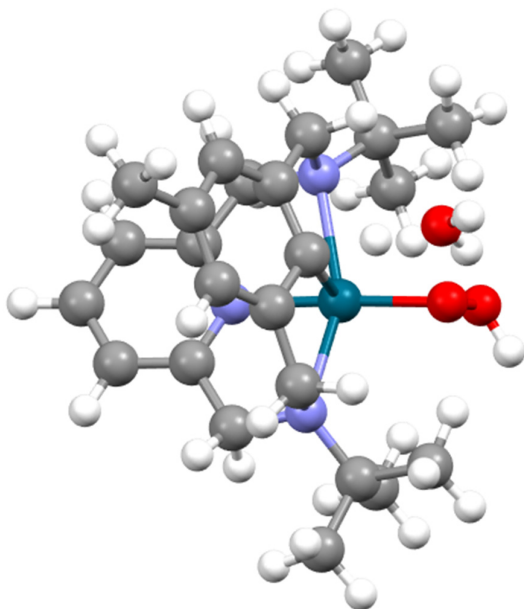
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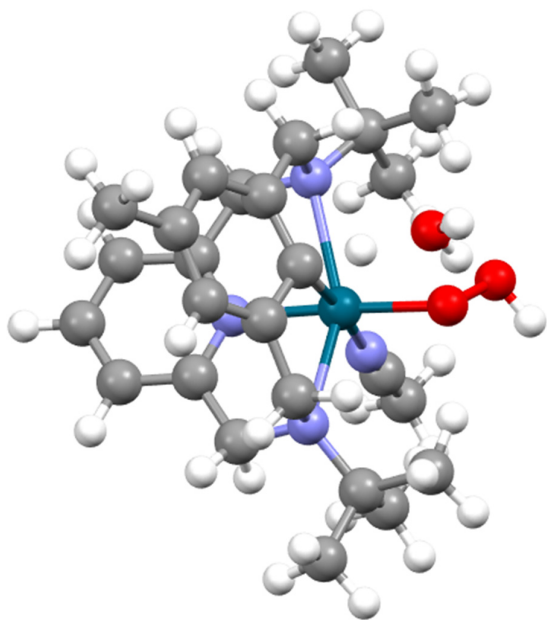
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C 0.6374722172 -1.732078842 1.5310918117
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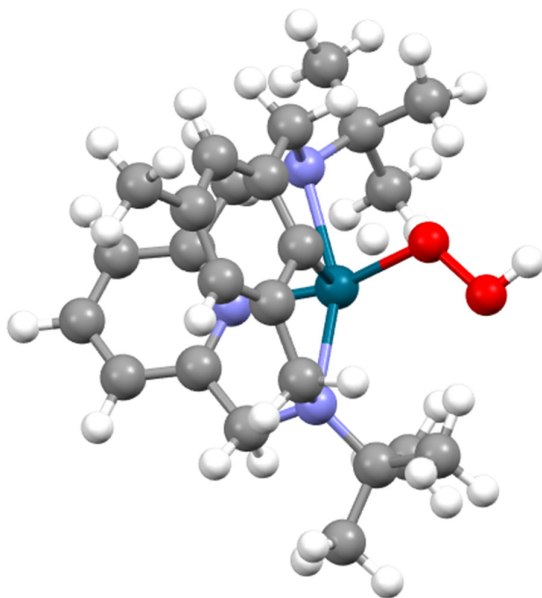
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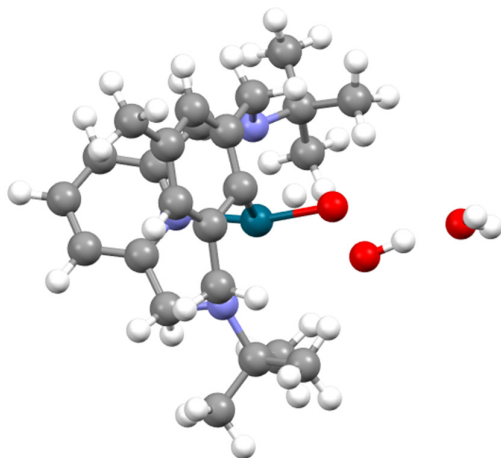
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