

1 Supporting information

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3 **Facile non-fullerene acceptors for efficient photocatalytic hydrogen**  
4 **evolution**

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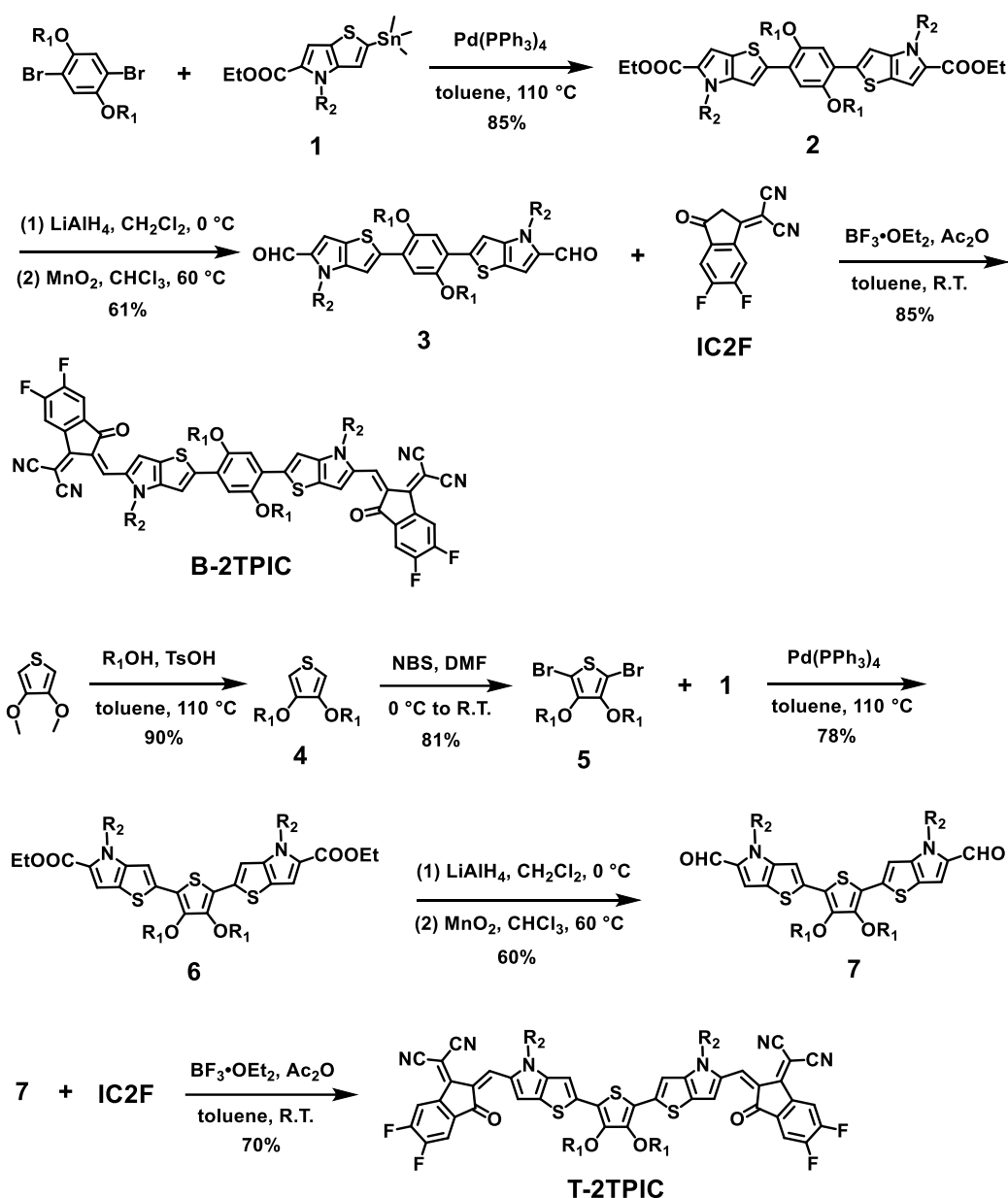
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## 1 Materials and synthesis



2

3 **Scheme S1.** Synthesis routes of B-2TPIC and T-2TPIC.

4

5 Unless stated otherwise, all the solvents and chemical reagents used were obtained

6 commercially and were used without further purification. Toluene and tetrahydrofuran

7 (THF) were distilled from sodium benzophenone under argon prior to use. 3,4-

8 Dimethoxythiophene was purchased from Bide Pharmatech Ltd. 1,4-Dibromo-2,5-

9 bis[(2-ethylhexyl)oxy]benzene was purchased from Shanghai Haohong Biological

10 Pharmaceutical Technology Co., Ltd. Compound **1**<sup>S1</sup> and 3-(1,1-dicyanomethylene)-

1 5,6-difluoro-1-indanone (IC2F)<sup>S2</sup> were synthesized according to our previous works.  
2 PM6 was purchased from Solarmer Material Inc. PDINN was purchased from HyperPV  
3 Technology Co., Ltd.

4

#### 5 *Synthesis of compound 2*

6 To a dry three-necked round flask was added compound **1** (0.33 M in toluene, 7.2  
7 mL, 2.4 mmol), 1,4-dibromo-2,5-bis[(2-ethylhexyl)oxy]benzene (490 mg, 1.00 mmol)  
8 and dry toluene (10 mL). After deoxygenated with argon for 20 min, Pd(PPh<sub>3</sub>)<sub>4</sub> (58 mg,  
9 0.050 mmol) was added and then deoxygenated with argon for another 20 min. The  
10 reaction mixture was stirred at reflux for 24 h. After cooling, the mixture was extracted  
11 with CH<sub>2</sub>Cl<sub>2</sub> (50 mL × 3) and the organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and  
12 filtered. After removing the solvent under reduced pressure, the residue was purified by  
13 column chromatography on silica gel using petroleum ether: CH<sub>2</sub>Cl<sub>2</sub> (6:1, v/v) as the  
14 eluent yielding yellow oil (880 mg, 83%). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 7.12 (s, 2H),  
15 7.03 (s, 2H), 7.01 (s, 2H), 4.41 (d, *J* = 7.6 Hz, 4H), 4.34 (q, *J* = 7.1 Hz, 4H), 3.81 (m,  
16 4H), 2.06–1.99 (m, 2H), 1.78–1.71 (m, 2H), 1.55–1.19 (m, 54H), 0.99–0.85 (m, 24H).  
17 MS (MALDI-TOF): *m/z* 1057.69 [MH]<sup>+</sup>.

18

#### 19 *Synthesis of compound 3*

20 To a dry three-necked round bottom flask were added compound **2** (0.53 g, 0.50  
21 mmol) and dry CH<sub>2</sub>Cl<sub>2</sub> (10 mL). The mixture was deoxygenated with argon for 20 min.  
22 A solution of LiAlH<sub>4</sub> (1.0 M in THF, 1.42 mL, 1.42 mmol) was added dropwise at 0 °C.  
23 The mixture was stirred at 0 °C for 2 h. A saturated NaOH aqueous solution (10 mL)  
24 was added and the mixture was extracted with EtOAc (30 mL × 3). The organic layer  
25 was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and filtered. After removing the solvent, the residue  
26 was dissolved in CHCl<sub>3</sub> again, and MnO<sub>2</sub> (1.09 g, 1.25 mmol) was added in a dry three-  
27 necked round bottom flask under argon. The mixture was deoxygenated with argon for  
28 20 min and was refluxed overnight. After cooling to room temperature, the reaction  
29 mixture was filtered through a pad of celite and the filtrate was dried over anhydrous

1 Na<sub>2</sub>SO<sub>4</sub> and filtered. After removing the solvent, the residue was purified by column  
2 chromatography on silica gel using petroleum ether: CH<sub>2</sub>Cl<sub>2</sub> (2:1, v/v) as eluent  
3 yielding yellow solid (296 mg, 61%). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 9.97 (s, 2H), 7.12  
4 (s, 2H), 7.03 (s, 2H), 7.01 (s, 2H), 4.41 (d, *J* = 7.6 Hz, 4H), 3.81 (m, 4H), 2.06–1.99 (m,  
5 2H), 1.78–1.71 (m, 2H), 1.55–1.19 (m, 48H), 0.99–0.85 (m, 24H). MS (MALDI-TOF):  
6 *m/z* 969.65 [MH]<sup>+</sup>.

7

#### 8 *Synthesis of compound B-2TPIC*

9 To a dry three-necked round bottom flask were added compound **3** (97 mg, 0.10  
10 mmol), IC2F (57 mg, 0.25 mmol), toluene (10 mL), acetic anhydride (0.5 mL) and  
11 BF<sub>3</sub>·Et<sub>2</sub>O (0.3 mL). The reaction mixture was stirred at room temperature for 30 min  
12 and then poured into methanol (50 mL). After filtered, the residue was purified by  
13 column chromatography on silica gel using petroleum ether: CH<sub>2</sub>Cl<sub>2</sub> (2:1, v/v) as eluent  
14 yielding black solid (118 mg, 85%). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 8.87 (s, 2H), 8.37  
15 (s, 2H), 8.34 (s, 2H), 7.60 (s, 2H), 7.51 (s, 2H), 7.11 (s, 2H), 4.21–4.16 (m, 4H), 4.10–  
16 4.07 (m, 4H), 1.97–1.91 (m, 4H), 1.44–1.16 (m, 48H), 1.07–1.03 (m, 6H), 0.97–0.94  
17 (m, 6H), 0.85–0.80 (m, 12H). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>): δ 184.88, 160.17, 152.97,  
18 151.54, 150.34, 135.98, 135.66, 134.74, 128.32, 128.04, 123.99, 119.24, 119.17, 115.90,  
19 115.10, 114.44, 114.30, 112.07, 111.95, 109.75, 71.84, 66.16, 50.75, 39.97, 39.69, 31.72,  
20 31.28, 31.01, 30.83, 29.63, 29.24, 28.50, 26.30, 24.31, 23.12, 23.00, 22.61, 14.15, 14.04,  
21 13.97, 11.27. HR-MS (MALDI-TOF): *m/z* 1393.70018 [MH]<sup>+</sup>.

22

#### 23 *Synthesis of compound 4*

24 To a dry three-necked round bottom flask were added 3,4-dimethoxythiophene  
25 (290 mg, 2.00 mmol), 2-ethylhexan-1-ol (1.04 g, 8.00 mmol), TsOH (35 mg, 0.20 mmol)  
26 and dry toluene (10 mL). After deoxygenated with argon for 30 min, the reaction  
27 mixture was stirred at 110 °C for 6 h. After cooling, the mixture was quenched with  
28 water and extracted with CH<sub>2</sub>Cl<sub>2</sub> (30 mL × 3). The organic layer was dried over  
29 anhydrous Na<sub>2</sub>SO<sub>4</sub> and filtered. After removing the solvent under reduced pressure, the

1 residue was purified by column chromatography on silica gel using petroleum ether:  
2 ethyl acetate (100:1, v/v) as eluent yielding colorless oil (612 mg, 90%). <sup>1</sup>H NMR (400  
3 MHz, CDCl<sub>3</sub>): δ 6.16 (s, 2H), 3.84 (d, *J* = 6.0 Hz, 4H), 1.75 (m, 2H), 1.50–1.31(m,  
4 16H), 0.93 (m, 12H).

5

#### 6 *Synthesis of compound 5*

7 To a dry three-necked round bottom flask was added a solution of compound **4**  
8 (510 mg, 1.50 mmol) in DMF (10 mL) at 0 °C. NBS (587 mg, 3.30 mmol) in DMF (5  
9 mL) was added dropwise to the solution under darkness. The reaction mixture was  
10 stirred at room temperature for 16 h before being poured into water. After being  
11 extracted with CH<sub>2</sub>Cl<sub>2</sub> (50 mL × 3), the organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>  
12 and filtered. After removing the solvent under reduced pressure, the residue was  
13 purified by column chromatography on silica gel using petroleum ether: CH<sub>2</sub>Cl<sub>2</sub> (50:1,  
14 v/v) as the eluent yielding colorless oil (605 mg, 81%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ  
15 3.94 (m, 4H), 1.29–1.65 (m, 18H), 0.93 (m, 12H).

16

#### 17 *Synthesis of compound 6*

18 To a dry three-necked round flask was added compound **1** (0.33 M in toluene, 7.2  
19 mL, 2.4 mmol), compound **5** (502 mg, 1.00 mmol) and dry toluene (10 mL). After  
20 deoxygenated with argon for 20 min, Pd(PPh<sub>3</sub>)<sub>4</sub> (58 mg, 0.050 mmol) was added and  
21 then deoxygenated with argon for another 20 min. The reaction mixture was stirred at  
22 reflux for 24 h. After cooling, the mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> (50 mL × 3) and  
23 the organic layer was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and filtered. After removing the  
24 solvent under reduced pressure, the residue was purified by column chromatography on  
25 silica gel using petroleum ether: CH<sub>2</sub>Cl<sub>2</sub> (6:1, v/v) as the eluent yielding yellow oil (830  
26 mg, 78%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.08 (s, 2H), 7.02 (s, 2H), 4.41 (m, 4H), 4.34  
27 (m, 4H), 3.94 (m, 4H), 2.05–2.00 (m, 2H), 1.91–1.85 (m, 2H), 1.65–1.23 (m, 54H),  
28 1.05–1.02 (m, 6H), 0.97–0.94 (m, 6H), 0.86–0.81 (m, 12H). MS (MALDI-TOF): *m/z*  
29 1063.66 [MH]<sup>+</sup>.

1 *Synthesis of compound 7*

2 To a dry three-necked round bottom flask were added compound **6** (0.53 g, 0.50  
3 mmol) and dry CH<sub>2</sub>Cl<sub>2</sub> (10 mL). The mixture was deoxygenated with argon for 20 min.  
4 A solution of LiAlH<sub>4</sub> (1.0 M in THF, 1.42 mL, 1.42 mmol) was added dropwise at 0 °C.  
5 The mixture was stirred at 0 °C for 2 h. A saturated NaOH aqueous solution (10 mL)  
6 was added and the mixture was extracted with EtOAc (30 mL × 3). The organic layer  
7 was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and filtered. After removing the solvent, the residue  
8 was dissolved in CHCl<sub>3</sub> again, and MnO<sub>2</sub> (1.09 g, 1.25 mmol) was added in a dry three-  
9 necked round bottom flask under argon. The mixture was deoxygenated with argon for  
10 20 min and was refluxed overnight. After cooling to room temperature, the reaction  
11 mixture was filtered through a pad of celite and the filtrate was dried over anhydrous  
12 Na<sub>2</sub>SO<sub>4</sub> and filtered. After removing the solvent, the residue was purified by column  
13 chromatography on silica gel using petroleum ether: CH<sub>2</sub>Cl<sub>2</sub> (2:1, v/v) as eluent  
14 yielding yellow solid (290 mg, 60%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 9.96 (s, 2H), 7.08  
15 (s, 2H), 7.02 (s, 2H), 4.41 (m, 4H), 3.94 (m, 4H), 2.05–2.00 (m, 2H), 1.91–1.85 (m,  
16 2H), 1.65–1.23 (m, 48H), 1.05–1.02 (m, 6H), 0.97–0.94 (m, 6H), 0.86–0.81 (m, 12H).  
17 MS (MALDI-TOF): *m/z* 975.61 [MH]<sup>+</sup>.

18

19 *Synthesis of compound T-2TPIC*

20 To a dry three-necked round bottom flask were added compound **7** (120 mg, 0.12  
21 mmol), IC2F (69 mg, 0.30 mmol), toluene (10 mL), acetic anhydride (0.5 mL) and  
22 BF<sub>3</sub>·Et<sub>2</sub>O (0.3 mL). The reaction mixture was stirred at room temperature for 30 min  
23 and then poured into methanol (50 mL). After filtered, the residue was purified by  
24 column chromatography on silica gel using petroleum ether: CH<sub>2</sub>Cl<sub>2</sub> (2:1, v/v) as eluent  
25 yielding black solid (126 mg, 75%). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>): δ 8.92 (s, 2H), 8.38  
26 (s, 2H), 8.32 (s, 2H), 7.59 (s, 2H), 6.85 (s, 2H), 4.14–4.08 (m, 8H), 2.05–2.00 (m, 2H),  
27 1.91–1.85 (m, 2H), 1.44–1.14 (m, 48H), 1.05–1.02 (m, 6H), 0.97–0.94 (m, 6H), 0.86–  
28 0.81 (m, 12H). <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>): δ 184.84, 159.97, 154.97, 154.79, 153.07,  
29 151.80, 148.74, 145.57, 135.97, 134.73, 128.87, 127.85, 121.12, 119.40, 119.23, 115.75,

1 115.03, 114.43, 114.28, 112.19, 112.07, 105.07, 66.36, 50.65, 40.43, 39.81, 31.74, 31.09,  
2 30.86, 30.19, 29.58, 29.10, 28.41, 26.18, 23.60, 23.18, 23.00, 22.63, 14.14, 14.09, 13.99,  
3 11.12. HR-MS (MALDI-TOF):  $m/z$  1399.65542 [MH]<sup>+</sup>.

4

## 5 **Characterizations**

6 The <sup>1</sup>H and <sup>13</sup>C NMR spectra were measured on Bruker AVANCE 400 MHz and  
7 600 MHz spectrometers. Mass spectra were measured on a Bruker Daltonics BIFLEX  
8 III MALDI-TOF Analyzer using MALDI mode. Solution (chloroform) and thin film  
9 (on quartz substrate) UV-vis absorption spectra were recorded on a JASCO V-570  
10 spectrophotometer. The steady-state photoluminescence and time-resolved  
11 photoluminescence spectra of films were measured using an FLS1000 spectrometer.  
12 The films were prepared by spin-coating the solution (15 mg mL<sup>-1</sup> in chloroform) at  
13 2000 rpm for 40 s on the quartz substrate.

14 Electrochemical measurements were carried out under nitrogen on a deoxygenated  
15 solution of tetra-*n*-butylammonium hexafluorophosphate (0.1 M) in CH<sub>3</sub>CN using a  
16 computer-controlled CHI660C electrochemical workstation, a glassy-carbon working  
17 electrode, a platinum-wire auxiliary electrode, and an Ag/AgCl as a reference electrode.  
18 The samples were first dissolved in chloroform, and the resulting solutions were drop-  
19 cast onto a glassy carbon working electrode to form thin films prior to the measurements.  
20 Potentials were referenced to ferrocenium/ferrocene (FeCp<sub>2</sub><sup>0/+</sup>) couple by using  
21 ferrocene as an internal standard. The HOMO and LUMO energy levels of the samples  
22 were estimated from onset redox potentials, assuming the absolute energy level of  
23 FeCp<sub>2</sub><sup>0/+</sup> to be 4.8 eV below vacuum. The HOMO and LUMO energy levels of the  
24 samples were calculated as:

$$25 E_{\text{HOMO}} = - (E_{\text{ox}}^{\text{onset}} + 4.8 \text{ eV}) \text{----- eqn S1}$$

$$26 E_{\text{LUMO}} = - (E_{\text{red}}^{\text{onset}} + 4.8 \text{ eV}) \text{----- eqn S2}$$

27 where  $E_{\text{ox}}^{\text{onset}}$  (onset potentials for oxidation) and  $E_{\text{red}}^{\text{onset}}$  (onset potentials for reduction)  
28 are defined as the difference between the respective onset potential of the sample and  
29 the redox potential of FeCp<sub>2</sub><sup>0/+</sup>.

1 Grazing incidence wide-angle X-ray scattering (GIWAXS) measurements were  
2 accomplished with a Xeuss 2.0 SAXS/WAXS laboratory beamline using a Cu X-ray  
3 source (8.05 keV, 1.54 Å) and a Pilatus3R 300K detector. The incidence angle is 0.2°.  
4 GIWAXS samples were prepared by spin coating B-2TPIC or T-2TPIC chloroform  
5 solution (12 mg mL<sup>-1</sup>) at 1500 rpm for 40 s on silicon substrate.

6 The hydrodynamic diameter and the zeta potentials of nanoparticles were measured  
7 by dynamic light scattering (Malvern Zetasizer ZS).

### 8 9 **Computational methodology**

10 Density functional theory (DFT) calculations were carried out at the B3LYP/6-  
11 31G(d) level in the Gaussian 16 package,<sup>S3</sup> where all alkyl side chains of B-2TPIC and  
12 T-2TPIC were replaced by methyl groups to simplify the calculations. Geometry  
13 optimizations were performed with full relaxation of all atoms in gas phase, following  
14 which vibration frequency calculations were used to check that the optimized structures  
15 had no imaginary frequency. Charge distribution of the molecules was examined by  
16 Mulliken population analysis. The surface electrostatic potential (ESP) mapping was  
17 obtained by using the Multiwfn software.<sup>S4, S5</sup>

### 18 19 **SCLC measurements**

20 Hole-only devices were fabricated for hole mobility measurements with the  
21 structure of ITO/PEDOT:PSS/active layer/MoO<sub>3</sub>/Ag. The ITO substrates were first  
22 scrubbed by detergent and then sonicated with deionized water, acetone and isopropanol  
23 subsequently, and dried in an oven. The ITO glass substrates were treated by UV-ozone  
24 for 20 min before use. PEDOT:PSS was spin-coated onto the ITO substrates at 4000  
25 rpm for 30 s. After annealing at 150 °C for 15 min in air, the active layers were formed  
26 by spin-coating a solution of B-2TPIC or T-2TPIC (18 mg mL<sup>-1</sup> in chloroform) at 2000  
27 rpm for 40 s on PEDOT:PSS in a glovebox. Then, the MoO<sub>3</sub> layer (ca. 5 nm) and Ag  
28 (ca. 90 nm) were successively evaporated onto the surface of the photoactive layer  
29 under vacuum (ca. 10<sup>-5</sup> Pa).

1 Electron-only devices were fabricated for electron mobility measurements with the  
2 structure ITO/ZnO/active layer/PDINN/Ag. The mixture of zinc acetate dihydrate (100  
3 mg), 2-methoxyethanol (973  $\mu\text{L}$ ) and ethanolamine (28.29  $\mu\text{L}$ ) was stirred overnight to  
4 prepare the ZnO precursor solution. After the pretreatment of the ITO substrate, ZnO  
5 layer (ca. 30 nm) was spin-coated onto the ITO substrate at 4000 rpm for 30 s from  
6 ZnO precursor solution. After annealing at 200  $^{\circ}\text{C}$  for 30 min, the active layers were  
7 formed by spin-coating a solution of B-2TPIC or T-2TPIC (18 mg  $\text{mL}^{-1}$  in chloroform)  
8 at 2000 rpm for 40 s on ZnO in a glovebox. Then, a thin PDINN layer ( $\sim 5$  nm) was  
9 coated on the active layer, followed by the deposition of Ag electrode (90 nm).

10 The active area of the device was 4  $\text{mm}^2$  and the thickness of the active layer was  
11 90 nm. The mobility was extracted by fitting the  $J$ - $V$  curves using space charge limited  
12 current (SCLC) method: <sup>S6</sup>

13  $J = \frac{9\epsilon_r\epsilon_0\mu V^2}{8L^3}$  ----- eqn S3

14 here,  $J$  refers to the current density,  $\mu$  is hole or electron mobility,  $\epsilon_r$  is relative dielectric  
15 constant of the transport medium, which is equal to 3,  $\epsilon_0$  is the permittivity of free space  
16 ( $8.85 \times 10^{-12}$  F  $\text{m}^{-1}$ ),  $V = V_{\text{appl}} - V_{\text{bi}}$ , where  $V_{\text{appl}}$  is the applied voltage to the device, and  
17  $V_{\text{bi}}$  is the built-in voltage due to the difference in work function of the two electrodes,  $L$   
18 is the thickness of the active layer.

## 20 Nanoparticle preparation

21 Individual stock solutions (0.5 mg  $\text{mL}^{-1}$ ) of B-2TPIC, T-2TPIC and PM6 were  
22 prepared in chloroform. The solutions were stirred at room temperature for 2 h. The  
23 precursor solutions of single-component nanoparticles (NPs) were prepared with the  
24 corresponding stock solution, while that of heterojunction NPs were prepared from the  
25 stock solutions by mixing them in the ratio of the desired NP composition. Then NP  
26 precursor solution (1 mL) was added to the different weight ratios of TEBS aqueous  
27 solution (4 mL) and then sonicated for 5 min with an ultrasonic processor (Sonics  
28 SCIENTZ-IID) to obtain the mini-emulsion. The mini-emulsion was evaporated to  
29 remove the chloroform, obtaining a surfactant-stabilized NP dispersion in water. Finally,

1 the dispersion was filtered (0.45- $\mu\text{m}$  mixed cellulose) to remove any large aggregates.  
2 The final composition was determined by the following: 2 mL methanol was added to  
3 1 mL samples for de-emulsification. Then 2 mL chloroform was added to extract the  
4 semiconductor material dissolved in chloroform. The solution was evaporated to  
5 remove the organic solvents and then 1 mL chloroform was added to the samples.  
6 Finally, the solution was measured by UV-vis absorption to determine the final weight  
7 of B-2TPIC, T-2TPIC, PM6:B-2TPIC or PM6:T-2TPIC NPs in the solutions.

8

### 9 **Hydrogen evolution**

10 The concentration of single-component (B-2TPIC or T-2TPIC) NPs and  
11 heterojunction (PM6:B-2TPIC or PM6:T-2TPIC) NPs was calibrated by UV-vis  
12 absorption. Then, different volumes of photocatalyst NPs solution were added to the  
13 prepared ascorbic acid (AA) aqueous solution (0.2 M) to obtain NPs solutions with  
14 different concentrations, and the total volume was maintained at 7.5 mL. The  
15 concentration of the suspensions with single-component (B-2TPIC or T-2TPIC) NPs  
16 and heterojunction (PM6:B-2TPIC or PM6:T-2TPIC) NPs for the HER was 6.67  $\mu\text{g mL}^{-1}$   
17 or 26.67  $\mu\text{g mL}^{-1}$ . According to the usage for corresponding reaction condition, a  
18 certain amount of  $\text{K}_2\text{PtCl}_6$  aqueous solution (1  $\text{mg mL}^{-1}$ ) was added to the above-  
19 mentioned NPs solutions with corresponding concentrations. The aging treatment of  
20 the  $\text{K}_2\text{PtCl}_6$  solution was performed according to the modified method reported in  
21 literature,<sup>S7</sup> and different aging treatments affect the initial demand for Pt loading ratios.  
22 For the HER under simulated solar light illumination (AM 1.5G, 100  $\text{mW cm}^{-2}$ ), the  
23 mixed reaction solution was added into a recirculating batch reactor (cross sectional  
24 area, 5.786  $\text{cm}^2$ ; penetration depth of light, 1.296 cm) kept at 5  $^\circ\text{C}$ . The recirculating  
25 batch reactor was purged with nitrogen several times to remove oxygen, and the  
26 pressure was set to 1.2 kPa. The suspensions with photocatalyst NPs were stirred and  
27 illuminated with a 300 W Xe lamp equipped with a mirror module (330–1100 nm) and  
28 AM 1.5G filtered. Multiple-channel reactors were applied to improve the efficiency of  
29 screening the hydrogen evolution conditions of single-component (B-2TPIC or T-

1 2TPIC) NPs and heterojunction (PM6:B-2TPIC or PM6:T-2TPIC) NPs. The  
2 photocatalyst NPs formed under various conditions were respectively added into each  
3 off-line reactors (cross-sectional area, 3.142 cm<sup>2</sup>; penetration depth of light, 2.387 cm)  
4 with 0.2 M AA in aqueous solution (7.5 mL), and then the reactors were purged with  
5 nitrogen several times to remove oxygen. The suspensions with photocatalyst NPs were  
6 stirred at room temperature and illuminated with the light source which covered the  
7 visible range (400–800 nm, 100 mW cm<sup>-2</sup>). All hydrogen evolution was evaluated by  
8 an all-glass automatic online trace gas analysis system (Labsolar 6A, PerfectLight) with  
9 an online gas chromatograph (Thermal conductivity detector, N<sub>2</sub> carrier). The multiple-  
10 channel photochemical reaction system is configured with an LED lamp (400–800 nm,  
11 100 mW cm<sup>-2</sup>) rather than the reference spectrum for standard tests of solar energy  
12 conversion systems (AM 1.5G, 100 mW cm<sup>-2</sup>). Meanwhile, the reactor has a light-  
13 concentrating effect leading to the actual irradiation intensity being higher than 100  
14 mW cm<sup>-2</sup>. Hence, all values from multiple-channel reactors are normalized to the  
15 maximum value in each set of parallel experiments to obtain the optimal conditions.  
16 Three parallel hydrogen evolution experiments of the champion PM6:B-2TPIC NPs  
17 were conducted under the same optimized conditions to verify reproducibility.

18

### 19 **EQE measurements**

20 EQE was measured by using PM6:B-2TPIC (26.67 μg mL<sup>-1</sup>; D:A, 4:6, w/w; 0.3  
21 wt% TEBS/H<sub>2</sub>O; 20 wt% Pt loading) or PM6:T-2TPIC (26.67 μg mL<sup>-1</sup>; D:A, 3:7, w/w;  
22 0.5 wt% TEBS/H<sub>2</sub>O; 16 wt% Pt loading) and 0.2 M AA aqueous solution (7.5 mL;  
23 cross-sectional area, 5.786 cm<sup>2</sup>; penetration depth of light, 1.3 cm). An all-glass  
24 automatic online trace gas analysis system (Labsolar 6A, PLS-SXE300+, PerfectLight)  
25 was used for the measurements, equipped with an online gas chromatograph featuring  
26 a thermal conductivity detector and N<sub>2</sub> as the carrier gas. Monochromatic light (400 nm,  
27 500 nm, 600 nm, 700 nm, 808 nm) was obtained from a 300 W xenon lamp fitted with  
28 corresponding bandpass filters which have a bandwidth of 15 nm. The sample was first  
29 illuminated under AM 1.5G light, 100 mW cm<sup>-2</sup> for 2 h to complete Pt photodeposition.

1 Then, the reactor was evacuated to remove all of the H<sub>2</sub> that evolved during this time.

2 The light source was fitted with a corresponding bandpass filter for EQE measurements.

3 The EQE was calculated using equations:<sup>S8</sup>

4 
$$\text{EQE} = 2 \times \frac{n_h}{n_p} \text{----- eqn S4}$$

5 
$$n_p = \frac{It\lambda}{N_Ahc} \text{----- eqn S5}$$

6 where  $n_h$  represents the number of moles of H<sub>2</sub> evolved per hour,  $n_p$  represents the total

7 number of incident photons per hour,  $I$  is the radiant power,  $t$  is the irradiation time

8 (excluding the induction time),  $\lambda$  is the light wavelength,  $N_A$  is the Avogadro constant,

9  $h$  is the Planck constant,  $c$  is the speed of light.

10

### 11 **Transmission electron microscope (TEM)**

12 High-resolution field emission TEM (Hitachi HT7700) was employed to

13 investigate the internal morphology of the PM6:B-2TPIC and PM6:T-2TPIC NPs. The

14 fabrication of NPs for TEM measurements was described in nanoparticle preparation

15 part. The specimen was prepared by placing 3.5  $\mu\text{L}$  of solution onto the copper TEM

16 grid and drying at room temperature. The TEM analysis was performed by setting the

17 microscope at an acceleration voltage of 100 kV.

18

### 19 **Transient absorption**

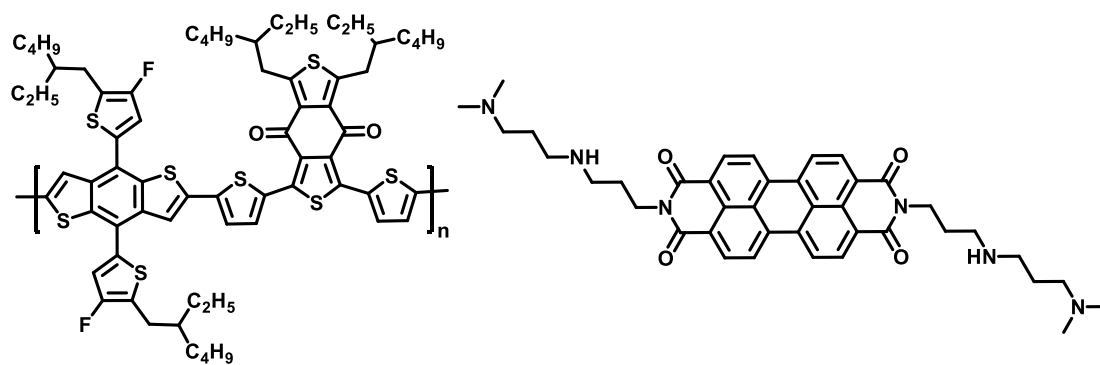
20 Transient absorption (TA) data were collected by using a pump and probe

21 femtosecond TA spectroscopy system (Vitara T-Legend Elite-TOPAS-Helios-EOS-

22 Omni). The samples were prepared as described in nanoparticle preparation part with a

23 concentration of 50  $\mu\text{g mL}^{-1}$ , ensuring strong absorption when excited by a pump laser

24 at 750 nm. Light in the ranges of 400–800 nm was served as the probe source.



**PM6**

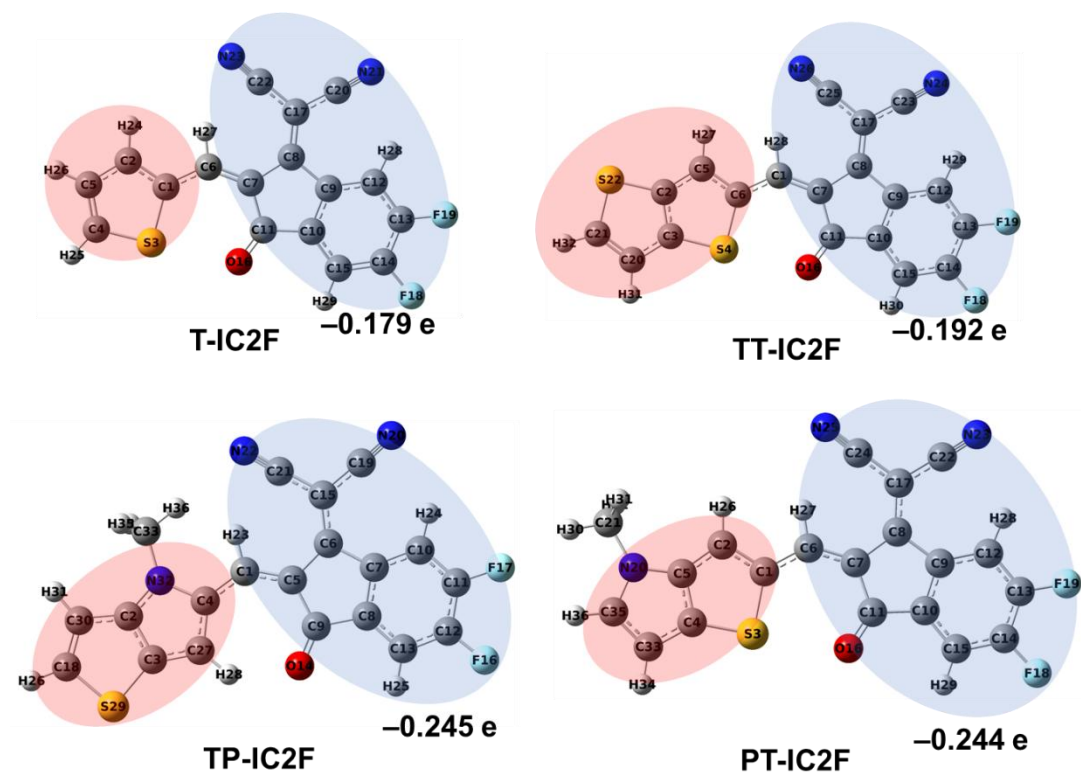
**PDINN**

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2 **Fig. S1** Chemical structures of PM6 and PDINN.

3

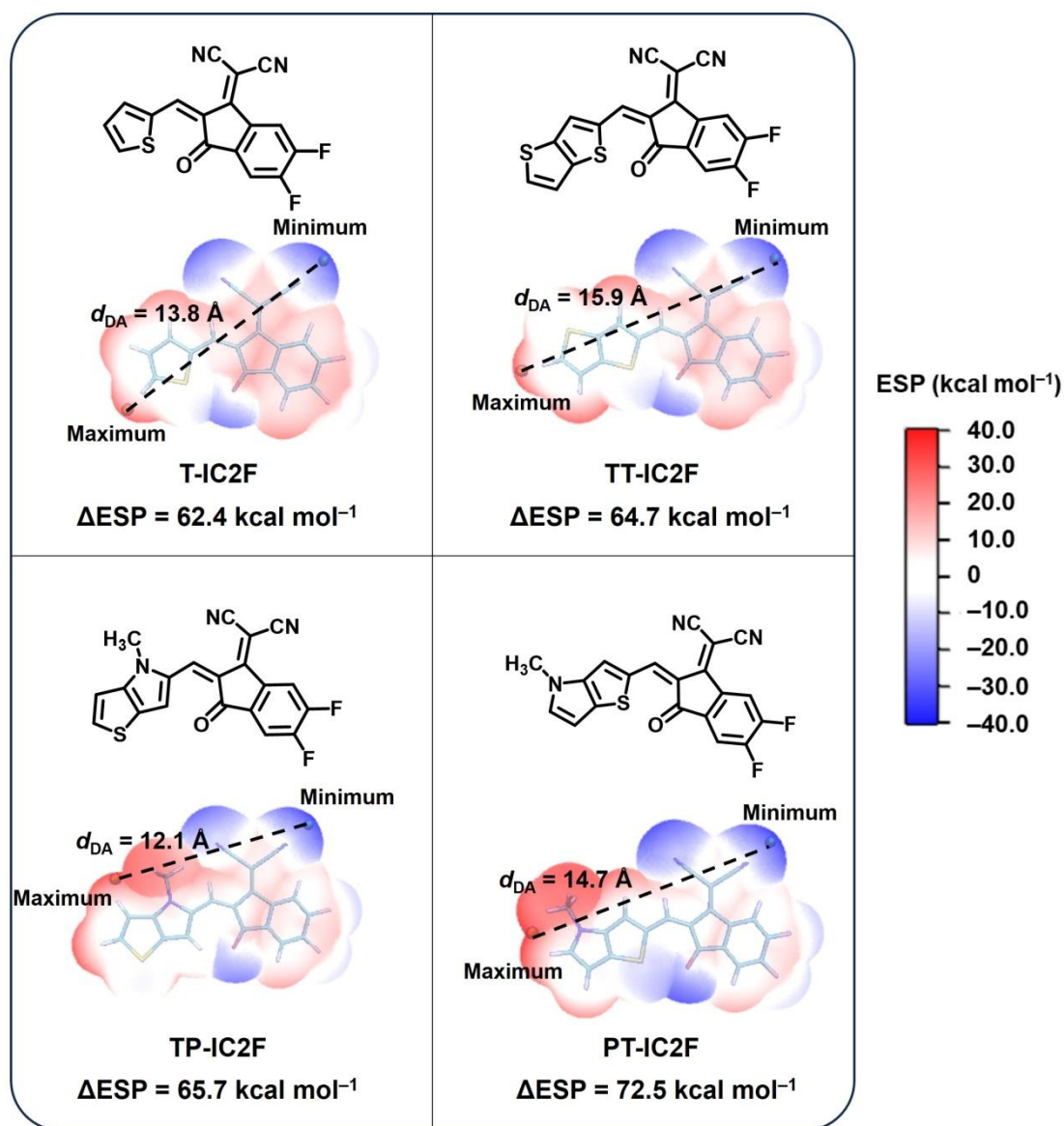
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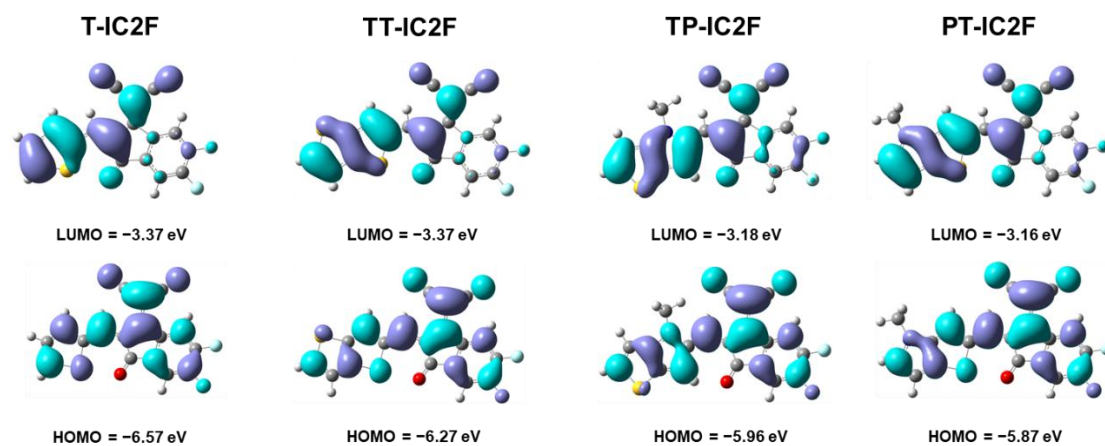
2 **Fig. S2** Mulliken population analysis of T-IC2F, TT-IC2F, TP-IC2F, and PT-IC2F.

3



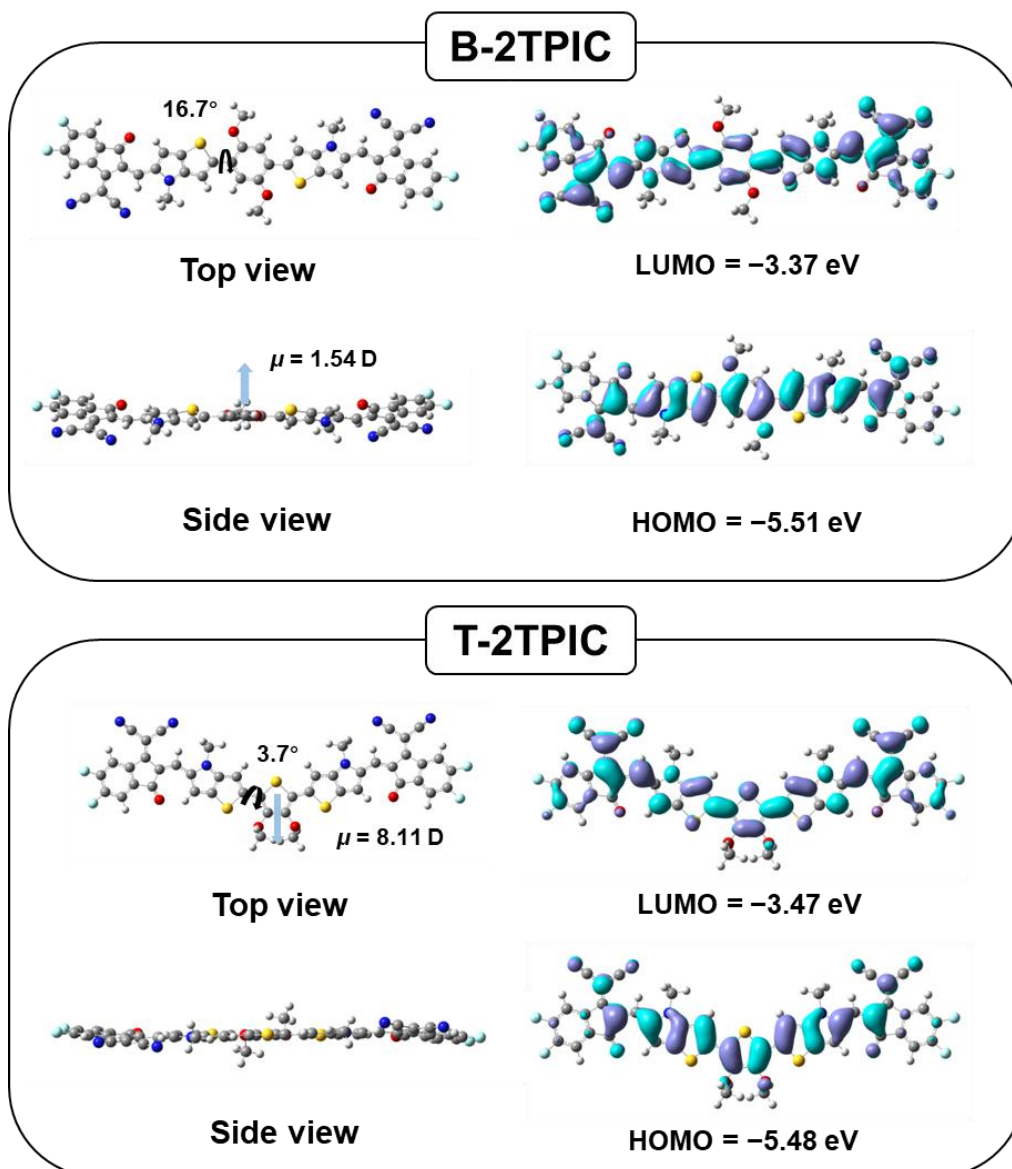
1  
 2 **Fig. S3** Chemical structures and surface ESP of T-IC2F, TT-IC2F, TP-IC2F, and PT-  
 3 IC2F. In ESP diagrams, the orange spheres represent the surface maximum points, and  
 4 the blue spheres represent the surface minimum points.

5



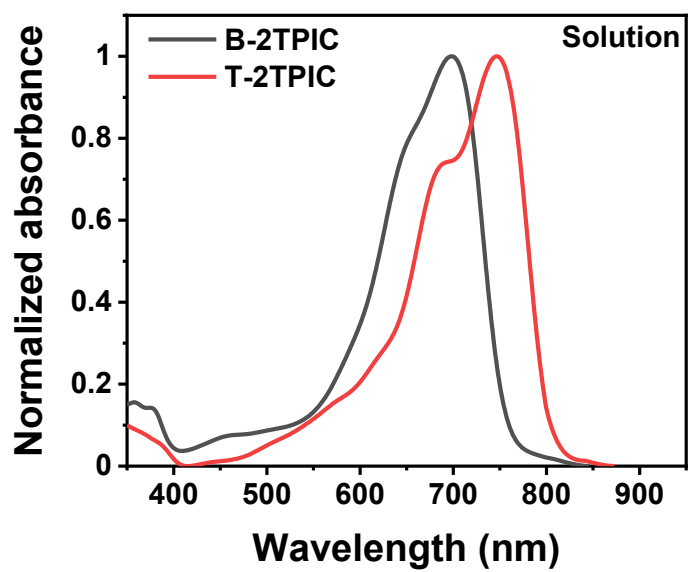
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**Fig. S4** The energy levels of T-IC2F, TT-IC2F, TP-IC2F and PT-IC2F. Calculated at the B3LYP/6-31G(d) level (isovalue = 0.02).



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**Fig. S5** The optimized geometry and calculated energy levels of B-2TPIC and T-2TPIC at the B3LYP/6-31G(d) level. To simplify the calculation process, the alkyl side chains were replaced by methyl groups.



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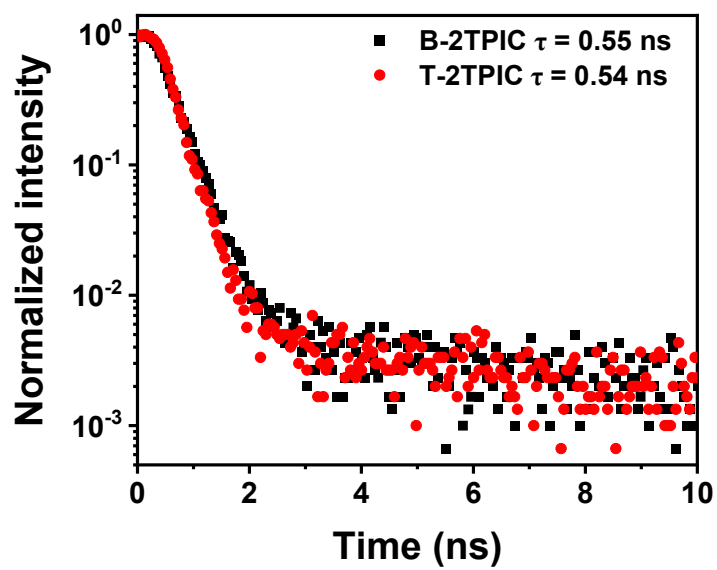
2 **Fig. S6** Normalized absorption spectra of B-2TPIC and T-2TPIC in chloroform  
3 solutions ( $10^{-6}$  M).

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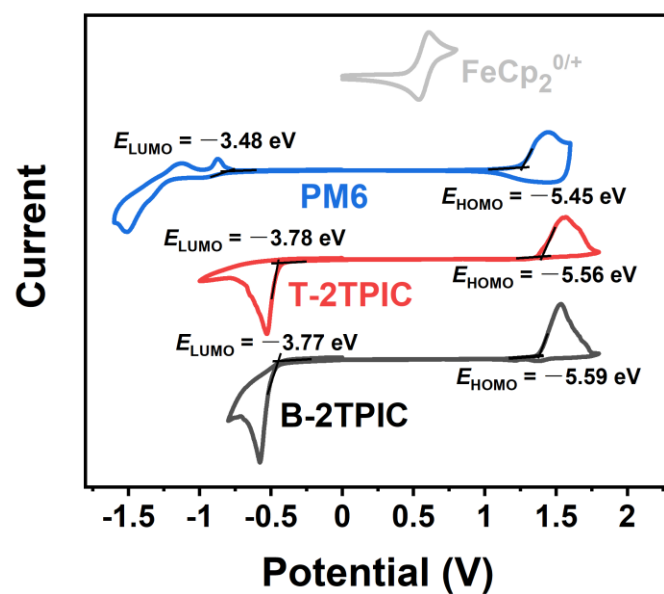
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2 **Fig. S7** TRPL spectra of B-2TPIC and T-2TPIC in films.

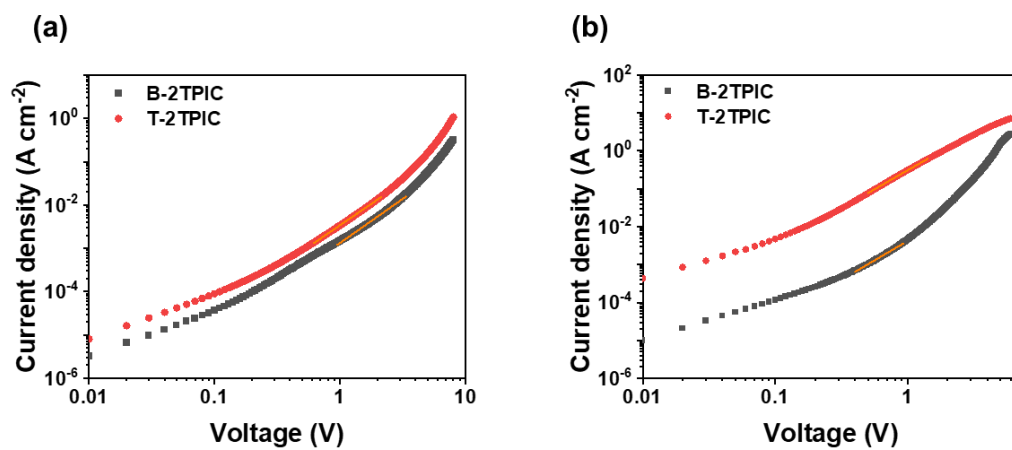


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2 **Fig. S8** Cyclic voltammograms of B-2TPIC, T-2TPIC and PM6 in  $\text{CH}_3\text{CN}/0.1 \text{ M}$   
 3  $[\text{Bu}_4\text{N}]^+[\text{PF}_6]^-$  at  $100 \text{ mV s}^{-1}$ , in which the horizontal scale refers to an Ag/AgCl  
 4 electrode.

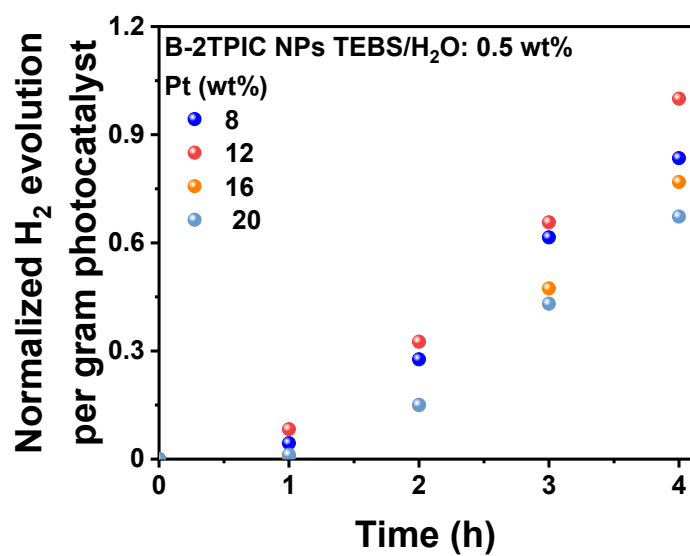
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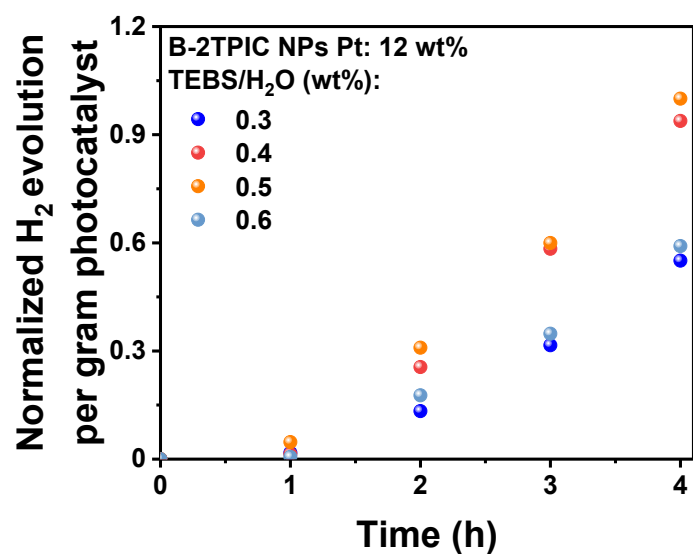
**Fig. S9**  $J$ - $V$  characteristics in dark for (a) hole-only and (b) electron-only devices based on B-2TPIC and T-2TPIC.



1

2 **Fig. S10** Normalized hydrogen evolution per gram photocatalyst versus time of B-  
 3 2TPIC NPs at a concentration of  $6.67 \mu\text{g mL}^{-1}$  with 0.5 wt% TEBS/H<sub>2</sub>O and varying  
 4 Pt loadings at room temperature, measured by a multiple-channel photochemical  
 5 reaction system.

6

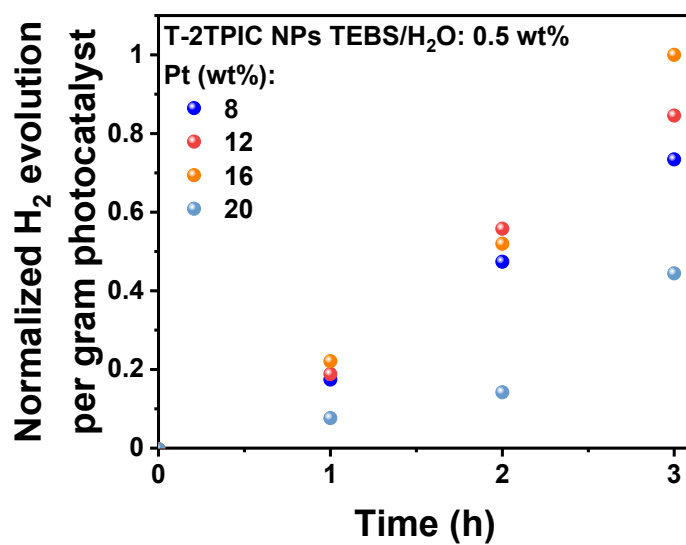


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2 **Fig. S11** Normalized hydrogen evolution per gram photocatalyst versus time of B-  
3 2TPIC NPs at a concentration of  $6.67 \mu\text{g mL}^{-1}$  with 12 wt% Pt loading at different  
4 weight ratios of surfactant (TEBS/H<sub>2</sub>O) at room temperature, measured by a multiple-  
5 channel photochemical reaction system.

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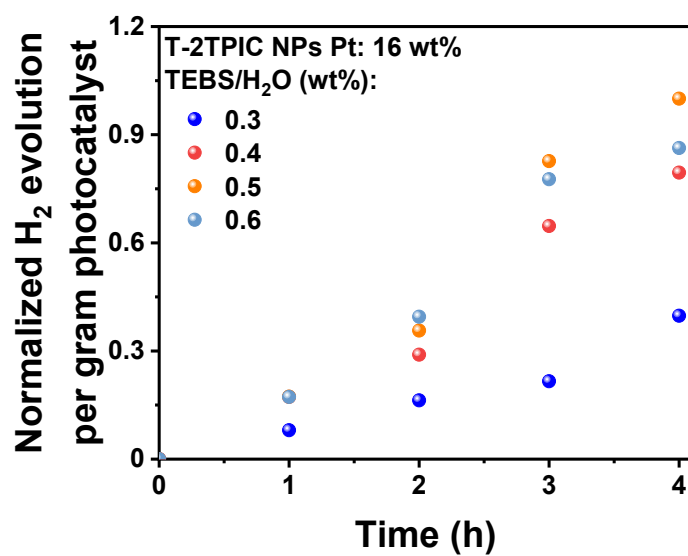
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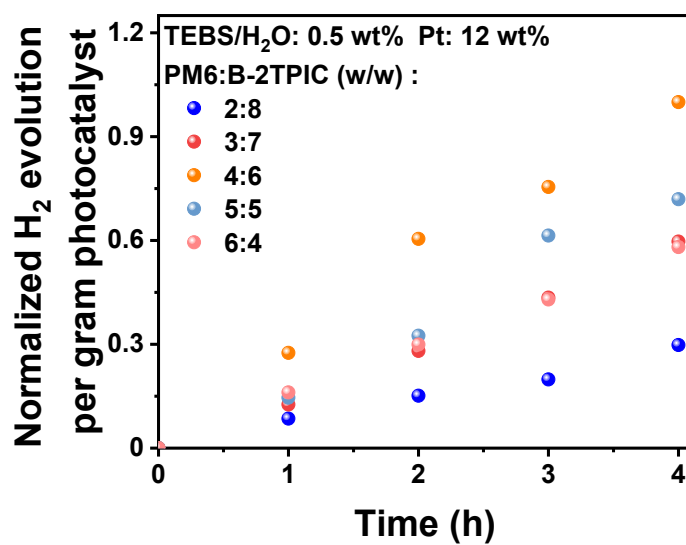
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2 **Fig. S12** Normalized hydrogen evolution per gram photocatalyst versus time of T-  
 3 2TPIC NPs at a concentration of  $6.67 \mu\text{g mL}^{-1}$  with 0.5 wt% TEBS/H<sub>2</sub>O and varying  
 4 Pt loadings at room temperature, measured by a multiple-channel photochemical  
 5 reaction system.

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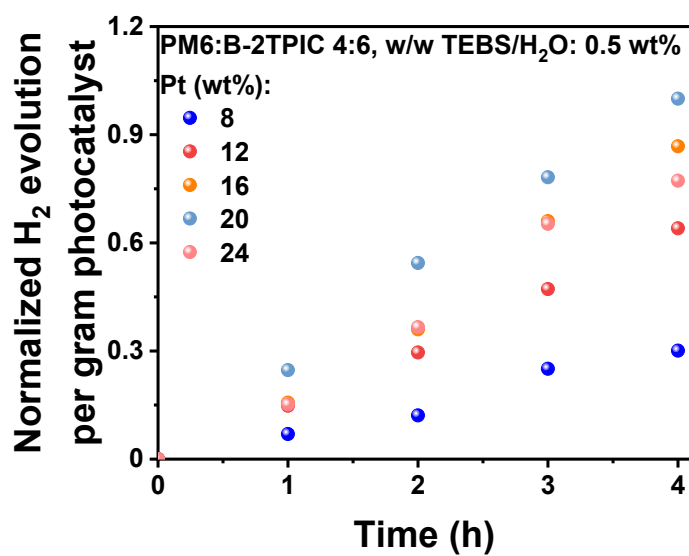
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2 **Fig. S13** Normalized hydrogen evolution per gram photocatalyst versus time of T-  
3 2TPIC NPs at a concentration of  $6.67 \mu\text{g mL}^{-1}$  with 16 wt% Pt loading at different  
4 weight ratios of TEBS/H<sub>2</sub>O at room temperature, measured by a multiple-channel  
5 photochemical reaction system.



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2 **Fig. S14** Normalized hydrogen evolution per gram photocatalyst versus time of  
 3 nanoparticles formed with different PM6:B-2TPIC weight ratios using 0.5 wt%  
 4 TEBS/H<sub>2</sub>O with 12 wt% Pt loading at a concentration of 6.67  $\mu\text{g mL}^{-1}$  at room  
 5 temperature, measured by a multiple-channel photochemical reaction system.

6

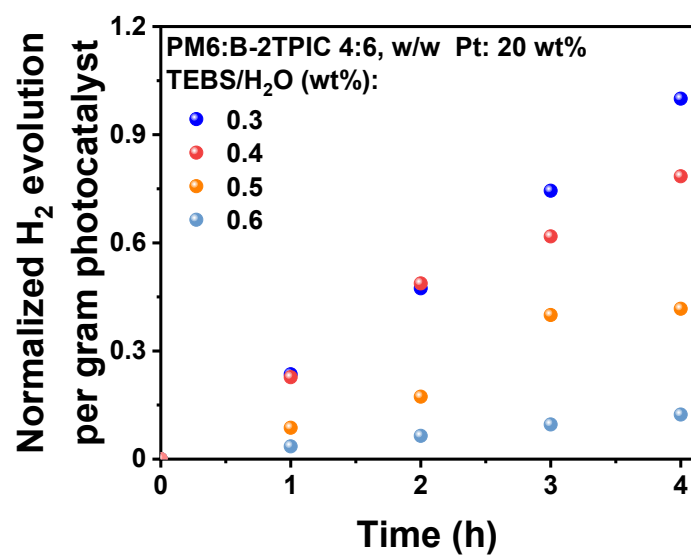


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2 **Fig. S15** Normalized hydrogen evolution per gram photocatalyst versus time of PM6:B-  
 3 2TPIC (4:6, w/w) NPs photocatalysts at a concentration of  $6.67 \mu\text{g mL}^{-1}$  with 0.5 wt%  
 4 TEBS/H<sub>2</sub>O and varying Pt loadings at room temperature, measured by a multiple-  
 5 channel photochemical reaction system.

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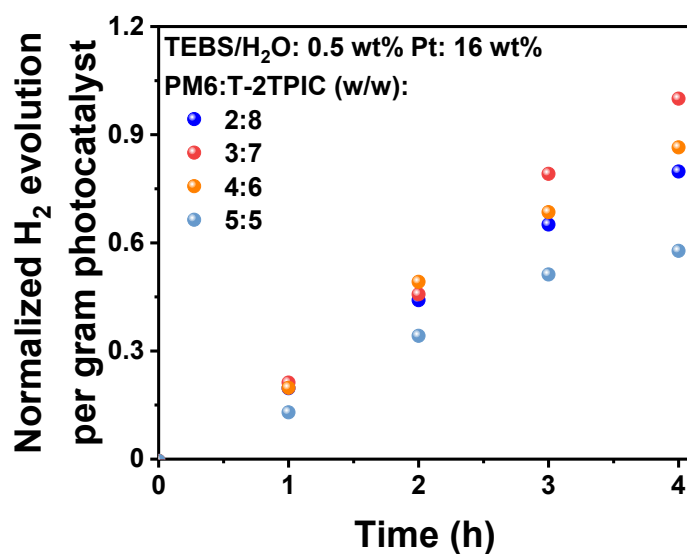
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2 **Fig. S16** Normalized hydrogen evolution per gram photocatalyst versus time of PM6:B-  
 3 2TPIC (4:6, w/w) NPs photocatalysts at a concentration of  $6.67 \mu\text{g mL}^{-1}$  with 20 wt%  
 4 Pt loading at different weight ratios of TEBS/H<sub>2</sub>O at room temperature, measured by a  
 5 multiple-channel photochemical reaction system.

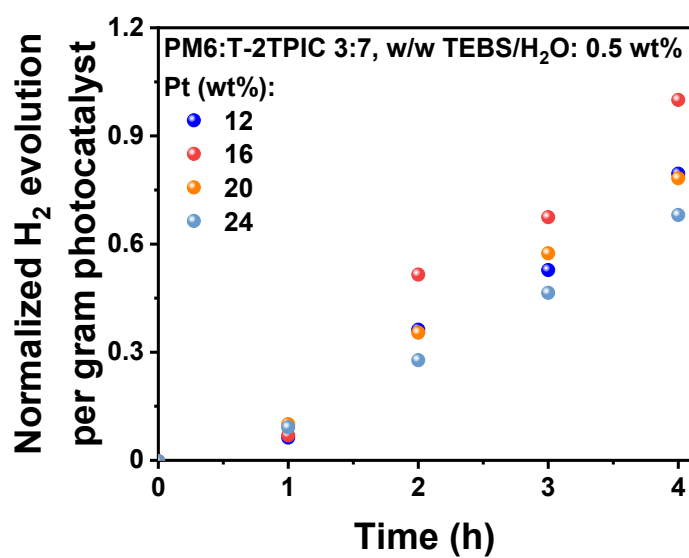
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2 **Fig. S17** Normalized hydrogen evolution per gram photocatalyst versus time of  
 3 nanoparticles formed with different PM6:T-2TPIC weight ratios using 0.5 wt%  
 4 TEBS/H<sub>2</sub>O with 16 wt% Pt loading at a concentration of 6.67  $\mu\text{g mL}^{-1}$  at room  
 5 temperature, measured by a multiple-channel photochemical reaction system.

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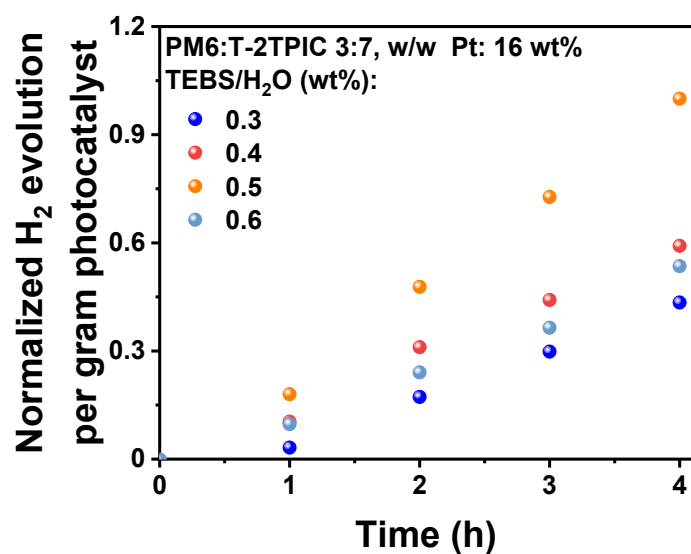


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2 **Fig. S18** Normalized hydrogen evolution per gram photocatalyst versus time of PM6:T-  
3 2TPIC (3:7, w/w) NPs photocatalysts at a concentration of  $6.67 \mu\text{g mL}^{-1}$  with 0.5 wt%  
4 TEBS/H<sub>2</sub>O and varying Pt loadings at room temperature, measured by a multiple-  
5 channel photochemical reaction system.

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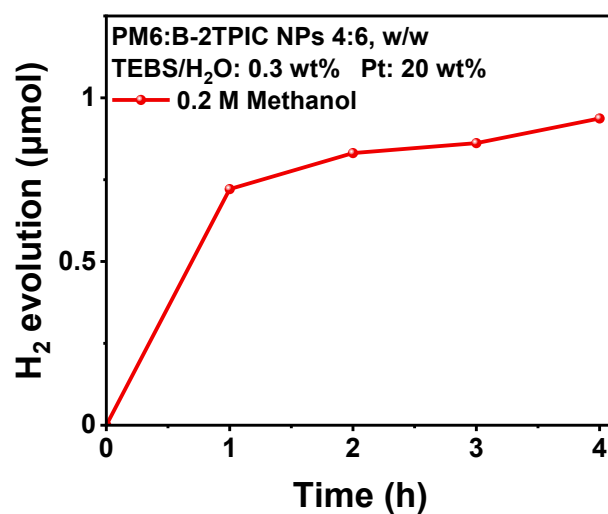


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2 **Fig. S19** Normalized hydrogen evolution per gram photocatalyst versus time of PM6:T-  
 3 2TPIC (3:7, w/w) NPs photocatalysts at a concentration of  $6.67 \mu\text{g mL}^{-1}$  with 16 wt%  
 4 Pt loading at different weight ratios of TEBS/H<sub>2</sub>O at room temperature, measured by a  
 5 multiple-channel photochemical reaction system.

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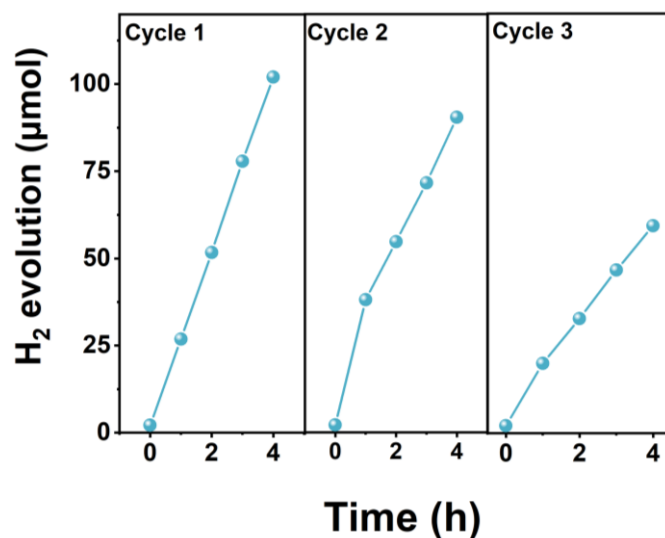
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2 **Fig. S20** Hydrogen evolution versus time of PM6:B-2TPIC NPs (D:A ratio: 4:6, w/w,  
3 0.3 wt% TEBS/H<sub>2</sub>O weight ratio and 20 wt% Pt loading) using methanol as the  
4 sacrificial agent at a concentration of 6.67 μg mL<sup>-1</sup>, under AM 1.5G, 100 mW cm<sup>-2</sup>.

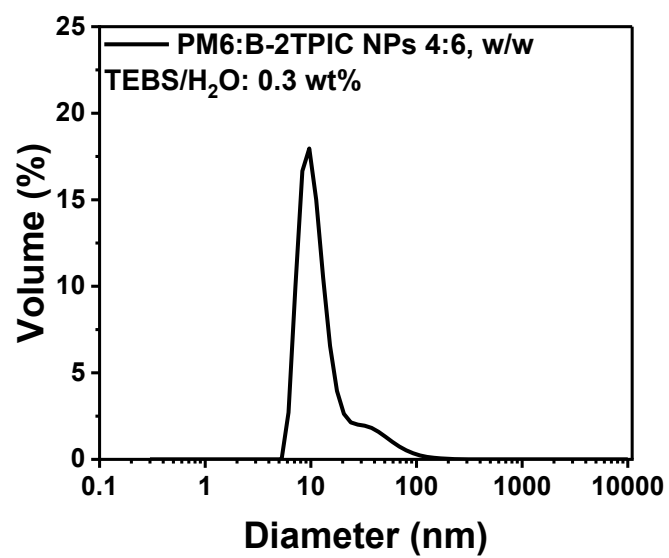
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2 **Fig. S21** Cycling stability of PM6:B-2TPIC NPs (D:A ratio: 4:6, w/w, 0.3 wt%  
 3 TEBS/H<sub>2</sub>O weight ratio and 20 wt% Pt loading) in 0.2 M AA at a concentration of 26.67  
 4 μg mL<sup>-1</sup>, under AM 1.5G, 100 mW cm<sup>-2</sup>. For recycling experiments, the equivalent  
 5 amount of ascorbic acid was added after the end of every cycle, according to the  
 6 consumption of ascorbic acid.

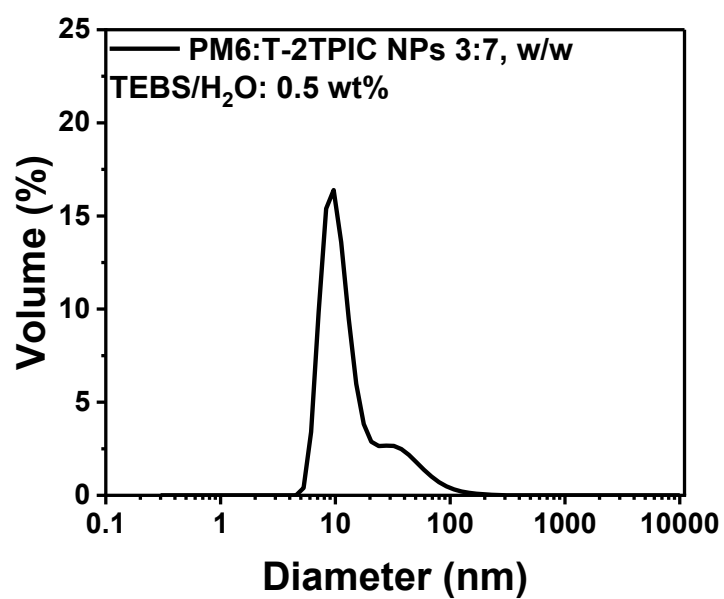
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2 **Fig. S22** DLS size distributions by volume of PM6:B-2TPIC (4:6, w/w) NPs using 0.3  
3 wt% TEBS/H<sub>2</sub>O.

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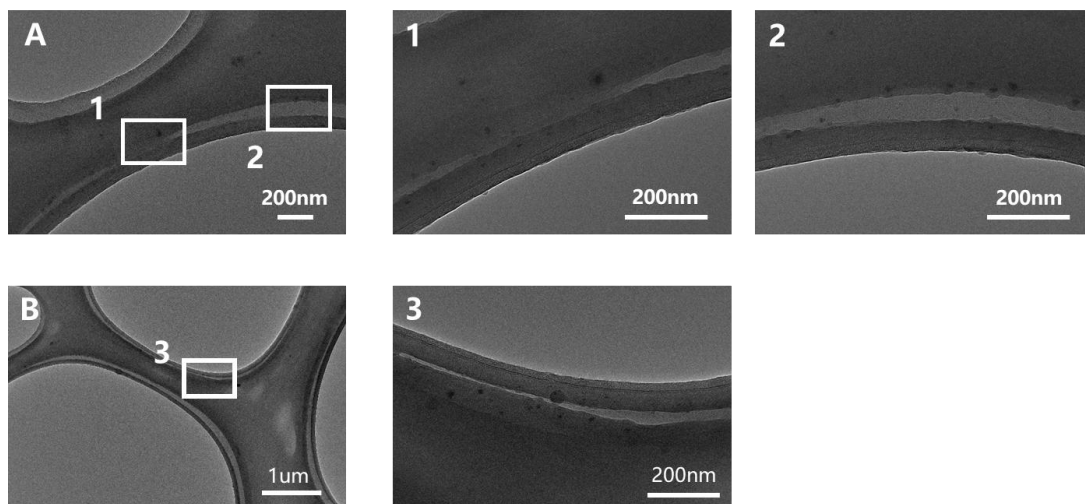


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2 **Fig. S23** DLS size distributions by volume of PM6:T-2TPIC (3:7, w/w) NPs using 0.5  
3 wt% TEBS/H<sub>2</sub>O.

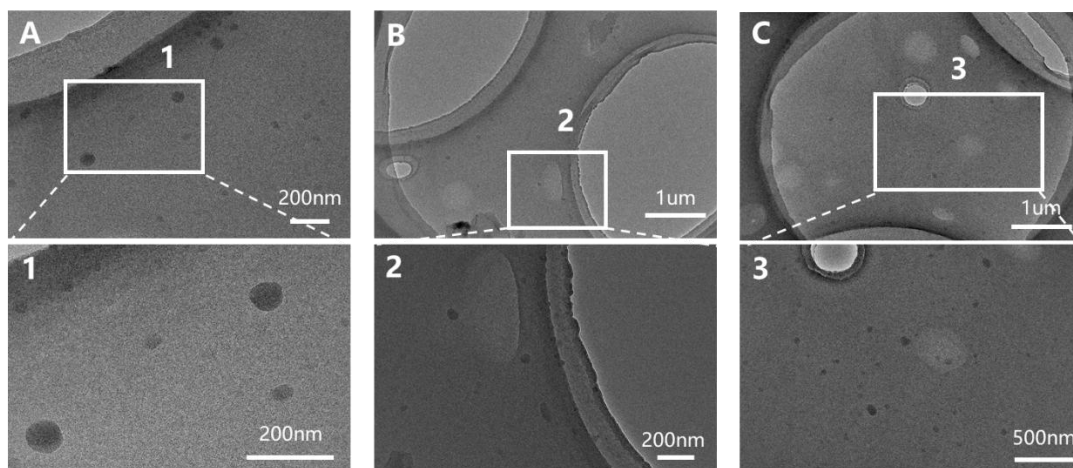
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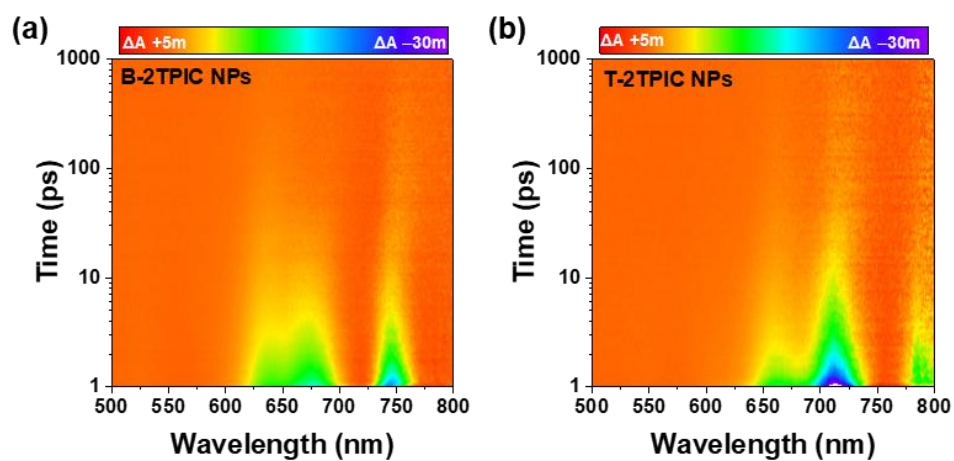
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**Fig. S24** TEM images of PM6:B-2TPIC (4:6, w/w) NPs using 0.3 wt% TEBS/H<sub>2</sub>O.



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**Fig. S25** TEM images of PM6:T-2TPIC (3:7, w/w) NPs using 0.5 wt% TEBS/H<sub>2</sub>O.

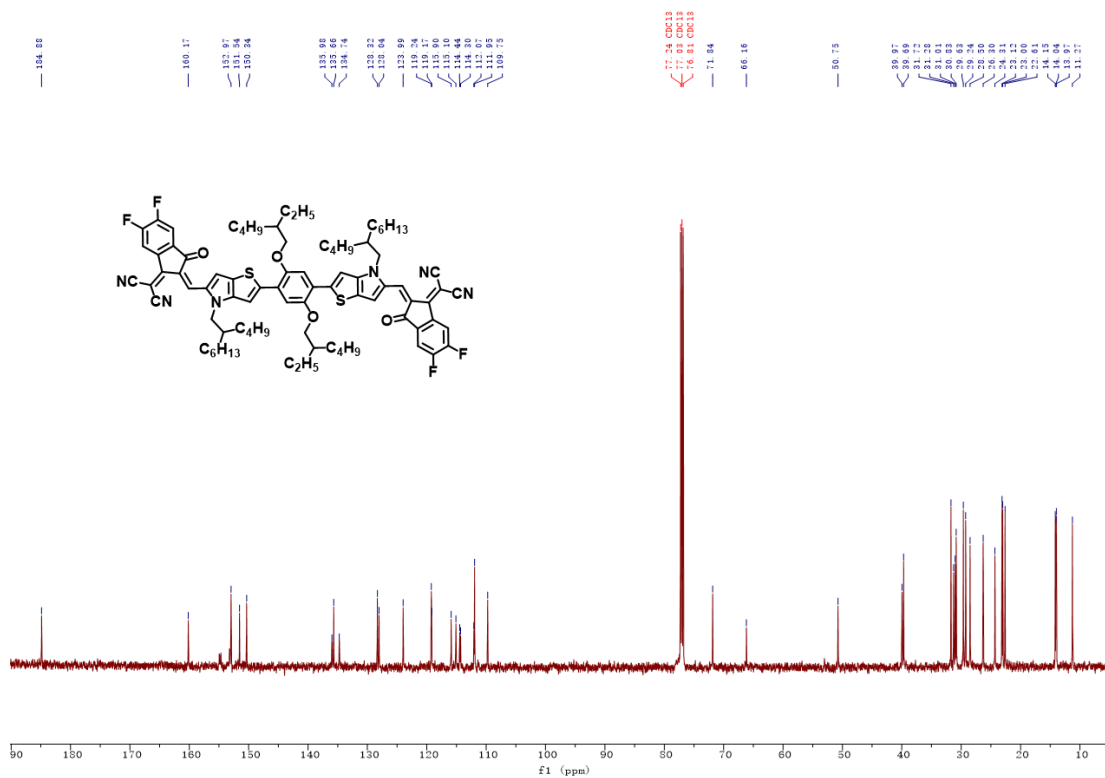


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2 **Fig. S26** The 2D transient absorption contours in the 500–800 nm range of (a) B-2TPIC

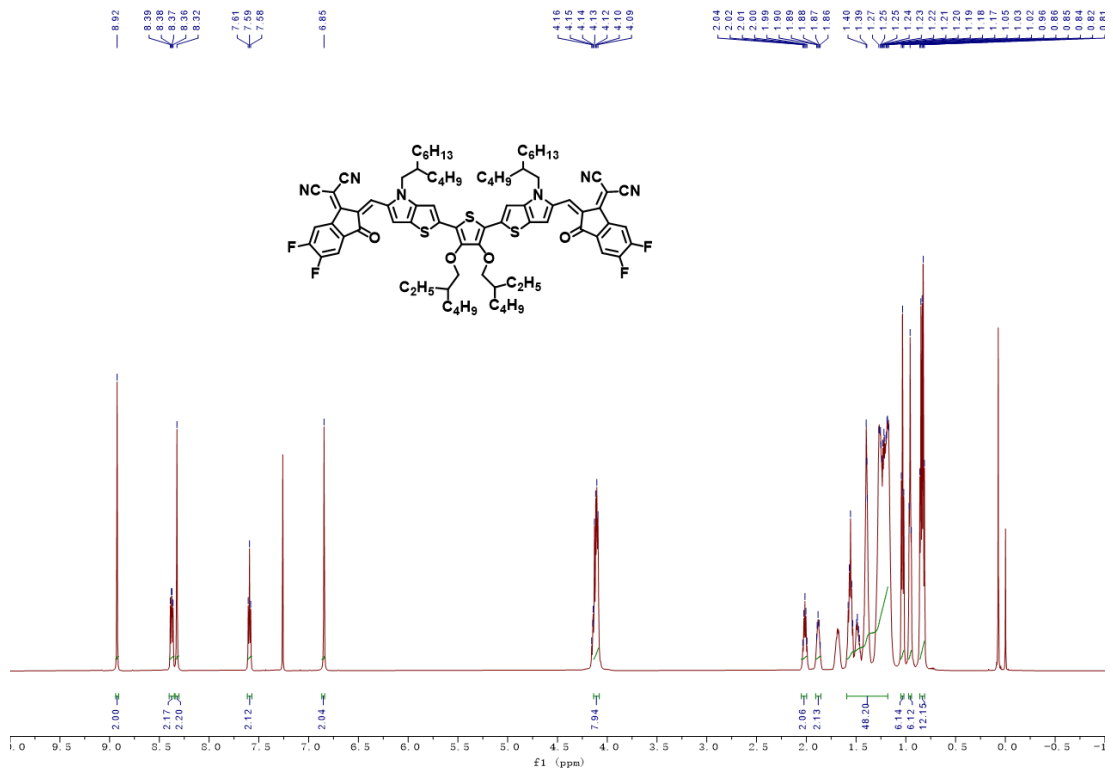
3 and (b) T-2TPIC NPs. Excited at 750 nm.





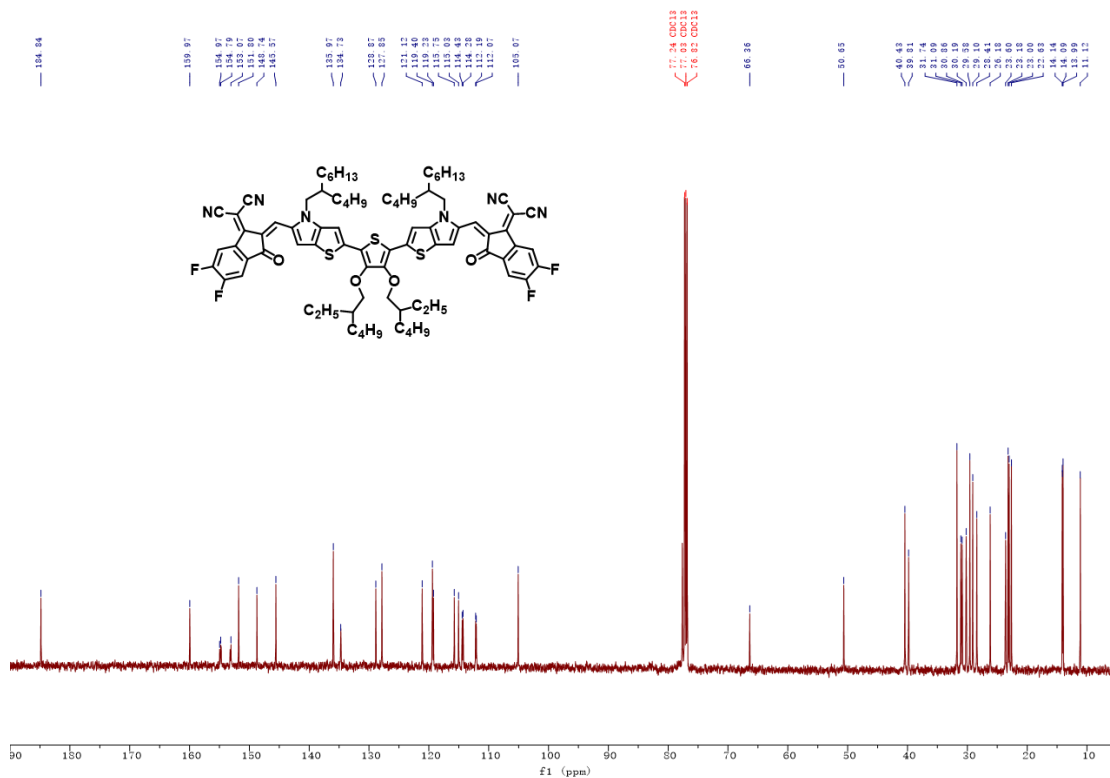
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**Fig. S28** <sup>13</sup>C NMR Spectrum of B-2TPIC.



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**Fig. S29** <sup>1</sup>H NMR Spectrum of T-2TPIC.



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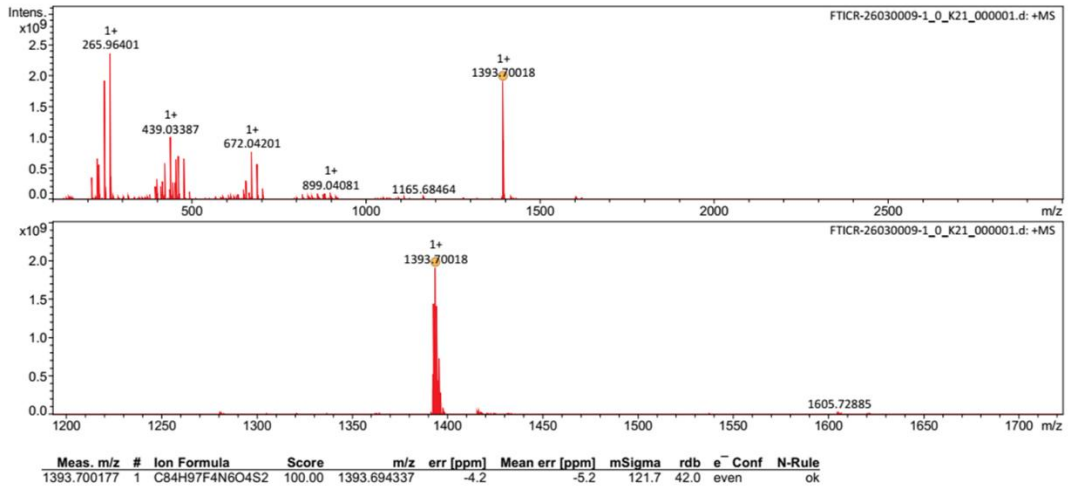
**Fig. S30**  $^{13}\text{C}$  NMR Spectrum of T-2TPIC.

# Peking University Mass Spectrometry Sample Analysis Report

## Analysis Info

Analysis Name FTICR-26030009-1\_0\_K21\_000001.d  
Sample B-2TPIC  
Comment MALDI, matrix CHCA

Acquisition Date 3/13/2026 11:50:41 AM  
Instrument Bruker Solarix XR FTMS  
Operator Peking University



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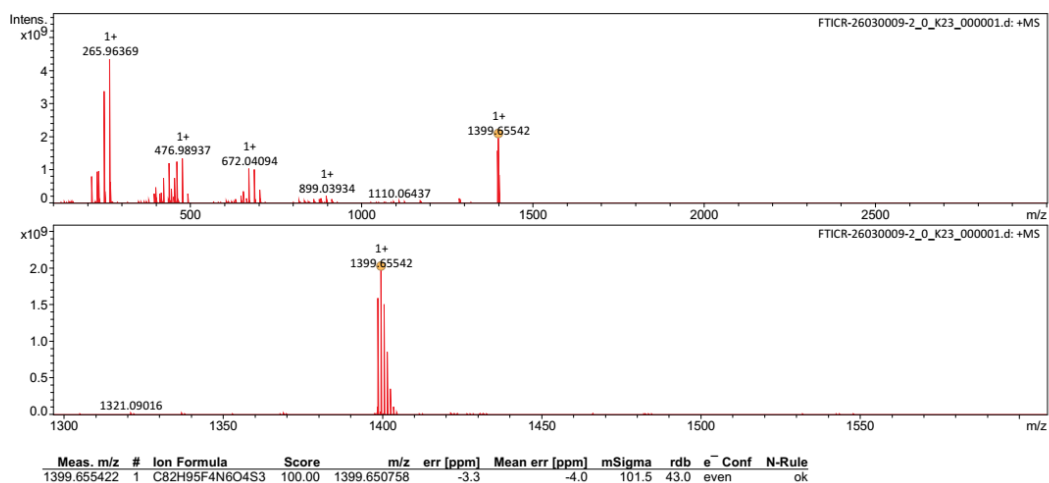
**Fig. S31** HR mass spectrum of B-2TPIC using the MALDI-TOF mode.

## Peking University Mass Spectrometry Sample Analysis Report

### Analysis Info

Analysis Name FTICR-26030009-2\_0\_K23\_000001.d  
 Sample T-2TPIC  
 Comment MALDI, matrix CHCA

Acquisition Date 3/13/2026 11:56:18 AM  
 Instrument Bruker Solarix XR FTMS  
 Operator Peking University



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**Fig. S32** HR mass spectrum of T-2TPIC using the MALDI-TOF mode.

1 **Table S1** Mulliken atomic charge population analysis of T-IC2F

atom	charge (e)	atom	charge (e)
C1	-0.135	O16	-0.486
C2	-0.123	C17	0.067
S3	0.371	F18	-0.267
C4	-0.346	F19	-0.267
C5	-0.098	C20	0.304
C6	-0.210	N21	-0.499
C7	0.042	C22	0.306
C8	0.098	N23	-0.501
C9	0.066	H24	0.175
C10	0.061	H25	0.190
C11	0.353	H26	0.160
C12	-0.274	H27	0.195
C13	0.339	H28	0.209
C14	0.335	H29	0.189
C15	-0.254		

1 **Table S2** Mulliken atomic charge population analysis of TT-IC2F

atom	charge (e)	atom	charge (e)
C1	-0.217	C17	0.066
C2	-0.132	F18	-0.268
C3	-0.123	F19	-0.267
S4	0.356	C20	-0.114
C5	-0.142	C21	-0.332
C6	-0.128	S22	0.285
C7	0.041	C23	0.303
C8	0.098	N24	-0.501
C9	0.066	C25	0.306
C10	0.062	N26	-0.504
C11	0.354	H27	0.185
C12	-0.274	H28	0.193
C13	0.338	H29	0.209
C14	0.334	H30	0.188
C15	-0.255	H31	0.169
O16	-0.487	H32	0.192

1 **Table S3** Mulliken atomic charge population analysis of TP-IC2F

atom	charge (e)	atom	charge (e)
C1	-0.267	C19	0.300
C2	0.344	N20	-0.506
C3	-0.161	C21	0.313
C4	0.347	N22	-0.522
C5	0.055	H23	0.158
C6	0.083	H24	0.208
C7	0.071	H25	0.185
C8	0.055	H26	0.188
C9	0.354	C27	-0.226
C10	-0.277	H28	0.221
C11	0.336	S29	0.270
C12	0.333	C30	-0.094
C13	-0.258	H31	0.160
O14	-0.498	N32	-0.576
C15	0.061	C33	-0.367
F16	-0.270	H34	0.183
F17	-0.269	H35	0.183
C18	-0.350	H36	0.235

1 **Table S4** Mulliken atomic charge population analysis of PT-IC2F

atom	charge (e)	atom	charge (e)
C1	-0.142	F19	-0.270
C2	-0.132	N20	-0.494
S3	0.332	C21	-0.330
C4	-0.135	C22	0.300
C5	0.304	N23	-0.506
C6	-0.221	C24	0.304
C7	0.038	N25	-0.512
C8	0.096	H26	0.175
C9	0.065	H27	0.189
C10	0.062	H28	0.207
C11	0.351	H29	0.186
C12	-0.276	H30	0.178
C13	0.336	H31	0.185
C14	0.332	H32	0.185
C15	-0.257	C33	-0.238
O16	-0.489	H34	0.155
C17	0.059	C35	0.067
F18	-0.271	H36	0.165

1 **Table S5** Summary of the GIWAXS parameters for the in-plane and out-of-plane  
 2 directions of the neat films.

acceptor	in-plane				out-of-plane			
	$q$ ( $\text{\AA}^{-1}$ )	FWHM	$d$ ( $\text{\AA}$ )	CL ( $\text{\AA}$ )	$q$ ( $\text{\AA}^{-1}$ )	FWHM	$d$ ( $\text{\AA}$ )	CL ( $\text{\AA}$ )
B-2TPIC	0.357	0.062	17.6	91.2	1.78	0.198	3.53	28.6
T-2TPIC	0.344	0.088	18.3	64.3	1.80	0.270	3.49	20.9

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1 **Table S6** Hydrogen evolution performance of advanced organic photocatalytic  
 2 nanoparticles.

photocatalyst (mass)	sacrificial agent (solvent volume)	Pt (wt%)	illumination	HER (mmol h <sup>-1</sup> g <sup>-1</sup> )	ref.
PTB7-Th:EH-IDTBR (2 mg)	0.2 M AA (20 mL)	10	Xe lamp 350- 800 nm	64	S9
PM6:Y6 (1 mg)	0.2 M AA (12 mL)	10	AM1.5G 100 mW cm <sup>-2</sup>	44	S10
PM6:PC <sub>71</sub> BM (2 mg)	0.2 M AA (12 mL)	5	AM1.5G 100 mW cm <sup>-2</sup>	75	S10
PM6:TPP (2 mg)	0.2 M AA (30 mL)	20	AM1.5G 100 mW cm <sup>-2</sup>	64	S11
PM6:Y6CO (0.2 mg)	0.2 M AA (7.5 mL)	28	AM1.5G 100 mW cm <sup>-2</sup>	183	S12
PM6:2FBP-4F (0.18 mg)	0.2 M AA (7.5 mL)	24	AM1.5G 100 mW cm <sup>-2</sup>	208	S13
PM6:B-2TPIC (0.2 mg)	0.2 M AA (7.5 mL)	20	AM1.5G 100 mW cm <sup>-2</sup>	152.8	this work

3

1 **Table S7** The experimental details used for the EQE measurements.

nanoparticle	wavelength (nm)	radiant power (mW cm <sup>-2</sup> )	radiation area (cm <sup>2</sup> )	time (s)	$n_h$ (10 <sup>-6</sup> mol)	EQE (%)
PM6:B-2TPIC	400	1.89	5.786	3600	1.67	2.5
	500	2.10	5.786	3600	3.13	3.4
	600	2.67	5.786	3600	5.73	4.1
	700	3.08	5.786	3600	3.98	2.1
	808	3.01	5.786	3600	3.62	1.7
PM6:T-2TPIC	400	1.89	5.786	3600	1.12	1.7
	500	2.10	5.786	3600	1.93	2.1
	600	2.67	5.786	3600	5.42	3.9
	700	3.08	5.786	3600	4.68	2.5
	808	3.01	5.786	3600	4.47	2.1

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