

SUPPORTING INFORMATION

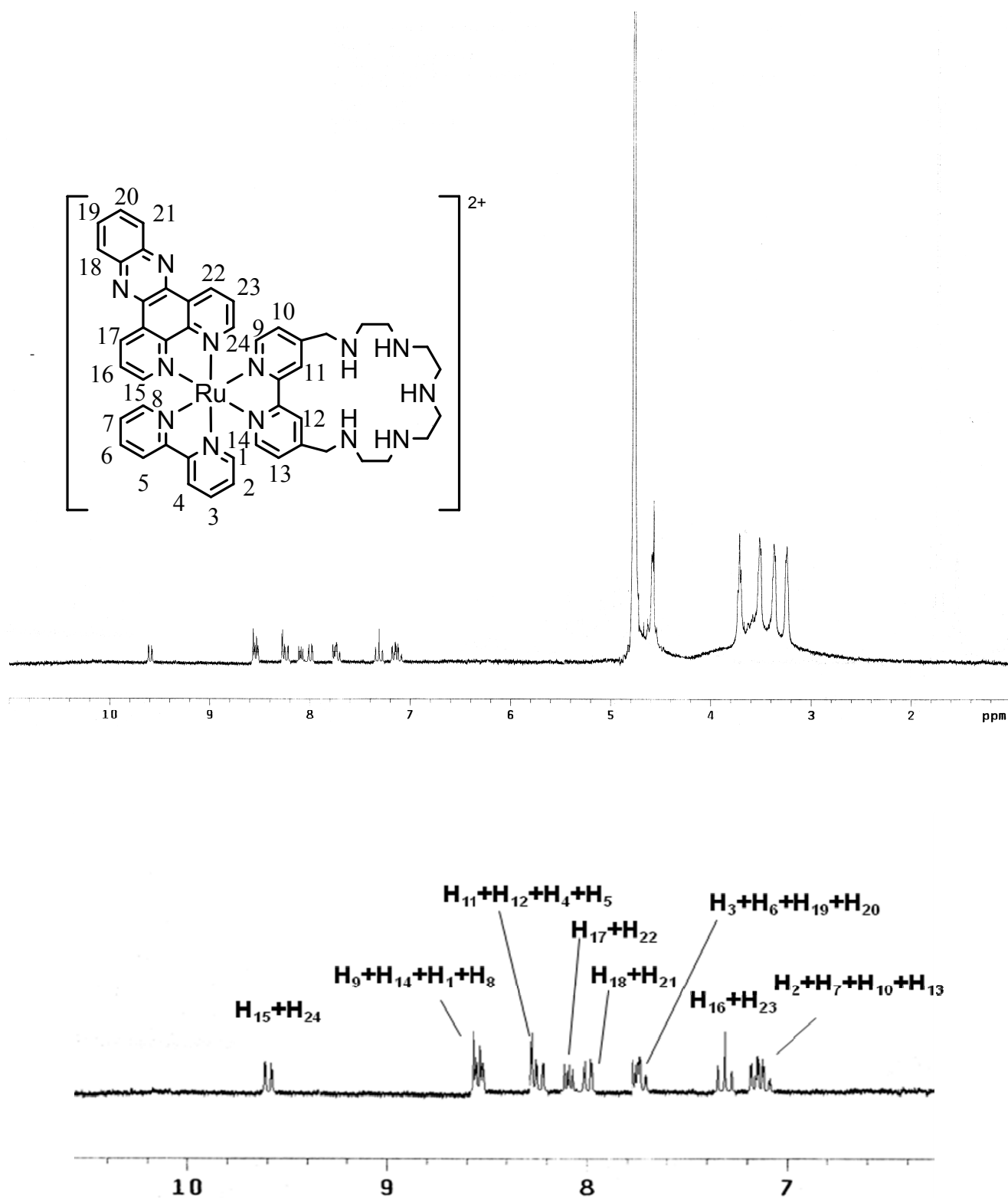


Figure S1. ^1H NMR spectrum of $D2^{2+}$ recorded in DMSO at 400 MHz

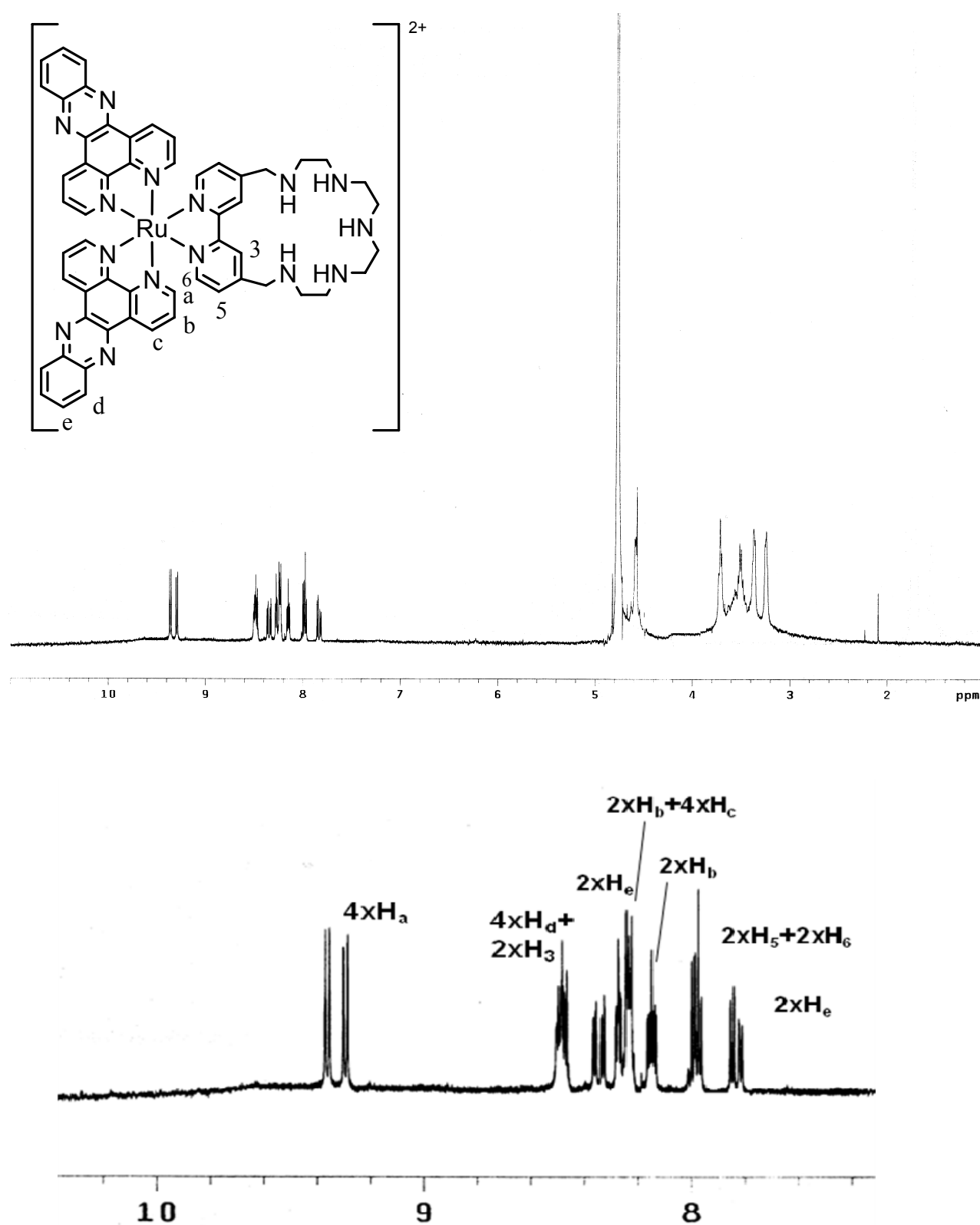


Figure S2. 1H NMR spectrum of $D3^{2+}$ recorded in DMSO at 400 MHz

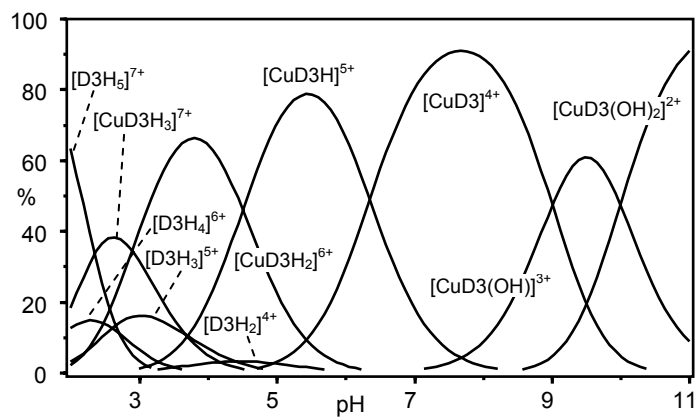


Figure S3. Distribution diagram of the Cu(II) complexes formed by $D3^{2+}$ in water. $[D3^{2+}] = [Cu(II)] = 1 \times 10^{-3}$.

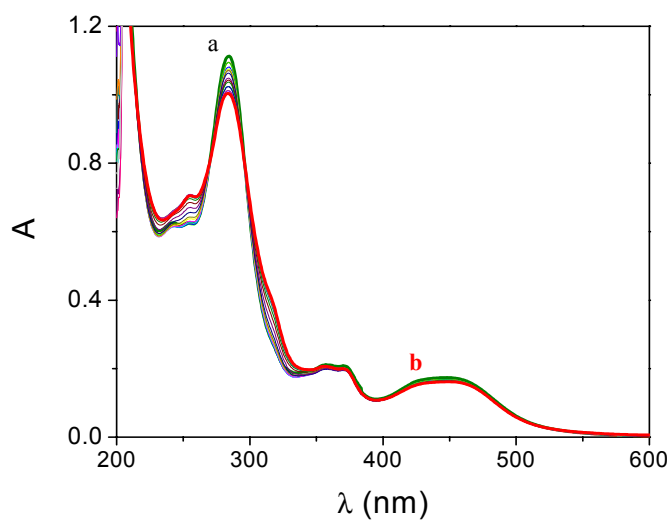


Figure S4. UV-Vis spectra for the $HD2^{3+}$ titration by copper(II). $C_{D2} = 1 \times 10^{-5}$ M, $I = 0.10$ M, $pH = 7.0$, $T = 25^\circ C$, $C_{Cu} = 0$ M (a), 1.1×10^{-4} M (b).

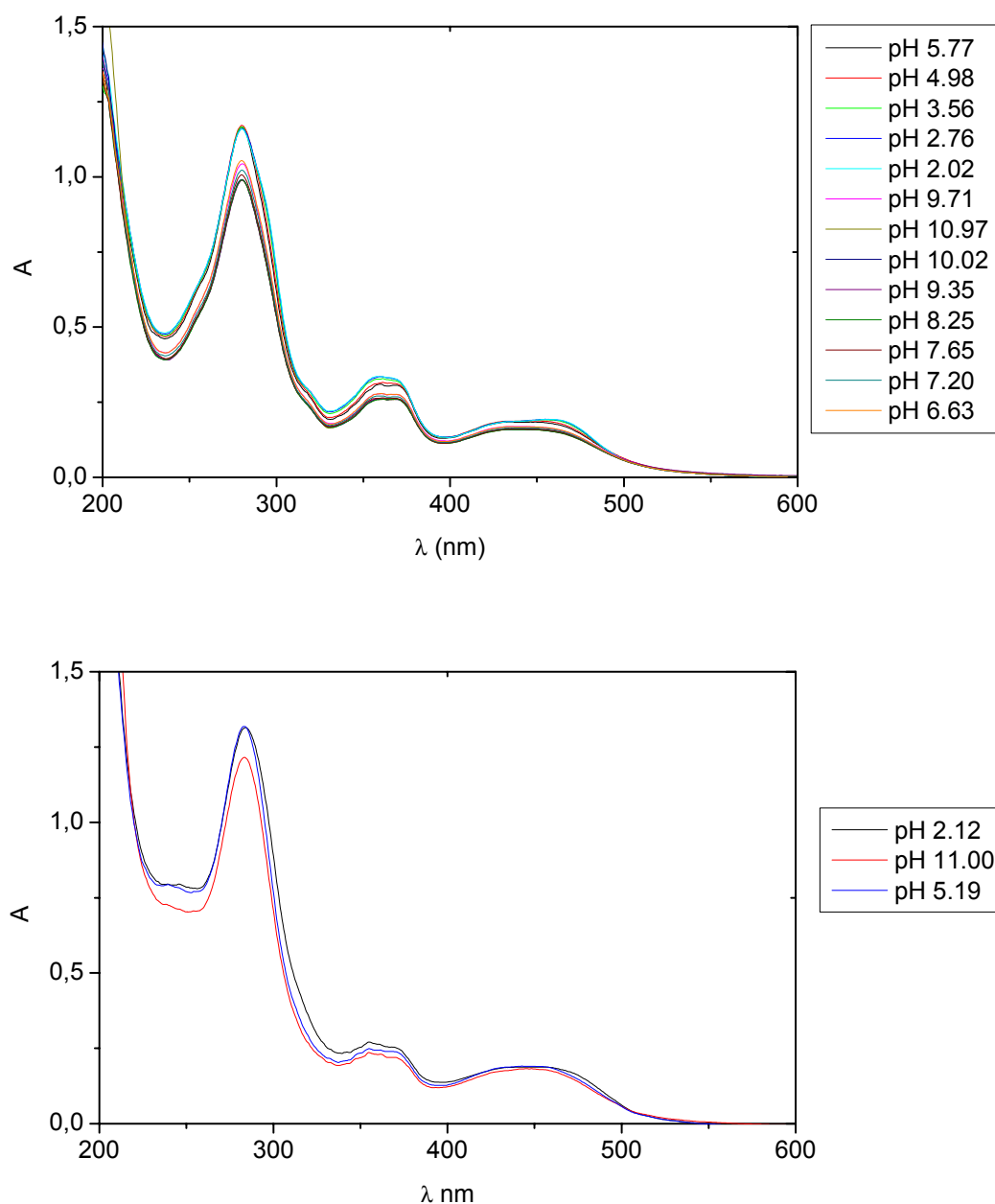


Figure S5. UV-vis absorption spectra of D3²⁺ (a) and D2²⁺ (b) recorded in water at different pH values and 25°C.

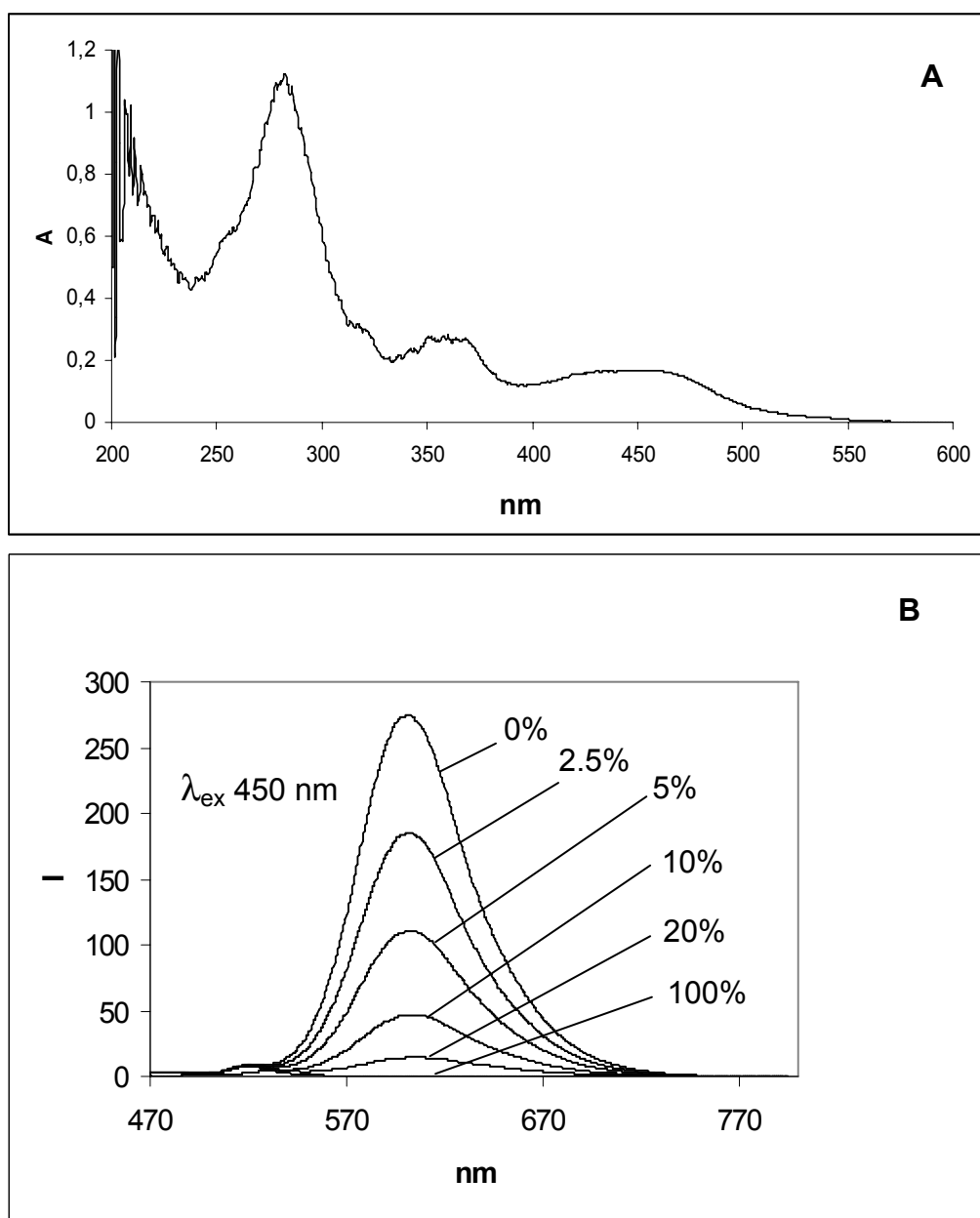


Figure S6. UV-Vis spectrum of $D3^{2+}$ in ethanol (A). Fluorescence emission spectra of $D3^{2+}$ in ethanol, water and ethanol/water mixtures (B). Percentages (v:v) are referred to water in ethanol. $[D3^{2+}] = 1 \times 10^{-5} M$, $T = 25^\circ C$.

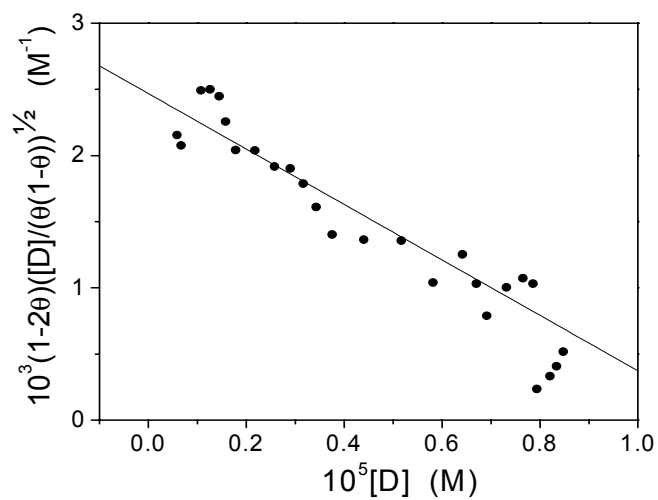


Figure S7. Analysis of fluorescence titration data for the DNA/CuD₂⁴⁺ system to equation (6).
I = 0.10 M, pH = 7.0, T = 25°C.

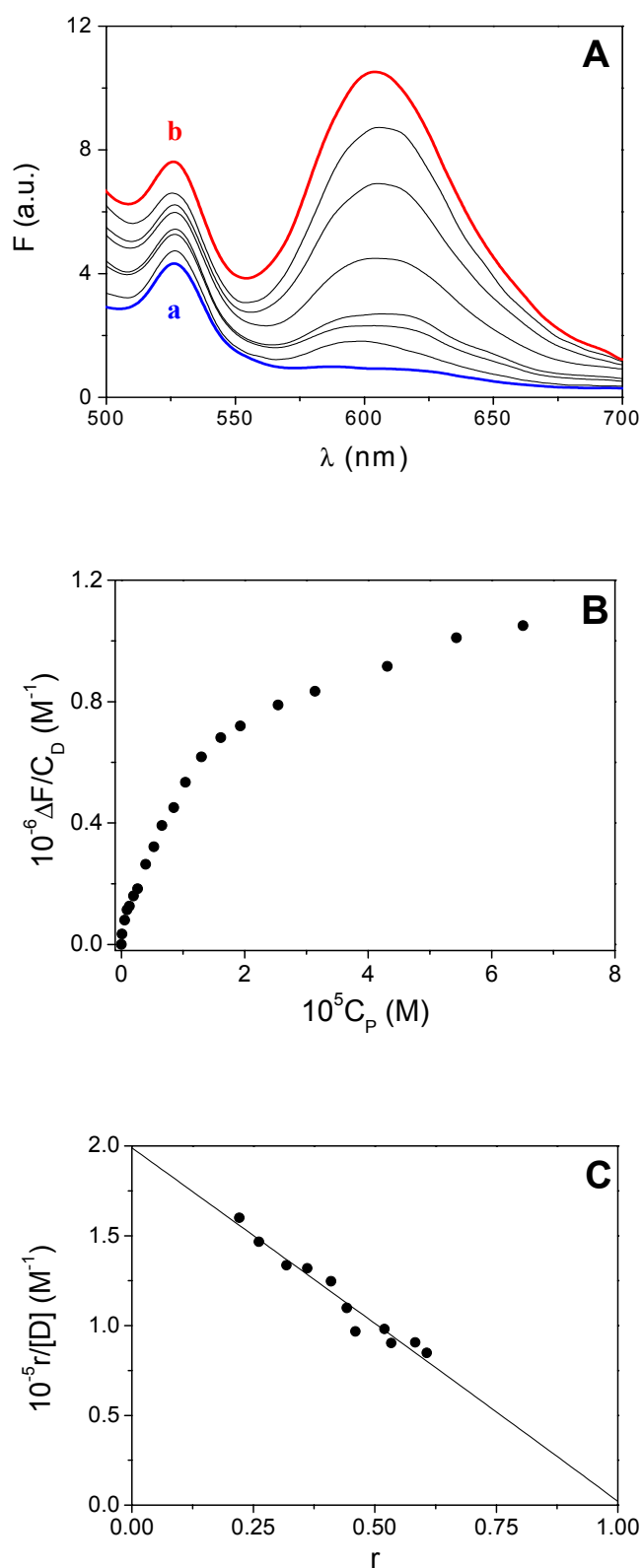


Figure S8. Spectrofluorometric titration of the DNA/HD3³⁺ system (A), relevant binding isotherm at λ_{em} = 600nm (B) and analysis of the fluorescence titration data to equation (5) (C). C_D = 8.8×10⁻⁶ M, C_P from 0 (a) to 6.5×10⁻⁵ M (b), λ_{ex} = 450nm, I = 0.10 M, pH = 7.0, T = 25°C.

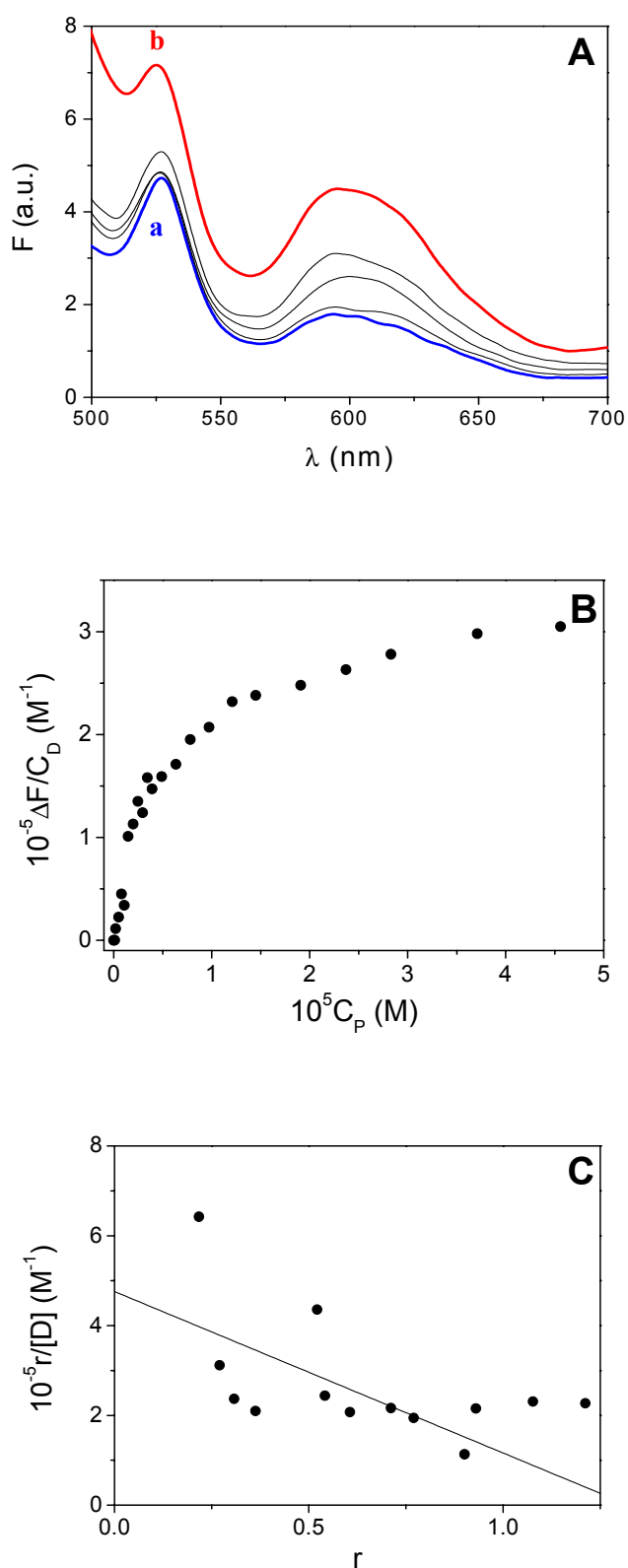


Figure S9. Spectrofluorometric titration of the DNA/CuD³⁺ system (A), relevant binding isotherm at $\lambda_{em} = 600\text{nm}$ (B) and analysis of the fluorescence titration data to equation (5) (C). $C_D = 8.8 \times 10^{-6}$ M, C_P from 0 (**a**) to 4.6×10^{-5} M (**b**), $\lambda_{ex} = 450\text{nm}$, $I = 0.10$ M, $\text{pH} = 7.0$, $T = 25^\circ\text{C}$.

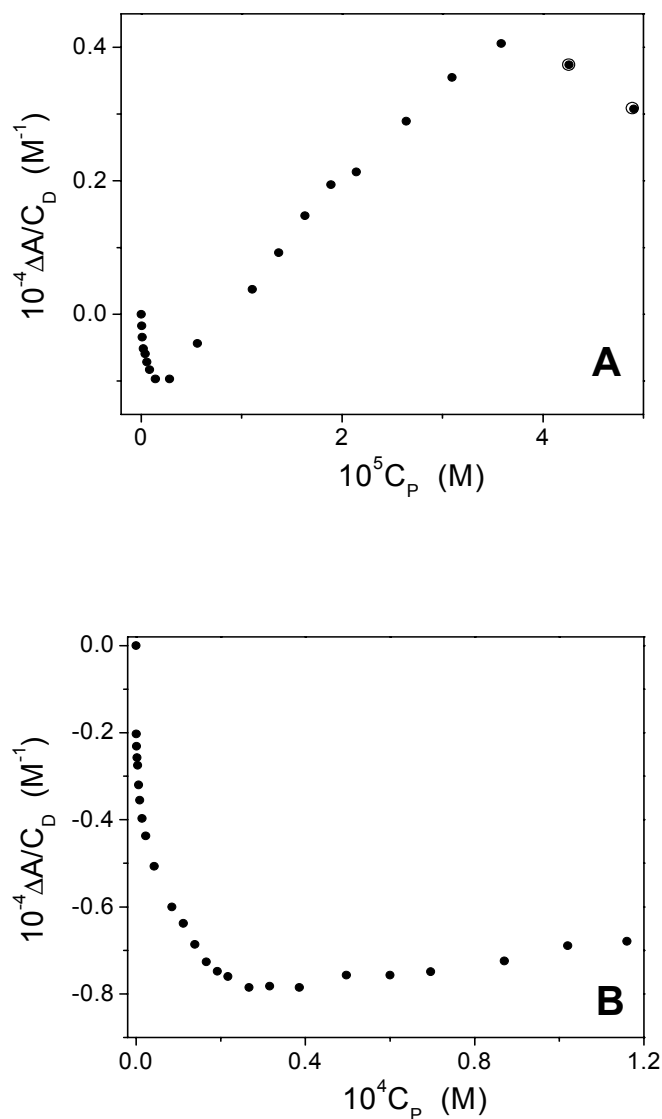


Figure S10. Binding isotherms from spectrophotometric titrations of the DNA/HD2³⁺ (A) and DNA/HD3³⁺ (B) systems under low added salt conditions; $C_D = 1.1 \times 10^{-5} \text{ M}$, $I = 0.003 \text{ M}$, $\text{pH} = 7.0$, $\lambda = 360 \text{ nm}$, $T = 25^\circ\text{C}$. Circled points in (A) are related to precipitation phenomena.

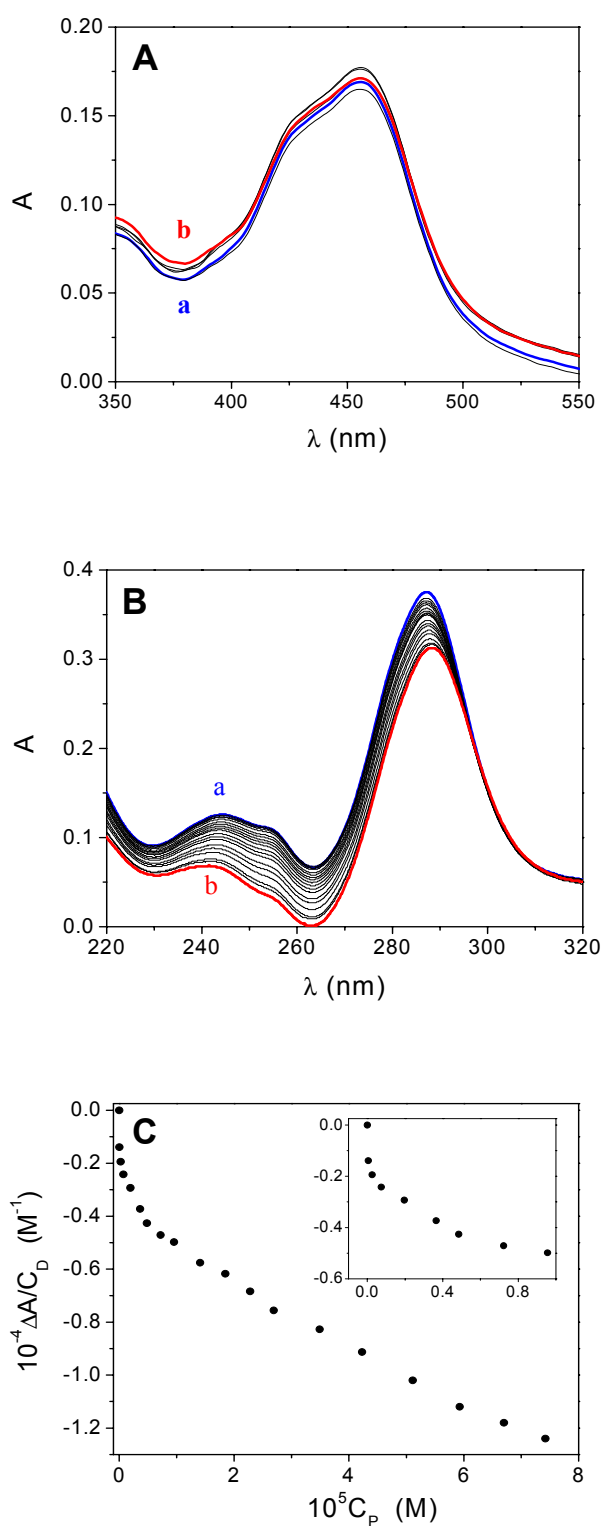


Figure S11. Spectrophotometric titrations of the DNA/H₂D1⁴⁺ system; I = 0.10 M, pH = 7.0, T = 25°C. (A) Visible range: C_D = 1.2×10⁻⁵ M, C_P from 0 (a) to 1.0×10⁻⁴ M (b). (B) UV range: C_D = 5.0×10⁻⁶ M, C_P from 0 M (a) to 7.4×10⁻⁵ M (b), differential titration (DNA added in both sample and reference cells). (C) Binding isotherm at 288nm from titration (B); the inset is an enlargement of the first part of the plot.

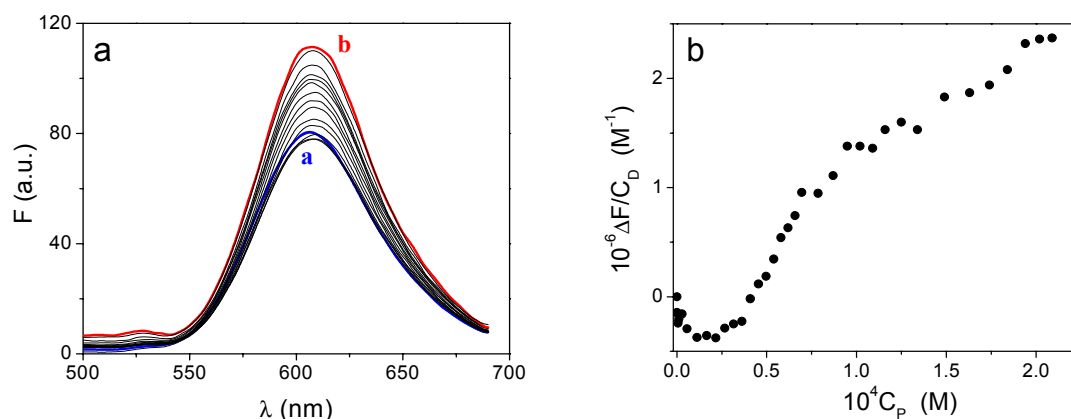


Figure S12. Fluorescence spectra for the DNA/ H_2D1^{4+} system recorded during titration (a) and relevant binding isotherm at 607nm (b). $C_D = 1.2 \times 10^{-5}$ M, $I = 0.10$ M, $pH = 7.0$, $T = 25^\circ C$, $\lambda_{ex} = 450$ nm, $C_P = 0$ M (a), 2.1×10^{-4} M (b).

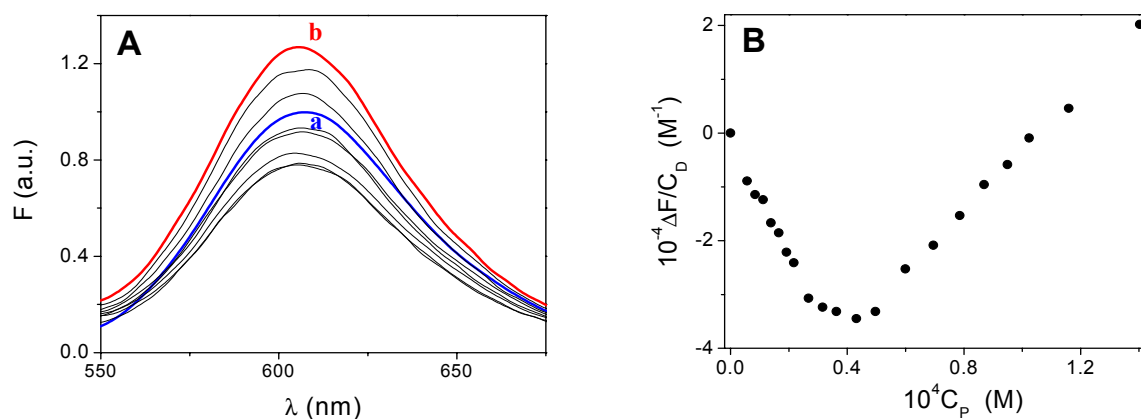


Figure S13. Spectrofluorometric titration of the DNA/ $CuD1^{4+}$ system; $I = 0.10$ M, $pH = 7.0$, $\lambda_{ex} = 450$ nm, $T = 25^\circ C$. (A) Collected spectra $C_D = 1.3 \times 10^{-5}$ M, C_P from 0 (a) to 1.4×10^{-4} M (b). (B) Binding isotherm at $\lambda_{em} = 607$ nm.

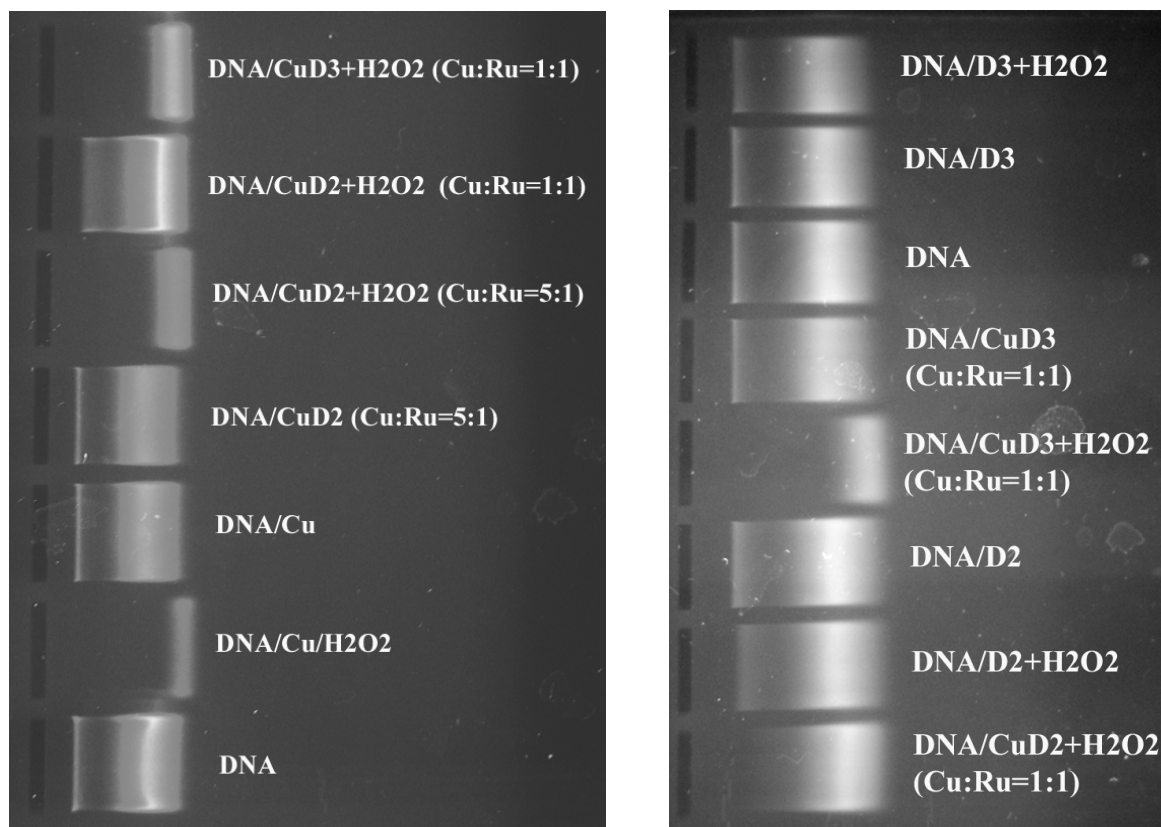


Figure S14. Electrophoresis experiments on the DNA/HD2³⁺, DNA/CuD2⁴⁺, DNA/HD3²⁺ and DNA/CuD3⁴⁺ systems, alone or in the presence of H₂O₂ (1.8×10⁻³M). C_{DNA} = 2.5×10⁻⁴M, Tris-acetate-EDTA buffer (pH = 7.4), 1% agarose gel with 1×10⁻⁶M Ethidium staining, 65V, 50mA.

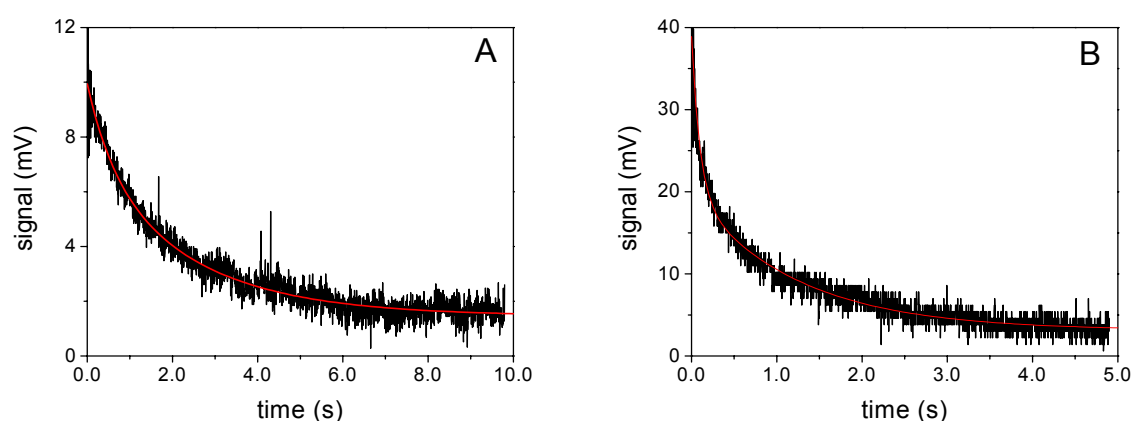


Figure S15. Stopped-flow curves for the DNA/HD2³⁺ system (A) and for the DNA/HD3²⁺ system (B) at I= 0.10 M, pH = 7.0, λ = 360nm, T = 25°C; the line is least-square bi-exponential data fitting. (A) C_D = 9.4×10⁻⁶ M, C_P = 9.5×10⁻⁵ M; (B) C_D = 1.0×10⁻⁵ M, C_P = 2.0×10⁻⁴ M.

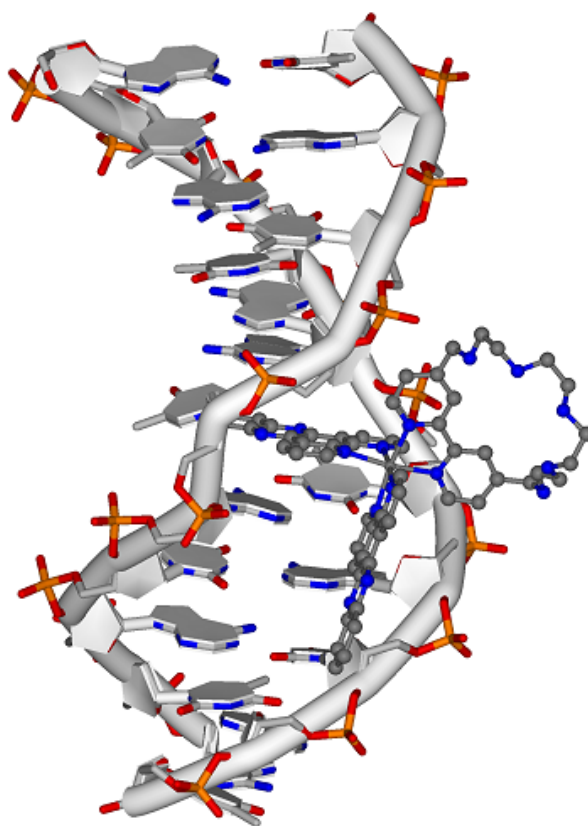


Figure S16. Calculated conformation for the ATATATATAT/ Δ -HD3³⁺ adduct.

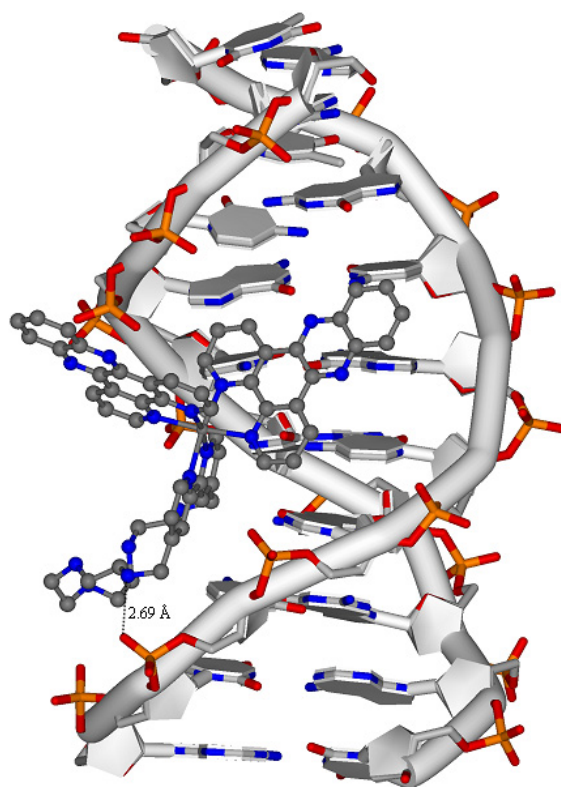


Figure S17. Calculated conformation for the ATCGCGCGAT/ Δ -HD3³⁺ adduct.

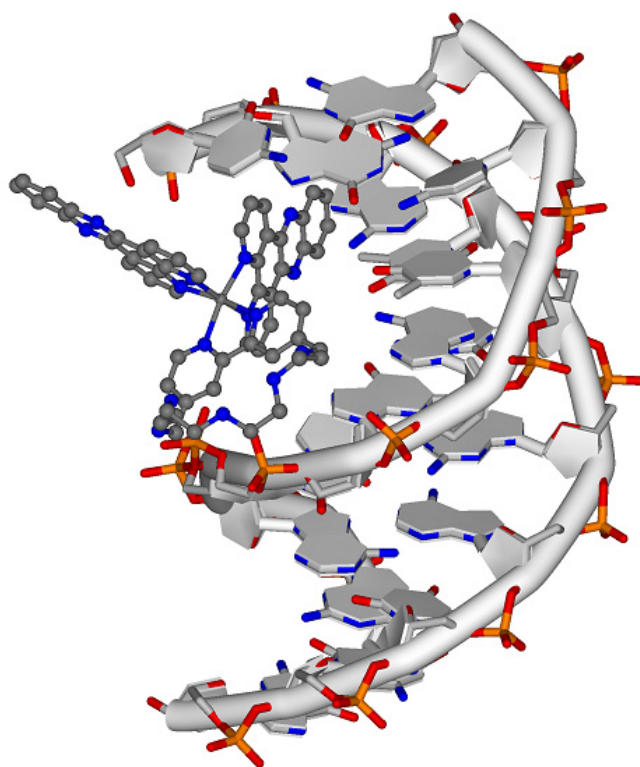


Figure S18. Calculated conformation for the CGATCGATCG/ Δ -HD3³⁺ adduct.

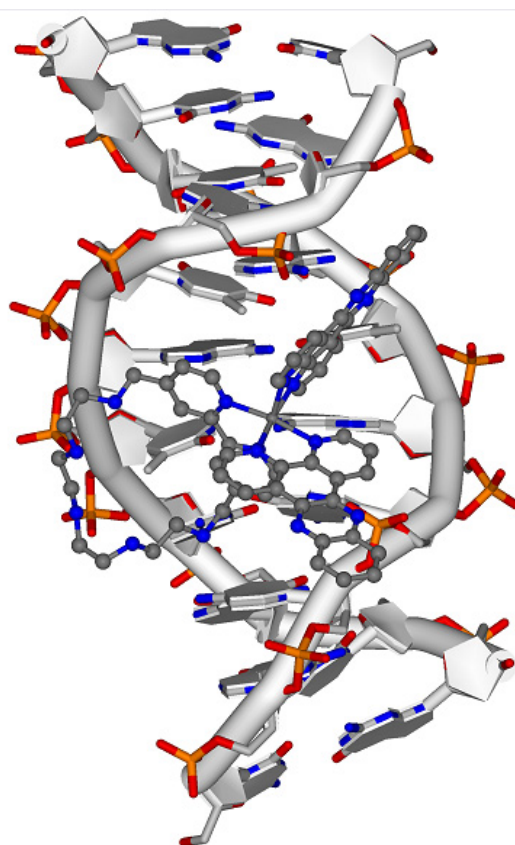


Figure S19. Calculated conformation for the CGCGATATCG/ Δ -HD3³⁺ adduct.

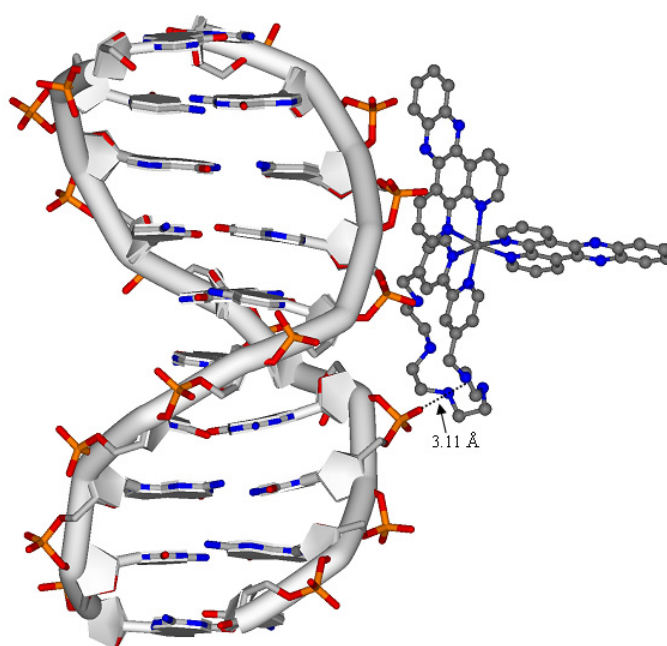


Figure S20. Calculated conformation for the GCGCGCGCGC/ Δ -HD3³⁺ adduct.

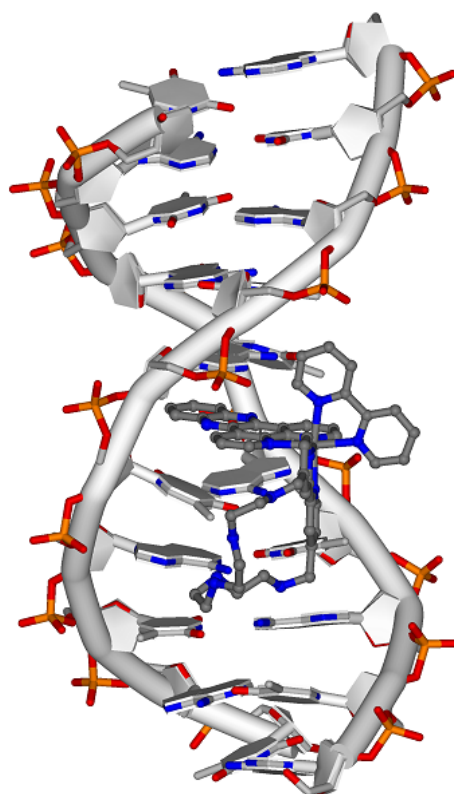


Figure S21. Calculated conformation for the ATATATATAT/ Δ -HD2³⁺ adduct.

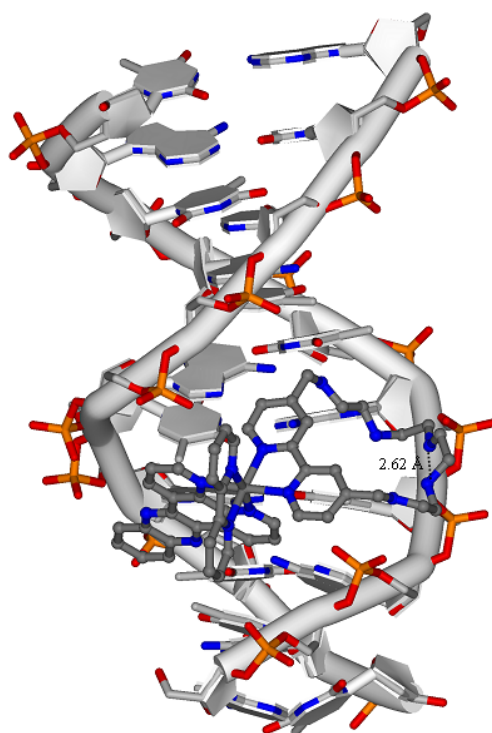


Figure S22. Calculated conformation for the ATATATATAT/ Δ -H₂D2⁴⁺ adduct.

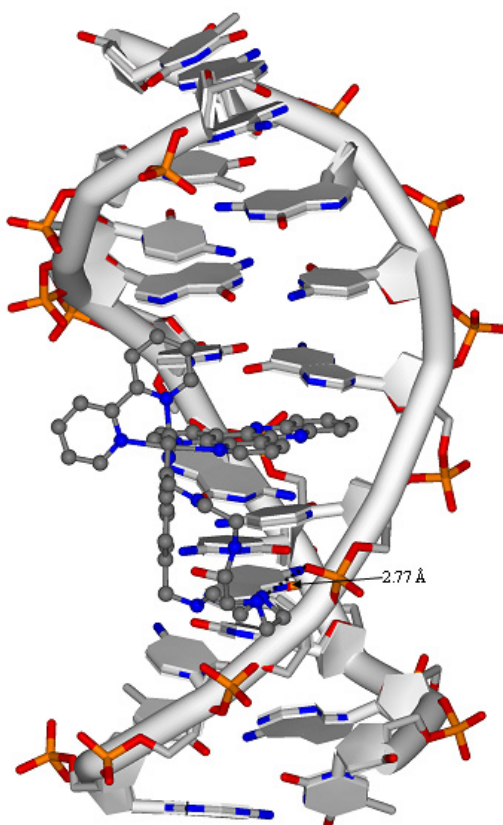


Figure S23. Calculated conformation for the ATCGCGCGAT/ Δ -H₂D2⁴⁺ adduct.

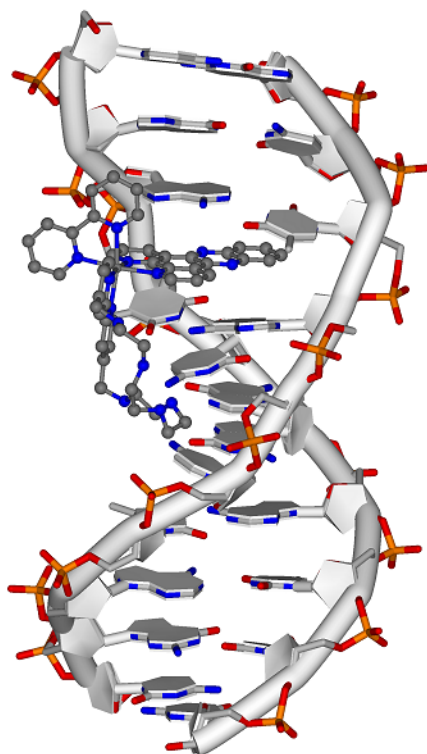


Figure S24. Calculated conformation for the CGATCGATCG/ Λ -H₂D2⁴⁺ adduct.

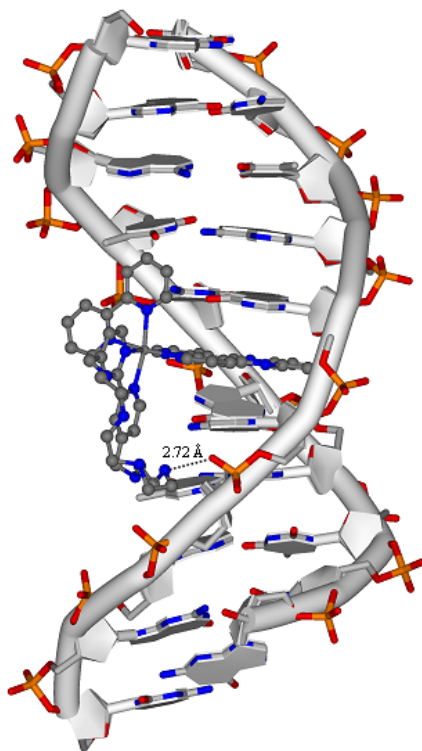


Figure S25. Calculated conformation for the CGATCGATCG/ Λ -HD2³⁺ adduct.

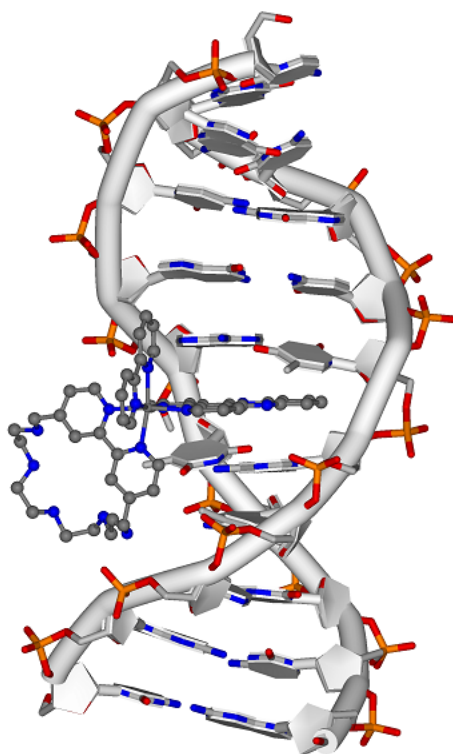


Figure S26. Calculated conformation for the CGCGATATCG/ Δ -H₂D2⁴⁺ adduct.

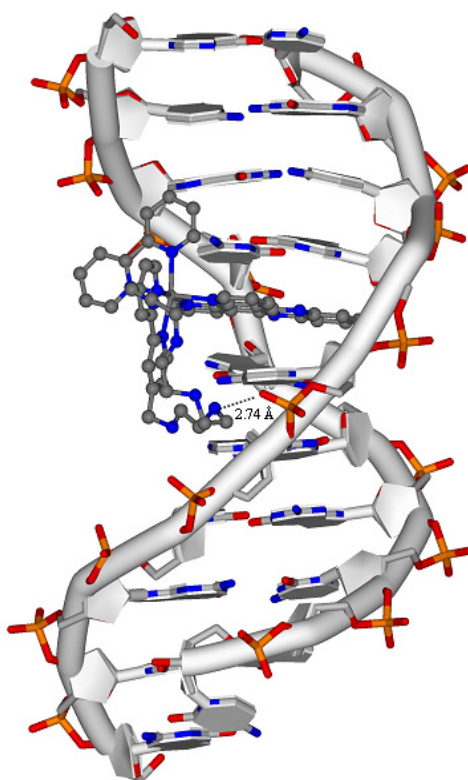


Figure S27. Calculated conformation for the GCGCGCGCGC/ Δ -H₂D2⁴⁺ adduct.