

Lanthanide-binding peptides with two pendant aminodiacetate arms: impact of the sequence on chelation

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1. pH-metric titrations

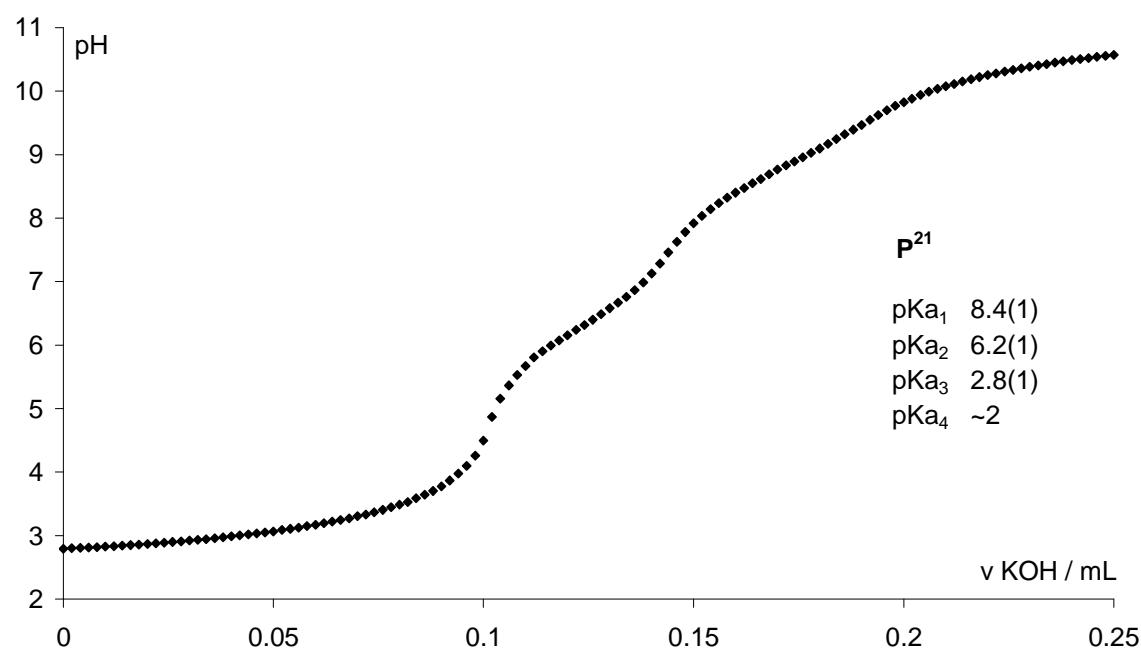
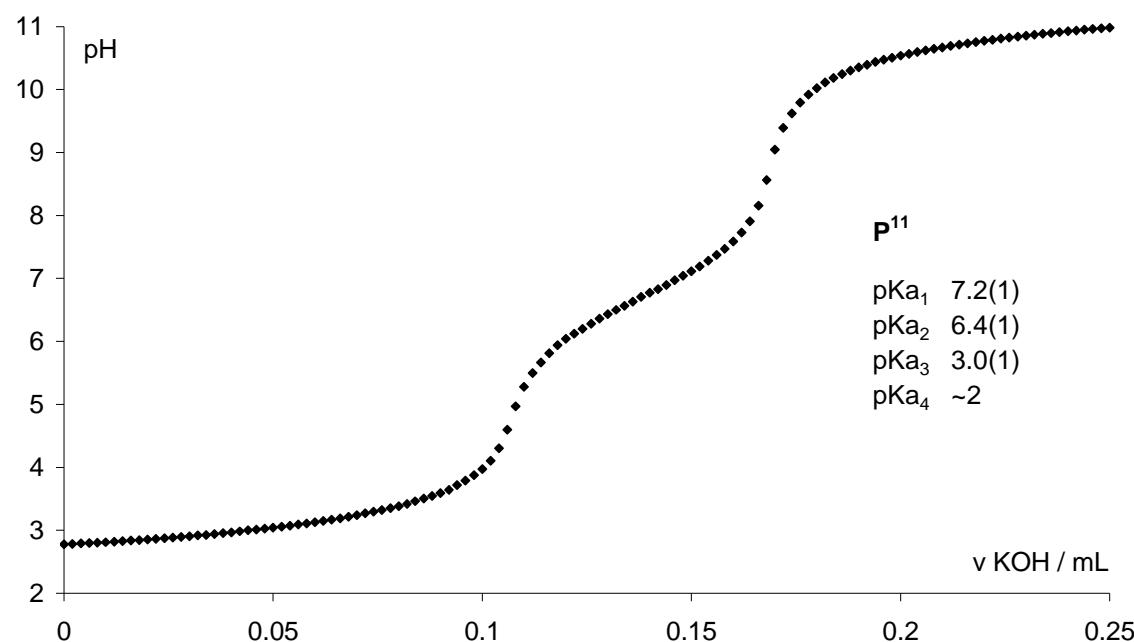


Figure S1. pH-metric titrations of \mathbf{P}^{11} (0.559 mM) and \mathbf{P}^{21} (0.826 mM) in 0.1 M KCl with 0.1 M KOH at 298 K.

2. ES-MS signature of the polynuclear species formed with \mathbf{P}^{21}

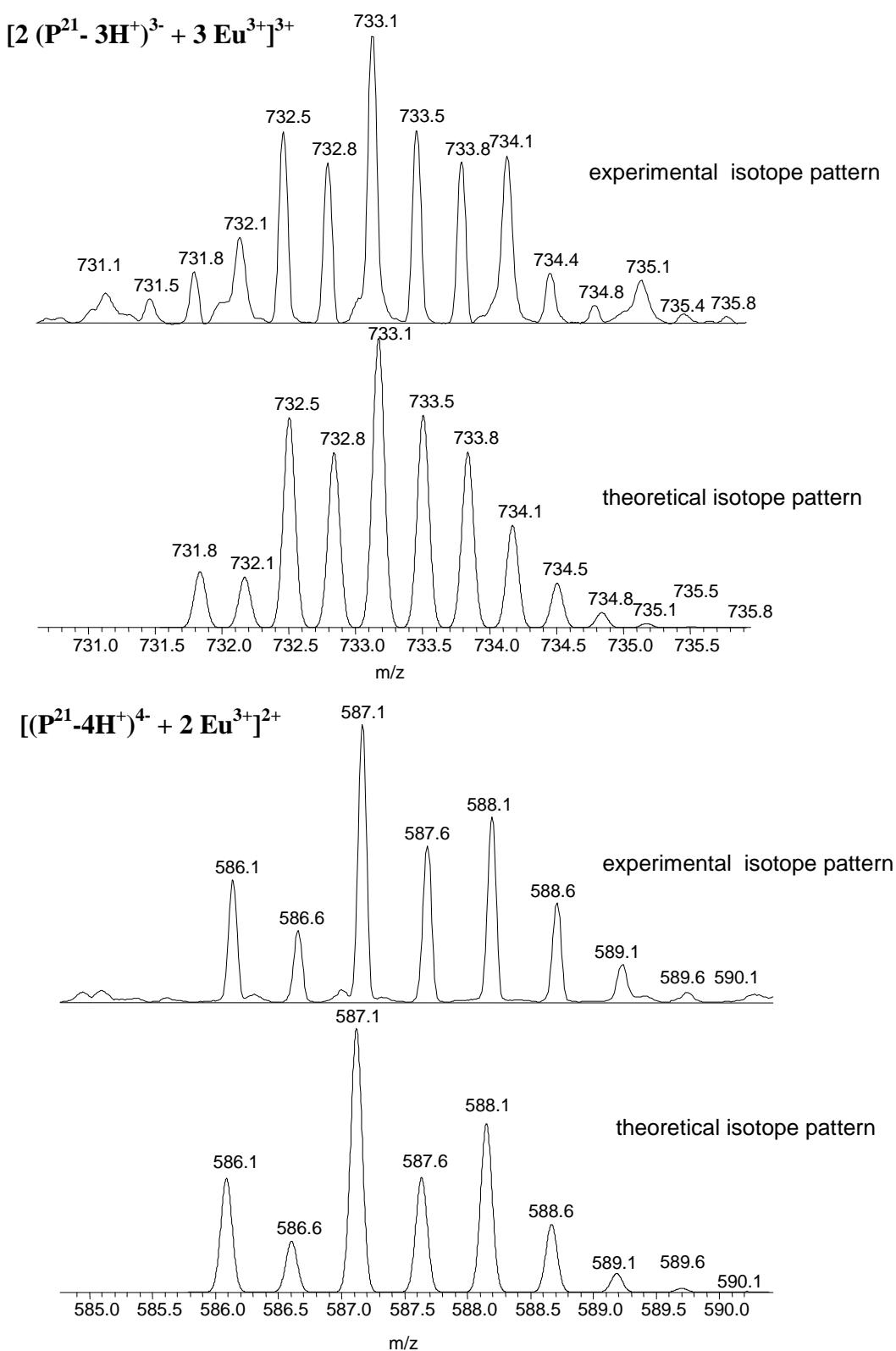


Figure S2. Experimental and theoretical isotope patterns of the polynuclear species formed with \mathbf{Eu}^{3+} and \mathbf{P}^{21} , detected by ES^+ -MS.

3. Competition experiments

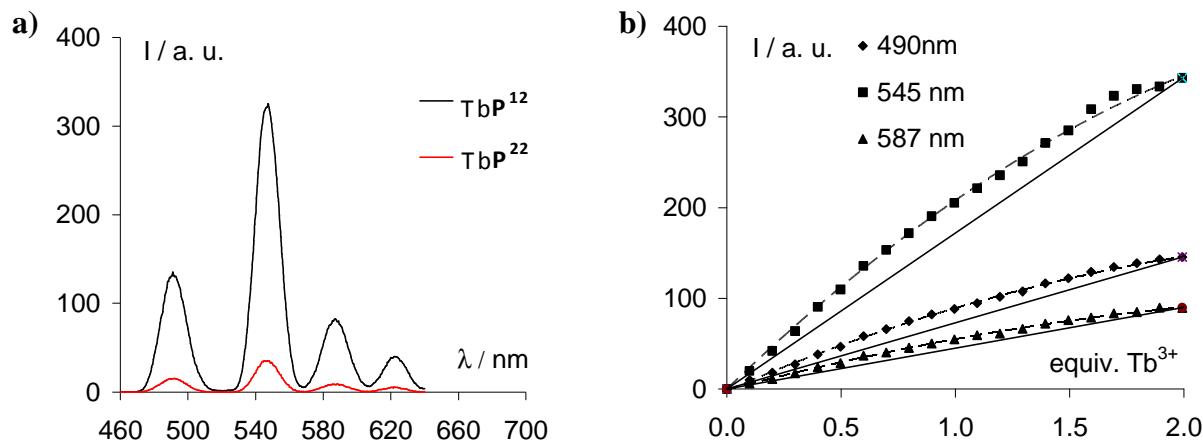


Figure S3. a) Emission spectra of 15 μM TbP^{12} and TbP^{22} in HEPES buffer (10 mM, pH = 7.0). b) Evolution of the Tb-centred luminescence during the titration of an equimolar mixture of P^{12} and P^{22} (15 μM) with TbCl_3 in HEPES buffer (10 mM, pH = 7.0). The emission spectra were recorded after 3 ms delay, during 5 ms gate time.

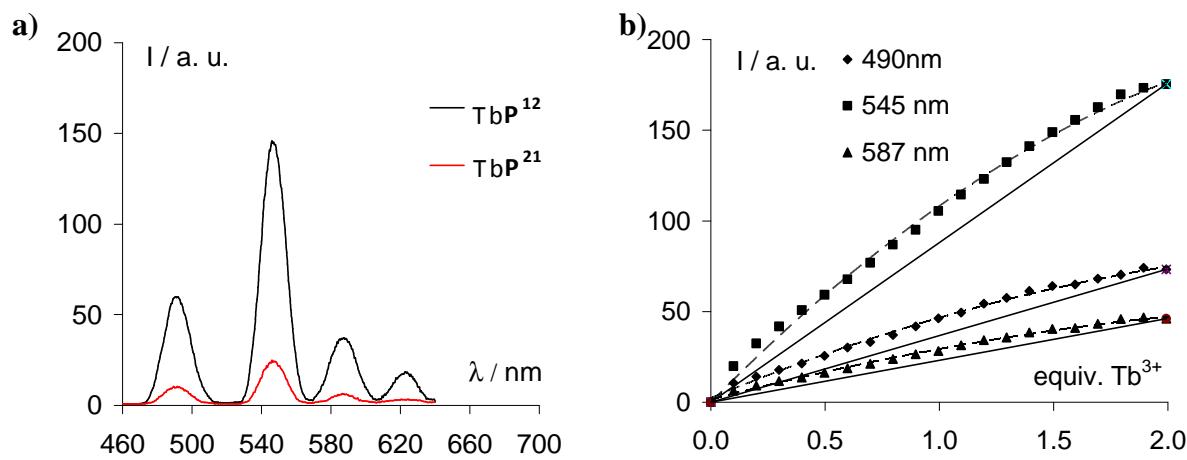


Figure S4. a) Emission spectra of 15 μM TbP^{12} and TbP^{21} in HEPES buffer (10 mM, pH = 7.0). b) Evolution of the Tb-centred luminescence during the titration of an equimolar mixture of P^{12} and P^{21} (15 μM) with TbCl_3 in HEPES buffer (10 mM, pH = 7.0). The emission spectra were recorded after 4 ms delay, during 5.5 ms gate time.

4. Supplementary luminescence experiments

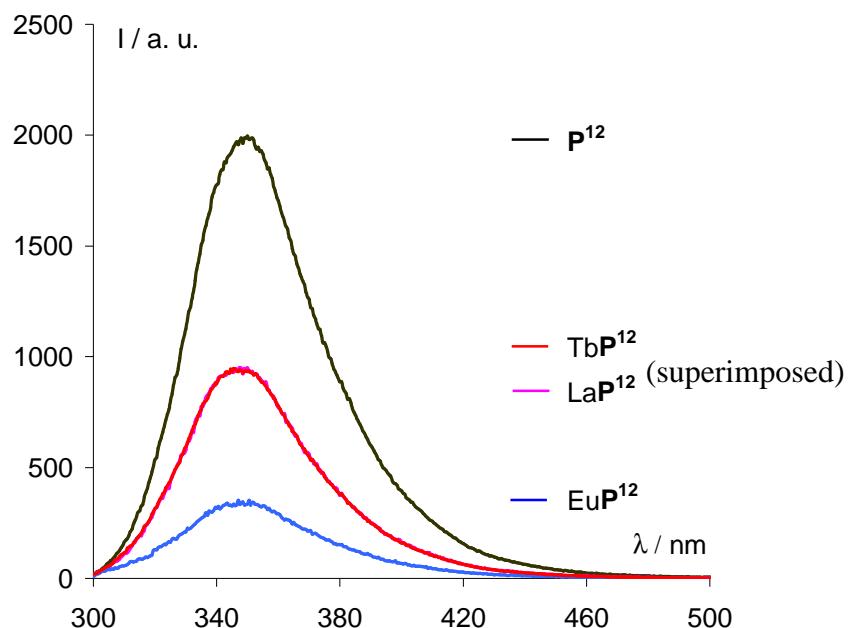


Figure S5. Fluorescence of tryptophan residue in \mathbf{P}^{12} and its Ln complexes upon excitation at 280 nm (40 μM , HEPES buffer 10 mM, pH = 7.0).

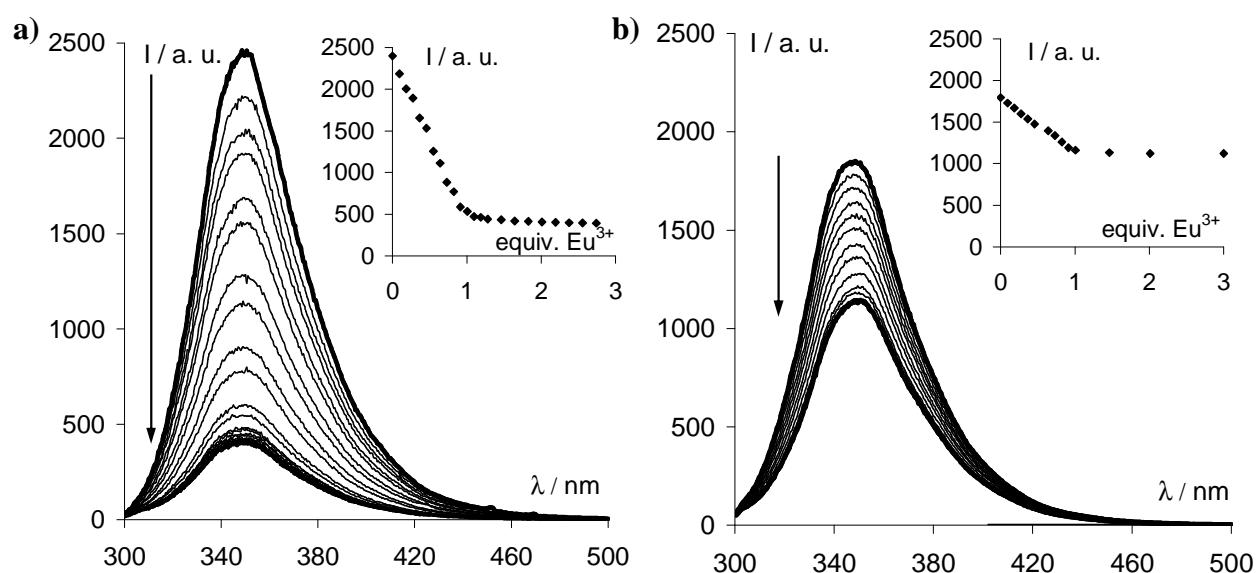


Figure S6. Titration of 51 μM (a) \mathbf{P}^{12} and (b) \mathbf{P}^{22} with EuCl_3 in HEPES buffer (10 mM, pH = 7.0). Evolution of the tryptophan fluorescence upon excitation at 280 nm.

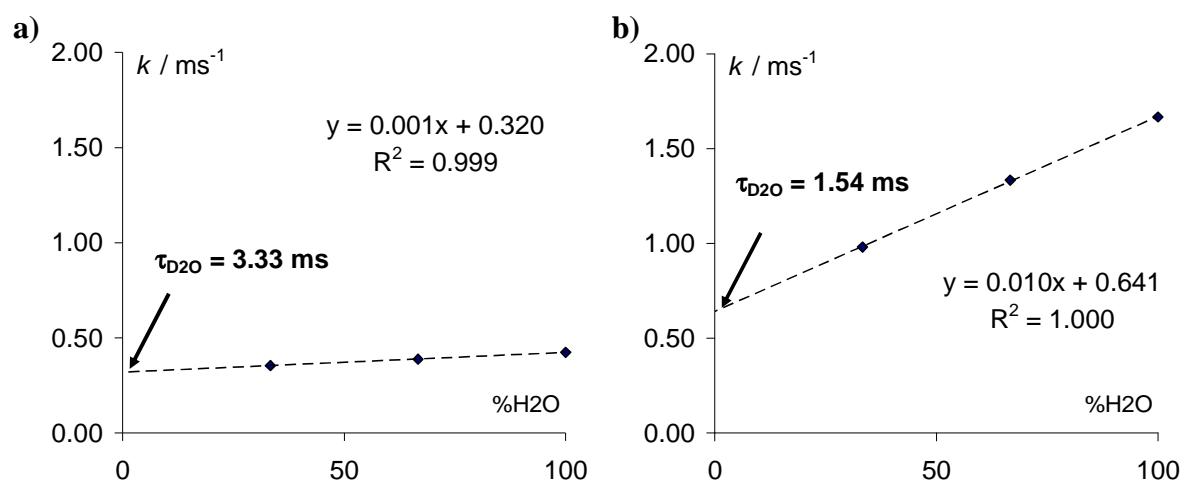


Figure S7. Relationship between the mole fraction of H_2O in $\text{H}_2\text{O}/\text{D}_2\text{O}$ mixtures and the radiative rate constant (k) for 30 μM (a) TbP^{12} and (b) EuP^{12} complexes in HEPES buffer (10 mM, pH = 7.0).

5. Supplementary NMR tables

Table S1. ^1H NMR (500 MHz) chemical shifts (δ / ppm) for **P¹¹** in $\text{H}_2\text{O}/\text{D}_2\text{O}$ v/v 9/1 at 298K, 0.99 mM, pH = 2.0. Signals assigned by COSY, TOCSY and ROESY 2D experiments.

Residue	HN	H α	H β	Others
Trp(1)	8.29	4.62	3.25, 3.21	$\text{H}\epsilon_1$: 10.18, $\text{H}\epsilon_3$: 7.60, $\text{H}\zeta_2$: 7.49, $\text{H}\delta_1$: 7.27, $\text{H}\eta_2$: 7.24, $\text{H}\zeta_3$: 7.15 Ac: 2.00
Ada ₁ (2)	8.40	4.75	3.43, 2.97	
Pro(3)		4.14	2.09, 1.89	$\text{H}\gamma$: 1.89, 1.79, $\text{H}\delta$: 3.34, 2.89
Gly(4)	8.47	3.99, 3.93		
Ada ₁ (5)	8.62	4.85	3.76, 3.50	
Gly(6)	8.53	3.89 (2H)		CONH ₂ : 7.46, 7.07 CH_2COOH : 3.94 (m, 4H), 3.84 (s, 4H)

Table S2. ^1H NMR (500 MHz) chemical shifts (δ / ppm) for **P¹²** in $\text{H}_2\text{O}/\text{D}_2\text{O}$ v/v 9/1 at 298K, 2.37 mM, pH = 2.4. Signals assigned by COSY, TOCSY and ROESY 2D experiments.

Residue	HN	H α	H β	Others
Trp(1)	8.30	4.64	3.24 (m, 2H)	$\text{H}\epsilon_1$: 10.19, $\text{H}\epsilon_3$: 7.63, $\text{H}\zeta_2$: 7.50, $\text{H}\delta_1$: 7.29, $\text{H}\eta_2$: 7.26, $\text{H}\zeta_3$: 7.17 Ac: 2.00
Ada ₁ (2)	8.45	4.75	3.44, 2.95	
Pro(3)		4.16	2.08, 1.90	$\text{H}\gamma$: 1.88, 1.79, $\text{H}\delta$: 3.35, 2.89
Gly(4)	8.50	3.94 (2H)		
Ada ₂ (5)	8.46	4.50	2.33, 2.13	$\text{H}\gamma$: 3.36 (m, 2H)
Gly(6)	8.50	3.94 (2H)		CONH ₂ : 7.48, 7.08 CH_2COOH : 3.90 (m, 4H), 3.83 (s, 4H)

Table S3. ^1H NMR (500 MHz) chemical shifts (δ / ppm) for \mathbf{P}^{21} in $\text{H}_2\text{O}/\text{D}_2\text{O}$ v/v 9/1 at 298K, 1.14 mM, pH = 2.5. Signals assigned by COSY, TOCSY and ROESY 2D experiments.

Residue	HN	H α	H β	Others
Trp(1)	8.26	4.46	3.27, 3.13	$\text{H}\epsilon_1$: 10.13, $\text{H}\epsilon_3$: 7.51, $\text{H}\zeta_2$: 7.47, $\text{H}\delta_1$: 7.28, $\text{H}\eta_2$: 7.21, $\text{H}\zeta_3$: 7.11 Ac: 2.00
Ada ₂ (2)	7.75	4.49	1.94, 1.71	$\text{H}\gamma$: 2.99(m, 2H)
Pro(3)		4.07	2.19, 1.87	$\text{H}\gamma$: 1.87 (m, 2H), $\text{H}\delta$: 3.37, 3.13
Gly(4)	8.43	3.96		
Ada ₁ (5)	8.67	4.87	3.77, 3.49	
Gly(6)	8.53	3.88		CONH ₂ : 7.45, 7.05 CH_2COOH : 3.89 (s 4H), 3.78 (s, 4H)

Table S4. ^1H NMR (500 MHz) chemical shifts (δ / ppm) for Lu \mathbf{P}^{12} in $\text{H}_2\text{O}/\text{D}_2\text{O}$ v/v 9/1 at 298K, 2.78 mM, pH = 7.0. Signals assigned by COSY, TOCSY and ROESY 2D experiments.

Residue	HN	H α	H β	Others
Trp(1)	8.25	4.59	3.30, 3.15	$\text{H}\epsilon_1$: 10.18, $\text{H}\epsilon_3$: 7.62, $\text{H}\zeta_2$: 7.51, $\text{H}\delta_1$: 7.26, $\text{H}\eta_2$: 7.24, $\text{H}\zeta_3$: 7.16 Ac: 2.00
Ada ₁ (2)	8.53	4.26	3.40, 3.00	
Pro(3)		4.34	2.15, 1.95	$\text{H}\gamma$: 1.95, 1.83, $\text{H}\delta$: 2.70, 2.50
Gly(4)	8.52	4.20, 4.00		
Ada ₂ (5)	8.53	nd*	3.2, 2.8	
Gly(6)	8.52	3.80 (2H)		CONH ₂ : 7.49, 7.10

* under the water signal.

Table S5. ^1H NMR (500 MHz) chemical shifts (δ / ppm) for La \mathbf{P}^{21} in $\text{H}_2\text{O}/\text{D}_2\text{O}$ v/v 9/1 at 278K, 4.3 mM, pH = 6.8. Signals assigned by COSY, TOCSY and ROESY 2D experiments.

Residue	HN	H α	H β	Others
Trp(1)	8.46	4.55	3.19 (2H)	$\text{H}\epsilon_1$: 10.27, $\text{H}\epsilon_3$: 7.56, $\text{H}\zeta_2$: 7.48, $\text{H}\delta_1$: 7.26, $\text{H}\eta_2$: 7.22, $\text{H}\zeta_3$: 7.12 Ac: 2.00
Ada ₂ (2)	8.32	4.97	2.09, 1.61	$\text{H}\gamma$: 3.19, 3.02
Pro(3)		4.06	2.12, 1.85	$\text{H}\gamma$: 1.85 (m, 2H), $\text{H}\delta$: 3.38, 3.13
Gly(4)	8.54	3.87		
Ada ₁ (5)	9.23	4.67	3.84 (2H)	
Gly(6)	8.71	4.12, 3.73		CONH ₂ : 7.44, 7.07