

Supporting Information

A novel quinoline-based two-photon fluorescent probe for detecting Cd²⁺ *in vitro* and *in vivo*

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1. Cell cytotoxicity

2. Spectroscopic properties of APQ and APQ-Cd²⁺ complex

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1. Cell cytotoxicity

To ascertain the cytotoxic effect, the MTT (5-dimethylthiazol-2-yl-2, 5-diphenyltetrazolium bromide) assay was performed as previously reported. HeLa cells were passed and plated to ~ 70% confluence in 96-well plates 24h before treatment. Prior to APQ treatment, the DMEM was removed and replaced with fresh DMEM, and aliquots of APQ stock solutions (5 mM DMSO) were added to obtain final concentrations of 10, 30, and 50 μ M. The treated cells were incubated for 24 h at 37 °C and under 5% CO₂. Subsequently, the cells were treated with 5 mg/mL MTT (40 μ L /well) and incubated for an additional 4 h (37°C, 5% CO₂). Then the cells were dissolved in DMSO (150 μ L/well), and the absorbance at 570 nm was recorded. The cell viability (%) was calculated according to the following Equation: Cell viability%=OD570(sample)/OD570(control)×100, where OD570 (sample) represents the optical density of the wells treated with various concentration of 6-MPQ and OD570(control) represents that of the wells treated with DMEM+10% FCS. percent cell survival values are relative to untreated control cells.

Table S1. Cytotoxicity Data of APQ (HeLa, 24 h)

Probe concentration	0 μ M	10 μ M	30 μ M	50 μ M
Cell survival %	97.37±6.7%	93.25±6.9%	52.75±7.10%	43.68±7.0%

2. Spectroscopic Properties of APQ

Fluorescence quantum yields were determined using the quinine sulfate (in 0.1 N H₂SO₄, Φ=0.55) as the standard (excited by 320 nm).

$$\Phi_U = \frac{\Phi_S(F_U A_S)}{F_S A_U}$$

The quantum yields are calculated by the following equation:

A_u and A_s are the UV absorption of analyte and the standard. F_u and F_s are integrated fluorescence emission of analyte and the standard.

The dissociation constant K_d is calculated by following equation:

$$\frac{F}{F_0} = 1 + \left(\frac{F_{\max}}{2F_0} - \frac{1}{2} \right) \left\{ 1 + \frac{C_M}{C_L} + \frac{1}{K_S C_L} - \left[\left(1 + \frac{C_M}{C_L} + \frac{1}{K_S C_L} \right)^2 - 4 \frac{C_M}{C_L} \right]^{\frac{1}{2}} \right\}$$

where F and F₀ are the fluorescence intensity of APQ in the presence and absence of Cd²⁺, C_M and C_L are the concentrations of Cd²⁺ and APQ; K_S is the stability constant. K_d is the reciprocal of K_S.

Table S2 Spectroscopic Properties of **APQ** and **APQ-Cd²⁺**

absorption maxima λ (nm), ε×10 ⁴ (M ⁻¹ cm ⁻¹)		emission maxima λ (nm)		fluorescence quantum yields Φ	
APQ	APQ+Cd ²⁺	APQ	APQ+Cd ²⁺	APQ	APQ+Cd ²⁺
27044	23980	480	520	0.057	0.2124

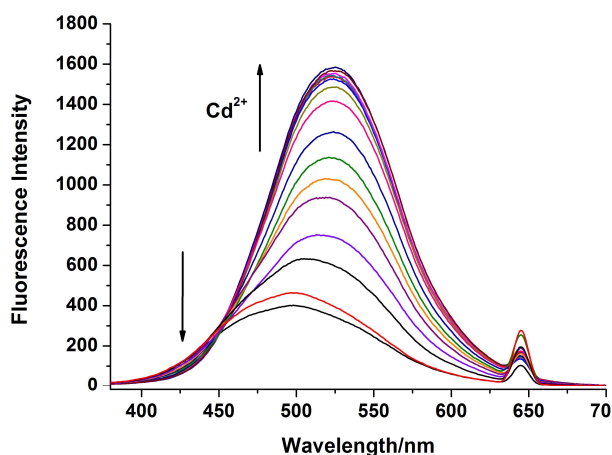


Fig S1. Fluorescence emission spectra of **APQ** (25 μM) with the excitation at 320 nm upon the titration of Cd²⁺ (0-1.5 equiv) in the methanol-water solutions (1:9, v/v, 50 mM HEPES buffer, pH = 7.4).

3. MALDI-TOF MS spectra of APQ、APQ-Cd²⁺ complex and APQ-Zn²⁺ complex

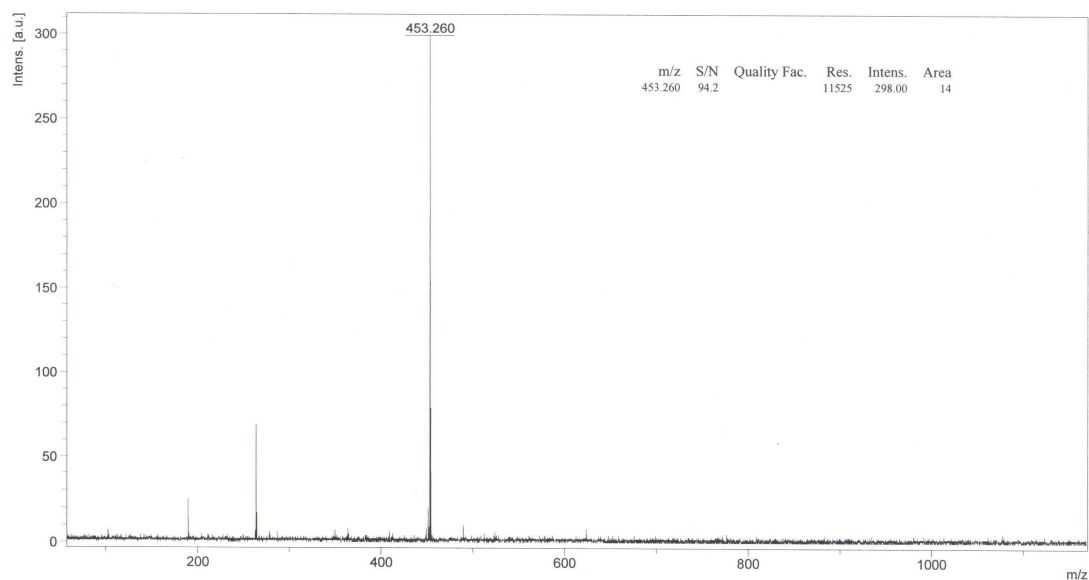


Fig S2. MALDI TOF MS of APQ (DCTB as matrix)

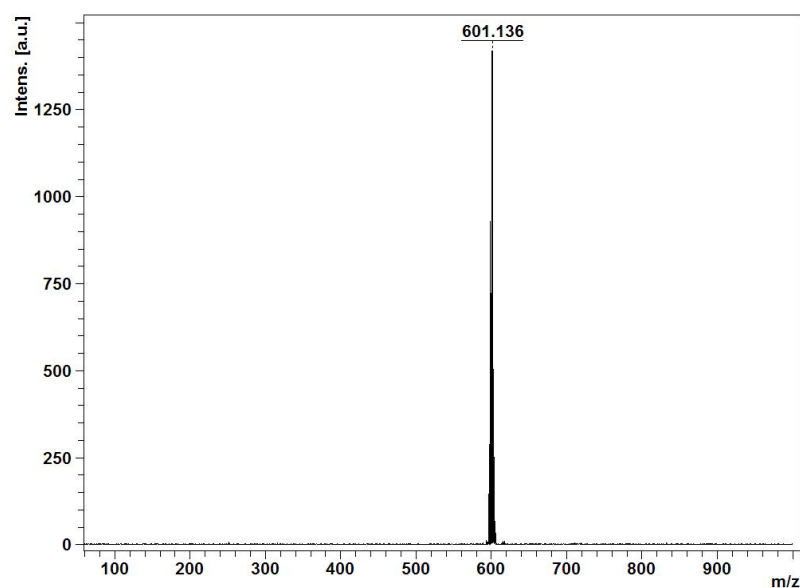


Fig S3. MALDI-TOF MS spectra of APQ-Cd²⁺ complex

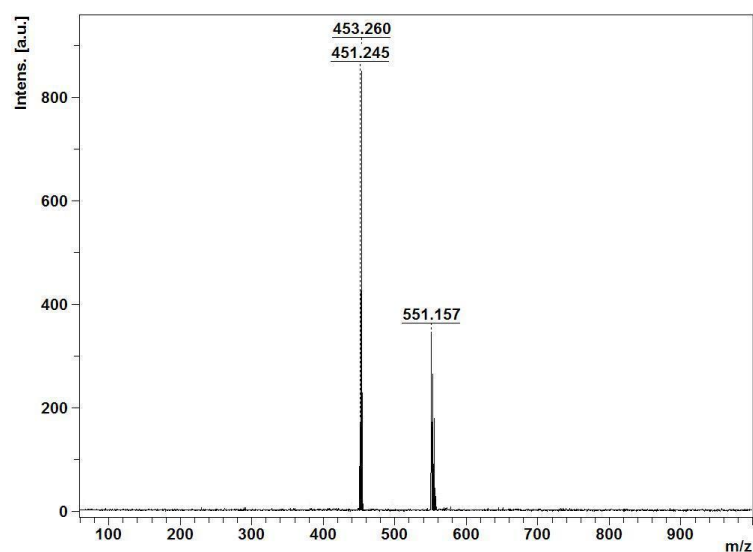


Fig S4. MALDI-TOF MS spectra of APQ-Zn²⁺ complex

4. ¹H NMR spectra of APQ and APQ-Cd²⁺ complex

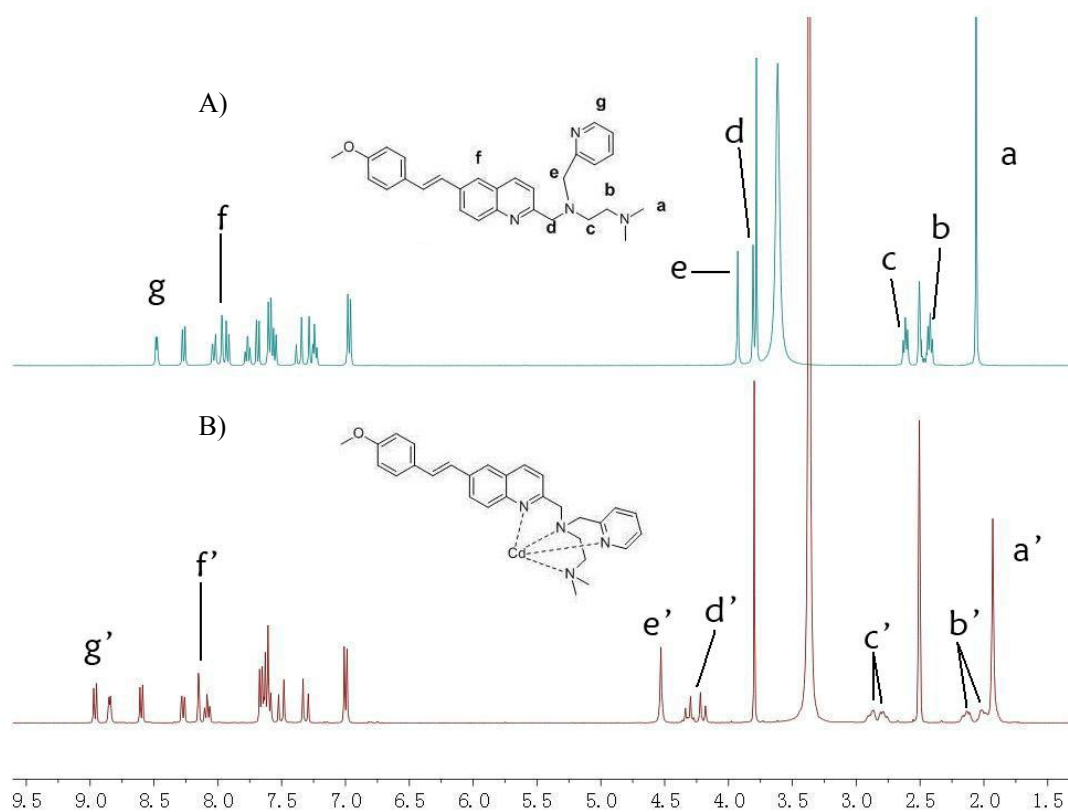
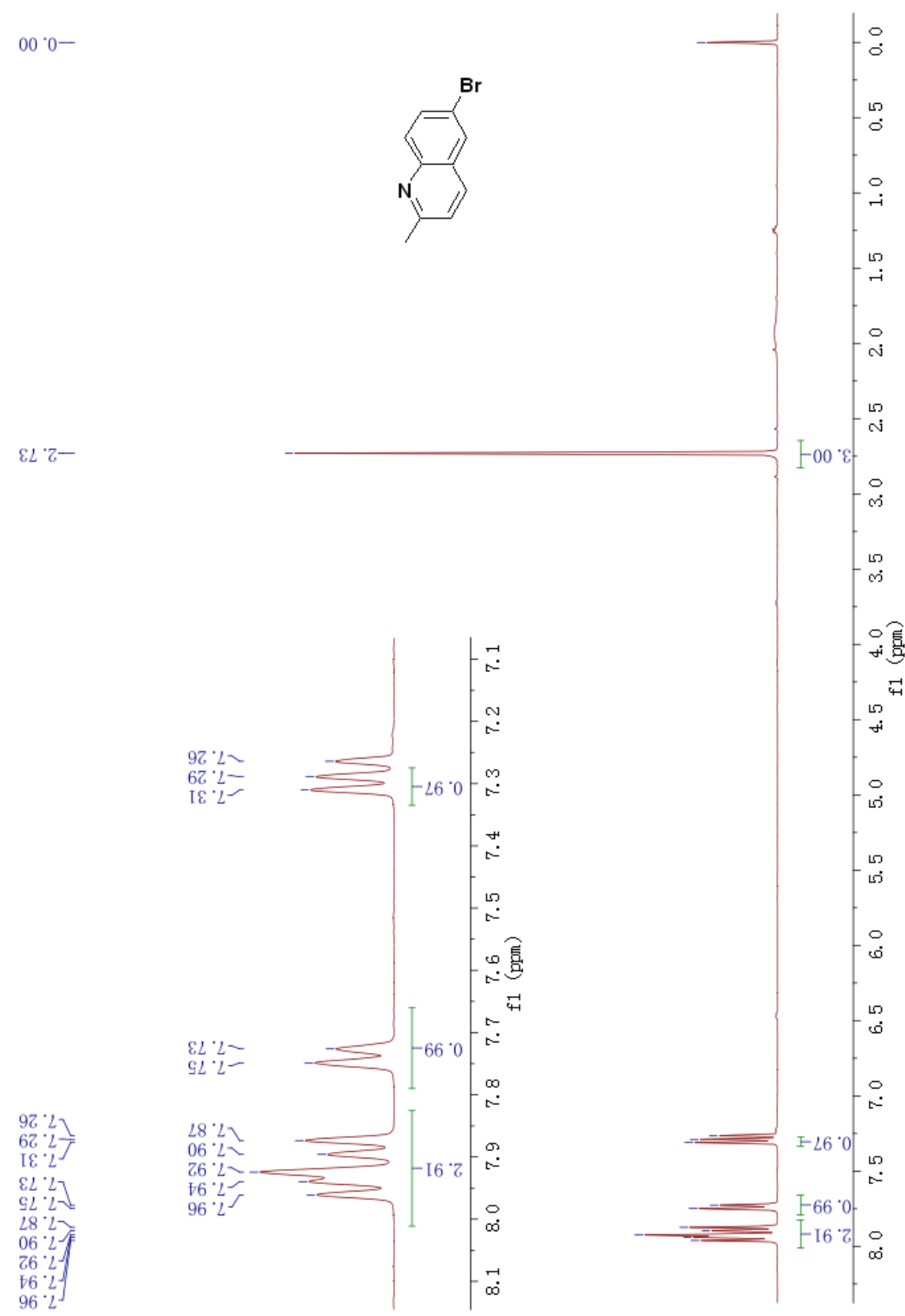
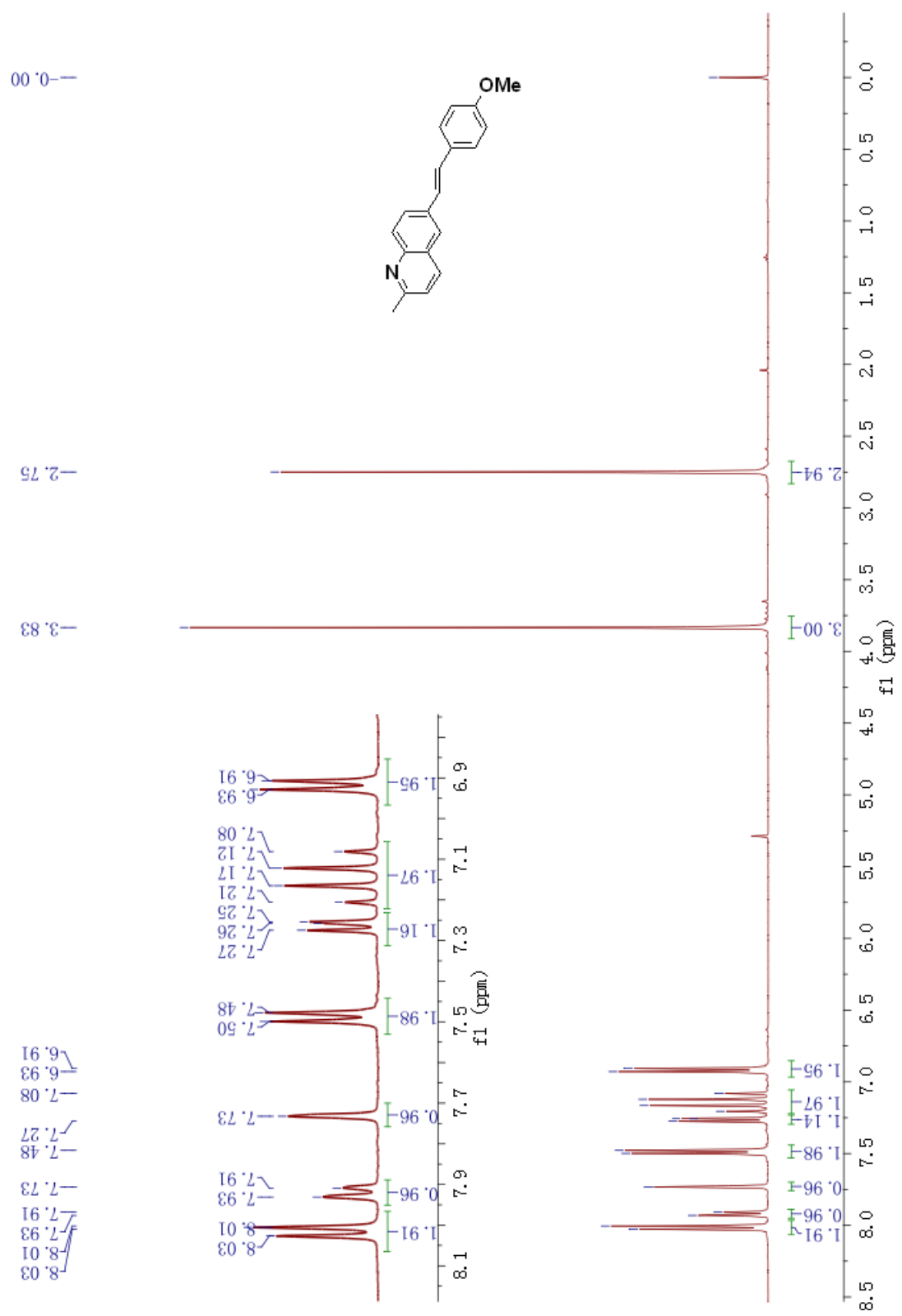
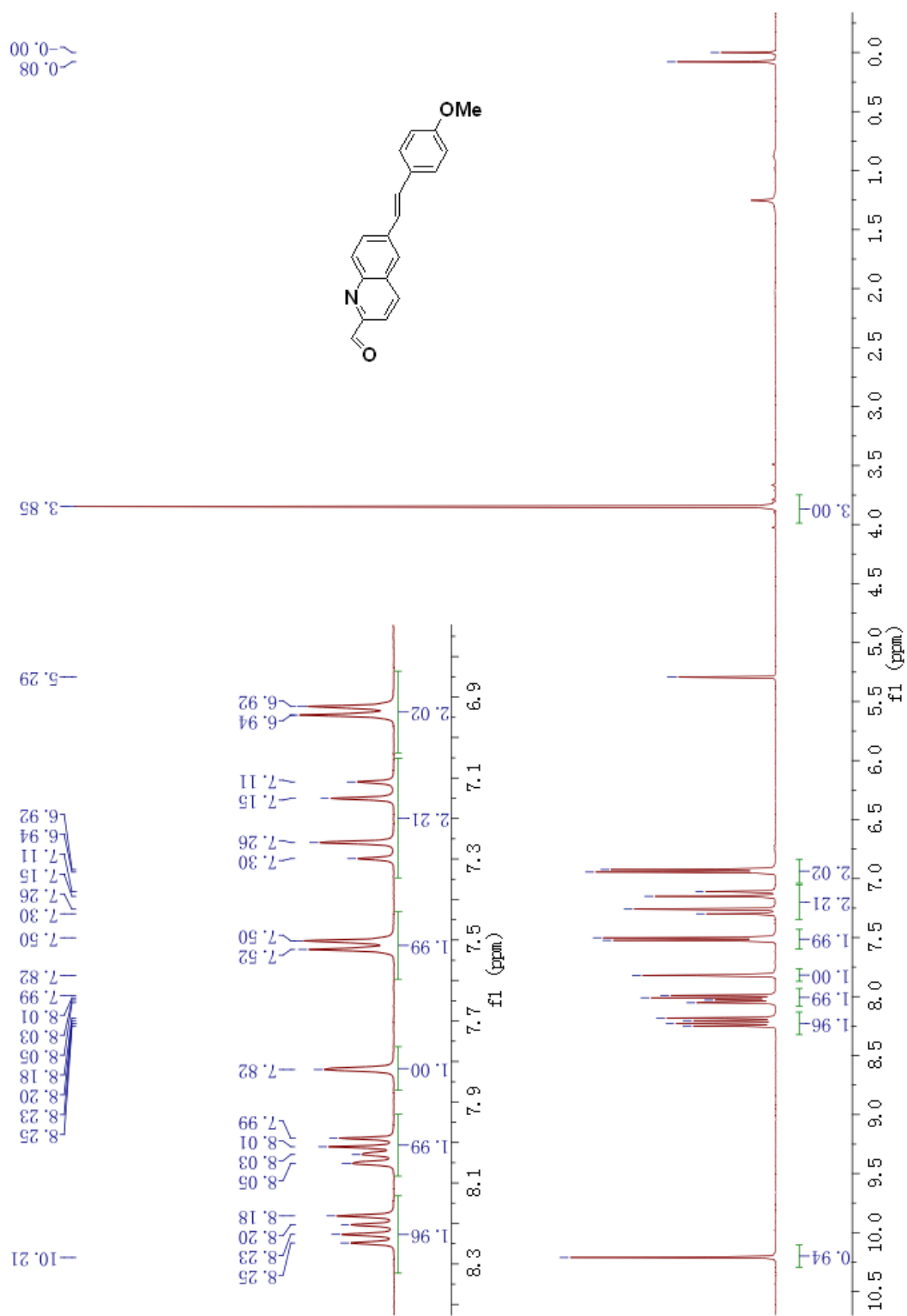


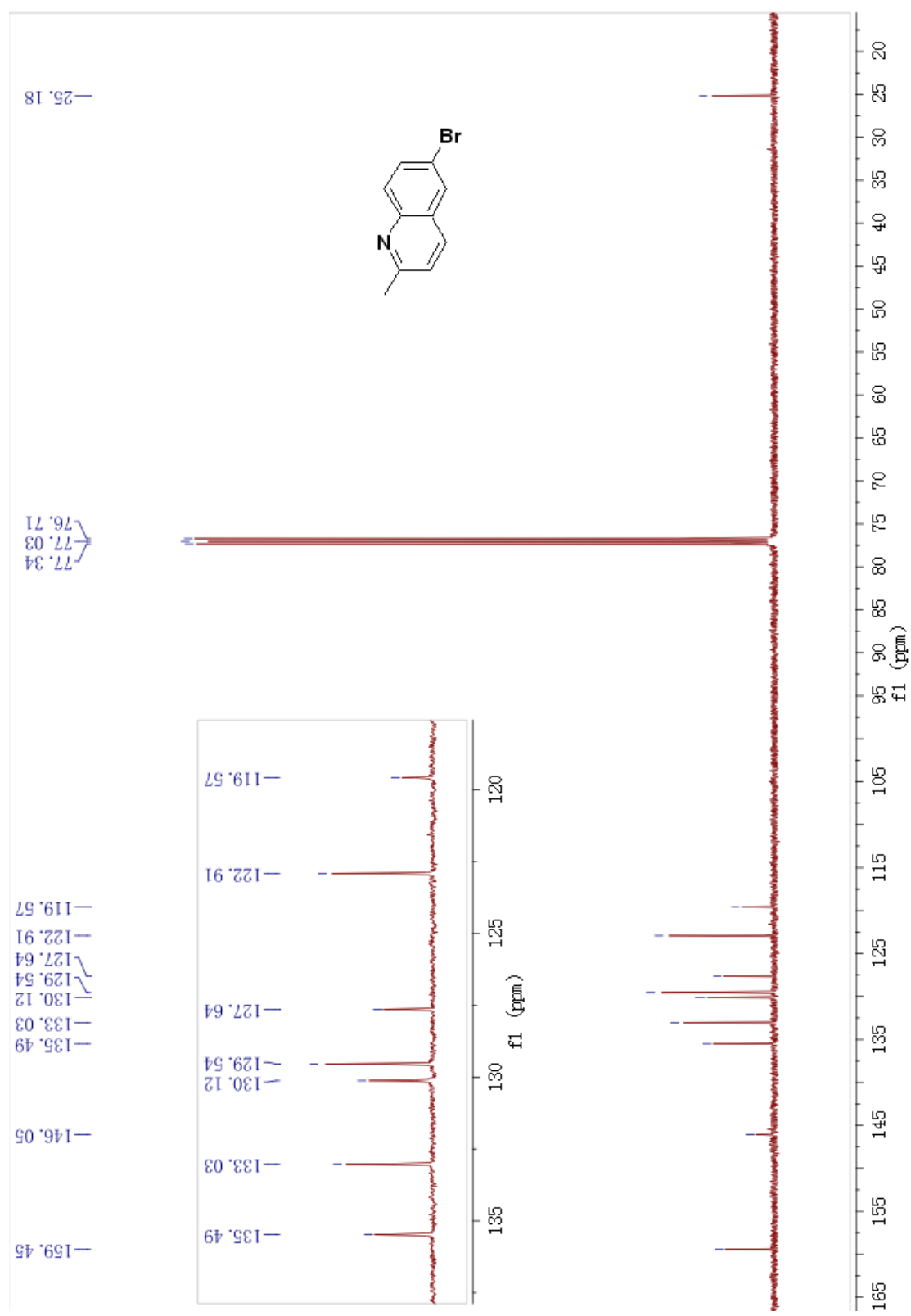
Fig S5. The ¹H-NMR spectra of the APQ (A) and APQ+1.2eq Cd²⁺ (B) in DMSO-d₆.

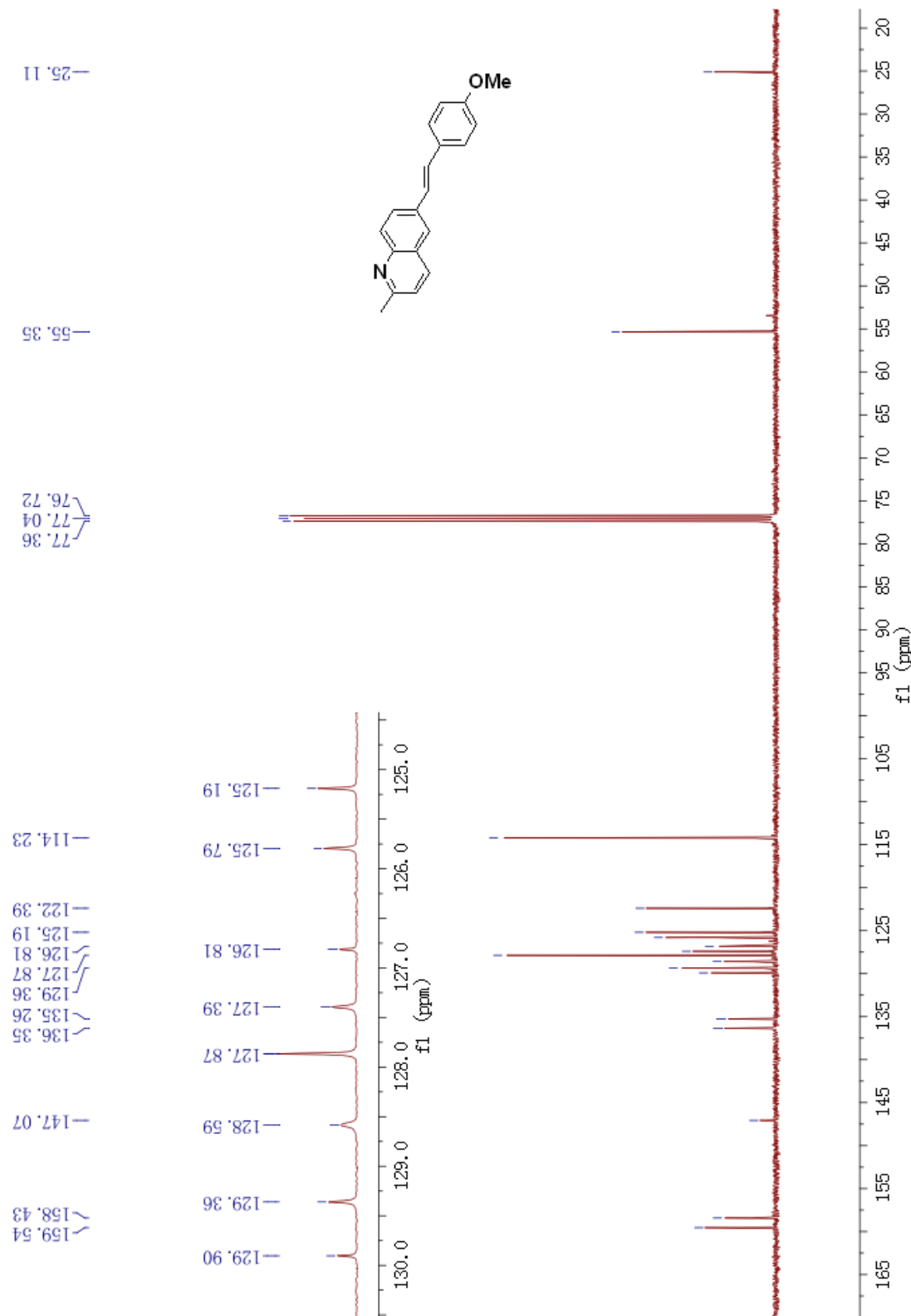
5. NMR spectra

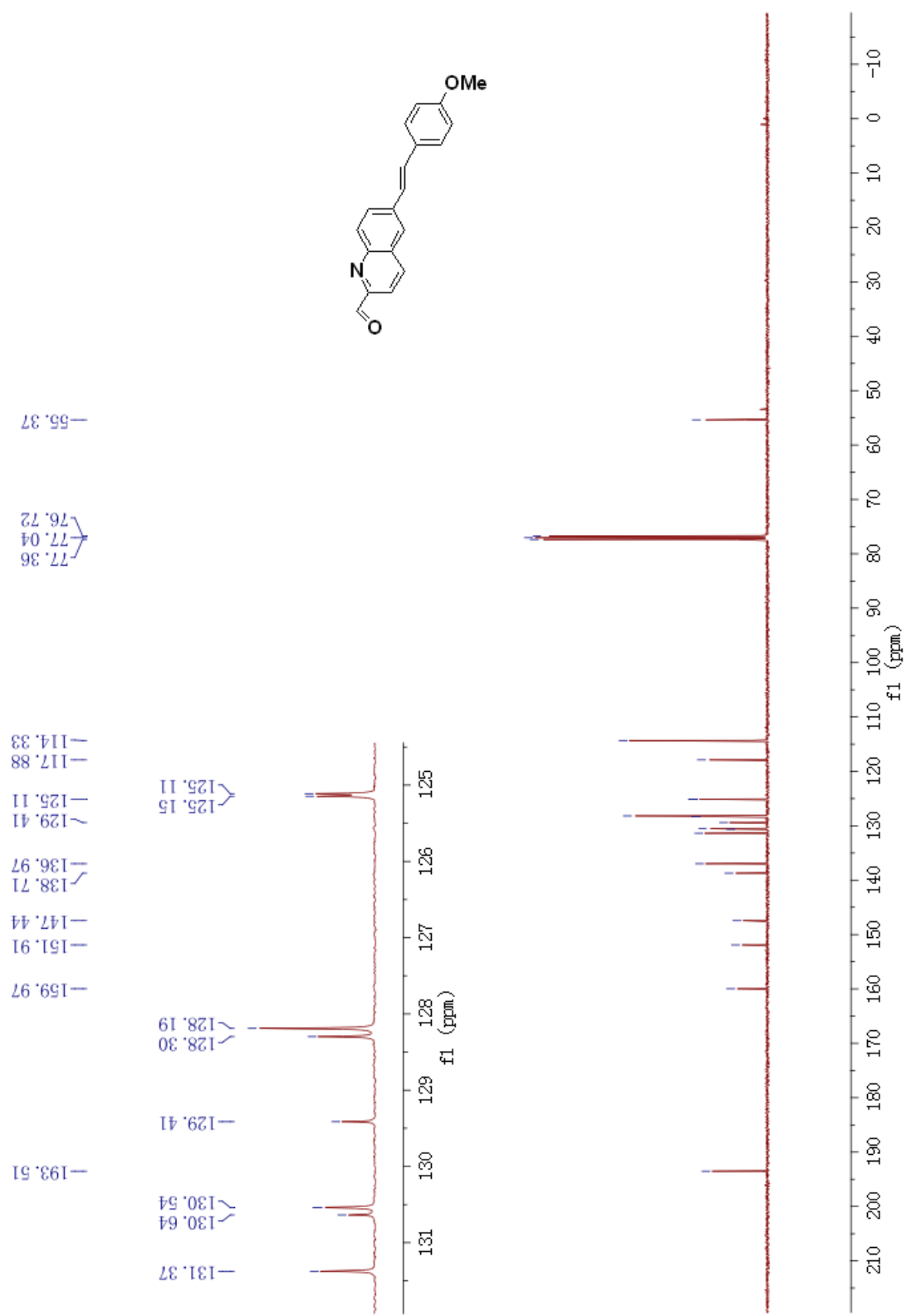


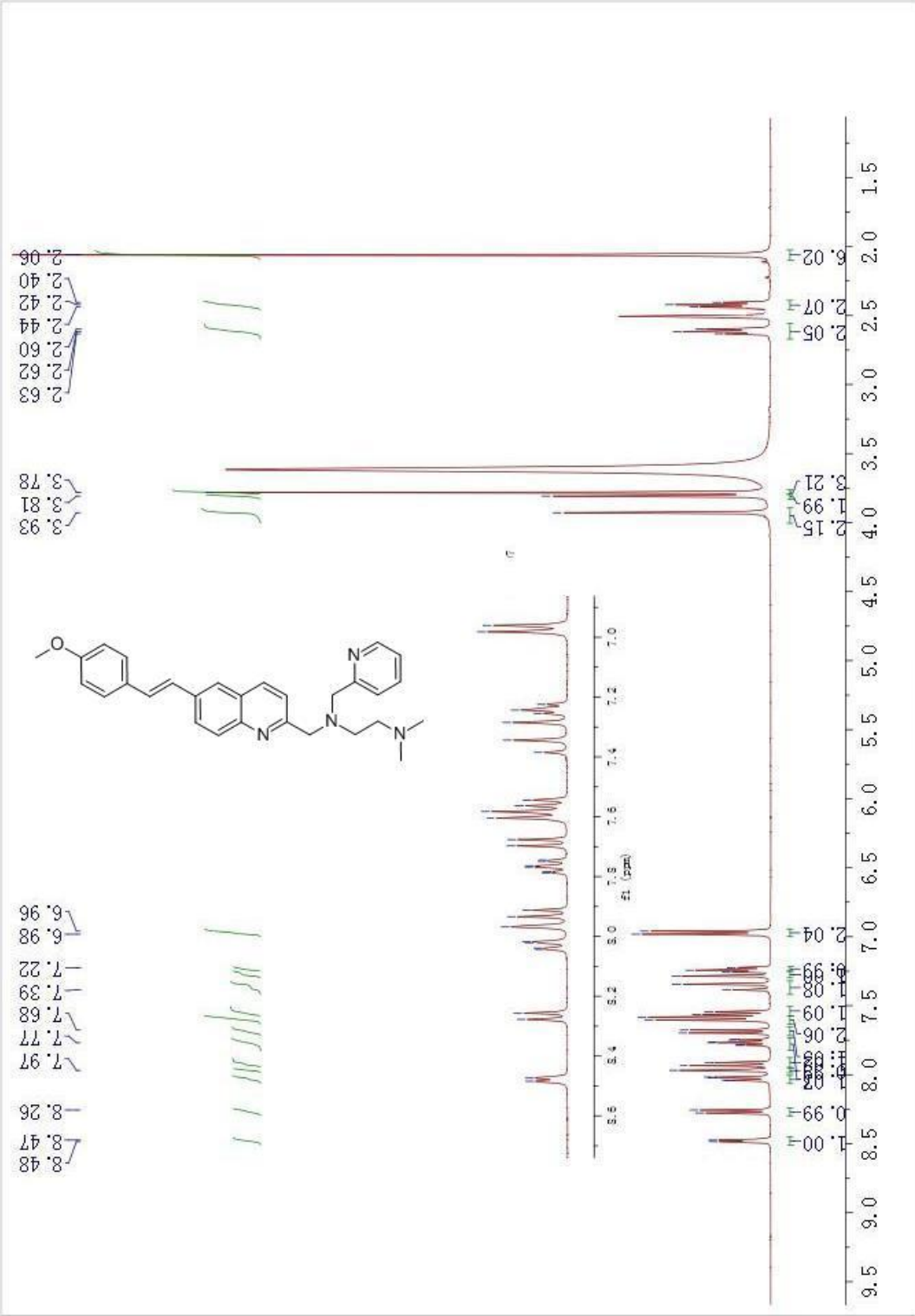


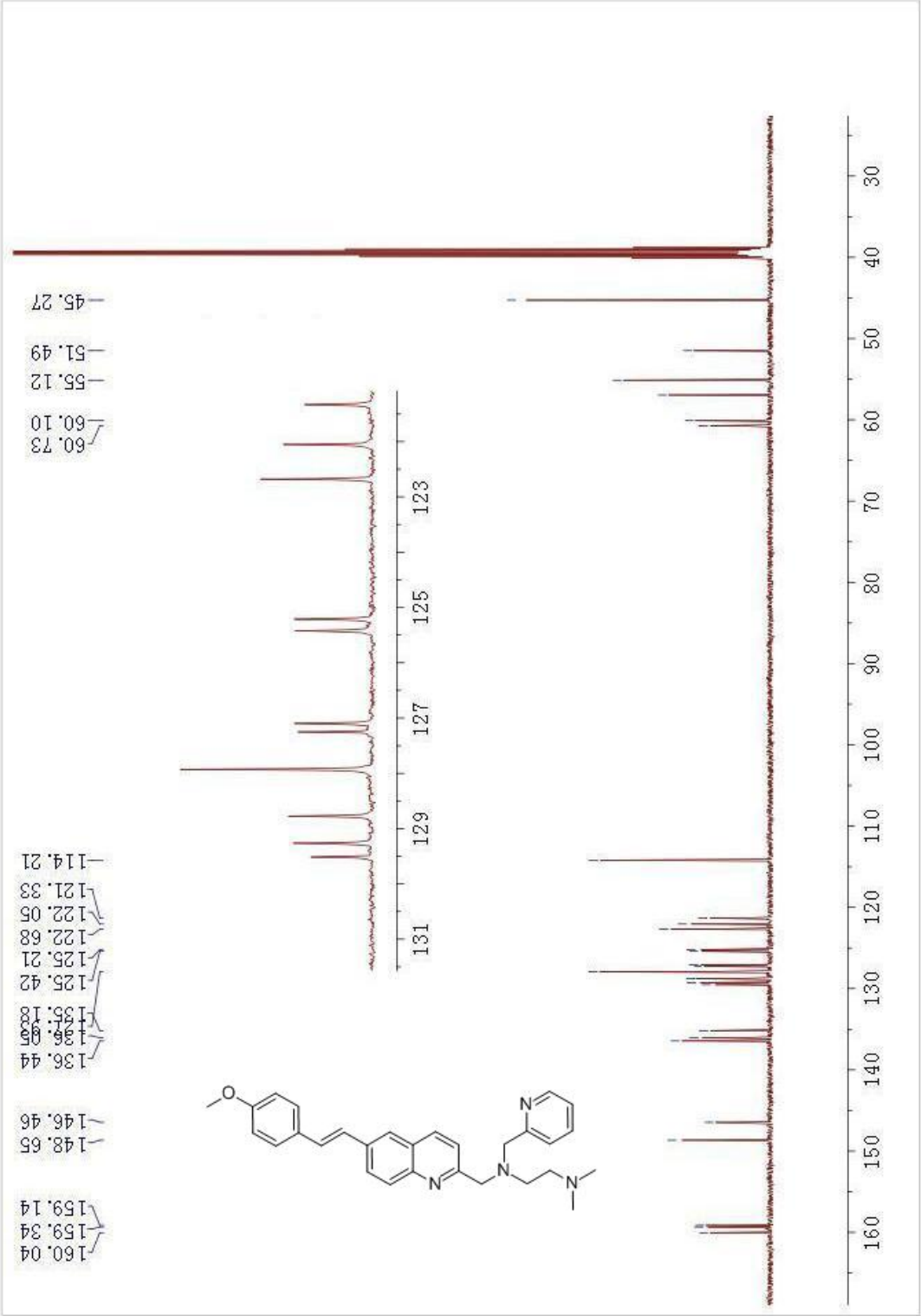


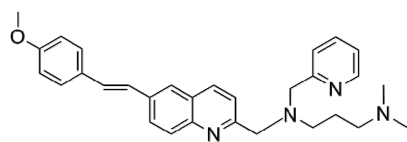






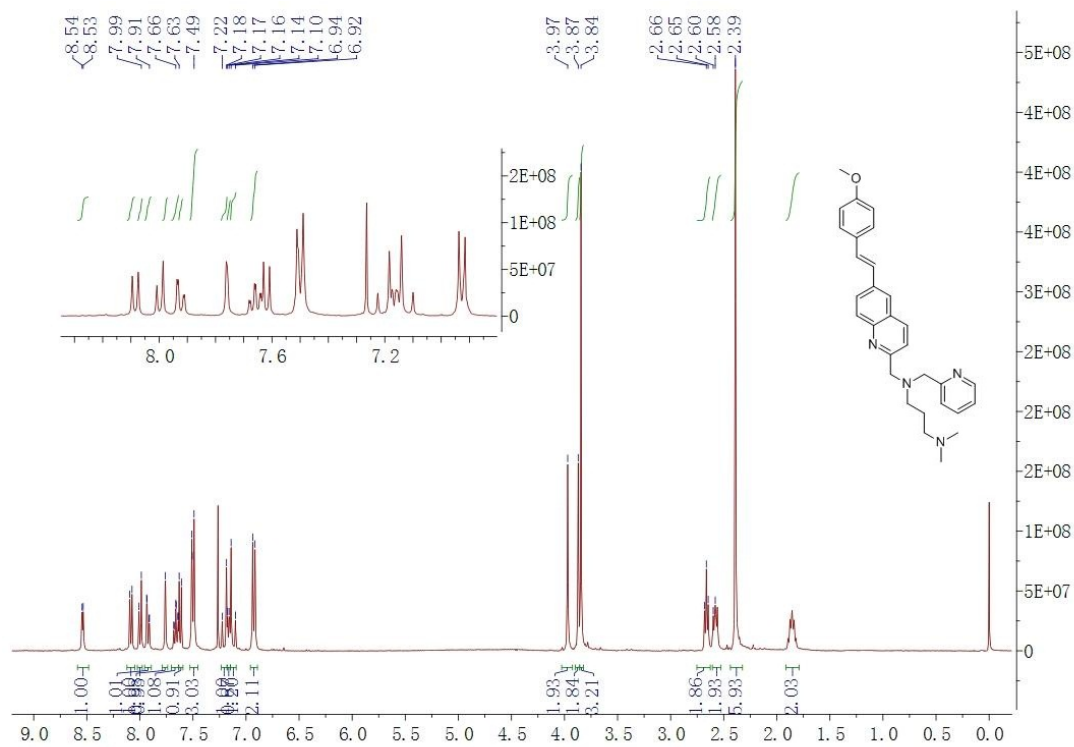






APPQ

¹HNMR



¹³CNMR

