ELECTRONIC SUPPLEMENTARY INFORMATION

Copper(II) Complexes of Quinoline Polyazamacroclyic Scorpiand-Type Ligands: X-Ray, Equilibrium and Kinetic Studies

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Figure S1. Distribution diagrams for the protonated species of L1.



Figure S2. Distribution diagrams for the protonated species of L2.







Figure S4. Distribution diagrams for the copper complexes of L2.

	Protonation process ^b			
NMR signal ^c	$L{\rightarrow} HL^+$	$HL^+ \rightarrow H_2 L^{2+}$	$H_2L^{2+} \rightarrow H_3L^{3+}$	$H_3L^{3+} \rightarrow H_4L^{4+}$
H1	0.32	0.26	0.51	-0.02
H2	0.46	0.31	0.21	-0.02
H3	0.61	0.19	0.02	-0.04
H4	0.36	0.17	0.16	-0.06
H5	0.21	0.13	0.52	0.07
H6	0.03	0.39	0.28	0.21
C1	-2.0	-0.9	0.6	0.0
C2	1.3	0.4	-1.0	-0.1
C3	-0.2	-1.5	-2.0	-0.1
C4	-0.3	-0.6	-1.8	-0.1
C5	-0.4	-0.6	-1.7	1.0
C6	0.8	-0.7	-2.8	-2.9

Table S1. Shift of the NMR $\Delta(\Delta\delta)$ signals in ppm observed upon formation of the different protonated forms of the L1 ligand.^a

^o The values are only approximate because the different protonated forms of the ligand usually do not reach 100% formation at any pH value. ^b The shifts in the signals were obtained from the spectra recorded at pD values where the different species reach its maximum concentration: 12.11 for L, 9.22 for HL⁺, 7.53 for H₂L²⁺, 4.10 for H₃L³⁺ and 0.93 for H₄L⁴⁺. ^c For simplicity, only the most relevant proton and carbon signals are included.

Table S2. Shift of the NMR signals observed upon formation of the different protonated forms of the L2 ligand.^a

	Protonation process ^b			
NMR signal ^c	$L \rightarrow HL^+$	$\mathrm{HL}^+ \rightarrow \mathrm{H}_2 \mathrm{L}^{2+}$	$H_2L^{2+} \rightarrow H_3L^{3+}$	$H_3L^{3+} \rightarrow H_4L^{4+}$
H1	0.43	0.42	0.33	-0.17
H2	0.55	0.33	0.24	-0.17
H3	0.59	0.14	0.08	-0.18
H4	0.41	0.11	0.17	-0.16
H5	0.25	0.03	0.60	-0.07
H6	0.00	0.08	0.71	0.11
C1	-1.4	-0.9	0.6	-0.3
C2	0.6	0.6	-0.8	-0.5
C3	-0.9	-2.1	-1.8	-0.3
C4	-0.1	-0.3	-3.8	-0.6
C5	-0.2	-0.5	-1.9	0.3
C6	0.1	-0.4	-0.8	-0.4

^a The values are only approximate because the different protonated forms of the ligand usually do not reach 100% formation at any pH value. ^b The shifts in the signals were obtained from the spectra recorded at pD values where the different species reach its maximum concentration: 12.38 for L, 9.17 for HL⁺, 7.36 for H₂L²⁺, 4.34 for H₃L³⁺, and 0.00 for H₄L^{4+, c} For simplicity, only the most relevant proton and carbon signals are included in the Table. Nevertheless, complete spectra can be found in ref 9.

Bond Distances (Å)		Bond Angles	
Cu1-N1	1.979(5)	N1-Cu1-N2	78.8(2)
Cu1-N2	2.309(6)	N1-Cu1-N3	91.8(2)
Cu1-N3	2.183(6)	N1-Cu1-N4	78.9(2)
Cu1-N4	2.276(6)	N1-Cu1-N5	175.0(3)
Cu1-N5	1.976(7)	N1-Cu1-N6	104.4(2)
Cu1-N6	2.173(5)	N2-Cu1-N3	81.1(2)
		N2-Cu1-N4	150.2(2)
		N2-Cu1-N5	98.9(3)
		N2-Cu1-N6	104.0(2)
		N3-Cu1-N4	80.0(2)
		N3-Cu1-N5	83.5(3)
		N3-Cu1-N6	163.7(2)
		N4-Cu1-N5	101.6(3)
		N4-Cu1-N6	100.5(2)
		N5-Cu1-N6	80.4(2)

 $\label{eq:solution} \textbf{Table S3}. \ Selected \ bond \ lengths (\AA) \ and \ angles \ (deg) \ for \ complex \ [Cu(L1)](ClO_4)_2 \cdot H_2O.$



Figure S5. Spectral changes recorded during the titration of L1. The spectra were recorded at the following pH values: 1.09, 1.56, 2.10, 2.47, 2.90, 3.61, 4.00, 4.53, 5.11, 5.55, 6.06, 6.56, 6.99, 7.51, 8.08, 8.67, 8.98, 9.46, 9.95, 10.50, and 11.03, and the arrows indicate the direction of changes when the pH is increased.



Figure S6. Spectral changes recorded during the titration of L2. The spectra were recorded at the following pH values: 2.05, 2.74,3.01,3.85,4.46, 5.15,5.81,6.87,8.18,8.94,9.62 and 10.12, and the arrows indicate the direction of changes when the pH is increased.