Electronic Supplementary Information

Non-conventional coordination of cavity-confined metal centres

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[†] Electronic supplementary information (ESI) available: Experimental details and CIF file for the reported complex. CCDC reference number 801259.

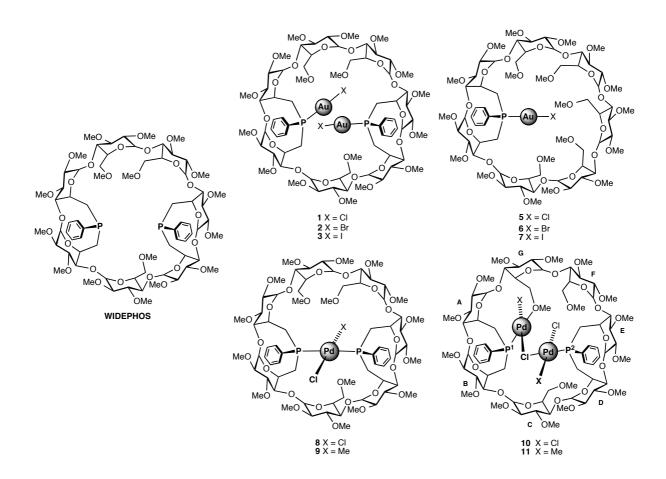
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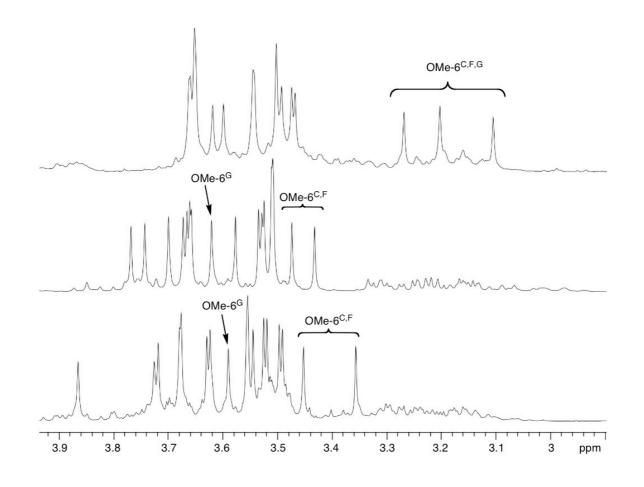


Fig. SI-1. Part of the ¹H NMR spectra of **WIDEPHOS** (top), complex **10** (middle) and complex **11** (bottom) recorded in CDCl₃ (298 K) showing the methoxy signals.

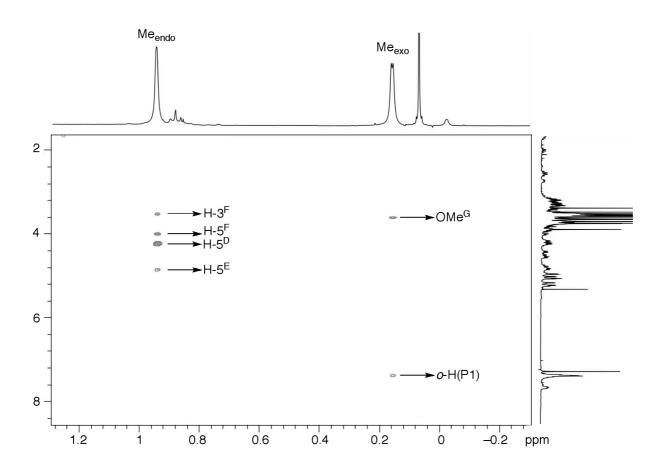
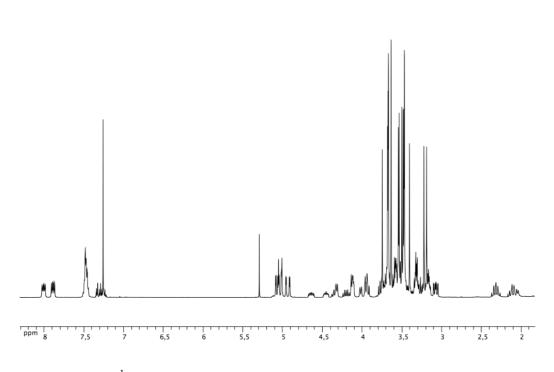
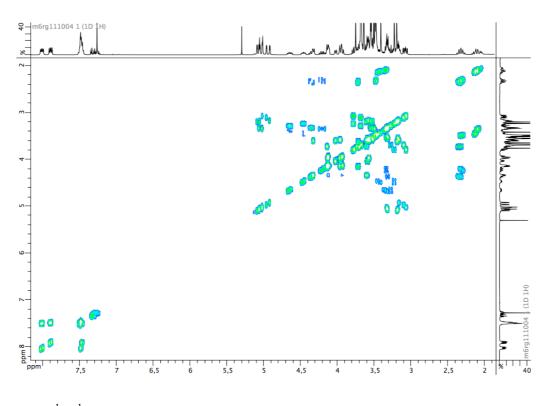


Fig. SI-2. Part of ¹H-¹H ROESY NMR spectrum of bridged dinuclear palladium(II) complex **11** recorded in CDCl₃ at 400.1 MHz.

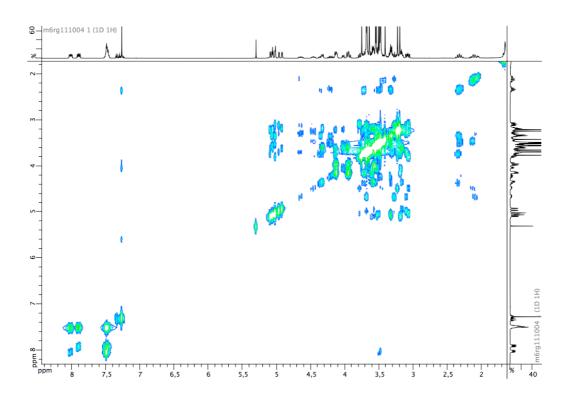
NMR spectra of complexes 1-3, 5-11



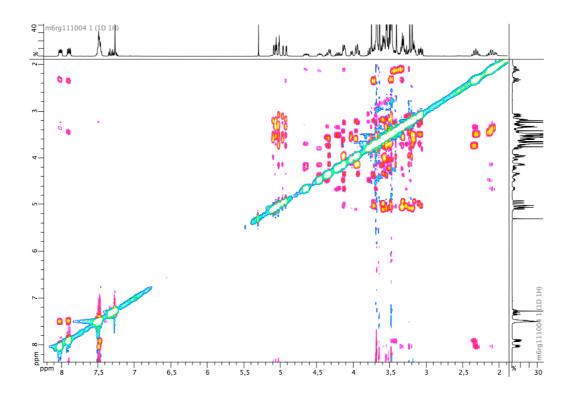
¹H NMR spectrum of **1** recorded in CDCl₃ at 500.1 MHz.



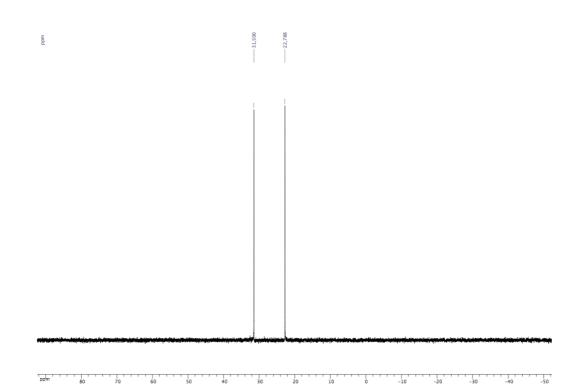
¹H-¹H COSY NMR spectrum of **1** recorded in CDCl₃ at 500.1 MHz.



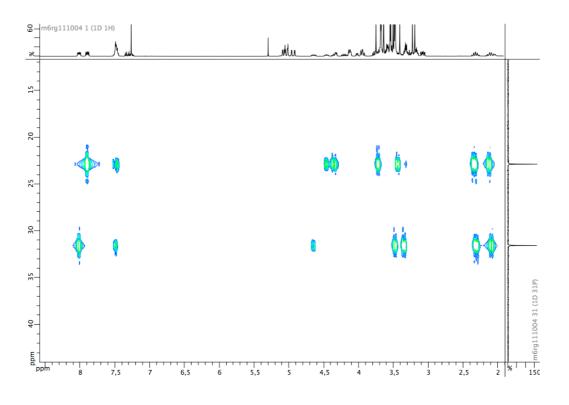
¹H-¹H TOCSY NMR spectrum of **1** recorded in CDCl₃ at 500.1 MHz.



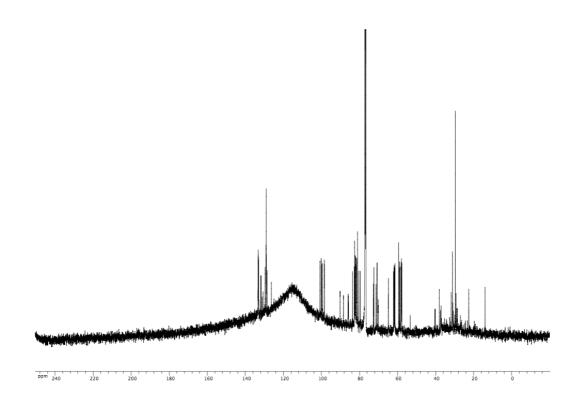
¹H-¹H ROESY NMR spectrum of **1** recorded in CDCl₃ at 500.1 MHz.



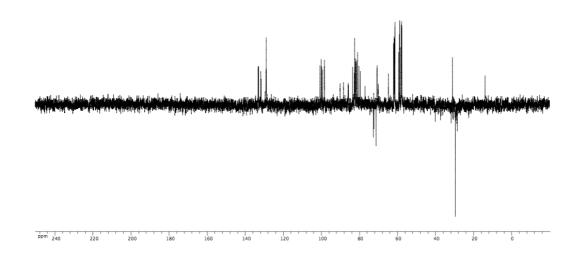
³¹P{¹H} NMR spectrum of **1** recorded in CDCl₃ at 121.5 MHz.



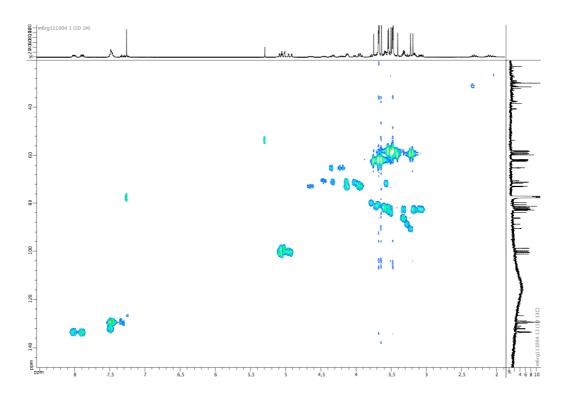
 $^{1}\text{H-}^{31}\text{P}\{^{1}\text{H}\}$ NMR spectrum of 1 recorded in CDCl₃ at 500.1 MHz.



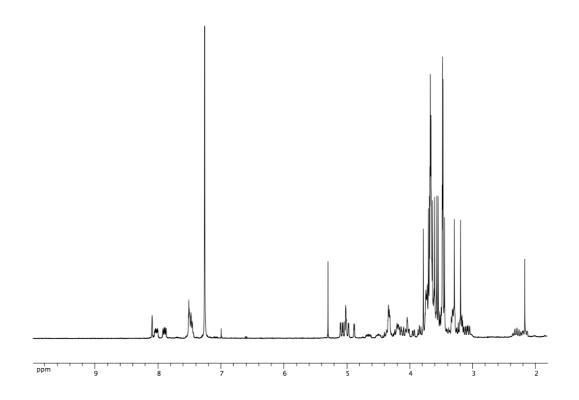
 $^{13}C\{^1H\}$ NMR spectrum of 1 recorded in CDCl3 at 125.8 MHz.



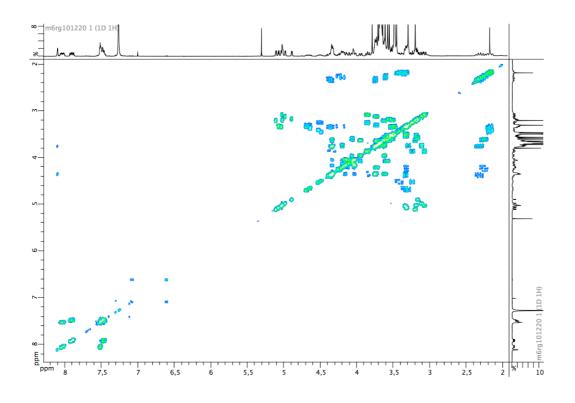
DEPT135 spectrum of 1 recorded in CDCl₃ at 125.8 MHz.



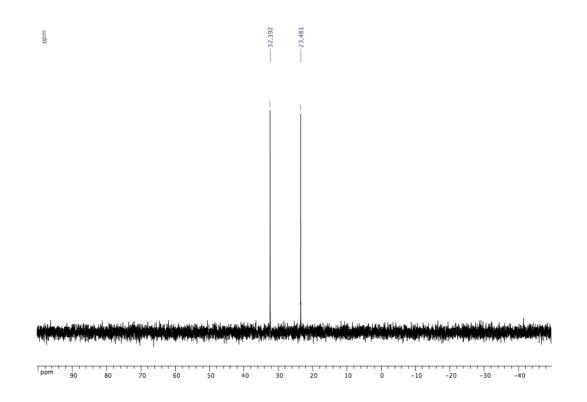
 1 H- 13 C{ 1 H} HMQC spectrum of 1 recorded in CDCl₃ at 500.1 MHz.



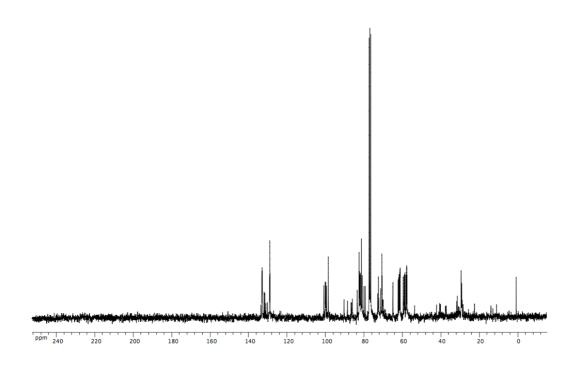
¹H NMR spectrum of **2** recorded in CDCl₃ at 400.1 MHz.



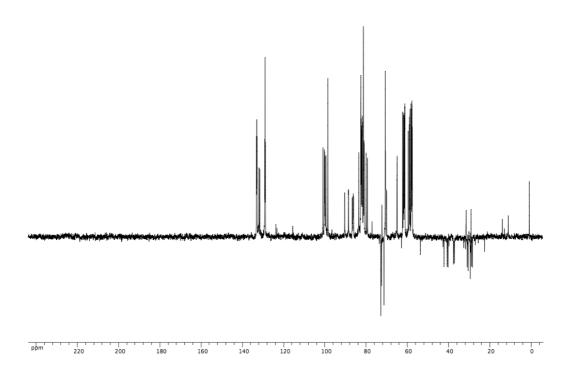
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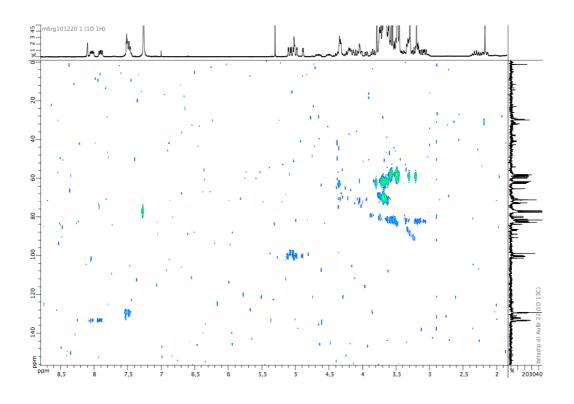
 $^{31}P{^{1}H}$ NMR spectrum of 2 recorded in CDCl₃ at 121.5 MHz.



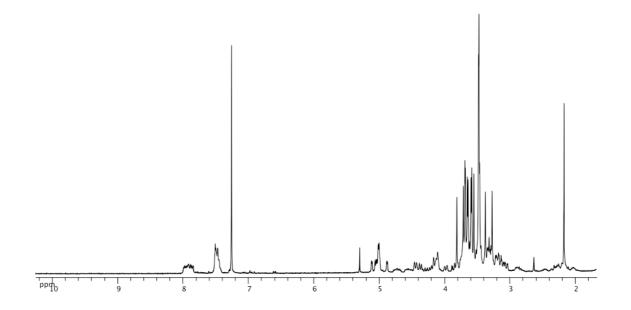
 $^{13}C\{^{1}H\}$ NMR spectrum of 2 recorded in CDCl₃ at 75.5 MHz.



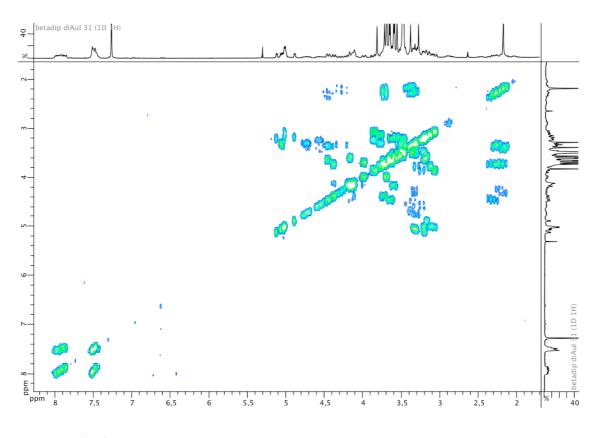
DEPT135 spectrum of 2 recorded in CDCl₃ at 75.5 MHz.



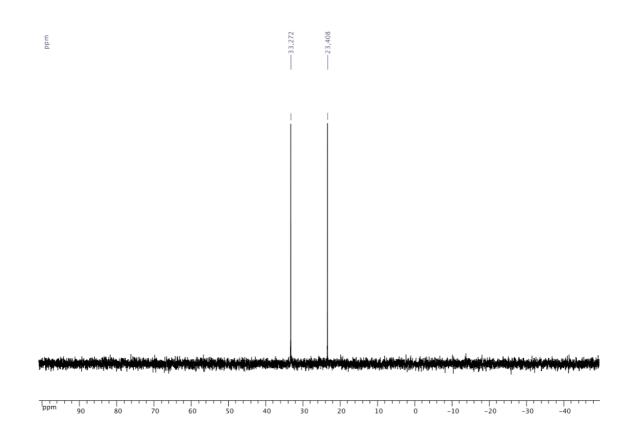
¹H-¹³C{¹H} HMQC spectrum of **2** recorded in CDCl₃ at 400.1 MHz.



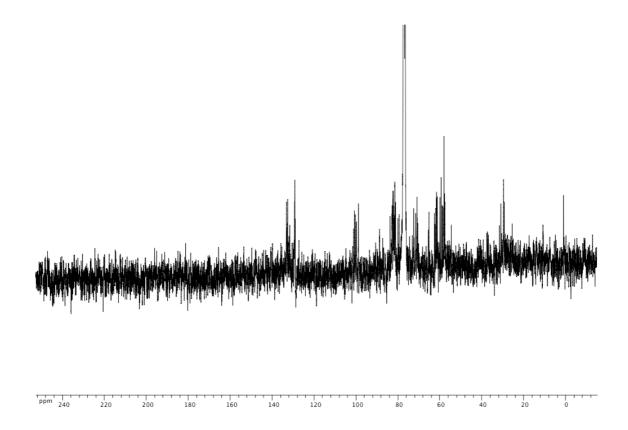
 1 H NMR spectrum of **3** recorded in CDCl₃ at 300.1 MHz.



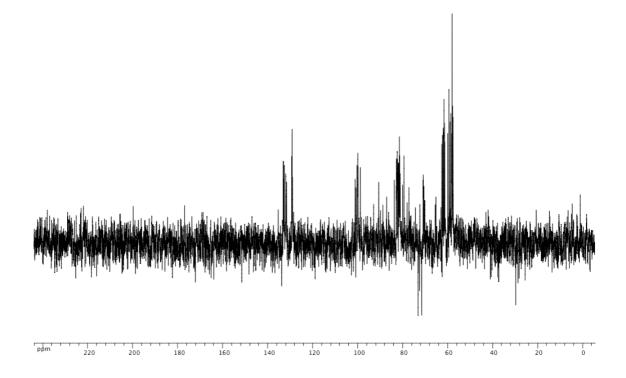
¹H-¹H COSY NMR spectrum of **3** recorded in CDCl₃ at 300.1 MHz.



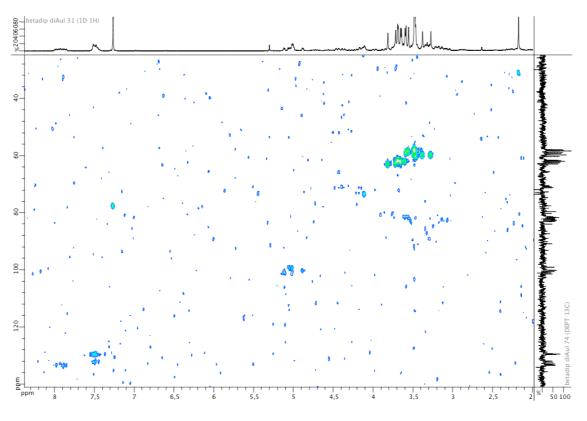
 $^{31}P\{^{1}H\}$ NMR spectrum of **3** recorded in CDCl₃ at 121.5 MHz.



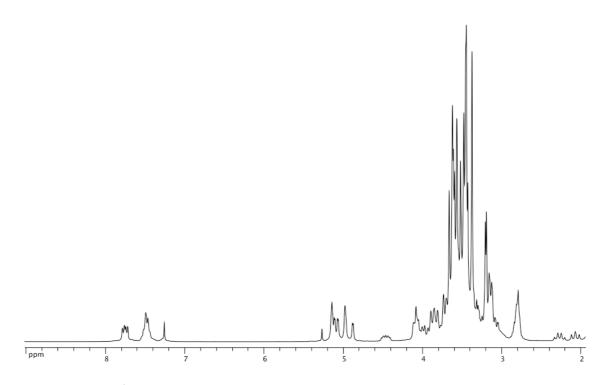
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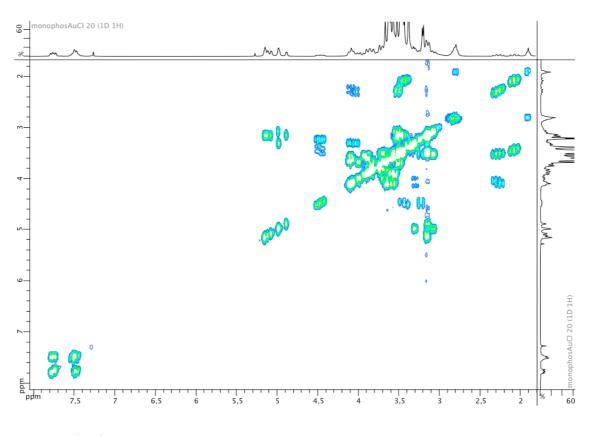
DEPT135 spectrum of $\mathbf{3}$ recorded in CDCl₃ at 75.5 MHz.



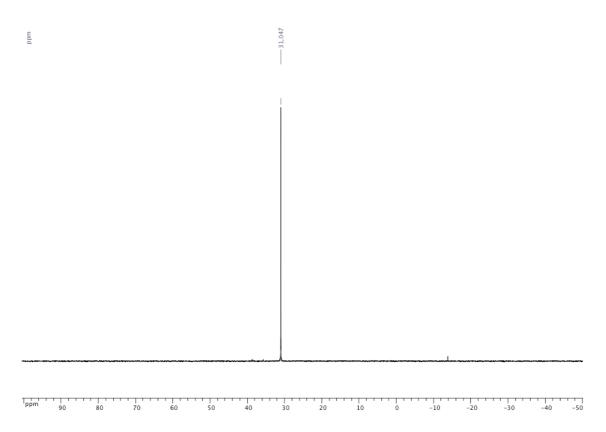
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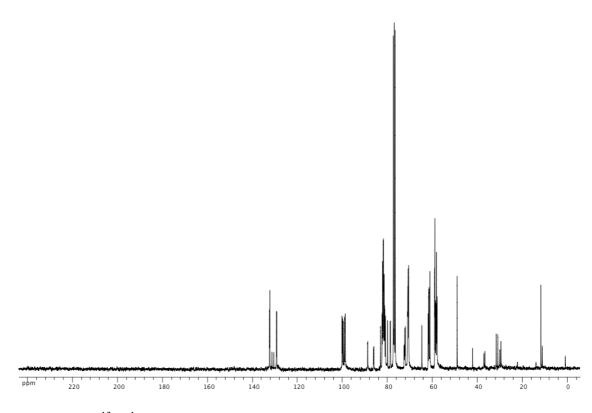
 ^1H NMR spectrum of **5** recorded in CDCl₃ at 300.1 MHz.



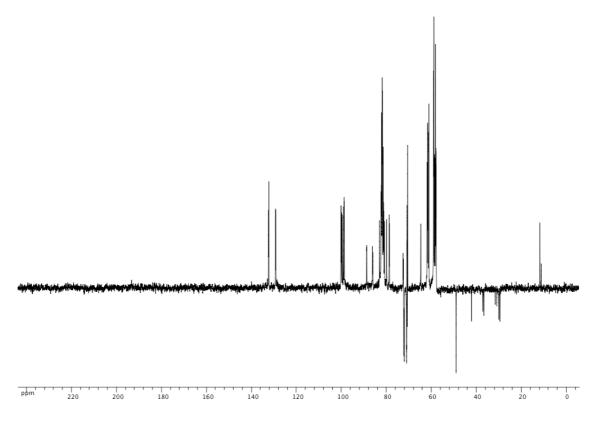
¹H-¹H COSY NMR spectrum of **5** recorded in CDCl₃ at 300.1 MHz.



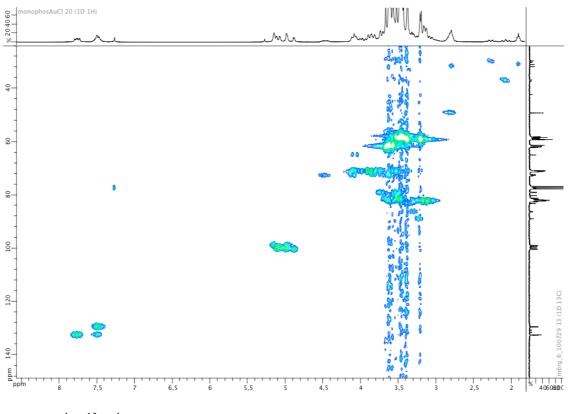
 $^{31}P\{^{1}H\}$ NMR spectrum of 5 recorded in CDCl₃ at 121.5 MHz.



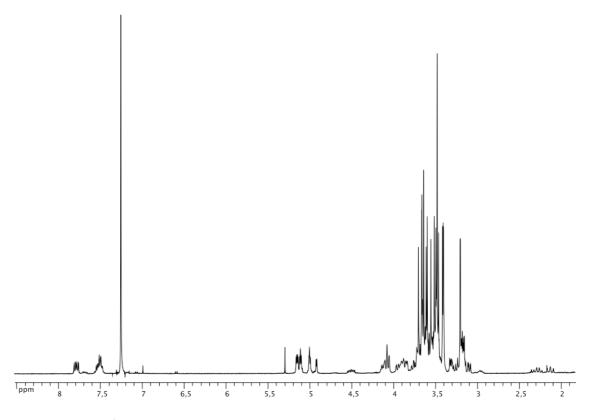
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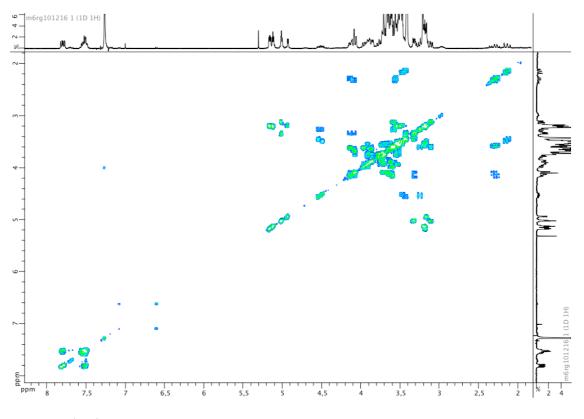
DEPT135 spectrum of 5 recorded in CDCl₃ at 75.5 MHz.



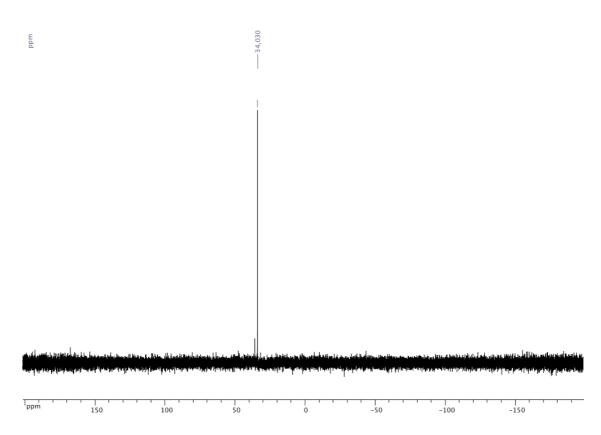
¹H-¹³C{¹H} HMQC spectrum of **5** recorded in CDCl₃ at 300.1 MHz.



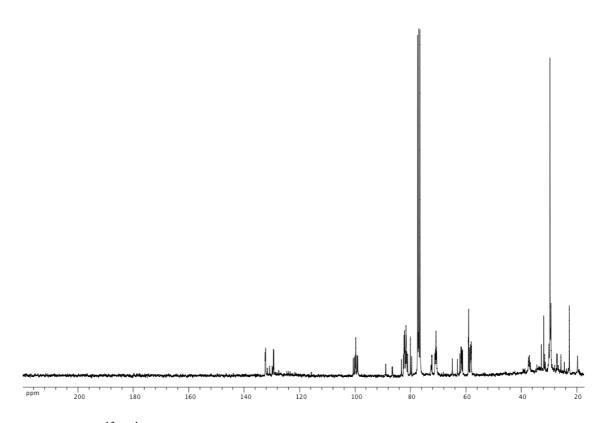
¹H NMR spectrum of **6** recorded in CDCl₃ at 400.1 MHz.



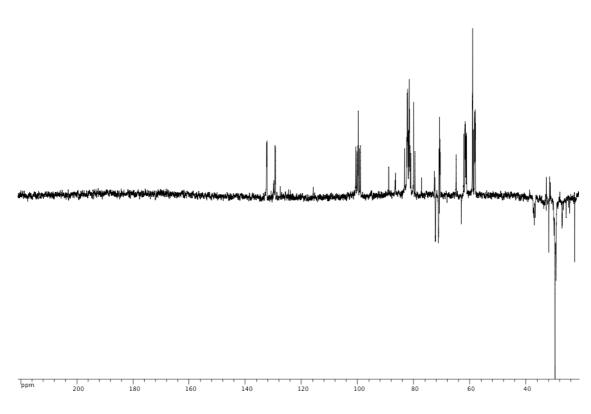
 $^1\text{H-}^1\text{H}$ COSY NMR spectrum of **6** recorded in CDCl₃ at 400.1 MHz.



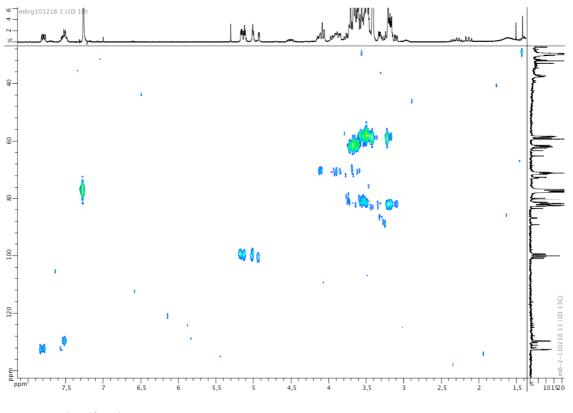
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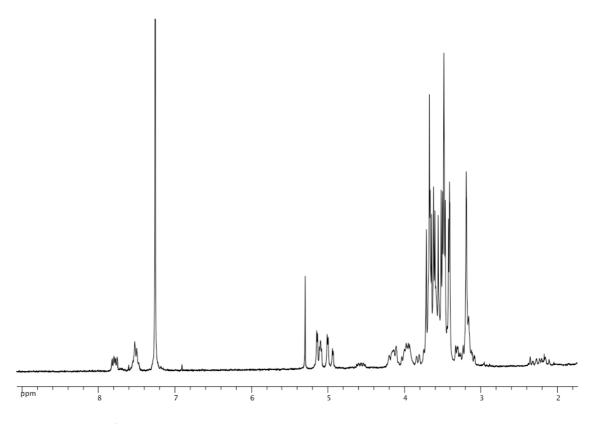
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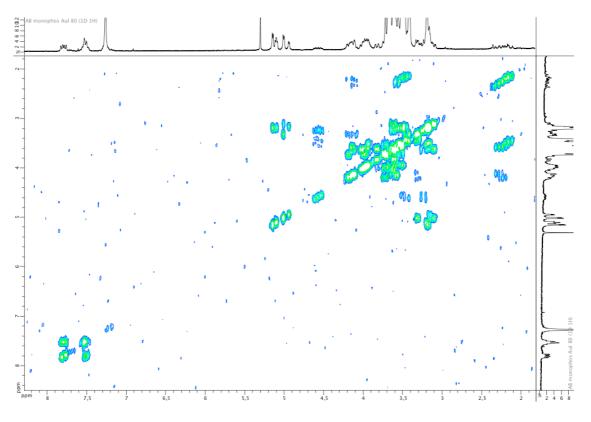
DEPT135 spectrum of 6 recorded in CDCl₃ at 75.5 MHz.



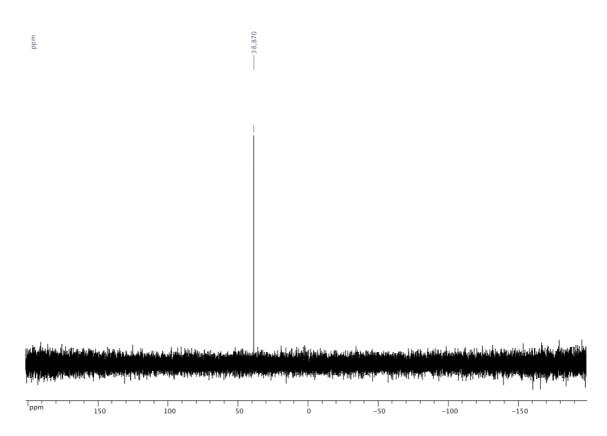
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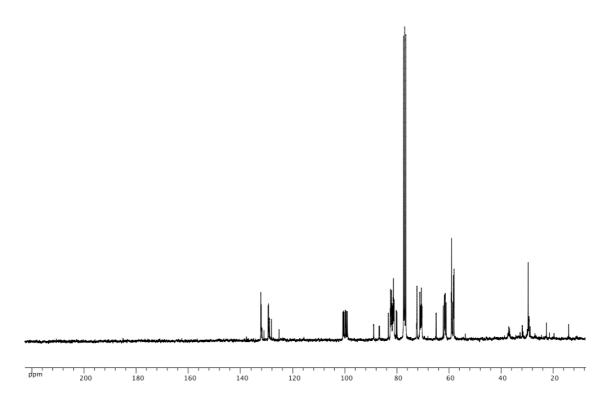
 $^1\mathrm{H}$ NMR spectrum of 7 recorded in CDCl3 at 300.1 MHz.



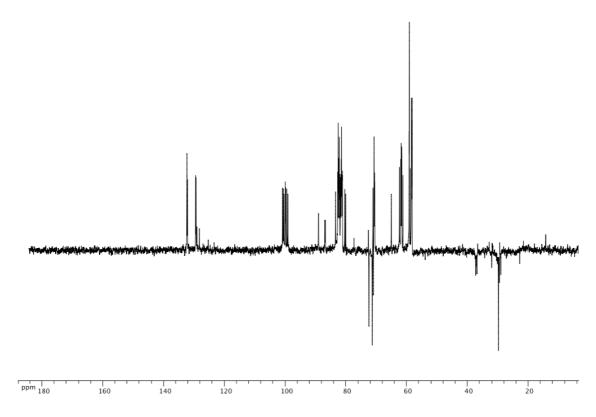
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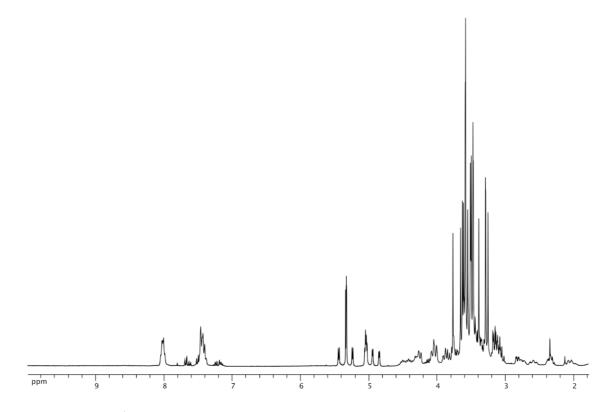
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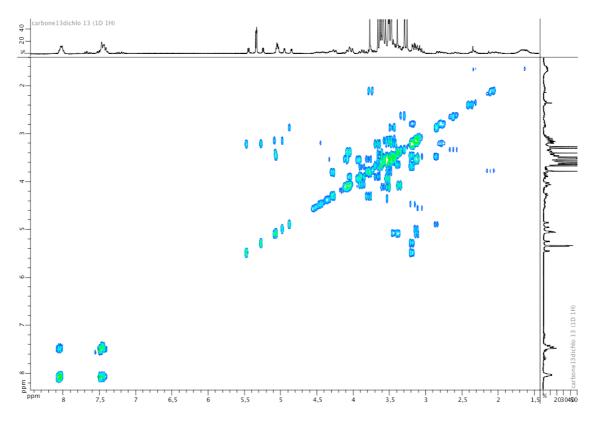
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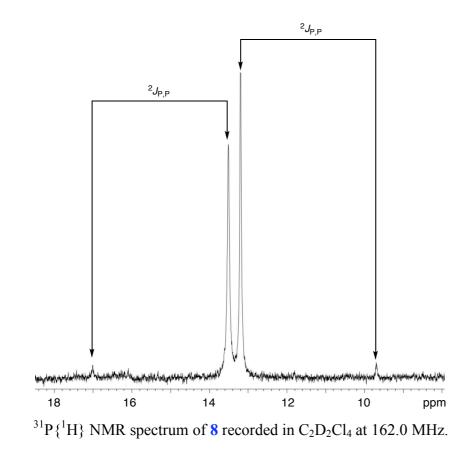
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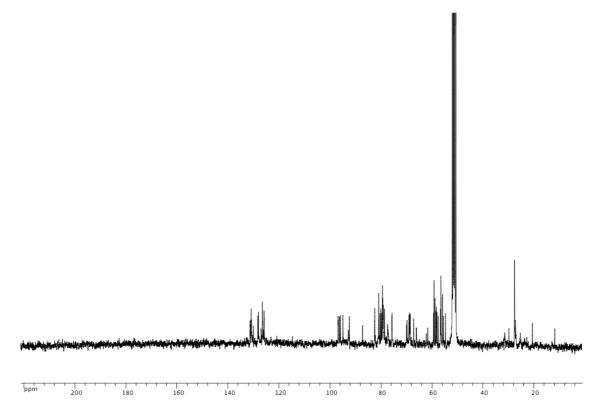


 $^1\mathrm{H}$ NMR spectrum of $\boldsymbol{8}$ recorded in CD_2Cl_2 at 300.1 MHz.

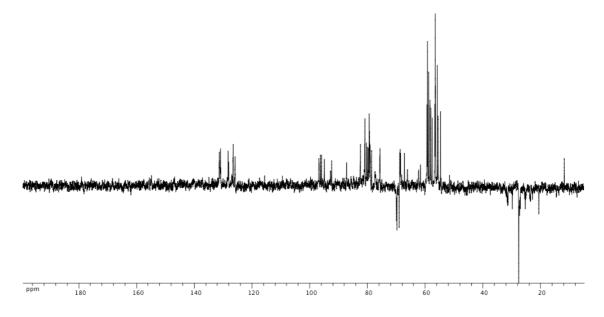


¹H-¹H COSY NMR spectrum of **8** recorded in CDCl₃ at 300.1 MHz.

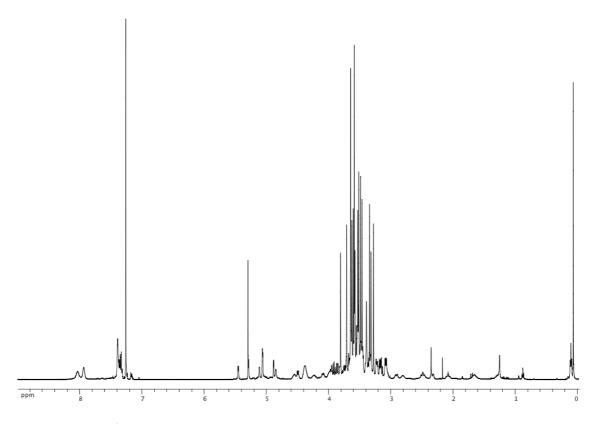




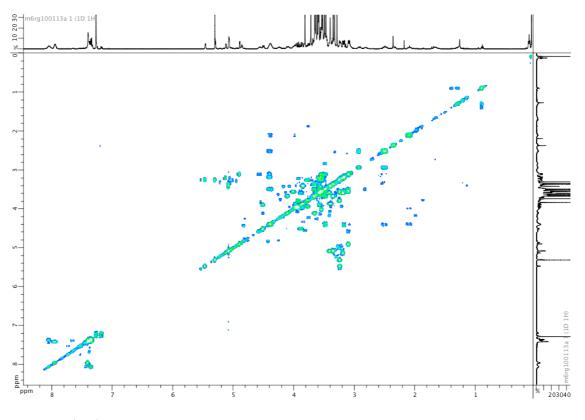
 $^{13}\mathrm{C}\{^{1}\mathrm{H}\}$ NMR spectrum of 8 recorded in CD₂Cl₂ at 75.5 MHz.



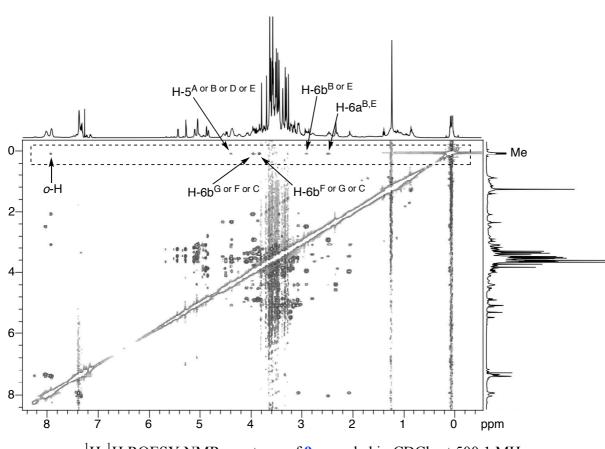
DEPT135 spectrum of 8 recorded in CD₂Cl₂ at 75.5 MHz.

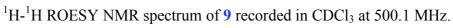


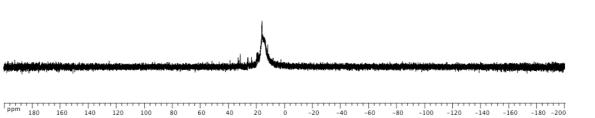
¹H NMR spectrum of **9** recorded in CDCl₃ at 500.1 MHz.



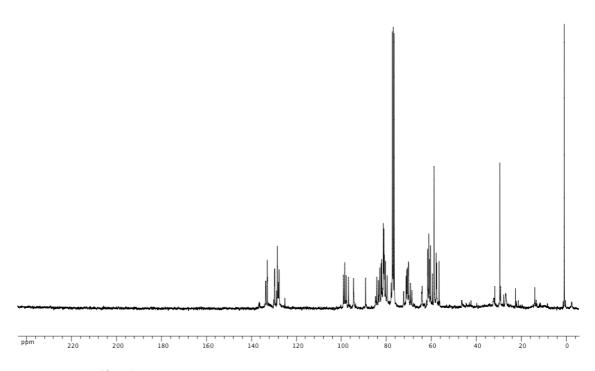
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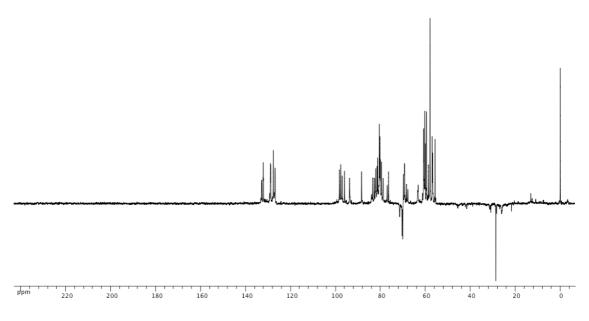




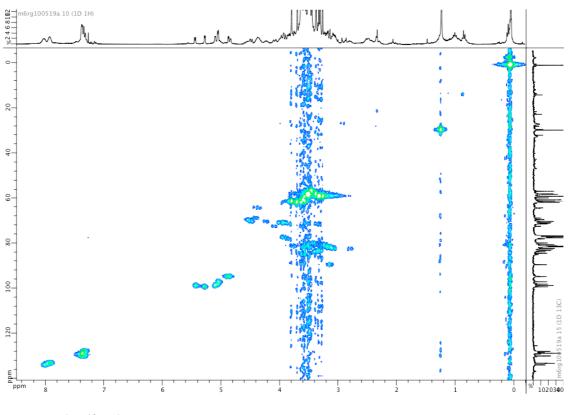
 $^{31}P\{^{1}H\}$ NMR spectrum of 9 recorded in CDCl₃ at 121.5 MHz (298 K).



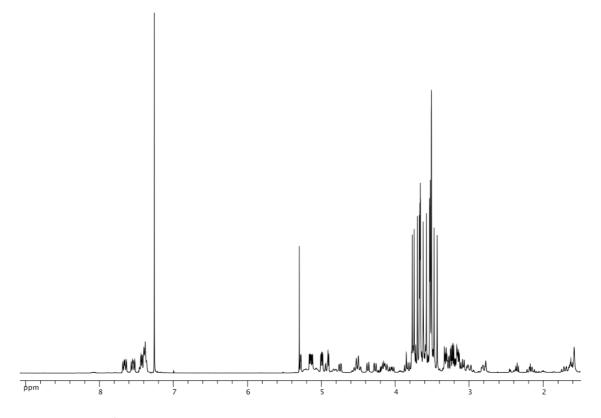
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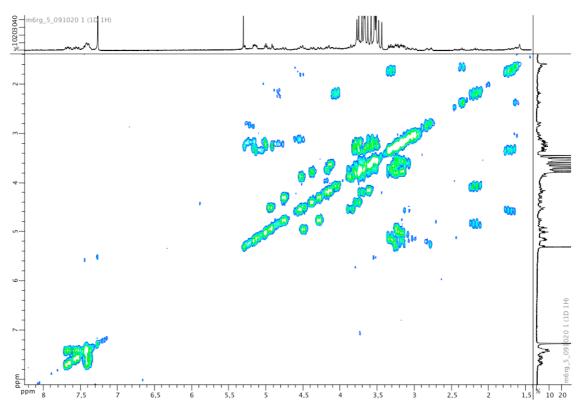
DEPT135 spectrum of 9 recorded in CDCl₃ at 75.5 MHz.



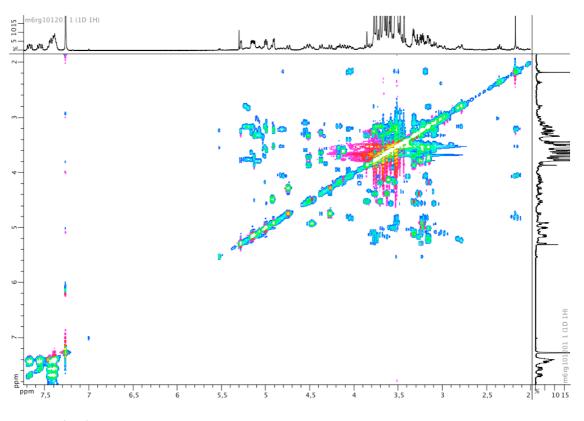
¹H-¹³C{¹H} HMQC spectrum of **9** recorded in CDCl₃ at 300.1 MHz.



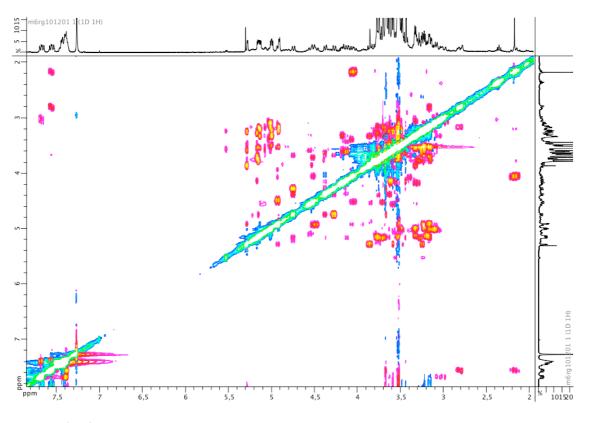
 1 H NMR spectrum of **10** recorded in CDCl₃ at 400.1 MHz.



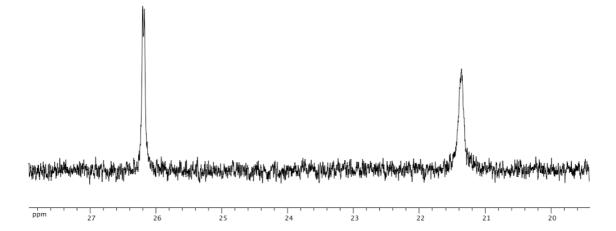
¹H-¹H COSY NMR spectrum of **10** recorded in CDCl₃ at 400.1 MHz.



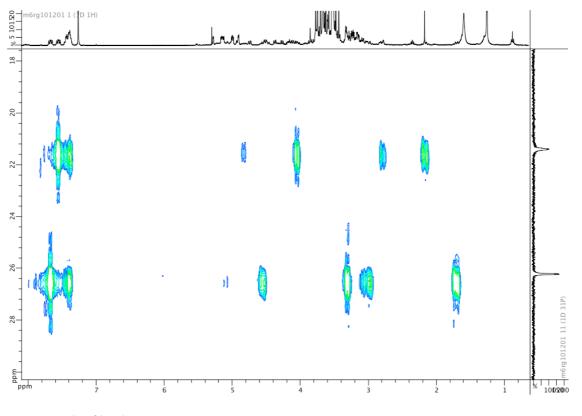
¹H-¹H TOCSY NMR spectrum of **10** recorded in CDCl₃ at 400.1 MHz.



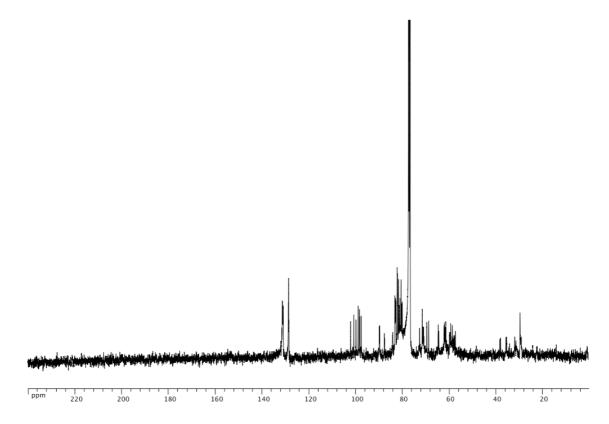
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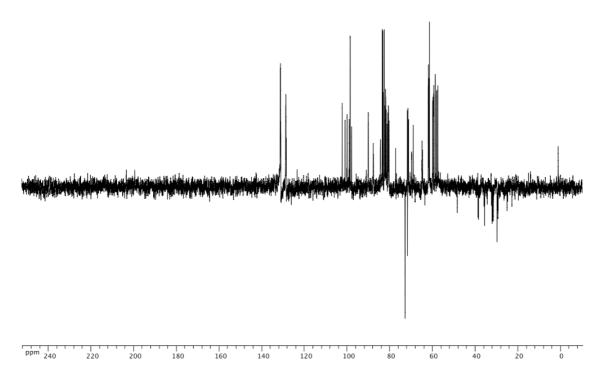
 $^{31}P{^{1}H}$ NMR spectrum of 10 recorded in CDCl₃ at 162.0 MHz (298 K).



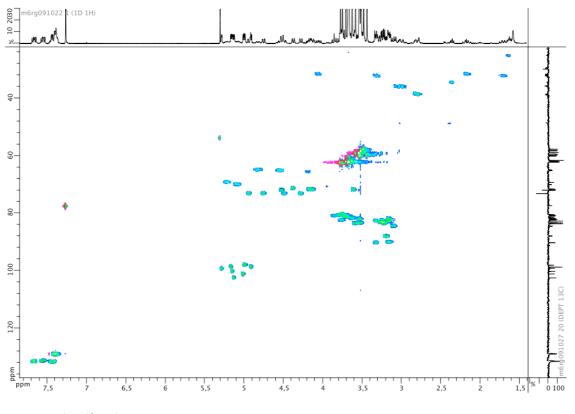
 ${}^{1}\text{H}-{}^{31}\text{P}\{{}^{1}\text{H}\}$ NMR spectrum of **10** recorded in CDCl₃ at 400.1 MHz.



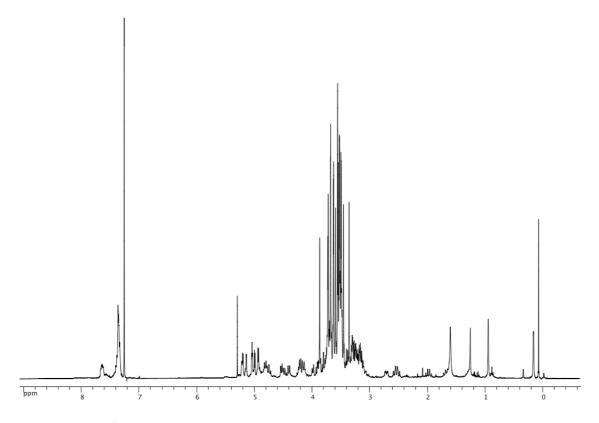
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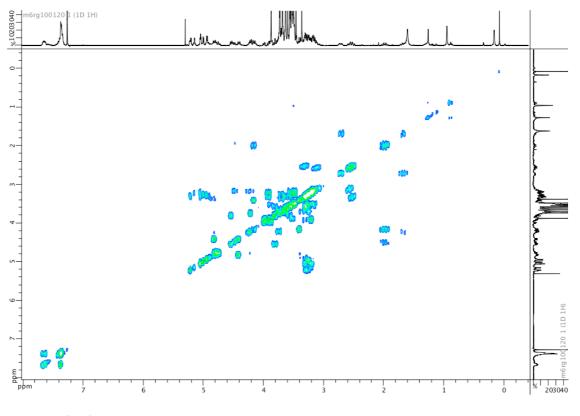
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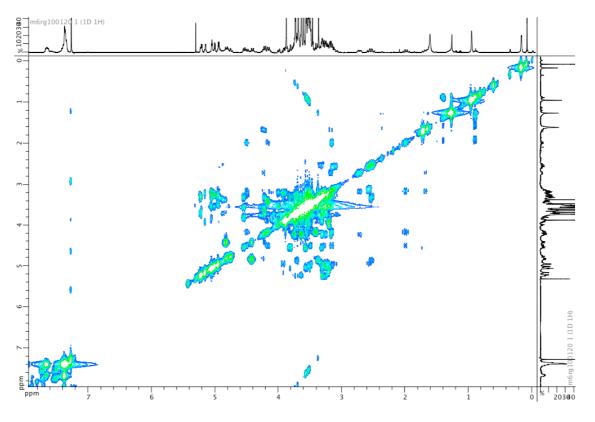
¹H-¹³C{¹H} HMQC spectrum of **10** recorded in CDCl₃ at 400.1 MHz.



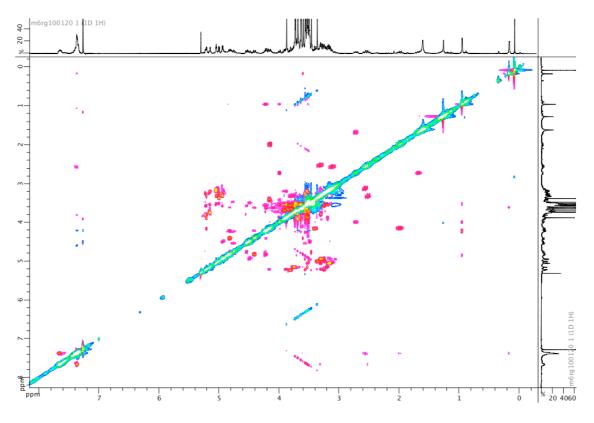
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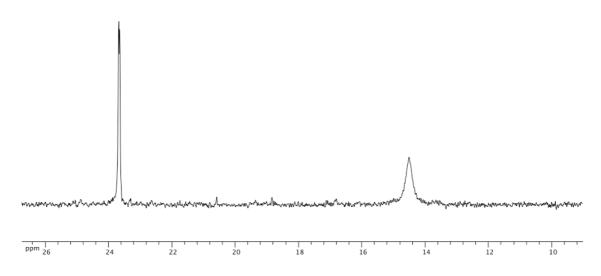
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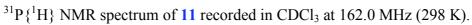


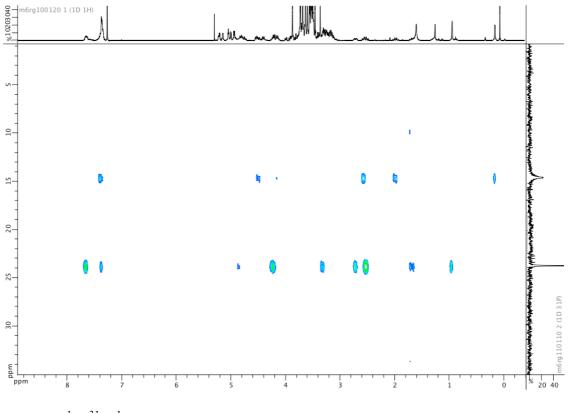
¹H-¹H TOCSY NMR spectrum of **11** recorded in CDCl₃ at 400.1 MHz.



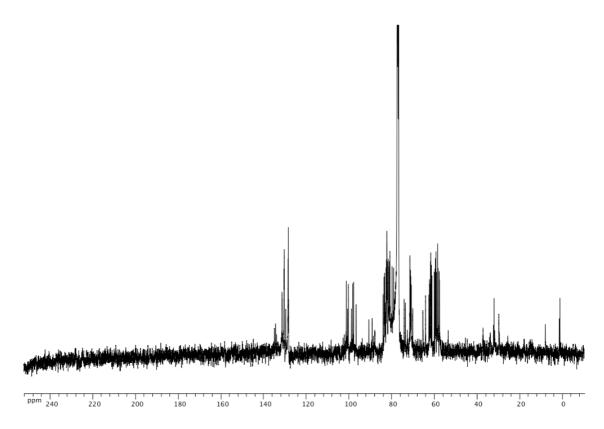
 $^{1}\text{H}\text{-}^{1}\text{H}$ ROESY NMR spectrum of **11** recorded in CDCl₃ at 400.1 MHz.



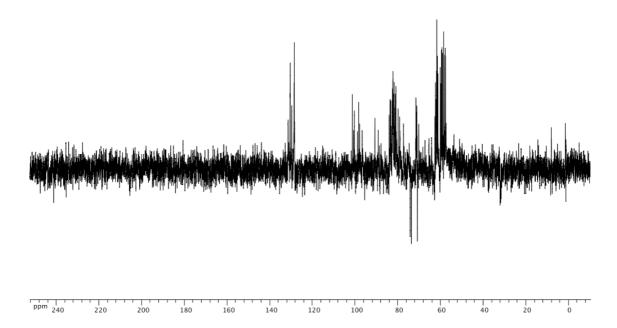




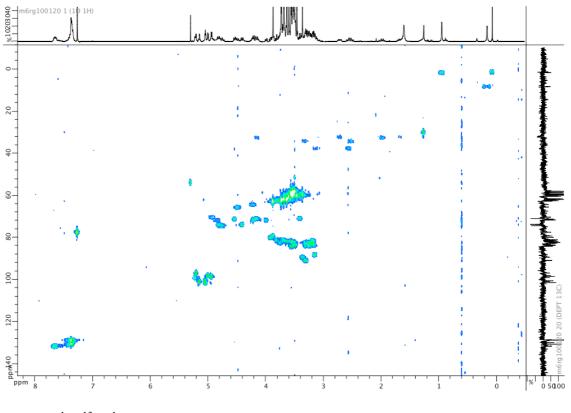
 ${}^{1}\text{H}-{}^{31}\text{P}\{{}^{1}\text{H}\}$ NMR spectrum of **11** recorded in CDCl₃ at 400.1 MHz.



 $^{13}C{^{1}H}$ NMR spectrum of 11 recorded in CDCl₃ at 100.6 MHz.



DEPT135 spectrum of 11 recorded in CDCl₃ at 100.6 MHz.



¹H-¹³C{¹H} HMQC spectrum of **11** recorded in CDCl₃ at 400.1 MHz.

General procedure for full assignment of the ¹H NMR signals of the glucose units.

The strategy applied for full structural assignment began with the differentiation between capped and non-capped C-6 carbon atoms by DEPT 135. These appear as two distinct sets of signals. The H-6 protons could then be identified using ${}^{1}\text{H}{}^{-13}\text{C}$ HMQC (Heteronuclear Multiple Quantum Coherence spectroscopy). By using TOCSY (TOtal Correlation SpectroscopY) and COSY (COrrelated SpectroscopY), each H-6 proton was correlated to the set of protons belonging to the same glucose residue. The connectivity between individual glucose units was then established via a ROESY (Rotating frame Overhauser Effect SpectroscopY) experiment showing the proximity between H-4_N and H-1_{N+1} protons (N and N+1 standing for neighbouring glucose moieties labeled in the alphabetical order).