

Bromide ion binding by a dinuclear gold(I) *N*-heterocyclic carbene complex: A spectrofluorescence and X-ray absorption spectroscopic study

Louise E. Wedlock, Jade B. Aitken, Susan J. Berners-Price and Peter J. Barnard

Electronic Supplementary Information

Contents

- 1. Table S1:** Constraints and restraints used in the MS EXAFS analysis of $1.(PF_6)_2$ in DMSO.
- 2. XFit Data Analysis –** Determinacy, Goodness-of-Fit and Monte-Carlo Error Analysis
- 3. Table S2:** Paths and importance factors from the analysis using MS model (Table 1), for the MS refinement of the EXAFS of $1.(PF_6)_2$.
- 4. 1H NMR spectrum of $[1](PF_6)_2$ in d_6 -DMSO**

Table S1 Constraints and restraints used in the MS EXAFS analysis of **1**.(PF₆)₂ in DMSO.

Mean displacement factor constraints (Å ²)	$\sigma_{(C1)}^2 = \sigma_{(C2)}^2$ $\sigma_{(N4)}^2 = \sigma_{(N5)}^2 = \sigma_{(N6)}^2 = \sigma_{(N7)}^2$ $\sigma_{(C8)}^2 = \sigma_{(C9)}^2 = \sigma_{(C10)}^2 = \sigma_{(C11)}^2$
Mean displacement factor restraints (Å ²)	$\sigma_i^2 > 0.0005 \{0.0001\}$ where <i>i</i> is all shells $\sigma_i^2 < 0.02 \{0.01\}$ where <i>i</i> is all shells
Bond length restraints (Å)	C(1) – Au(0) \sim 2.03 {0.05} C(2) – Au(0) \sim 2.03 {0.05} C(1) – N(5) \sim 1.38 {0.05} C(5) – C(9) \sim 1.40 {0.05} C(8) – C(9) \sim 1.35 {0.05} C(8) – N(4) \sim 1.40 {0.05} N(4) – C(1) \sim 1.38 {0.05} C(2) – N(6) \sim 1.38 {0.05} N(6) – C(11) \sim 1.40 {0.05} C(10) – C(11) \sim 1.35 {0.05} C(10) – N(7) \sim 1.40 {0.05} N(7) – C(2) \sim 1.38 {0.05}
Bond angle restraints (°)	Au(0) – C(1) – N(5) \sim 127 {5} Au(0) – C(1) – N(4) \sim 127 {5} C(1) – N(5) – C(9) \sim 110 {5} N(5) – C(1) – N(4) \sim 106 {5} N(5) – C(9) – C(8) \sim 107 {5} C(9) – C(8) – N(4) \sim 107 {5} C(8) – N(4) – C(1) \sim 110 {5} Au(0) – C(2) – N(6) \sim 127 {5} Au(0) – C(2) – N(6) \sim 127 {5} C(2) – N(6) – C(11) – N(6) \sim 110 {5} N(6) – C(2) – N(7) \sim 106 {5} N(6) – C(11) – C(10) \sim 107 {5} C(11) – C(10) – N(7) \sim 107 {5} C(10) – N(7) – C(2) \sim 110 {5}
Symmetry restraints	$z_i = 0$, where <i>i</i> is all shells except Au(3) $x_{Au(3)} = 0$ $y_{Au(3)} = 0$ $x_{N(4)} = x_{N(5)}$ $x_{N(6)} = x_{N(7)}$ $x_{C(8)} = x_{C(9)}$ $x_{C(10)} = x_{C(11)}$ $x_{C(1)} = -x_{C(2)}$ $x_{N(4)} = -x_{N(7)}$ $x_{N(5)} = -x_{N(6)}$ $x_{C(8)} = -x_{C(10)}$ $x_{C(9)} = -x_{C(11)}$ $y_{C(1)} = y_{C(2)}$ $y_{N(4)} = -y_{N(5)}$ $y_{N(6)} = -y_{N(7)}$ $y_{C(8)} = -y_{C(9)}$ $y_{C(10)} = -y_{C(11)}$ $y_{N(6)} = y_{N(5)}$ $y_{N(7)} = y_{N(4)}$ $y_{C(10)} = y_{C(8)}$ $y_{C(11)} = y_{C(9)}$
Occupancy (<i>N</i>) restraints	$N_i = 1$, where <i>i</i> is all shells

XFit Data Analysis – Determinacy, Goodness-of-Fit and Monte-Carlo Error Analysis

Determinacy

The number of parameters being fitted, p , compared to the number of independent information data points (independent points in the EXAFS plus the number of independent structural parameters), N_i , was calculated to give the degree of determinacy N_i/p . If this ratio is < 1 , then the model is considered to be underdetermined and a unique fit is not possible. In all cases the ratio was > 1 and, hence, the models were overdetermined. The value of N_i is given by:¹

$$N_i = 2(\Delta r)(\Delta k)/\pi + \Sigma[D(N-2)+1] \quad (1)$$

where D is the number of dimensions in which the refinement takes place and N is the number of atoms in the unit.²

Goodness-of-Fit (Residual)

The method of determining the goodness of fit was through an R value where R is given by:

$$R = (\chi^2/\chi^2_{\text{calculated}=0})^{1/2} \quad (2)$$

where χ^2 is the quantity minimized during the refinement and $\chi^2_{\text{calculated}=0}$ is the value of χ^2 when the calculated EXAFS is uniformly 0.³ Residual R values of $\leq 20\%$ were considered reasonable for MS models, and relatively high ($>20\%$) values of R are explained by the exclusion of multiple-scattering contributions on the SS models.^{2,4}

Monte-Carlo Error Analysis

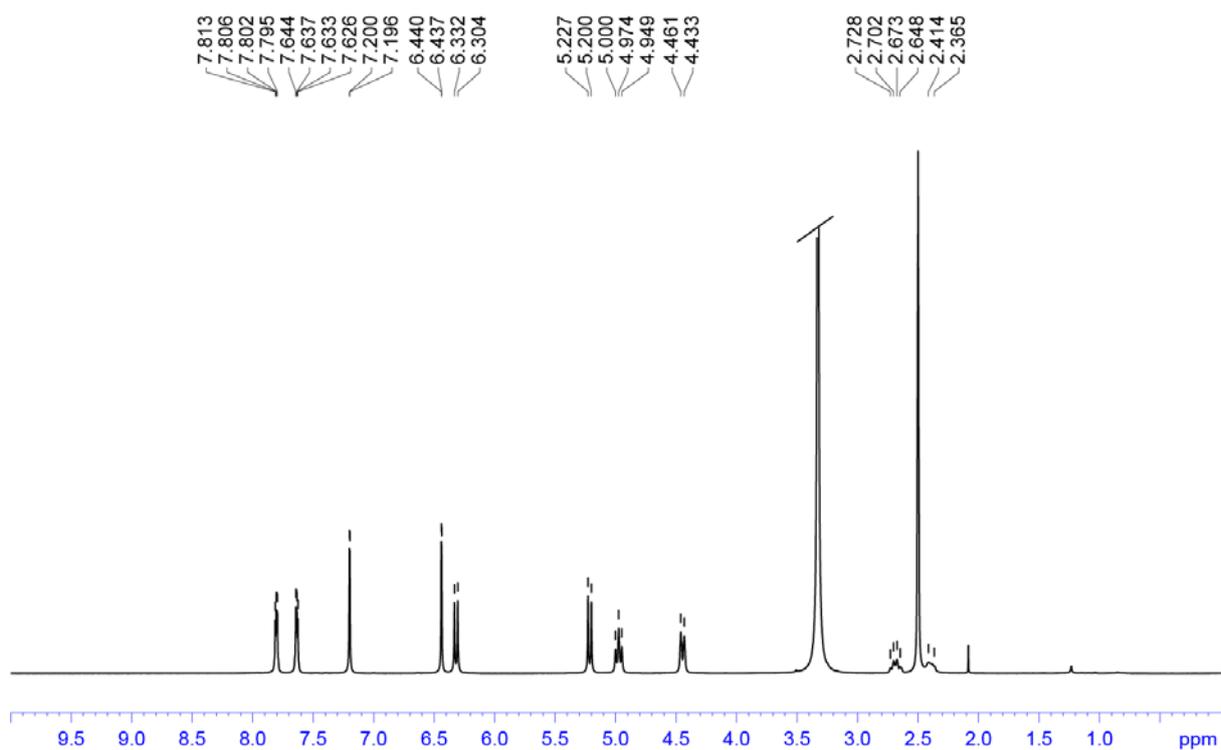
Monte Carlo analyses were conducted to estimate the rms deviations in final parameters arising from the noise in the data. Two consecutive sets of 16×16 Monte-Carlo cycles were calculated and the resulting rms errors were combined with systematic errors to determine the final error estimates. The probable errors in the Au-C and Au•••Au bond lengths were estimated as $[\sigma_r^2 + \sigma_s^2]^{1/2}$, where σ_r and σ_s represent contributions from the random and systematic errors, respectively. The random (statistical) errors due to noise in the data were estimated by Monte Carlo calculations,³ and the systematic errors were assigned a conservative consensus value, 0.02 \AA .⁵

The following applies to tables of multiple scattering pathways presented in the supporting information: ^aThe number of legs represents the path travelled by the photoelectron originating from and returning to the XAFS absorber Au(0). ^bThe total distance travelled by the photoelectron (R_{eff}) is twice the value of R_{as} . ^cThe importance factor (given to 2 d.p.), represents the percent contribution of a path relative to the strongest path Au(0) → C(1) → Au(0), including contribution from the Debye-Waller Factors. All pathways have a maximum of 5 legs per MS pathway, curve and plane wave filters of 3% and 2%, respectively, $R_{\text{eff}} \leq 10$ Å.

Table S2 Paths and importance factors from MS analysis (Table 1), for the MS refinement of the EXAFS of **1**.(PF₆)₂.

Atoms in MS Pathway	Legs ^a	Deg	R_{as} (Å) ^b	Importance Factor ^c
Au(0) → C(1) → Au(0)	2	2	2.03	100.00%
Au(0) → Au(3) → Au(0)	2	1	3.02	20.45%
Au(0) → N(4) → Au(0)	2	4	3.06	100.00%
Au(0) → N(6) → C(2) → Au(0)	3	8	3.23	100.00%
Au(0) → C(2) → N(6) → C(2) → Au(0)	4	4	3.40	30.40%
Au(0) → C(2) → C(1) → Au(0)	3	2	4.06	17.00%
Au(0) → C(1) → Au(0) → C(2) → Au(0)	4	2	4.06	32.47%
Au(0) → C(1) → Au(0) → C(1) → Au(0)	4	2	4.06	9.57%
Au(0) → N(4) → N(5) → Au(0)	3	4	4.16	6.03%
Au(0) → C(10) → Au(0)	2	4	4.25	28.73%
Au(0) → C(9) → C(1) → Au(0)	3	8	4.28	64.64%
Au(0) → C(2) → N(6) → N(7) → Au(0)	4	8	4.33	21.17%
Au(0) → C(1) → C(9) → C(1)	3	4	4.30	42.92%
Au(0) → C(10) → N(7) → Au(0)	3	8	4.36	43.09%
Au(0) → N(4) → C(8) → C(1) → Au(0)	4	8	4.38	55.71%
Au(0) → N(7) → C(2) → N(7) → Au(0)	4	4	4.43	24.06%
Au(0) → N(5) → C(1) → N(4) → Au(0)	4	4	4.43	5.63%
Au(0) → C(1) → N(4) → N(5) → C(1) → Au(0)	5	4	4.50	11.14%
Au(0) → N(6) → C(11) → N(6) → Au(0)	4	4	4.46	22.57%
Au(0) → C(8) → N(4) → C(1) → Au(0)	4	8	4.53	39.22%
Au(0) → C(1) → C(9) → N(5) → C(1) → Au(0)	5	8	4.55	41.91%
Au(0) → C(1) → N(4) → C(1) → N(5) → Au(0)	5	8	4.60	12.52%
Au(0) → N(4) → C(1) → N(4) → C(1) → Au(0)	5	8	4.60	28.98%
Au(0) → N(5) → C(9) → N(5) → C(1) → Au(0)	5	8	4.63	36.46%
Au(0) → C(9) → N(4) → Au(0)	3	8	4.77	9.88%
Au(0) → C(2) → C(11) → N(7) → Au(0)	4	8	4.79	10.43%
Au(0) → N(7) → C(10) → N(6) → Au(0)	4	8	4.87	4.55%
Au(0) → C(9) → C(8) → Au(0)	3	4	4.92	5.45%
Au(0) → C(8) → N(5) → C(1) → Au(0)	4	8	4.94	11.53%
Au(0) → C(11) → C(10) → C(2) → Au(0)	4	8	4.95	13.32%

[1](PF₆)₂



¹H NMR Spectrum of [1](PF₆)₂ in d₆-DMSO

References

1. N. Binsted, R. W. Strange and S. S. Hasnain, *Biochemistry*, 1992, **31**, 12117-12125.
2. A. Levina, R. S. Armstrong and P. A. Lay, *Coord. Chem. Rev.*, 2005, **249**, 141-160.
3. P. J. Ellis and H. C. Freeman, *J. Synchrotron Radiat.*, 1995, **2**, 190-195.
4. A. M. Rich, R. S. Armstrong, P. J. Ellis, H. C. Freeman and P. A. Lay, *Inorg. Chem.*, 1998, **37**, 5743-5753.
5. S. J. Gurman, *J. Synchrotron Radiat.*, 1995, **2**, 56-63.