

Supporting Information

Evaluation of Arene Ruthenium(II) *N*-heterocyclic Carbene Complexes as Organometallics Interacting with Thiol and Selenol Containing Biomolecules

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1) Computational Chemistry

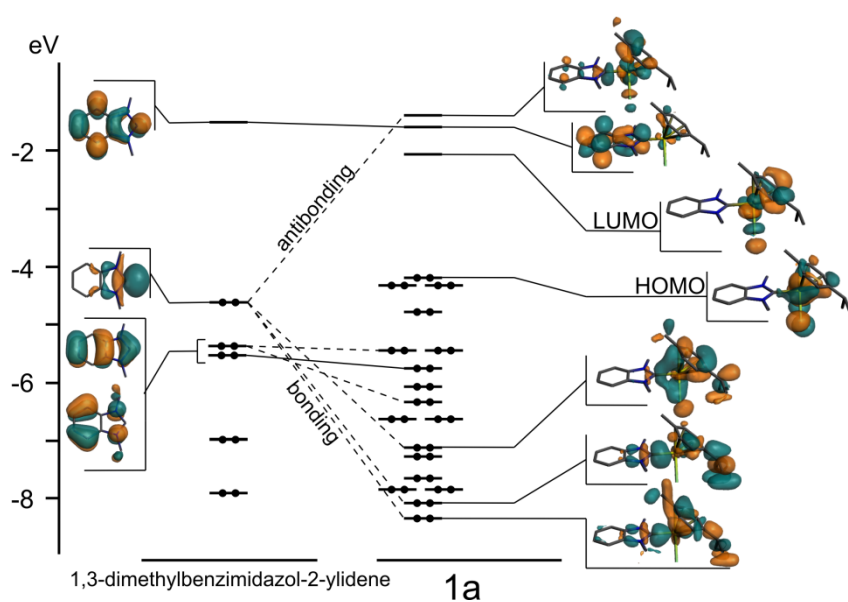


Figure S1. Molecular orbital diagram denoting the (*p*-cymene)(NHC)RuCl₂ interaction, where contribution larger than 20% to the **1a** molecular orbitals are shown. The s-donor interaction exhibits the larger bonding/antibonding gap reflecting its main role in such interaction. This diagram is representative and can be extended to **2a**, **3a** and **4a**.

	1a	2a	3a	4a
Distances (Å)				
Ru-NHC	2.061	2.062	2.090	2.063
Ru(η^6 - <i>p</i> -cymene)	1.796	1.797	1.769	1.796
Ru-Cl	2.451	2.455	2.457	2.458
BDE (kcal/mol)				
Ru-NHC	61.34	64.01	59.46	62.54
Ru(η^6 - <i>p</i> -cymene)	58.44	56.46	57.61	59.36
Ru-Cl (each)	126.58	125.15	124.72	124.90
Pop. Analysis				
Ru(II)	+0.475	+0.478	+0.474	+0.467
NHC	+0.343	+0.349	+0.333	+0.314
<i>p</i> -cymene	+0.183	+0.172	+0.188	+0.214
[2Cl] ²⁻	-1.001	-0.999	-0.995	-0.995

Table S1. Selected distances, bond dissociation energies and population analysis for **1a** - **4a**

Theoretical considerations concerning hydrolysis

Compounds **1a** - **4a** in aqueous medium can undergo hydrolysis leading to a ligand exchange involving the chlorido ligands to form the respective aquo-complexes, namely $[(p\text{-cymene})\text{RuCl}(\text{H}_2\text{O})(\text{NHC})]^+$, and $[(p\text{-cymene})(\text{NHC})\text{Ru}(\text{H}_2\text{O})_2]^{2+}$, where the structures vary slightly from the chlorido counterparts (see table S2 and figures S2, S3). Interestingly, a similar theoretical evaluation of the BDE for the involved arene and NHC ligand in the aquo-complexes reveals an enhanced bonding strength amounting to values in the 71 - 99 kcal/mol range for both organometallic fragments driven by the difference in the Cl^- and H_2O ligands. The latter point suggests that the $\text{Ru}(\eta^6\text{-}p\text{-cymene})$ or Ru-NHC interaction can be modulated by the presence of different ligands into the coordination sphere of Ru(II) , becoming more labile under the presence of ligands with stronger interaction towards the ruthenium center.

$[(p\text{-cymene})\text{Ru}(\text{H}_2\text{O})_2(\text{NHC})]^{2+}$	1a'	2a'	3a'	4a'
Distances (Å)				
Ru-NHC	2.037	2.041	2.041	2.052
$\text{Ru}(\eta^6\text{-}p\text{-cymene})$	1.714	1.706	1.714	1.700
Ru-OH ₂	2.161	2.176	2.157	2.170
BDE (kcal/mol)				
Ru-NHC	85.46	85.89	88.27	98.96
$\text{Ru}(\eta^6\text{-}p\text{-cymene})$	96.77	91.36	89.43	84.20
Ru-OH ₂ (each)	23.79	21.69	22.12	25.03

$[(p\text{-cymene})\text{RuCl}(\text{H}_2\text{O})(\text{NHC})]^+$	1a''	2a''	3a''	4a''
Distances (Å)				
Ru-NHC	2.070	2.084	2.105	2.078
$\text{Ru}(\eta^6\text{-}p\text{-cymene})$	1.807	1.750	1.795	1.803
Ru-OH ₂	2.272	2.275	2.271	2.276
Ru-Cl	2.451	2.455	2.451	2.441
BDE (kcal/mol)				
Ru-NHC	73.13	72.96	71.46	75.72
$\text{Ru}(\eta^6\text{-}p\text{-cymene})$	78.69	77.04	76.47	77.87
Ru-OH ₂	20.01	20.09	19.51	19.57
Ru-Cl	216.47*	213.68*	212.37*	209.04*

Table S2. Selected distances, bond dissociation energies for the di-aquo and mono-aquo complexes derived from **1a** - **4a**, namely **1a'** - **4a'** and **1a''** - **4a''** respectively.

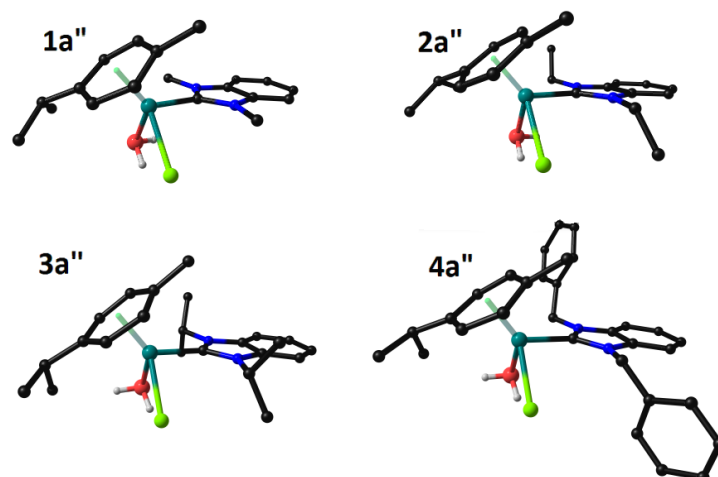


Fig S2: Optimized structures (minimum energy conformation) of the mono-aquo counterparts of **1a- 4a**

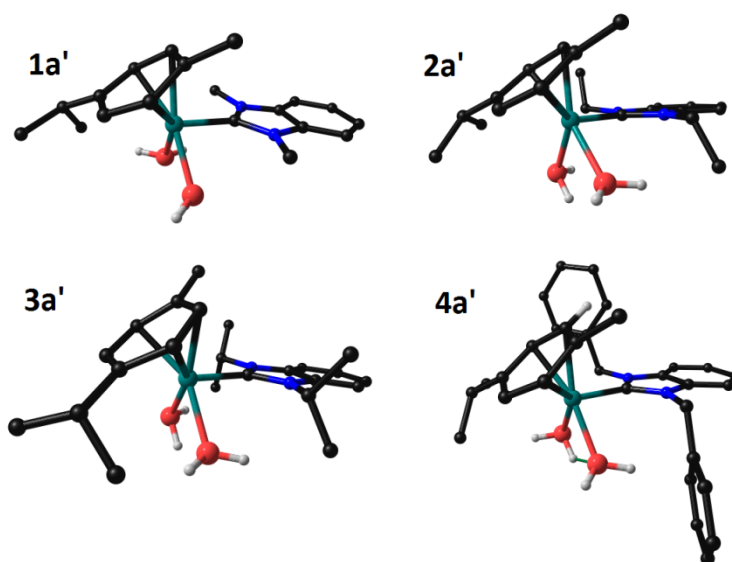


Fig S3: Optimized structures of the di-aquo counterparts of **1a- 4a**

2) Stability in Solution (UV-Vis spectroscopy)

Complex **4a** was dissolved in a 0.5% solution of acetonitrile in water and its UV-Vis spectra were monitored for 10 hours showing no changes in shape and absorption (figure S4). The spectrum of the ligand **4** shows an absorption band between 235 and 295 nm, while for complex **4a** a band is observed at 230 nm and another more intense between 280 and 300 nm corresponding to a LLCT. Due to these spectral differences between it is possible to distinguish between the free ligand **4** and the ruthenium complex **4a**. Exchange of the chlorido ligands from ruthenium arene complexes by water molecules has been reported in the literature.^{1,2} This would involve spectral changes in the range of 300-350 nm. However, in the case of **4a** this wavelength range shows absorbances of the respective NHC ligand, which hamper an evaluation of possible Cl⁻/ water exchanges by this technique.

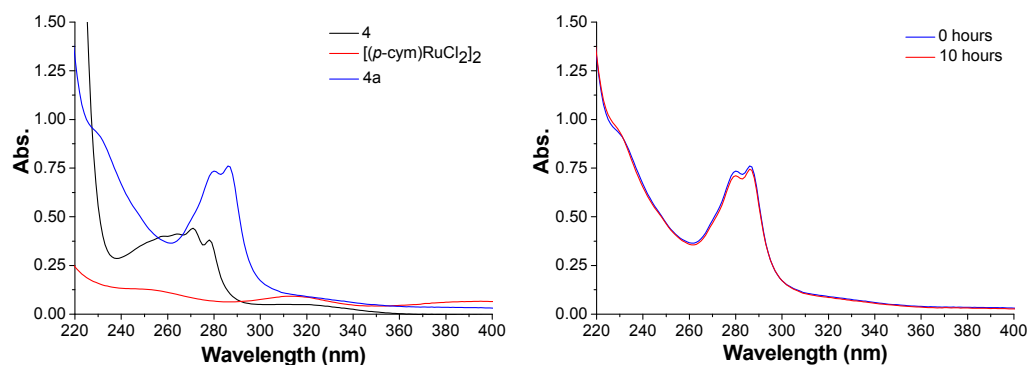


Figure S4. (right) UV-Vis spectra of ligand **4**, complex **4a** and $[(p\text{-cym})\text{RuCl}_2]_2$ in a 0.5% acetonitrile solution in H_2O ; (left) UV-Vis spectra of complex **4a** at time 0 and after 10 hours in a 0.5% acetonitrile solution in H_2O .

3) Interaction with *N*-acetylcysteine (^1H NMR spectroscopy)

Complex **1a** was incubated with *N*-acetylcysteine in $\text{DMSO-}d_6$ over 24 hours and ^1H NMR spectra were measured. For assigning the structural changes spectra of *N*-acetylcysteine, **1**³ and **1a** in $\text{DMSO-}d_6$ were measured. The spectrum of *N*-acetylcysteine is shown in figure S5, the spectra of **1** and **1a** (see figure S6) were evaluated as follows:

1: ^1H NMR ($\text{DMSO-}d_6$): (ppm) 9.64 (s, 1H, N-CH-N), 8.02 (dd, 2H, $^3J_{\text{HH}}=3.2$ Hz, $^3J_{\text{HH}}=6.3$ Hz, $\text{ArH}_{5/6}$), 7.71 (dd, 2H, $^3J_{\text{HH}}=3.2$ Hz, $^3J_{\text{HH}}=6.3$ Hz, $\text{ArH}_{4/7}$), 4.08 (s, 6H, -NCH₃).

1a: ^1H NMR ($\text{DMSO-}d_6$): (ppm) 7.81 - 7.72 (m, 2H, C_6H_4), 7.50 - 7.43 (m, 2H, C_6H_4), 6.45 (d, 1H, MeC_6H_4 -), 6.29 (d, 1H, MeC_6H_4 -), 6.18 (m, 2H, MeC_6H_4 -), 4.10 (s, 6H, -NCH₃), 3.34 (hept, 1H, CHMe_2 , $^3J_{\text{HH}}=7.0$ Hz), 2.07 (s, 3H, $\text{C}_6\text{H}_4\text{CH}_3$), 1.13 (d, 6H, $(\text{CH}_3)_2\text{CHC}_6\text{H}_4$ -, $^3J_{\text{HH}}=7.0$ Hz).

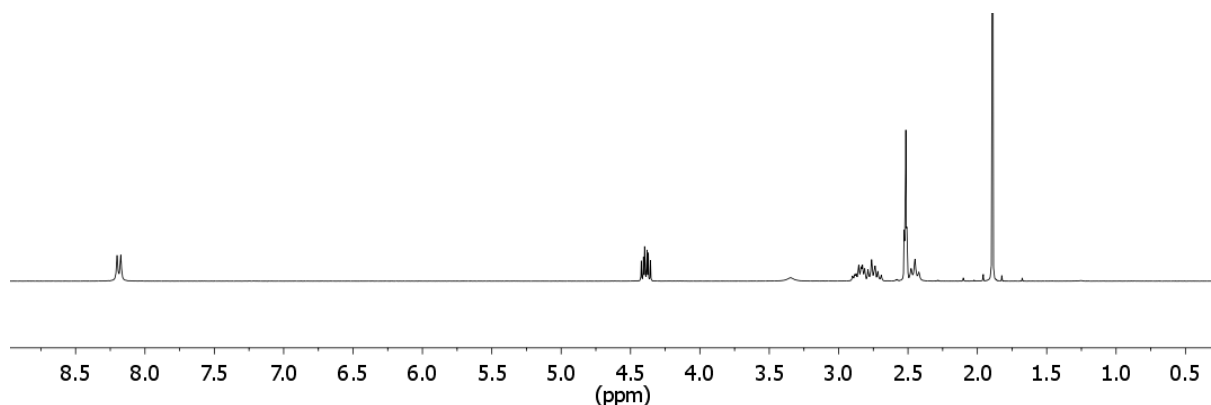


Figure S5 ^1H NMR spectrum of *N*-acetylcysteine in $\text{DMSO-}d_6$.

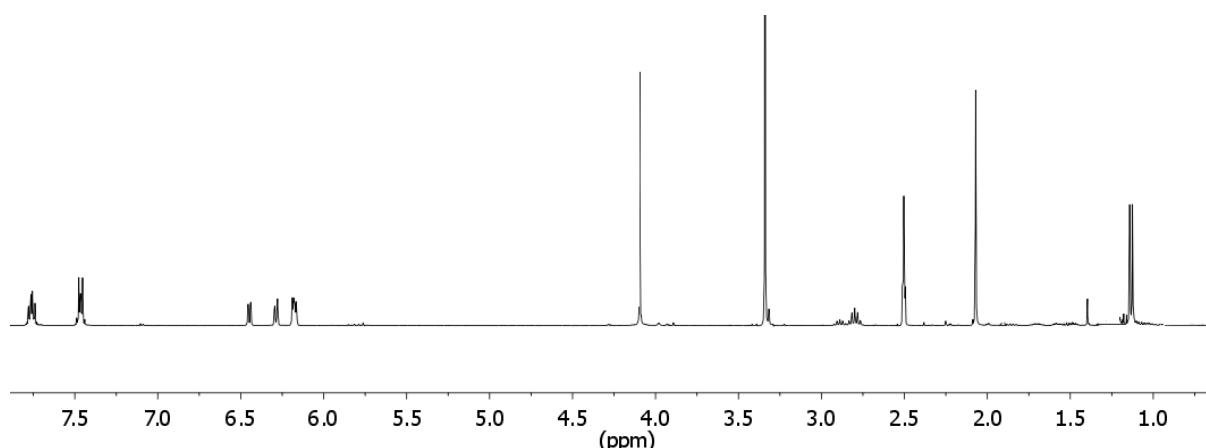


Figure S6 ^1H NMR spectrum of compound **1a** in $\text{DMSO}-d_6$

Upon exposure of the mixture of **1a** and *N*-acetylcysteine the following spectral changes were noted after 24 h (see figure S7 for the spectra after 5 min and 24 hours):

A signal at 9.62 emerged, which corresponds to the protonated C^2 of the released NHC ligand. The aromatic signals of the benzimidazolylidene core were shifted to 8.00 and 7.71 ppm and the singlet of the NCH_3 fragment moved slightly to 4.08 confirming the release of **1** from **1a**. The aromatic signals of the *p*-cymene ligand in the region of 6.1 - 6.5 ppm merged to a signal at 7.10 ppm. *N*-acetylcysteine was added in excess and resulted in additional emerging signals at 8.86 and 8.34 ppm (NH), 4.47 ppm (CH), 2.8-3.2 ppm (CH_2), as well as 1.96 and 1.86 (CH_3) indicating the formation of chelate complexes with ruthenium.

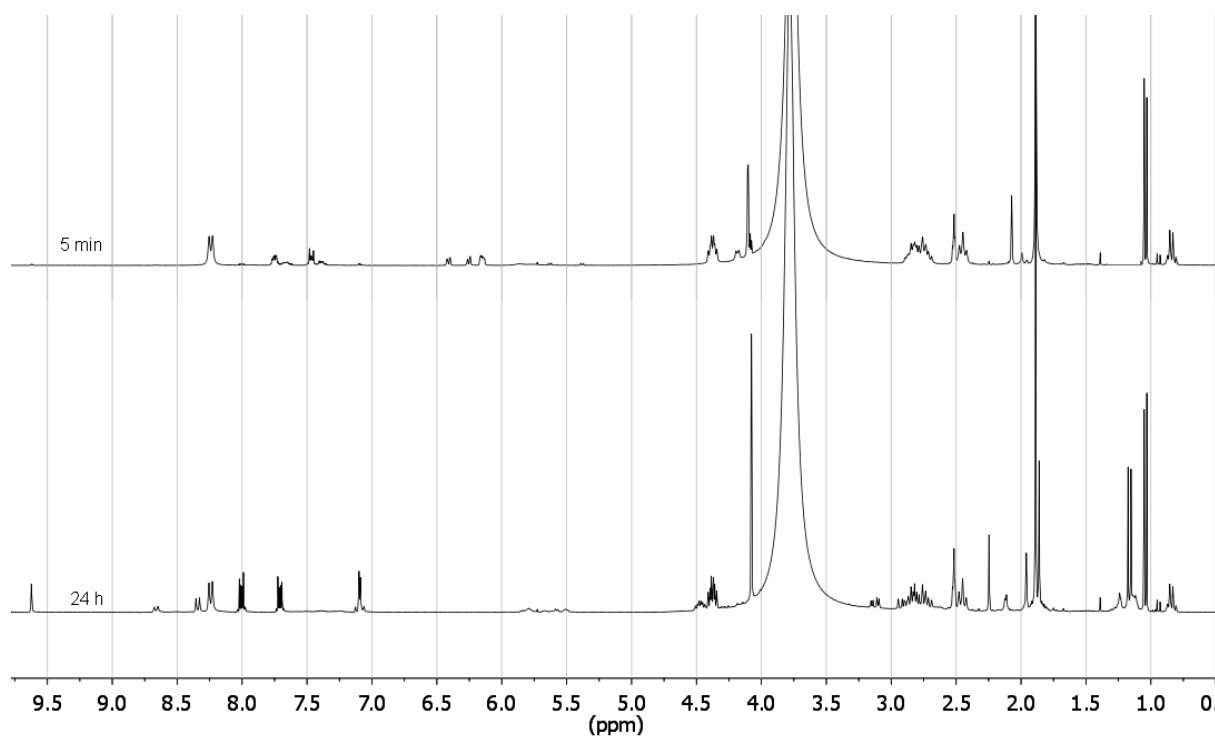


Figure S7 ^1H NMR spectrum of compound **1a** and *N*-acetyl-L-cysteine in $\text{DMSO}-d_6$ after 5 minutes and 24 hours.

3) MS/MS spectra after incubation with peptides

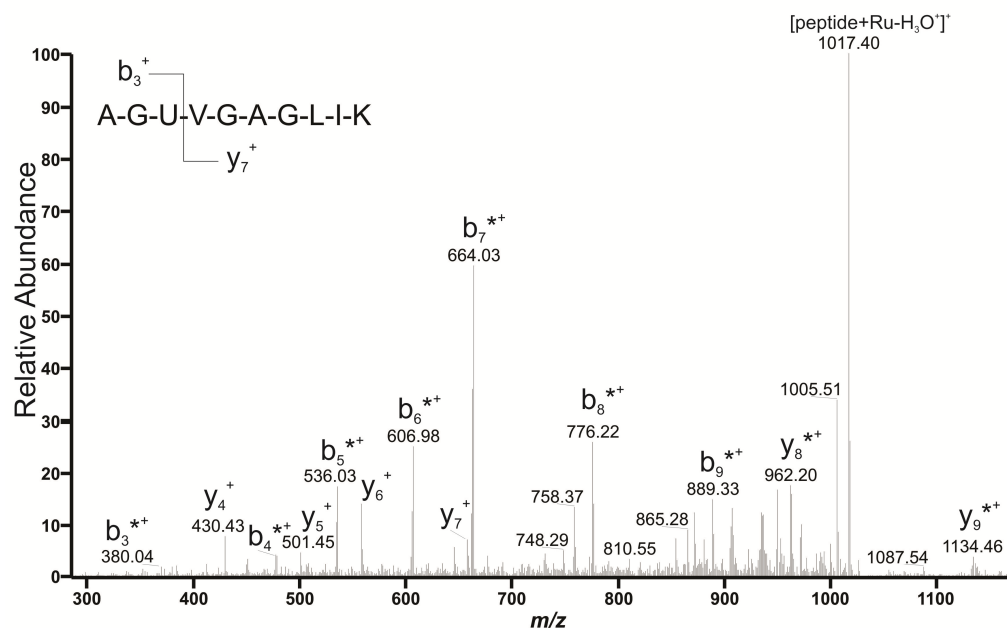


Figure S8: MS/MS spectrum of the molecular ion $[\text{peptide} + \text{Ru}]^+$ at m/z 1036 after a 4 h incubation at 37 °C of a 5:1 mixture of complex **4a** with the peptide H-AGUVGAGLIK-OH. The asterisk (*) represents the ruthenium atom.

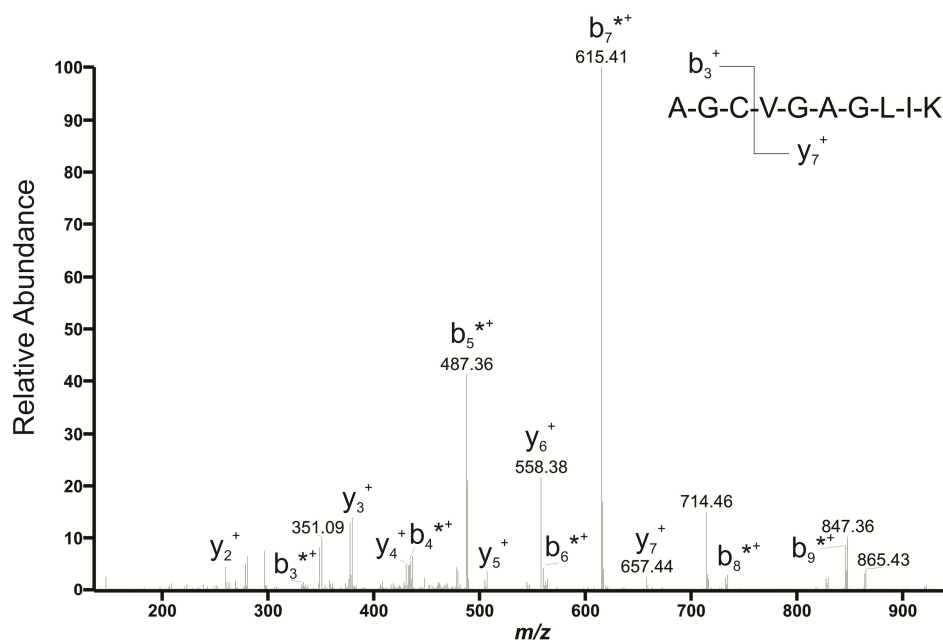


Figure S9: MS/MS spectrum of the molecular ion $[\text{peptide} + \text{Ru}]^{2+}$ at m/z 497 after a 1 h incubation at 37 °C of a 5:1 mixture of complex **1a** with the peptide H-AGCVGAGLIK-OH. The asterisk (*) represents the ruthenium atom.

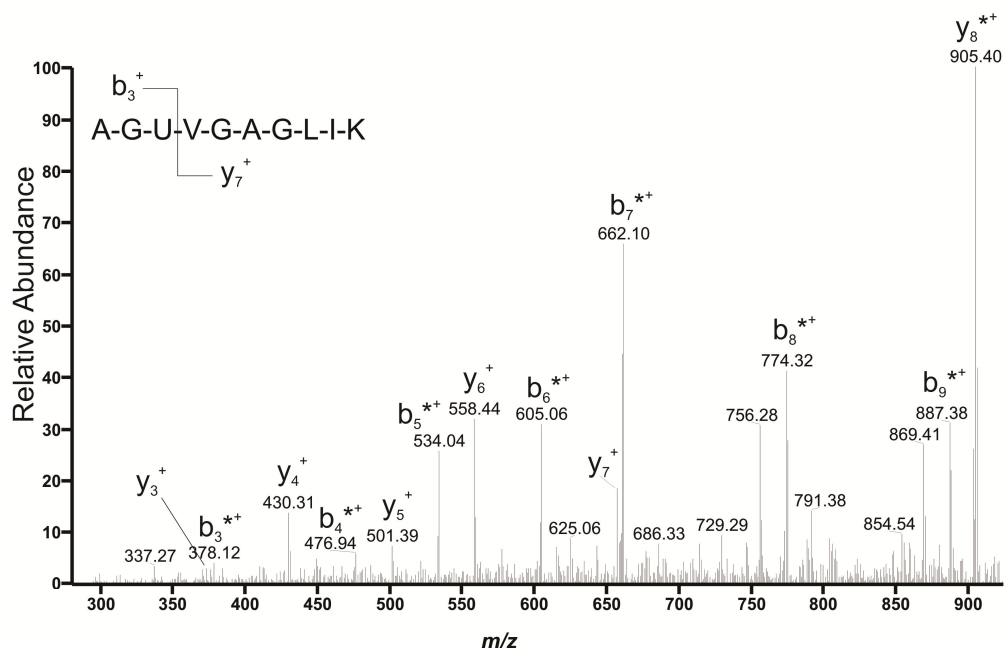


Figure S10: MS/MS spectrum of the molecular ion [peptide + Ru]⁺ at m/z 1034 after a 1 h incubation at 37 °C of a 5:1 mixture of complex **1a** with the peptide H-AGUVGAGLIK-OH. The asterisk (*) represents the ruthenium atom.

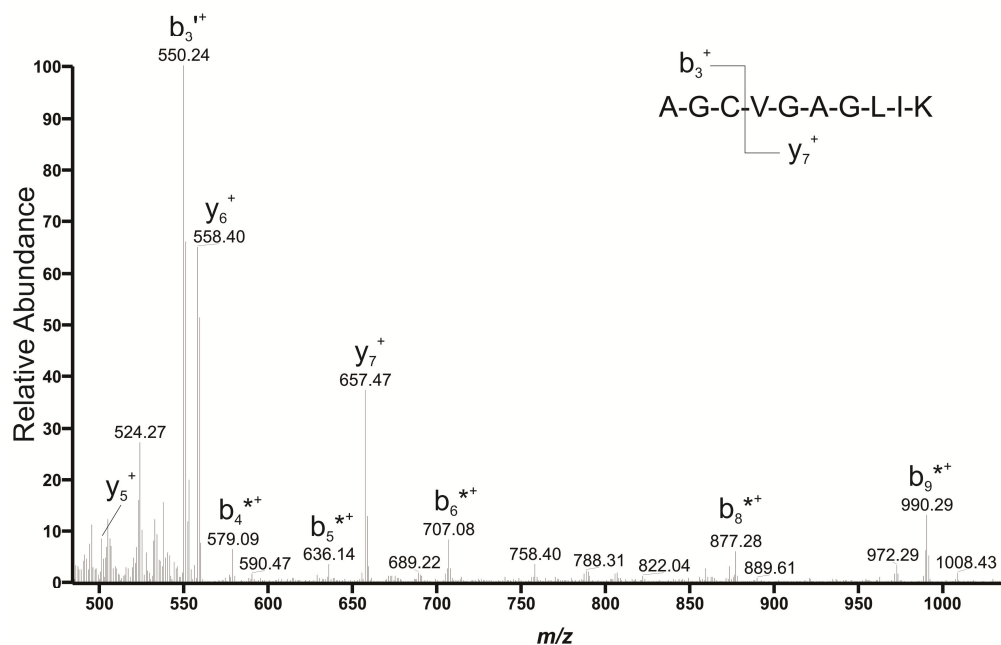


Figure S11: MS/MS spectrum of the molecular ion [peptide + **1a***]²⁺ at m/z 569 after a 1 h incubation at 37 °C of a 1:1 mixture of complex **1a** with the peptide H-AGCVGAGLIK-OH. The asterisk (*) represents the fragment ion [**1a** – (*p*-cymene) – 2Cl]⁺. Only the b_3 -ion is differentially indicated since it could be characterized as [b_3 + **1a** – (*p*-cymene)]⁺.

5. References (1) Morris, R. E.; et al. *J. Med. Chem.* **2001**, *44*, 3616-3621. (2) Scolaro, C. et al. *J. Inorg. Biochem.* **2008**, *102*, 1743–1748. (3) Rubbiani, R. et al. *J. Med. Chem.* **2010**, *53*, 8608–8618.

