

Supporting Information for

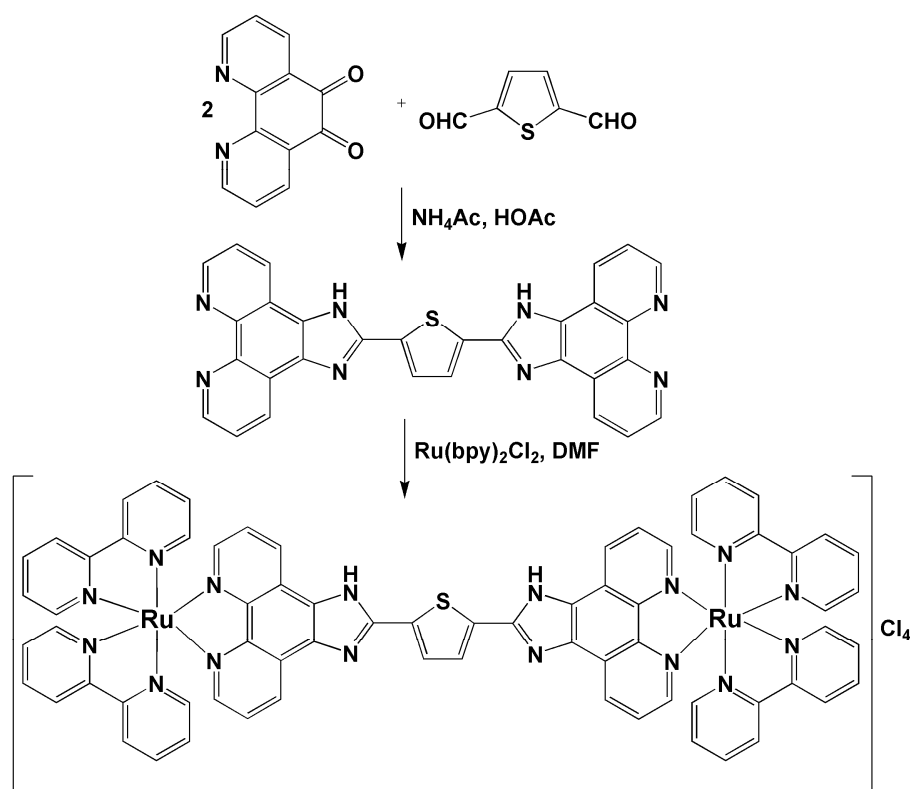
pH luminescence switch, DNA binding and photocleavage, and cytotoxicity of a dinuclear ruthenium complex

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Scheme S1. The synthetic route to **1Cl₄**

1. Synthesis of 2-(5-(1*H*-imidazo[4,5-*f*][1,10]phenanthrolin-2-yl)thiophen-2-yl)-1*H*-imidazo[4,5-*f*][1,10]phenanthroline (**H₂bipt**)

To a solution of 1,10-phenanthroline-5,6-dione (0.42 g, 2.0 mmol) and ammonium acetate (3.0 g, 40 mmol) in 40 mL glacial acetic acid were added thiophene-2,5-dicarbaldehyde (0.14 g, 1.0 mmol). The reaction mixture was refluxed at 80 °C for 1 h under the protection of nitrogen. After cooling, the solution was neutralized to pH = 7 with concentrated NH₃ solution and filtered. The yellow precipitate was collected, washed with distilled water and diethyl ether, and dried in vacuo, giving yield of 67%. The **H₂bipt** ligand is sparingly soluble in common organic solvents, so it was not characterized by ¹H NMR.

2. Synthesis of [Ru₂(bpy)₄(**H₂bipt**)]Cl₄ **1Cl₄**

A mixture of *cis*-Ru(bpy)₂Cl₂•2H₂O (0.104 g, 0.2 mmol) and **H₂bipt** (0.052 g, 0.1 mmol) in 8 mL DMF was refluxed at 110 °C for 12 h under nitrogen. After cooling to room temperature, the solution was filtrated and the filtrate was collected. The purification was carried out by column chromatography on Al₂O₃ followed by recrystallized from CH₃CN-diethyl ether, giving **1Cl₄** as a red solid with yield of 46%. ¹H NMR (400 MHz, DMSO-*d*₆) δ/ppm: 9.06 (s, 4H), 8.90 (dd, *J*₁ = 8 Hz, *J*₂ = 15 Hz, 8H), 8.21 (t, *J* = 8 Hz, 4H), 8.10 (t, *J* = 8 Hz, 4H), 7.95 (s, 2H), 7.88 (d, *J* = 5 Hz, 6H), 7.83 (d, *J* = 6 Hz, 6H), 7.61 (dd, *J*₁ = 5 Hz, *J*₂ = 10 Hz, 8H), 7.37 (t, *J* = 6 Hz, 4H). Anal. Found: C, 48.71; N, 13.76; H, 5.00. Calc. for C₇₀H₄₈N₁₆C₁₄Ru₂•4.3HCON(CH₃)₂•13H₂O: C, 48.83; N, 13.90; H, 5.10. Anal. Calc. for MALDI-TOF MS: *m/z* 336.8 ([M-4Cl]⁴⁺); Found : *m/z* 336.9 ([M-4Cl]⁴⁺). IR (KBr, cm⁻¹): 3427, 3072, 1602, 1444, 1384, 1191, 809, 768, 725.

Some equations used in this paper.

$$1/T_m^0 - 1/T_m = (R/\Delta H_m) \ln(1 + KL)^{1/s} \quad (1)$$

T_m^0 and T_m are the melting temperatures of DNA, in the absence and the presence of the ruthenium complex, respectively. ΔH_m is the enthalpy of DNA melting (per base pair) taken as 6.9 kcal mol⁻¹, R is the gas constant, K is the DNA binding constant at T_m , L is the free complex concentration (approximated by the total complex concentration at T_m , and s is the binding site size.

$$\ln(K_1/K_2) = (\Delta H^0/R)[(T_1 - T_2)/T_1 T_2] \quad (2)$$

$$\Delta G_T^0 = -RT \ln K \quad (3)$$

$$\Delta G_T^0 = \Delta H^0 - T\Delta S^0 \quad (4)$$

K_1 and K_2 are the DNA-binding constants of the complex at temperatures T_1 and T_2 , respectively. ΔG_T^0 , ΔH^0 , and ΔS^0 are the changes of the standard free energy, standard enthalpy, and standard entropy of binding of the complex to DNA, respectively.

The ¹O₂ generation quantum yields (Φ_Δ) of the complexes were calculated according to following Eqs. (5) and (6).

$$-\Delta[\text{DPBF}]/t = (I_0 - I_t)/t = I_{\text{in}} \Phi_{\text{ab}} \Phi_\Delta \Phi_r \quad (5)$$

$$k/k_s = \Phi_{\text{ab}} \Phi_\Delta / \Phi_{\text{ab}}^s \Phi_\Delta^s \quad (6)$$

I_{in} is the incident monochromatic light intensity, Φ_{ab} is the light absorbing efficiency of the photosensitizer, Φ_r is the reaction quantum yield of ¹O₂ with DPBF, t is the irradiation time, I_0 and I_t are the luminescence intensity of DPBF before and after irradiation, k is the slope and superscript s stands for the standard complex [Ru(bpy)₃]²⁺.

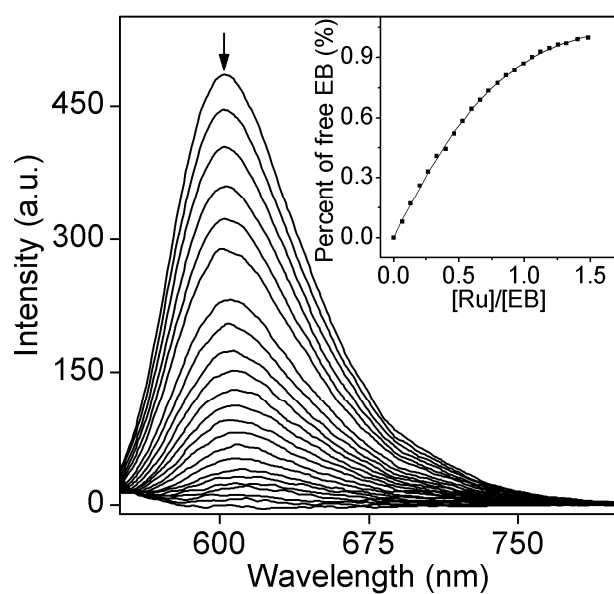


Fig. S1 Changes in emission spectra ($\lambda_{\text{ex}} = 537 \text{ nm}$) of EB ($10 \mu\text{M}$) bound to DNA ($50 \mu\text{M}$) in the presence of ICl_4 . Inset: plot of percentage of free EB vs. $[\text{Ru}]/[\text{EB}]$.

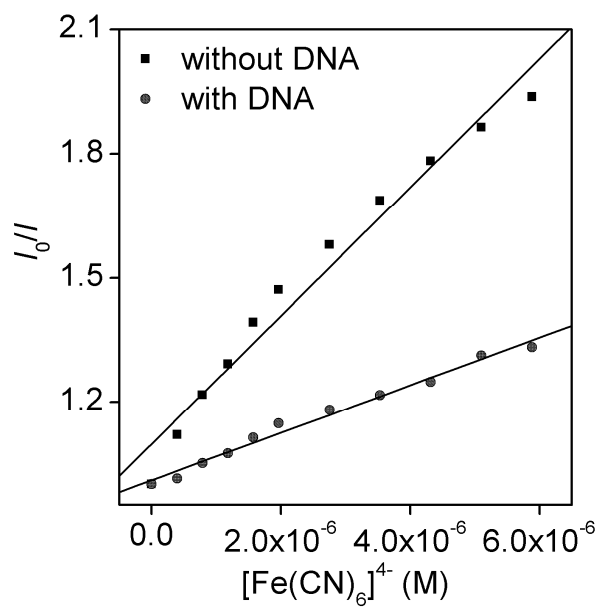


Fig. S2 Emission quenching of **1**Cl₄ (5 μM) upon increasing concentrations of [Fe(CN)₆]⁴⁻ in the absence and the presence of DNA (50 μM).

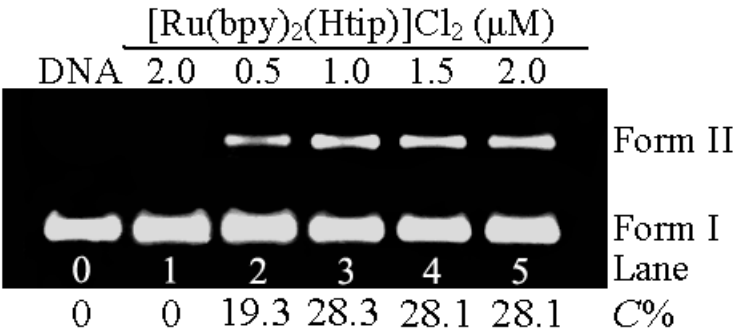


Fig. S3 Photoactivated cleavage of pUC 18 DNA in the presence of different concentrations of $[\text{Ru}(\text{bpy})_2(\text{Htip})]\text{Cl}_2$ after irradiation at 360 nm (16 mW/cm²) for 2 h.

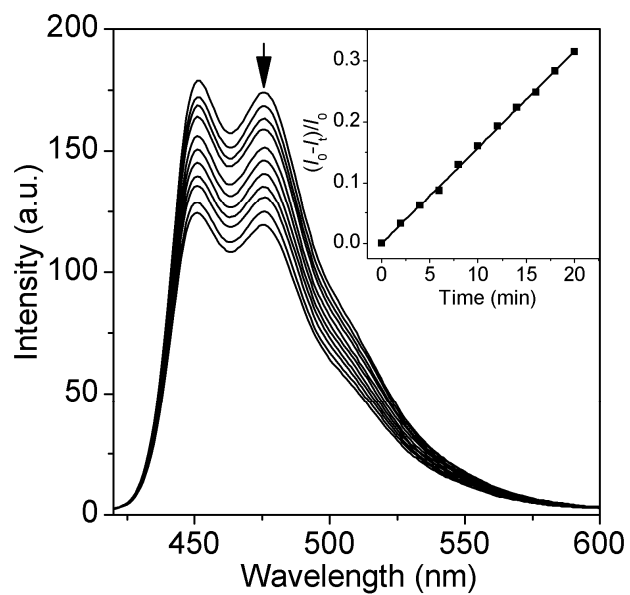


Fig. S4 The emission spectra changes of DPBF (20 μM)–[Ru(bpy)₃]²⁺ (5 μM) system upon irradiation at 480 nm. Inset: the DPBF consumption percentage as a function of irradiation time in the air-equilibrated methanol solution of [Ru(bpy)₃]²⁺.

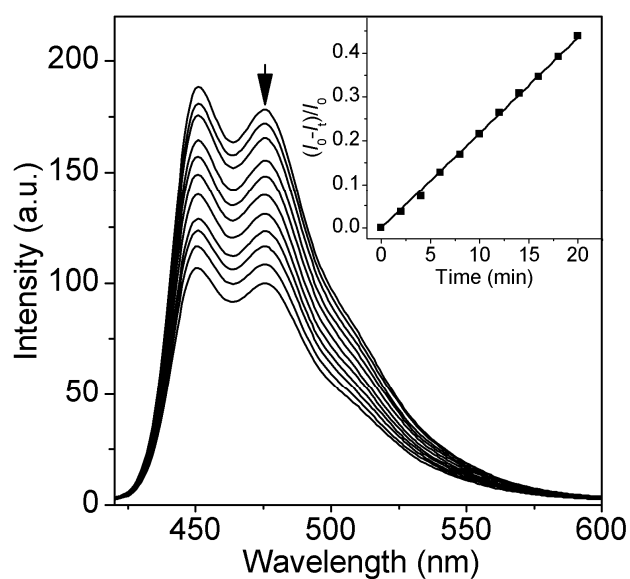


Fig. S5 The emission spectra changes of DPBF (20 μM)-9Cl₂ (5 μM) system upon irradiation at 480 nm. Inset: the DPBF consumption percentage as a function of irradiation time in the air-equilibrated methanol solution of 9Cl₂.

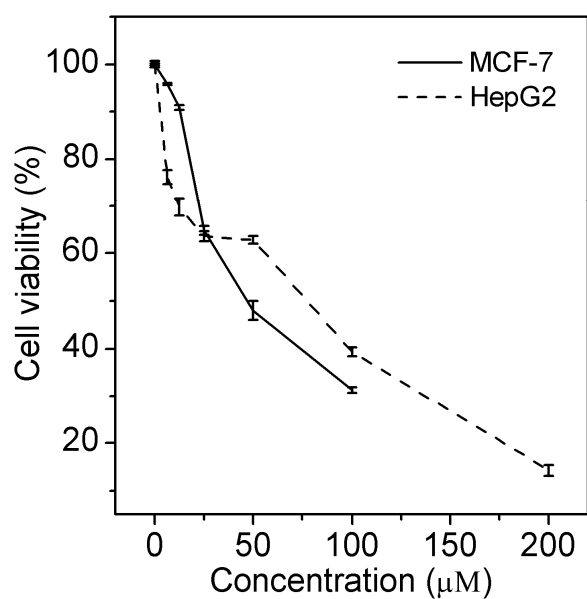


Fig. S6 Cell viability of 1Cl_4 on tumor (MCF-7 and HepG2) cell proliferation in vitro.

Table S1 Computed selective bond lengths (nm), bond angles A_m and A_{co} ($^{\circ}$)*, dihedral angles and some frontier molecular orbital energies (a.u.)

Complex	Ru-N _m	Ru-N _{co}	C-C(N) _m	C-C(N) _{co}	A_m	A_{co}	Dihedral angles	Frontier molecular orbital energies	Ref.
9	0.2124	0.2115	0.1397	0.1389	78.46	77.70	179.88	-0.255(LUMO+2)	Z.S. Li, H.X. Yang, A.G. Zhang, H. Luo, K.Z. Wang, <i>Inorg. Chim. Acta</i> , 2011, 370 , 132.
1	0.2127	0.2118	0.1393	0.1389	78.10	77.66	179.42(N1-C2-C3-S4) 179.40(S4-C5-C6-N7)	-0.327(LUMO)	This work

*Subscript “m” stands for the main ligands of Hip, Htip and H₂bip, and “co” represents for the co-ligand of bpy.