Electronic Supplementary Information

Synthesis and Characterization of Conjugated Polyoxometalate-Porphyrin Copolymers Obtained from a Dawson-Type Polyoxophosphovanadotungstate †

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General Remarks

Reagents and chemicals were purchased from commercial sources and used as received. The starting $TBA_5H_4[P_2V_3W_{15}O_{62}]$ was prepared as described in the literature.¹

¹H NMR [¹³C NMR] spectra were recorded at room temperature with a 300 MHz [75 MHz] Bruker 300 UltraShield Spectrometer equipped with a QNP probe, a 400 Mhz [100MHz] Bruker Avance 400 spectrometer equipped with a QNP probe, or with a 500 MHz Bruker Avance 500 UltraShield equipped with a BBI ATM probe. Chemical shifts are given in ppm, referenced to TMS ($\delta = 0$ ppm) using the solvent signals ($\delta = 1.94$ ppm for CHD₂CN [$\delta = 1.32$ ppm for CD₃CN] and $\delta = 7.26$ ppm for CHCl₃ [$\delta = 77.16$ ppm for CDCl₃] for calibration. Coupling constants (*J*) are given in Hertz (Hz).

Mass spectrometry experiments have been carried out on an electrospray-time of flight instrument (Waters, LCT Premier XE) for organic compounds and on an electrospray-ion trap instrument (Bruker, Esquire 3000) for polyoxometalate hybrids. The 50 μ mol·L⁻¹ solutions of POMs were infused using a syringe pump (160 μ L·h⁻¹). The negative ion mode was used with capillary high voltage 3500 V. The orifice/skimmer voltage difference was set to 45 V to avoid decomposition of the POMs. The low-mass-cutoff (LMCO) of the ion trap was set to 80 Th.

Elemental analyses were carried out by ICSN (CNRS, Gif, France).

IR spectra were recorded from a Bruker Tensor 27 ATR diamond PIKE spectrophotometer for organic compounds, and Bio-Rad FTS 165 (KBr) for polyoxometalates hybrids.

Cyclovoltammograms [and electropolymerisation experiments] were recorded at room temperature with a potentiostat model 273A from EG&G Princeton Applied Resarch using a conventional three electrode set-up. The working electrode was glassy carbon [ITO plates, Aldrich, 8-12 Ω /square, surface of about 1 cm²], the counter electrode was platinum wire, and the reference electrode was a saturated calomel electrode (SCE) connected through a salt bridge. The supporting electrolyte was tetrabutylammonium hexafluorophosphate (TBAPF₆) 0.1 M, and analyte concentration was 0.5 mM. The solutions were deaerated thoroughly by bubbling argon through the solution and kept under argon atmosphere during the whole experiment.

The $POM(py)_2$ hybrids reported here have a high tendency to stick to the electrode, consequently the electrode necessitate to be washed after each scan.

UV-visible absorption spectra in solution were recorded either with a double beam Perkin-Elmer Lambda 9 spectrophotometer operated at a resolution of 1 nm while those on ITO electrode were recorded in transmission mode.

Atomic Force Microscopy (AFM) was performed directly on the surface of the ITO using a Dimension 3100 (Veeco) in the tapping mode under ambient conditions. Silicon cantilevers (Veeco probes) with a spring constant of 300 N/m and a resonance frequency in the range of 120-139 kHz were used. The scanning rate was 1.0 Hz.

X-ray Photoelectron Spectroscopy (XPS) experiments were carried out on a RBD upgraded PHI-5000C ESCA system (Perkin-Elmer) with MgKR radiation ($h\Box$ =1253.6 eV) or Al KR radiation ($h\nu$ = 1486.6 eV). In general, the X-ray anode was run at 250W and the high voltage was kept at 14.0 kV with a detection angle at 54°. The pass energy was fixed at 23.5, 46.95, or 93.90 eV to ensure sufficient resolution and sensitivity. The base pressure of the analyzer chamber was about 5.10⁻⁸ Pa. The sample was directly pressed to a self-supported disk (10 x 10 mm) and mounted on a sample holder then transferred into the analyzer chamber. The whole spectra (0-1100 eV) and the narrow spectra of all the elements with higher resolution were both recorded by using RBD 147 interface (RBD Enterprises, U.S.A.) through the Auger Scan 3.21 software. Binding energies were calibrated by using the containment carbon (C1s = 284.6 eV). The data analysis was carried out by using the RBD Auger Scan 3.21 software provided by RBD Enterprises or XPS Peak4.1 provided by Raymund W.M. Kwok (The Chinese University of Hongkong, China)

¹ R. G. Finke, B. Rapko, R. J. Saxton, P. J. Domaille, J. Am. Chem. Soc. 1986, 108, 2947-2960

Ligand Synthesis



2-propenoic acid, 3-(3,5-dibromophenyl)-methyl ester : To a solution of 3,5-dibromobenzadehyde (1.5 g, 5.68 mmol) in CH₂Cl₂ (45 mL), methyl(triphenylphosphoranylidene)acetate (2.58 g, 7.57 mmol) was added. The solution was stirred under argon for 7h. The reaction was monitored by TLC. The solvent was removed *in vacuo* and the crude product was purified by column chromatography on silica gel (CH₂Cl₂: Hept = 1:1) to give the product as white solid (82.3 mg, 0.218 mmol) in 95% yield. mp : 142°C.

¹H NMR (300 MHz, CDCl₃) δ : 7.66 (s, 1H), 7.57(s, 2H), 7.52 (d, J = 16Hz, 1H), 6.42 (d, J = 16 Hz, 1H), 3.81 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ: 166.6 (C=O), 141.6 (CH=C), 137.9 (C_{q-Ar}), 135.4 (CH_{Ar}), 129.6 (CH_{Ar}), 123.5 (C_{q-Ar}), 120.7 (CH=C), 52.0 (CH₃).

IR (ATR, cm⁻¹): 3073, 2950, 1721, 1639, 1582, 1547, 1432, 1412, 1368, 1316, 1308, 1291, 1261, 1207, 1190, 1176, 1102, 1012, 984, 889, 849, 800, 742, 682, 656.

Elemental Analysis: Calc C 37.54%, H 2.52%, N 0%. Found C 37.78%, H 2.57%, N 0%.

evb3,3 : In a dry Schlenk tube, Pd(PPh₃)₄ (101.1 mg, 0.088 mmol) was solubilised in freshly distilled and degassed triethylamine (15mL) at 80°C . 2-propenoic acid, 3-(3,5-dibromophenyl)-methyl ester (280 mg, 0.875 mmol), 3-ethynylpyridine (255 mg, 2.48 mmol) and CuI (33 mg, 0.17 mmol) were successively added, and the resulting mixture was heated to 80°C under argon overnight. The solvent was removed *in vacuo* and the crude brown powder was purified by column chromatography on silica gel (CH₂Cl₂:MeOH = 97:3) to give **evb3,3** as a pale yellow solid (315 mg, 0.86 mmol) in 99% yield. mp: 142°C.

¹H NMR (500 MHz, CDCl₃) δ : 8.79 (d, J = 1.3Hz, 2H), 8.59 (dd, J = 1.5 Hz, J = 4.9 Hz, 2H), 7.83 (dt, J = 1.5Hz, J = 7.7 Hz, 2H), 7.74 (s, 1H), 7.66 (m, 3H), 7.32 (dd, J = 4.9 Hz, J = 7.7 Hz, 2H), 6.52 (d, J = 15.9 Hz, 1H), 3.83 (s, 3H).

¹³C NMR (125 MHz, CDCl₃) δ : 166.9 (C=O), 152.3 (CH_{py}), 149.0 (CH_{py}), 142.7 (CH=C), 138.6 (CH_{py}), 135.8 (CH_{Ar}), 135.3 (C_q), 131.0 (CH_{Ar}), 123.9 (C_q), 123.2 (CH_{py}), 120.0 (CH=C), 119.9 (C_q), 90.8 (C=C), 87.4 (C=C), 51.9 (CH₃).

IR (ATR, cm⁻¹): 3050, 2923, 2853, 2213, 1718, 1640, 1592, 1561, 1478, 1437, 1408, 1377, 1300, 1266, 1208, 1190, 1173, 1120, 1093, 1040, 1021, 979, 922, 856, 800, 722, 695, 670. TOF MS (ES+) calc for $C_{24}H_{16}N_2O_2$ -H⁺: 365.1285, found : 365.1285.

evb4,4 and evb4,Br: In a dry Schlenk tube, 4-ethynylpyridine hydrochloride (295 mg, 2.1 mmol) was solubilized in dry and degassed DMF (30 mL). To this dark blue suspension was added compound 2-propenoic acid, 3-(3,5-dibromophenyl)-methyl ester (225 mg, 0.70 mmol) and freshly distilled and deaerated triethylamine (15mL), followed by $PdCl_2(PPh_3)_2$ (49.4 mg, 0.07 mmol) and CuI (26.8 mg, 0.14 mmol). The mixture was heated under argon at 75°C for 4 days. Solvents were removed *in vacuo* and the crude product was purified by column chromatography on silica gel (gradient elution from

Hept:EA = 6:4 to 3:7) to give **evb4,4** as a pale yellow solid (143.9 mg, 0.86 mmol) in 56% yield and **evb4,Br** (50.4 mg, 0.15 mmol) in 21% yield.

evb4,4:

mp:197°C.

¹H NMR (300 MHz, CDCl₃) δ : 8.64 (d, J = 5.1 Hz, 4H), 7.74 (t, J = 1.4 Hz, 1H), 7.69 (d, J = 1.3 Hz, 2H), 7.65 (d, J = 16 Hz, 1H), 7.39 (d, J = 4.5 Hz, 4H), 6.52 (d, J = 16 Hz, 1H), 3.83 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ : 166.8 (C=O), 149.9 (CH_{py}), 142.4 (CH=C), 136.1 (CH_{Ar}), 135.4 (C_q), 131.5 (CH_{Ar}), 130.7 (C_q), 125.6 (CH_{py}), 123.6 (C_q), 120.2 (CH=C), 91.8 (C=C), 88.1 (C=C), 52.0 (CH₃).

IR (ATR, cm⁻¹):.3053, 2923, 2853, 2217, 1711, 1635, 1594, 1538, 1488, 1459, 1439, 1413, 1377, 1328, 1304, 1260, 1221, 1189, 1172, 1091, 1012, 922, 867, 814, 801, 742, 696, 672.

TOF MS (ES+) calc for $C_{24}H_{16}N_2O_2-H^+$: 365.1285 , found: 365.1288.

evb4,Br

mp:157°C.

¹H NMR (300 MHz, CDCl₃) δ : 8.62 (d, J = 5.8 Hz, 2H), 7.67 (t, J = 1.5 Hz, 1H), 1.64 (t, J = 1.5 Hz, 1H), 7.57 (m, 2H), 7.36 (d, J = 6Hz, 2H), 6.46 (d, J = 16 Hz, 1H), 3.81 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ : 166.7 (C=O), 149.9 (CH_{py}), 142.0 (CH=C), 136.6 (C_q), 135.6 (CH_{Ar}), 131.3 (CH_{Ar}), 130.6 (C_q), 129.9 (CH_{Ar}), 125.6 (CH_{py}), 124.7 (C_q), 122.9 (C_q), 120.4 (CH=C), 91.3 (C=C), 88.4 (C=C), 52.0 (CH₃).

IR (ATR, cm⁻¹):3037, 2950, 2219, 1718, 1641, 1594, 1537, 1540, 1489, 1436, 1413, 1329, 1286, 1218, 1203, 1175, 1112, 1037, 990, 925, 854, 819, 743, 670.

TOF MS (ES+) calc for $C_{17}H_{12}BrNO_2$ -H⁺: 342.0125 , found: 342.0145.

evb3,4:

In a dry Schlenk tube under argon compound **evb4,Br** (63.5 mg, 0.19 mmol) and Pd(PPh₃)₄ (55 mg, 0.048 mmol) were solubilised in distilled and deaerated triethylamine (4 mL) and dry DMSO(0.5 mL) at 80°C. 3-ethynylylpyridine (57.4 mg, 0.056 mmol), CuI (7.1 mg, 0.037 mmol) and triethylamine (3mL) were added and the mixture was heated at 80°C for 24h. The clear orange solution was evaporated *in vacuo* and the crude product was purified by column chromatography on silica gel (CH₂Cl₂:MeOH = 98:2) to give evb3,4 in presence of an impurity.

¹H NMR (500 MHz, CDCl₃) δ: 8.79 (m, 1H), 8.64 (m, 2H), 8.59 (m, 1H), 7.82 (d, J= 7.6Hz, 1H), 7.74 (s, 1H), 7.68 (m, 3H), 7.4 (d, J = 5.5 Hz, 2H), 7.32 (m, 1H), 6.52 (d, J = 16.1 Hz, 1H), 3.83 (s, 3H).



dvb3,3: In a 25mL round bottom flask is added evb3,3 (150 mg, 0.41 mmol), 2-Amino-2-ethyl-1,3propanediol (73.48 mg, 0.62 mmol) solution in CH2Cl2/DMSO (4.5/0.2 mL) and K2CO3 (56.8 mg, 0.41 mmol). The resulting mixture was heated to reflux overnight. The mixture was diluted with CH2Cl2 and MeOH and K2CO3 was removed by centrifugation. The clear supernatant was evaporated in vacuo and the residue was purified by column chromatography on silica gel (gradient elution from CH2Cl2:MeOH = 100:0 to 95:5) to give dvb3,3 as a waxy solid (149 mg, 0.33 mmol) in 80% yield.

¹H NMR (300 MHz, CDCl₃) δ : 8.73 (d, J = 1.3Hz, 2H), 8.54 (dd, J = 1.5 Hz, J = 4.9 Hz, 2H), 7.78 (dt, J = 1.9Hz, J = 7.9 Hz, 2H), 7.64 (m, 1H), 7.58 (m, 2H), 7.51 (d, J = 15.5 Hz), 7.29 (m, 2H), 6.53 (d, J = 15.5 Hz, 1H), 3.90 (d, J = 11.6 Hz, 2H), 3.69 (d, J = 11.6 Hz, 2H), 1.77 (q, J = 7.6 Hz, 2H), 0.92 (t, J = 7.6 Hz, 3H).

¹³C NMR (75 MHz, CDCl₃) δ : 166.3 (C=O), 152.3 (CH_{py}), 149.0 (CH_{py}), 139.4 (CH=C), 138.8 (CH_{py}), 135.6 (C_q), 135.5 (CH_{Ar}), 131.0 (CH_{Ar}), 123.8 (C_q), 123.3 (CH_{py}), 123.0 (CH=C), 120.1 (C_q), 91.2 (C=C), 87.3 (C=C), 65.9 (CH₂OH), 61.8 (C_q), 25.5 (CH₂), 7.8 (CH₃).

IR (ATR, cm⁻¹): 3277, 2930, 2399, 2216, 1726, 1660, 1618, 1477, 1421, 1407, 1337, 1230, 1187, 1044, 1024, 976, 856, 804, 731.

TOF MS (ES+) calc for $C_{28}H_{25}N_3O_3-H^+$: 452.1969, found: 452.1970.

dvb4,4: In 25mL round bottom flask is added **evb4,4** (112 mg, 0.31 mmol), 2-Amino-2-ethyl-1,3propanediol (54.9 mg, 0.46 mmol) solution in CH₂Cl₂/DMSO (5.5/0.2 mL) and K₂CO₃ (42.4 mg, 0.31 mmol). The resulting mixture was heated to reflux 4 days. The mixture was diluted with CH₂Cl₂ and MeOH and K₂CO₃ was removed by centrifugation. The clear supernatant was evaporated *in vacuo* and the residue was purified by column chromatography on silica gel (gradient elution from CH₂Cl₂:MeOH = 100:0 to 95:5) to give **dvb4,4** as a waxy solid (94.6 mg, 0.21 mmol) in 68% yield.

¹H NMR (300 MHz, CDCl₃) δ : 8.52 (s, 4H), 7.74 (t, J = 1.4 Hz, 1H), 7.62 (m, 3H), 7.46 (d, J = 15.6 Hz, 1H), 7.34 (d, J = 5.6 Hz, 4H), 6.55 (d, J = 15.6 Hz, 1H), 3.77 (d, J = 11.6 Hz, 2H), 3.60 (d, J = 11.6 Hz, 2 H), 1.73 (q, J = 7.5 Hz, 2H), 0.86 (t, J = 7.5 Hz, 3H).

¹³C NMR (75 MHz, CDCl₃) δ : 166.4 (C=O), 149.5 (CH_{py}), 138.7 (CH=C), 135.8 (C_q), 135.6 (CH_{Ar}), 131.4 (CH_{Ar}), 131.3 (C_q), 125.8 (CH_{py}), 123.4 (C_q), 123.3 (CH=C), 92.4 (C=C), 87.7 (C=C), 65.0 (CH₂OH), 61.7 (C_q), 25.0 (CH₂), 7.7 (CH₃).

IR (ATR, cm⁻¹): 3273, 2927, 2217, 1663, 1623, 1595, 1539, 1411, 1338, 1222, 1026, 999, 858, 821 TOF MS (ES+) calc for $C_{28}H_{25}N_3O_3$ -H⁺: 452.1969, found: 452.2006.

dvb3,4: In 25mL round bottom flask is added **evb3,4** (43.1 mg, 0.12 mmol), 2-Amino-2-ethyl-1,3propanediol (21.1 mg, 0.18 mmol) solution in CH₂Cl₂/DMSO (5.5/0.2 mL) and K₂CO₃ (16.3 mg, 0.12 mmol). The resulting mixture was heated to reflux 4 days. The mixture was diluted with CH₂Cl₂ and MeOH and K₂CO₃ was removed by centrifugation. The clear supernatant was evaporated *in vacuo* and the residue was purified by column chromatography on silica gel (gradient elution from CH₂Cl₂:MeOH = 100:0 to 95:5) to give **dvb3,4** as a waxy solid (39.7 mg, 0.088 mmol) in 73% yield.

¹H NMR (300 MHz, CDCl₃) δ : 8.66 (s, 1H, 3py), 8.49 (m, 3H, 3py(1H) + 4py(2H)), 7.79 (d, J = 7.9 Hz, 1H, 3py), 7.62 (m, 3H, CH_{Ar}), 7.46 (d, J = 15.6 Hz, 1H, CH=C), 7.36 (d, J = 4.71, 2H, 4py), 7.29 (m, 1H, 3py), 6.57 (d, J = 15.6 Hz, 1H), 3.74 (d, J = 11.7 Hz, 2H, CH₂OH), 3.58 (d, J = 11.7 Hz, 2H, CH₂OH), 1.72 (q, J = 7.4 Hz, 2H, CH₂), 0.85 (t, J = 7.4 Hz, 3H, CH₃).

¹³C NMR (75 MHz, CDCl₃) δ : 166.6 (C=O), 151.8 (CH_{3py}), 149.4 (CH_{4py}), 148.6 (CH_{3py}), 139.0 (CH_{3py}), 138.9 (CH=C), 135.8 (C_q), 135.5 (CH_{Ar}), 131.5 (C_q), 131.2 (CH_{Ar}), 131.2 (CH_{Ar}), 125.9 (CH_{4py}), 123.8 (C_q), 123.5 (CH_{3py}), 123.2 (CH=C),120.2 (C_q), 92.6 (C=C), 91.1 (C=C), 87.6 (C=C), 87.1 (C=C), 64.9 (CH₂OH), 61.7 (C_q), 24.9 (CH₂), 7.6 (CH₃).

IR (ATR, cm⁻¹): 3301, 2949, 2216, 1660, 1625,1599, 1413,1339, 1232, 1017.

TOF MS (ES+) calc for $C_{28}H_{25}N_3O_3$ -H⁺: 452.1969, found: 452.1986.

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tvb3,3: To a solution of **evb3,3** (77.8 mg, 0.21 mmol) in DCM (2mL), is added 2-Amino-2-hydroxyméthyl-1,3-propanediol (38.8 mg, 0.32 mmol), DMSO (0.1 mL) and K₂CO₃ (29.5 mg, 0.21 mmol). The resulting mixture was heated to reflux overnight. The mixture was diluted with CH₂Cl₂ and MeOH and K₂CO₃ was removed by centrifugation. The clear supernatant was evaporated *in vacuo* and the residue was purified by column chromatography on silica gel (gradient elution from CH₂Cl₂:MeOH = 100:0 to 95:5) to give **tvb3,3** as a waxy solid (66.6 mg, 0.088 mmol) in 69% yield. ¹H NMR (300 MHz, CDCl₃) δ : 8.71 (s, 2H), 8.51 (dd, J = 1Hz, J = 4.8 Hz, 2H), 7.84 (dt, J = 1.8 Hz, J = 7.9 Hz, 2H), 7.67 (m, 1H), 7.64 (m, 2H), 7.52 (d, J = 15.6 Hz, 1H), 7.33 (m, 2H), 6.62 (d, J = 15.6Hz, 1H), 3.71 (s, 6H).

¹³C NMR (75 MHz, CDCl₃) δ: 166.9 (C=O), 151.8 (CH_{py}), 148.5 (CH_{py}), 139.5 (CH=C), 139.0 (CH_{py}), 135.5 (C_q), 135.4 (CH_{Ar}), 130.9 (CH_{Ar}), 131.0 (CH_{Ar}), 123.6 (C_q), 123.4 (CH_{3py}), 122.6 (CH=C), 120.4 (C_q), 91.1 (C=C), 86.9 (C=C), 62.6 (C_q), 62.0 (CH₂OH).

IR (ATR, cm⁻¹): 3287, 2961, 1662, 1622, 1560, 1478, 1410, 1258, 1016, 860, 794 TOF MS (ES+) calc for $C_{28}H_{25}N_3O_3$ -H⁺: 454.1762, found: 454.1765.

Hybrids synthesis:

General Procedure :

A solution of $TBA_5H_4[P_2V_3W_{15}O_{62}]$ (300 mg, 0.056 mmol, 1eq.), the diol-amide ligand (1.4 eq.), and APTS (5.82 mg, 0.034 mmol, 0.6 eq.) in DMAc (0.6 mL) is heated at 80°C for 1h by microwave irradiation. The light orange solution turned brown. After removal *in vacuo* of the solvent, the brownish oil is solubilized in acetonitrile, precipitated by addition of diethyl ether and centrifuged for 10 min. The recovered yellow solid is solubilized in acetonitrile, and TBA resin is added to the solution for 30 min. The resin is then filtered off, and the orange solution is reduced *in vacuo*, precipitated by diethyl ether, and centrifuged for 10 min. The recovered solid is solubilized in a minimum amount of acetonitrile, precipitated by addition of EtOH and centrifuged for 30 min. The recovered solid is solubilized in acetonitrile, precipitated by diethyl ether, centrifuged, then triturated with diethyl ether, centrifuged and dried under vacuum. The ethanol supernatant is evaporated, and the oil is precipitated by addition of small amount of ethanol, followed by centrifugation, and two diethyl ether precipitation/centrifugation steps.



POM-dvb3,3: prepared with 72% yield.

IR (KBr, cm⁻¹): 3457, 2962, 2873, 1637, 1594, 1483, 1087, 953, 902, 817, 738.

³¹P NMR (162 MHz, CD₃CN) δ: - 6.99, -13.06.

¹H NMR (400 MHz, CD₃CN) δ : 9.6 (br, 1H, N**H**), 8.79 (s, 2H, H_{py}), 8.73 (d, 1H, J=15.2 Hz, C<u>H</u>=CH), 8.60 (d, 2H, J=4.0 Hz, H_{py}), 8.29 (s, 2H, H_{Ar}), 8.11 (d, 2H, J= 7.64 Hz, H_{py}), 7.83 (s, 1H, H_{Ar}), 7.53 (dd, 2H, J=4.9Hz, J= 7.6 Hz, H_{py}), 7.16 (d, 1H, J= 15.2Hz, CH=C<u>H</u>), 5.61 (d, 2H, J=12.7 Hz, C**H**₂OV), 5.48 (d, 2H, J=12.7Hz, C**H**₂OV), 3.17 (m, 43,1H, ⁺N(C**H**₂CH₂CH₂CH₃)₄), 1.86 (q, 2H, J=7.2Hz, C**H**₂-C**H**₃), 1.65 (m, 43.3H, ⁺N(CH₂CH₂CH₂CH₃)₄), 1.43 (q, 43.4H, J = 7.24Hz, ⁺N(CH₂CH₂CH₂CH₃)₄), 1.13 (t, 3H, J= 7.2Hz, CH₂-C**H**₃), 1.00 (t, 64.8H, J=7.24Hz, ⁺N(CH₂CH₂CH₂CH₂CH₂)₄).

¹³C NMR (100 MHz, CD₃CN) δ : 181.7 (C=O), 154.2 (CH=CH), 152.7 (C_{py}), 150.3 (C_{py}), 140.3 (C_{py}), 136.9 (CH_{Ar}), 136.5 (C_q), 134.3 (CH_{Ar}), 125.2 (C_q), 124.9 (C_{py}), 120.6 (C_q), 117.4 (CH=CH), 91.7 (C=C), 89.8 (CH₂OV), 88.9 (C=C), 64.8 (C_q), 59.3 (⁺N(CH₂CH₂CH₂CH₃)₄), 29.5 (CH₂-CH₃), 24,5 (⁺N(CH₂CH₂CH₂CH₃)₄), 20.4 (⁺N(CH₂CH₂CH₂CH₂CH₃)₄), 14.0 (⁺N(CH₂CH₂CH₂CH₃)₄), 7.52 (CH₂-CH₃) Anal. calc. for TBA₅(C₂₈H₂₃N₃O₆₂P₂V₃W₁₅).(Et₂O)_{0.7} (5630,04 g.mol⁻¹): C 23.64, H 3.76, N 1,99 ; found : C 23.64, H 3.76, N 2.00.

Electrospray mass spectrometry:

Entry	Charge	Simulated m/z	Observed m/z	Relative Intensity (%)	Composition
1	3-	1617.2	1617.4	18	$TBA_2(C_{28}H_{23}N_3O_{62}P_2V_3W_{15})$
2	4-	1152.3	1152.4	93	$TBA(C_{28}H_{23}N_3O_{62}P_2V_3W_{15})$
3	4-	1212.9	1212.9	11	$TBA_2 (C_{28}H_{23}N_3O_{62}P_2V_3W_{15}) R.$
4	5-	873.4	873.2	100	$(C_{28}H_{23}N_3O_{62}P_2V_3W_{15})$

(\mathbf{R} = reduced form of the POM)



POM-dvb4,4 : prepared with 70% yield.

IR (KBr, cm⁻¹): 2962, 2874, 2216, 1632, 1595, 1484, 1087, 953, 903, 816, 737.

³¹P NMR (162MHz, CD₃CN) δ: -7.54, -13.51.

¹H NMR (400 MHz, CD₃CN) δ: 8.66 (d, 4H, J=4.9 Hz, H_{py}), 8.29 (m, 3H, H_{Ar} + C**H**=CH), 7.73 (s, 1H, H_{Ar}), 7.58 (d, 4H, J = 4.9 Hz, H_{py}), 6.96 (d, 1H, J = 15,7Hz, CH=C**H**), 5.55 (d, 2H, J = 11.9 Hz, C**H**₂OV), 5.41 (d, 2H, J=11.9 Hz, C**H**₂OV), 3.19 (m, 51H, ⁺N(C**H**₂CH₂CH₂CH₃)₄), 1.66 (m, 53H, ⁺N(CH₂C**H**₂CH₂CH₃)₄ + C**H**₂-CH₃), 1.43 (q, 51H, J = 7.0Hz, ⁺N(CH₂CH₂CH₂CH₃)₄), 1.13 (t, 3H, J = 7.1Hz, CH₂-C**H**₃), 1.00 (t, 75H, J=7.0Hz, ⁺N(CH₂CH₂CH₃)₄).

¹³C NMR (100 MHz, CD₃CN) δ : 151,1 (C_{py}), 151.0 (CH=CH), 138.5 (C_q), 135.2 (CH_{Ar}), 134,1 (CH_{Ar}), 131.5 (C_q), 126.6 (CH_{py}), 124.4 (C_q), 116.8 (CH=CH), 93.2 (C=C), 90.3 (CH₂OV), 88.9 (C=C), 63.1 (C_q), 59.3 (⁺N(CH₂CH₂CH₂CH₃)₄), 32.6 (CH₂-CH₃), 24,4 (⁺N(CH₂CH₂CH₂CH₂CH₃)₄), 20.4 (⁺N(CH₂CH₂CH₃)₄), 14.0 (⁺N(CH₂CH₂CH₂CH₃)₄), 8.02 (CH₂-CH₃).

Anal. calc. for $TBA_5(C_{28}H_{23}N_3O_{62}P_2V_3W_{15})$: C 23.25, H 3.67, N 2.01 ; found: C 23.35, H 3.58, N 2.05 Electrospray mass spectrometry:

Entry	Charge	Simulated m/z	Observed m/z	Relative Intensity (%)	Composition
1	3-	1697.9	1697.9	12	$TBA_3(C_{28}H_{23}N_3O_{62}P_2V_3W_{15})$ R
2	3-	1617.2	1617.0	20	$TBA_2(C_{28}H_{23}N_3O_{62}P_2V_3W_{15})$
3	4-	1212.9	1212.9	54	$TBA_2 (C_{28}H_{23}N_3O_{62}P_2V_3W_{15}) \mathbf{R}$
4	4-	1152.3	1152.1	100	$TBA(C_{28}H_{23}N_3O_{62}P_2V_3W_{15})$
5	4-	1092.0	1088.3	27	$H(C_{28}H_{23}N_3O_{62}P_2V_3W_{15})$
6	5-	873.4	873.4	74	$(C_{28}H_{23}N_3O_{62}P_2V_3W_{15})$

 $(\mathbf{R} = \text{reduced form of the POM})$



POM-dvb3,4 prepared with 46% yield.

IR (KBr, cm⁻¹): 3449, 2962, 2874, 1634, 1595, 1483, 1087, 953, 902, 816, 737.

³¹P NMR (162 MHz, CD₃CN) δ: -7.26; -13.3.

¹H NMR (500 MHz, CD₃CN) δ : 8.78 (s, 1H, H_{3py}), 8.67 (m, 3H, J = 4.5 Hz, C**H**=CH + H_{4py}), 8.59 (d, 1H, J = 3.65 Hz, H_{3py}), 8.54 (s, 1H, H_{Ar}), 8.28 (s, 1H, H_{Ar}), 8.10 (d, 1H, J = 7.65, H_{3py}), 7.82 (s, 1H, H_{Ar}), 7.59 (d, 2H, J = 4.25 Hz, H_{4py}), 7.52 (m, 1H, H_{3py}), 5.60 (d, 2H, J = 12.8 Hz, C**H**₂OV), 5.46 (d, 2H, J = 12.2 Hz, , C**H**₂OV), 3.18 (m, 42H, ⁺N(C**H**₂CH₂CH₂CH₃)₄), 1.86 (m, 2H, C**H**₂-CH₃), 1.65 (m, 42H, ⁺N(CH₂CH₂CH₂CH₃)₄), 1.13 (m, 3H, CH₂-C**H**₃), 1.00 (t, 62H, J=7.0Hz, ⁺N(CH₂CH₂CH₂C**H**₃)₄).

¹³C NMR (100 MHz, CD₃CN) δ : 181.6 (CO), 154.1 (CH=CH), 152.7 (C_{3py}), 151.2 (C_{4py}), 150.2 (C_{3py}), 140.2 (C_{3py}), 136.9 (CH_{Ar}), 136.5 (C_{qAr}-CH=CH), 135.3 (CH_{Ar}), 134.1 (CH_{Ar}), 131.4 (C_{q4py}), 126.6 (C_{4py}), 125.2 (C_{qAr=-3py}), 124.9 (C_{3py}), 124.8 (C_{qAr=-4py}), 120.6 (C_{q3py}), 118.3 (CH=CH, masked), 92.8 (C=C), 91.6 (C=C), 89.8 (CH₂OV), 89.4 (C=C), 88.9 (C=C), 64.8 (Cq), 29.5 (CH₂-CH₃), 7.56 (CH₂-CH₃).

Anal. calc. for TBA₅(C₂₈H₂₃N₃O₆₂P₂V₃W₁₅) : C 23.25, H 3.67, N 2.01 ; found: C 23.26, H 3.65, N 1.87.

Electrospray mass spectrometry:

Entry	Charge	Simulated m/z	Observed m/z	Relative Intensity (%)	Composition
1	3-	1617.2	1617.9	25	$TBA_2(C_{28}H_{23}N_3O_{62}P_2V_3W_{15})$
2	4-	1152.3	1152.6	100	$TBA(C_{28}H_{23}N_3O_{62}P_2V_3W_{15})$
3	4-	1092.0	1091.3	7.6	$H(C_{28}H_{23}N_3O_{62}P_2V_3W_{15})$
3	5-	873.4	872.9	98	$(C_{28}H_{23}N_3O_{62}P_2V_3W_{15})$



POM-tvb3,3 prepared in 40% yield.

IR (KBr, cm⁻¹): 3455, 2962, 2873, 1632, 1483, 1086, 950, 911, 818, 734.

³¹P NMR (162 MHz, CD₃CN) δ: -7.45; -13.54.

¹H NMR (400 MHz, CD₃CN) δ : 8.75 (s, 2H, H_{py}), 8.58 (d, 2H, J=3.8 Hz, H_{py}), 7.91 (m, 2H, H_{py}), 7.71 (m, 3H, H_{Ar}), 7.50 (d, 1H, J=15.7 Hz, C**H**=CH), 7.40 (dd, 2H, J=4.9Hz, J= 7.7 Hz, H_{py}), 6.66 (d, 1H, J=15.6 Hz, CH=C**H**), 6.42 (s, 1H, N**H**), 5.73 (s, 6H, C**H**₂OV), 3.8 (m, 59H, ⁺N(C**H**₂CH₂CH₂CH₃)₄), 1.65 (quint, 59H, J= 7.7 Hz, ⁺N(CH₂C**H**₂CH₂CH₃)₄), 1.43 (sext, 59H, J = 7.3Hz, ⁺N(CH₂CH₂CH₂CH₂CH₃)₄), 0.99 (t, 89H, J=7.3Hz, ⁺N(CH₂CH₂CH₂CH₂CH₃)₄).

¹³C NMR (100 MHz, CD₃CN) δ: 166.0 (C=O), 152.9 (C_{py}), 150.2 (C_{py}), 139.6 (C_{py}), 138.7 (CH=CH), 137.5 (C_{qAr}-CH=CH), 135.6 (CH_{Ar(1H)}), 131.7 (CH_{Ar(2H)}), 125.3 (CH=CH), 124.7 (C_{qAr}-3py), 124.5 (C_{py}), 120.5 (C_{q3py}), 91.4 (C=C), 88.1 (C=C), 86.9 (CH₂OV), 59.3 (⁺N(CH₂CH₂CH₂CH₃)₄), 55.6 (C_q), 29.5 (CH₂-CH₃), 24,5 (⁺N(CH₂CH₂CH₂CH₃)₄), 20.5 (⁺N(CH₂CH₂CH₂CH₃)₄), 14.0 (⁺N(CH₂CH₂CH₂CH₃)₄).

Anal. calc. for $TBA_{5.7}H_{0.3}(C_{27}H_{20}N_3O_{63}P_2V_3W_{15})$ (5749.2 g.mol⁻¹): C 24.69, H 3.95, N 2.12 ; found: C 24.36, H 4.01, N 2.07.

Electrospray mass spectrometry:

Entry	Charge	Simulated m/z	Observed m/z	Relative Intensity (%)	Composition
1	3-	1698.2	1698.0	11	$TBA_{3}(C_{27}H_{20}N_{3}O_{63}P_{2}V_{3}W_{15})$
2	4-	1213.1	1212.9	68	$TBA_2(C_{27}H_{20}N_3O_{63}P_2V_3W_{15})$
3	4-	1152.8	1152.3	5	$TBAH(C_{27}H_{20}N_3O_{63}P_2V_3W_{15})$
4	4-	1092.5	1091.3	18	$H_2(C_{27}H_{20}N_3O_{63}P_2V_3W_{15})$
5	5-	922.0	921.7	100	$TBA(C_{27}H_{20}N_3O_{63}P_2V_3W_{15})$
6	5-	873.8	874.2	5	$H(C_{27}H_{20}N_3O_{63}P_2V_3W_{15})$

NMR Spectra

dvb3,3:



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Electronic Supplementary Material (ESI) for Dalton Transactions This journal is O The Royal Society of Chemistry 2013





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Electronic Supplementary Material (ESI) for Dalton Transactions This journal is O The Royal Society of Chemistry 2013

POM-dvb3,3:







POM-dvb4,4:







POM-dvb3,4:







POM-tvb3,3:









Fig S1 UV-Vis absorption spectra of a) POM-dvb3,3 , b) POM-dvb3,4 , c) POM-dvb3,4 , d) POM-tvb3,3 compared to $TBA_5H_4[P_2V_3W_{15}O_{62}]$ and their corresponding non-grafted ligand. Concentration is 6.25 10^{-6} M in CH₃CN.

Fabrication of Films

Copolymer Poly-ZnOEP-POM

Electropolymerization was performed under an argon atmosphere in a 0.1 mol L^{-1} solution of tetrabutylammonium hexafluorophosphate in 1,2-C₂H₄Cl₂/CH₃CN (7:3) containing 0.25 mmol L^{-1} ZnOEP and 0.25 mmol L^{-1} POM(py)₂ using iterative scans. Cyclic scanning (0.2 V s⁻¹) was applied at potentials between -1.30 and 1.90 V versus SCE or between 0 and 1.90 V versus SCE. The starting potential of the first scan and the ending potential value of the final scan were 0.0 and 0.5 V, respectively. Thus, at the end of the electropolymerization, the porphyrin subunits (ZnOEP) of the polymers were not oxidized (neutral form). The direction of the first scan was cathodic in order to record the eventual presence of reduction signals attributed to the pyridinium unit(s) before polymer formation. Polymers were obtained during the reverse anodic scan when the potential reached the oxidation potential value of the porphyrin ligand corresponding to the dications formation. After electrolysis, the working electrodes were washed five times with 10 mL of CH₃CN to remove traces of the conducting salt present on the deposited films.



CV evolution during electropolymerization



Fig S2 Cyclic voltammograms recorded during the electropolymerization of ZnOEP in the presence of a), b) **POM-dvb3,3**, c) and d) **POM-dvb3,4**, e) and f) **POM-dvb4,4**, g and h) **POM-tvb3,3** in 1,2-C2H4Cl2/CH3CN (7:3) (NBu4)PF6 0.1mol L⁻¹. Working electrode: ITO. S = 1 cm2. Scan rate: 0.1 V.s⁻¹. (\leftarrow) Start of the scan. Cyclic scanning was applied at potentials between -1.30 (-1.35 V for **POM-dvb4,4**) and 1.90 V *vs.* SCE (left) or between -0.05 V and 1.90 V *vs.* SCE (right).

Permeability



Fig. S3 Cyclic voltammograms of 1 mM of $K_3Fe(CN)_6$, in 0.5 M Na₂SO₄ of modified ITO electrode with a) poly-POM-dvb3,3-ZnOEP b) poly-POM-dvb3,4-ZnOEP, c) poly-POM-dvb4,4-ZnOEP, d) poly-POM-tvb3,3-ZnOEP film (full line) and of noncoated ITO electrode (dotted black line). $v=100 \text{ mV s}^{-1}$.

Reduction potentials explanation



Fig. S4 Tentative explanation of the poly-POM-ZnOEP reduction potential in term of π -conjugation and ligand geometry.

UV-vis spectra of poly-POM-ZnOEP



Fig S5 Normalized UV-vis spectra of a) **poly-POM-dvb3,3-ZnOEP**, b) **poly-POM-dvb3,4-ZnOEP**, c) **poly-POM-dvb4,4-ZnOEP**, and d) **poly-POM-tvb3,3-ZnOEP** copolymers deposited on ITO electrode after 25 iterative scans between -0.05 V and 1.90 V *vs*. SCE (plain line), and in DMF solution (dotted line). For comparison, normalized UV-Vis spectrum of ZnOEP in DMF is plotted on each graph.

UV-vis evolution during electropolymerization



Fig. S6 UV-visible absorption spectra of a) **poly-POM-dvb3,3-ZnOEP**, b) **poly-POM-dvb3,4-ZnOEP**, c) **poly-POM-dvb4,4-ZnOEP** and d) **poly-POM-tvb3,3-ZnOEP** (onto ITO) with different numbers of iterative scans (between -0.05 V and 1.90 V *vs.* SCE). Only one side is recovered by ITO.

XPS



Fig. S7-A XPS spectra of the modified ITO electrodes with **poly-POM-dvb3,4-ZnOEP** obtained after 25 iterative scans between -0.05 V and 1.90 V *vs.* SCE.) Global XPS spectra, and C1s, O1s, N1s, P 2p3, W 4f7, V 2p3 region.



Fig. S7-B XPS spectra of the modified ITO electrodes with **poly-POM-dvb4,4-ZnOEP** obtained after 25 iterative scans between -0.05 V and 1.90 V *vs.* SCE.) Global XPS spectra, and C1s, O1s, N1s, P 2p3, W 4f7, V 2p3 region.

AFM





Fig S8 Tapping mode AFM topography of a) and b) **poly-POM-dvb3,3-ZnOEP**, c) and d) **poly-POM-dvb3,4-ZnOEP**, e) and f) **poly-POM-dvb4,4-ZnOEP**, g) and h) **poly-POM-tvb3,3-ZnOEP** copolymer films deposited on ITO electrode after 25 iterative scans between -0.05 V and 1.90 V *vs*. SCE. Section analyses of the marked white line on the images are reported under each image.