

## Supplementary Information

### Appendix I

#### (A) ESI-MS spectra (Figs. S1.1-S1.4)

Figure S1.1 ESI-MS spectrum of  $[\text{Cu}(\text{phen})(\text{L-Threo})(\text{H}_2\text{O})]\text{NO}_3$  1

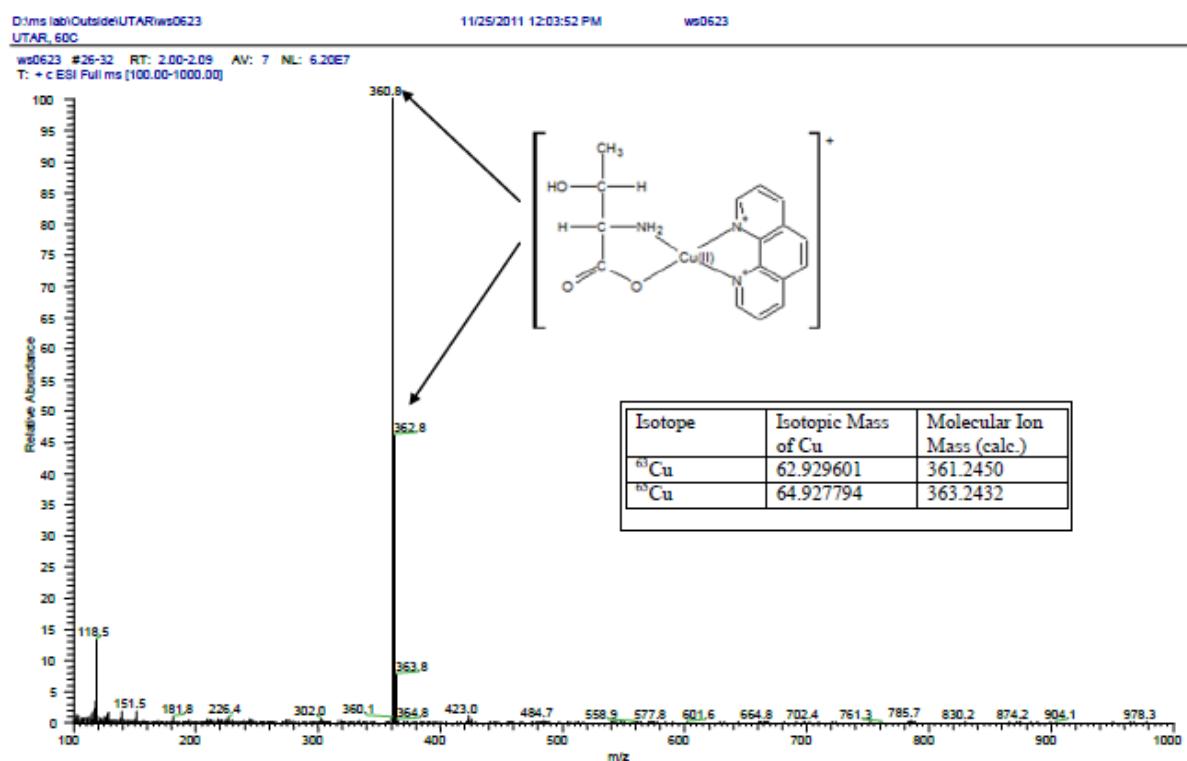


Figure S1.2 ESI-MS spectrum of  $[\text{Cu}(\text{phen})(\text{D-Threo})(\text{H}_2\text{O})]\text{NO}_3$  2

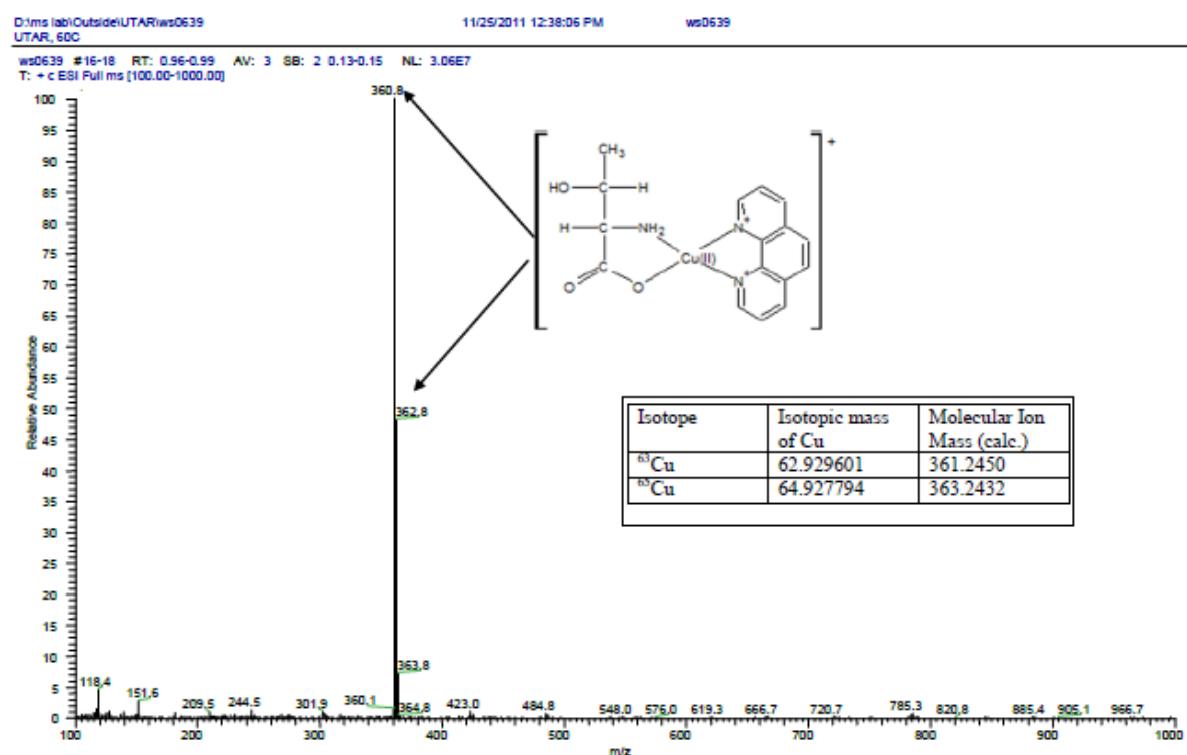


Figure S1.3 ESI-MS spectrum of L-[Cu(phen)(5MeOCA)(H<sub>2</sub>O)]NO<sub>3</sub> 3

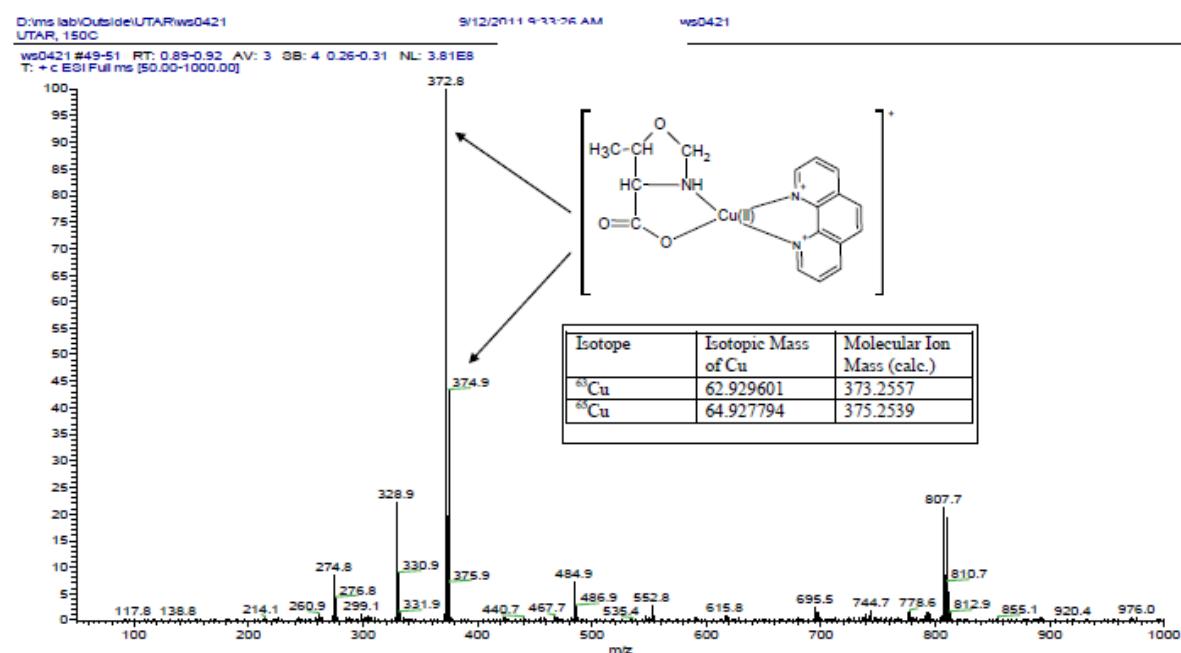
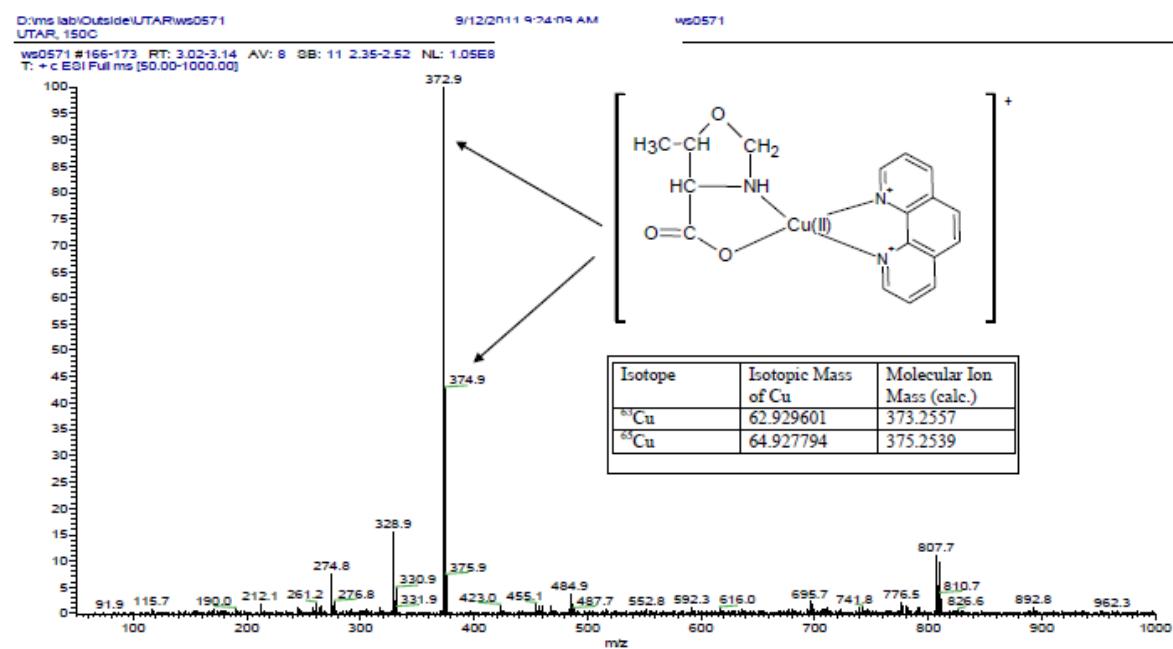
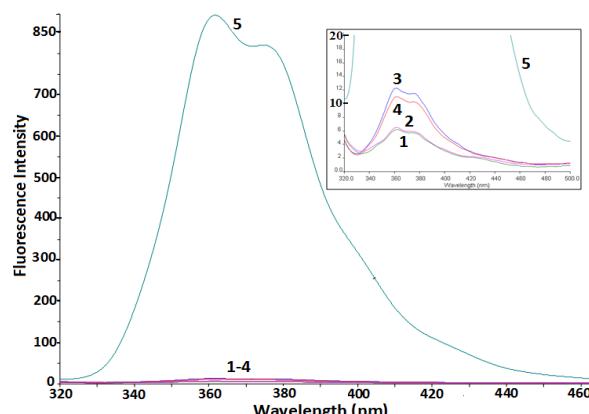


Figure 1.4 ESI-MS spectrum of D-[Cu(phen)(5MeOCA)(H<sub>2</sub>O)]NO<sub>3</sub> 4





Supplementary Fig. S2 Fluorescence spectra of  $1 \times 10^{-5}$  M **1 – 5** in water-methanol (1:1 v/v). Insert shows clearly the FL spectra of **1 – 4**.

(B) Restriction Enzyme inhibition by CuCl<sub>2</sub>, [Cu(phen)Cl<sub>2</sub>], and copper(II) complexes **1-4** (Figs. S3.1-S3.6)

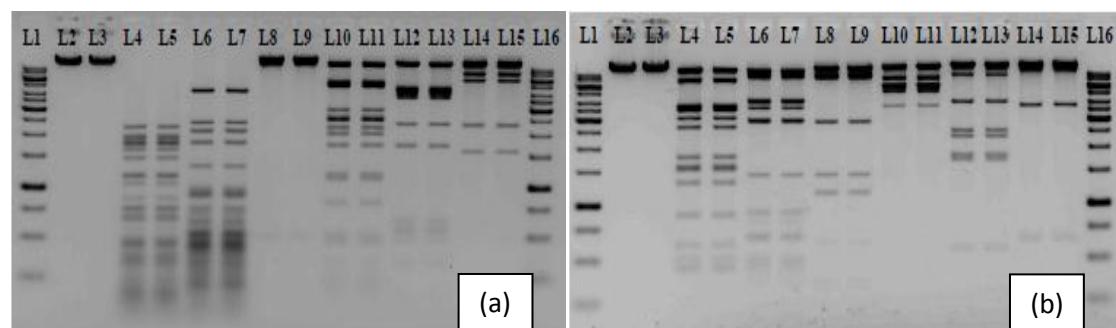


Figure S3.1 RE inhibition by CuCl<sub>2</sub>. Electrophoresis results of incubating  $\lambda$  DNA (0.5  $\mu$ g/ $\mu$ L) with 5 unit of restriction enzyme in the presence or absence of 50  $\mu$ M CuCl<sub>2</sub> for 2 hours at 37°C.

(a) Lane 1, 1kb DNA ladder; Lane 2,  $\lambda$  DNA alone (0.5  $\mu$ g); Lane 3,  $\lambda$  DNA + 50  $\mu$ M CuCl<sub>2</sub>; Lane 4,  $\lambda$  DNA + 5 unit Tsp 509I (control); Lane 5,  $\lambda$  DNA + 5 unit Tsp 509I + 50  $\mu$ M CuCl<sub>2</sub>; Lane 6,  $\lambda$  DNA + 5 unit Hae III (control); Lane 7,  $\lambda$  DNA + 5 unit Hae III + 50  $\mu$ M CuCl<sub>2</sub>; Lane 8,  $\lambda$  DNA + 5 unit Sal I (control); Lane 9,  $\lambda$  DNA + 5 unit Sal I + 50  $\mu$ M CuCl<sub>2</sub>; Lane 10,  $\lambda$  DNA + 5 unit Pst I (control); Lane 11,  $\lambda$  DNA + 5 unit Pst I + 50  $\mu$ M CuCl<sub>2</sub>; Lane 12,  $\lambda$  DNA + 5 unit Pvu II (control); Lane 13,  $\lambda$  DNA + 5 unit Pvu II + 50  $\mu$ M CuCl<sub>2</sub>; Lane 14,  $\lambda$  DNA + 5 unit Sca I (control); Lane 15,  $\lambda$  DNA + 5 unit Sca I + 50  $\mu$ M CuCl<sub>2</sub>; Lane 16, 1kb DNA ladder.

(b) Lane 1, 1kb DNA ladder; Lane 2,  $\lambda$  DNA alone (0.5  $\mu$ g); Lane 3,  $\lambda$  DNA + 50  $\mu$ M CuCl<sub>2</sub>; Lane 4,  $\lambda$  DNA + 5 unit Ssp I (control); Lane 5,  $\lambda$  DNA + 5 unit Ssp I + 50  $\mu$ M CuCl<sub>2</sub>; Lane 6,  $\lambda$  DNA + 5 unit Ase I (control); Lane 7,  $\lambda$  DNA + 5 unit Ase I + 50  $\mu$ M CuCl<sub>2</sub>; Lane 8,  $\lambda$  DNA + 5 unit Mun I (control); Lane 9,  $\lambda$  DNA + 5 unit Mun I + 50  $\mu$ M CuCl<sub>2</sub>; Lane 10,  $\lambda$  DNA + 5 unit EcoR I (control); Lane 11,  $\lambda$  DNA + 5 unit EcoR I + 50  $\mu$ M metal salt; Lane 12,  $\lambda$  DNA + 5 unit NdeI (control); Lane 13,  $\lambda$  DNA + 5 unit NdeI + 50  $\mu$ M CuCl<sub>2</sub>; Lane 14,  $\lambda$  DNA + 5 unit Bst 11071 (control); Lane 15,  $\lambda$  DNA + 5 unit Bst 11071 + 50  $\mu$ M CuCl<sub>2</sub>; Lane 16, 1kb DNA ladder.

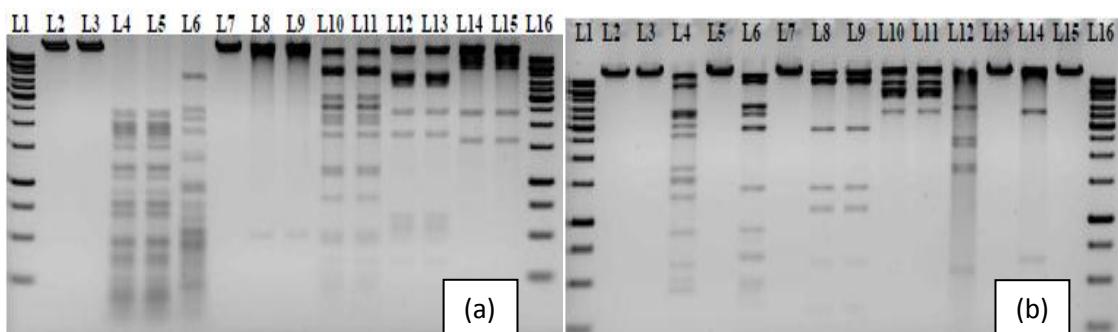


Figure S3.2 RE inhibition by  $[\text{Cu}(\text{phen})\text{Cl}_2]$ . Electrophoresis results of incubating  $\lambda$  DNA ( $0.5 \mu\text{g}/\mu\text{L}$ ) with 5 unit of restriction enzyme in the presence or absence of  $50 \mu\text{M}$   $[\text{Cu}(\text{phen})\text{Cl}_2]$  for 2 hours at  $37^\circ\text{C}$ .

(a) Lane 1, 1kb DNA ladder; Lane 2,  $\lambda$  DNA alone ( $0.5 \mu\text{g}$ ); Lane 3,  $\lambda$  DNA +  $50 \mu\text{M}$   $[\text{Cu}(\text{phen})\text{Cl}_2]$ ; Lane 4,  $\lambda$  DNA + 5 unit Tsp 509I (control); Lane 5,  $\lambda$  DNA + 5 unit Tsp 509I +  $50 \mu\text{M}$   $[\text{Cu}(\text{phen})\text{Cl}_2]$ ; Lane 6,  $\lambda$  DNA + 5 unit Hae III (control); Lane 7,  $\lambda$  DNA + 5 unit Hae III +  $50 \mu\text{M}$   $[\text{Cu}(\text{phen})\text{Cl}_2]$ ; Lane 8,  $\lambda$  DNA + 5 unit Sal I (control); Lane 9,  $\lambda$  DNA + 5 unit Sal I +  $50 \mu\text{M}$   $[\text{Cu}(\text{phen})\text{Cl}_2]$ ; Lane 10,  $\lambda$  DNA + 5 unit Pst I (control); Lane 11,  $\lambda$  DNA + 5 unit Pst I +  $50 \mu\text{M}$   $[\text{Cu}(\text{phen})\text{Cl}_2]$ ; Lane 12,  $\lambda$  DNA + 5 unit Pvu II (control); Lane 13,  $\lambda$  DNA + 5 unit Pvu II +  $50 \mu\text{M}$   $[\text{Cu}(\text{phen})\text{Cl}_2]$ ; Lane 14,  $\lambda$  DNA + 5 unit Sca I (control); Lane 15,  $\lambda$  DNA + 5 unit Sca I +  $50 \mu\text{M}$   $[\text{Cu}(\text{phen})\text{Cl}_2]$ ; Lane 16, 1kb DNA ladder.

(b) Lane 1, 1kb DNA ladder; Lane 2,  $\lambda$  DNA alone ( $0.5 \mu\text{g}$ ); Lane 3,  $\lambda$  DNA +  $50 \mu\text{M}$   $[\text{Cu}(\text{phen})\text{Cl}_2]$ ; Lane 4,  $\lambda$  DNA + 5 unit Ssp I (control); Lane 5,  $\lambda$  DNA + 5 unit Ssp I +  $50 \mu\text{M}$   $[\text{Cu}(\text{phen})\text{Cl}_2]$ ; Lane 6,  $\lambda$  DNA + 5 unit Ase I (control); Lane 7,  $\lambda$  DNA + 5 unit Ase I +  $50 \mu\text{M}$   $[\text{Cu}(\text{phen})\text{Cl}_2]$ ; Lane 8,  $\lambda$  DNA + 5 unit Mun I (control); Lane 9,  $\lambda$  DNA + 5 unit Mun I +  $50 \mu\text{M}$   $[\text{Cu}(\text{phen})\text{Cl}_2]$ ; Lane 10,  $\lambda$  DNA + 5 unit EcoR I (control); Lane 11,  $\lambda$  DNA + 5 unit EcoR I +  $50 \mu\text{M}$   $[\text{Cu}(\text{phen})\text{Cl}_2]$ ; Lane 12,  $\lambda$  DNA + 5 unit NdeI (control); Lane 13,  $\lambda$  DNA + 5 unit NdeI +  $50 \mu\text{M}$   $[\text{Cu}(\text{phen})\text{Cl}_2]$ ; Lane 14,  $\lambda$  DNA + 5 unit Bst 11071 (control); Lane 15,  $\lambda$  DNA + 5 unit Bst 11071 +  $50 \mu\text{M}$   $[\text{Cu}(\text{phen})\text{Cl}_2]$ ; Lane 16, 1kb DNA ladder.

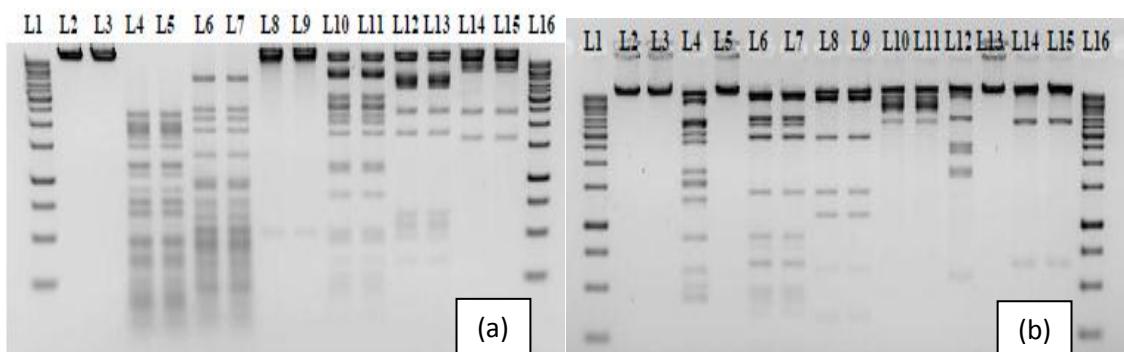


Figure S3.3 RE inhibition by **1**. Electrophoresis results of incubating λ DNA (0.5 µg/µL) with 5 unit of restriction enzyme in the presence or absence of 50 µM **1** for 2 hours at 37°C.

(a) Lane 1, 1kb DNA ladder; Lane 2, λ DNA alone (0.5 µg); Lane 3, λ DNA + 50 µM **1**; Lane 4, λ DNA + 5 unit Tsp 509I (control); Lane 5, λ DNA + 5 unit Tsp 509I + 50 µM **1**; Lane 6, λ DNA + 5 unit Hae III (control); Lane 7, λ DNA + 5 unit Hae III + 50 µM **1**; Lane 8, λ DNA + 5 unit Sal I (control); Lane 9, λ DNA + 5 unit Sal I + 50 µM **1**; Lane 10, λ DNA + 5 unit Pst I (control); Lane 11, λ DNA + 5 unit Pst I + 50 µM **1**; Lane 12, λ DNA + 5 unit Pvu II (control); Lane 13, λ DNA + 5 unit Pvu II + 50 µM **1**; Lane 14, λ DNA + 5 unit Sca I (control); Lane 15, λ DNA + 5 unit Sca I + 50 µM **1**; Lane 16, 1kb DNA ladder.

(b) Lane 1, 1kb DNA ladder; Lane 2, λ DNA alone (0.5 µg); Lane 3, λ DNA + 50 µM **1**; Lane 4, λ DNA + 5 unit Ssp I (control); Lane 5, λ DNA + 5 unit Ssp I + 50 µM **1**; Lane 6, λ DNA + 5 unit Ase I (control); Lane 7, λ DNA + 5 unit Ase I + 50 µM **1**; Lane 8, λ DNA + 5 unit Mun I (control); Lane 9, λ DNA + 5 unit Mun I + 50 µM **1**; Lane 10, λ DNA + 5 unit EcoR I (control); Lane 11, λ DNA + 5 unit EcoR I + 50 µM **1**; Lane 12, λ DNA + 5 unit NdeI (control); Lane 13, λ DNA + 5 unit NdeI + 50 µM **1**; Lane 14, λ DNA + 5 unit Bst 11071 (control); Lane 15, λ DNA + 5 unit Bst 11071 + 50 µM **1**; Lane 16, 1kb DNA ladder.

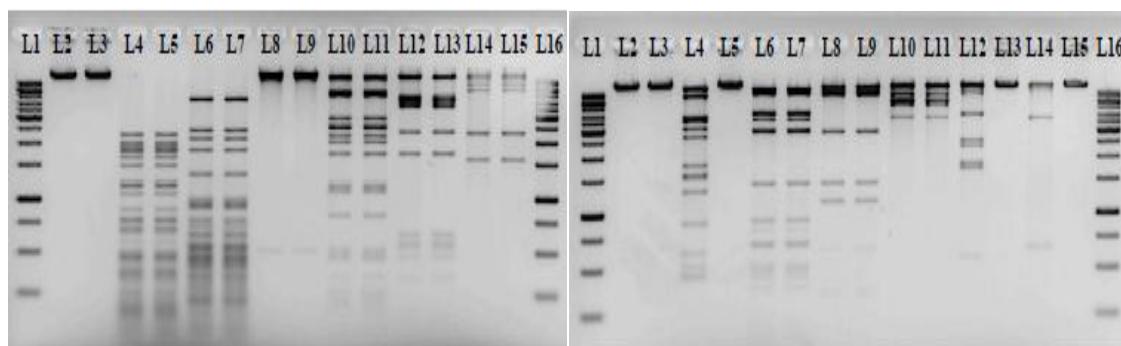


Figure S3.4 RE inhibition by **2**.

Electrophoresis results of incubating  $\lambda$  DNA (0.5  $\mu$ g/ $\mu$ L) with 5 unit of restriction enzyme in the presence or absence of 50  $\mu$ M **2** for 2 hours at 37°C.

(a) Lane 1, 1kb DNA ladder; Lane 2,  $\lambda$  DNA alone (0.5  $\mu$ g); Lane 3,  $\lambda$  DNA + 50  $\mu$ M **2**; Lane 4,  $\lambda$  DNA + 5 unit Tsp 509I (control); Lane 5,  $\lambda$  DNA + 5 unit Tsp 509I + 50  $\mu$ M **2**; Lane 6,  $\lambda$  DNA + 5 unit Hae III (control); Lane 7,  $\lambda$  DNA + 5 unit Hae III + 50  $\mu$ M **2**; Lane 8,  $\lambda$  DNA + 5 unit Sal I (control); Lane 9,  $\lambda$  DNA + 5 unit Sal I + 50  $\mu$ M **2**; Lane 10,  $\lambda$  DNA + 5 unit Pst I (control); Lane 11,  $\lambda$  DNA + 5 unit Pst I + 50  $\mu$ M **2**; Lane 12,  $\lambda$  DNA + 5 unit Pvu II (control); Lane 13,  $\lambda$  DNA + 5 unit Pvu II + 50  $\mu$ M **2**; Lane 14,  $\lambda$  DNA + 5 unit Sca I (control); Lane 15,  $\lambda$  DNA + 5 unit Sca I + 50  $\mu$ M **2**; Lane 16, 1kb DNA ladder.

(b) Lane 1, 1kb DNA ladder; Lane 2,  $\lambda$  DNA alone (0.5  $\mu$ g); Lane 3,  $\lambda$  DNA + 50  $\mu$ M **2**; Lane 4,  $\lambda$  DNA + 5 unit Ssp I (control); Lane 5,  $\lambda$  DNA + 5 unit Ssp I + 50  $\mu$ M **3**; Lane 6,  $\lambda$  DNA + 5 unit Ase I (control); Lane 7,  $\lambda$  DNA + 5 unit Ase I + 50  $\mu$ M **3**; Lane 8,  $\lambda$  DNA + 5 unit Mun I (control); Lane 9,  $\lambda$  DNA + 5 unit Mun I + 50  $\mu$ M **2**; Lane 10,  $\lambda$  DNA + 5 unit EcoR I (control); Lane 11,  $\lambda$  DNA + 5 unit EcoR I + 50  $\mu$ M **2**; Lane 12,  $\lambda$  DNA + 5 unit NdeI (control); Lane 13,  $\lambda$  DNA + 5 unit NdeI + 50  $\mu$ M **2**; Lane 14,  $\lambda$  DNA + 5 unit Bst 11071 (control); Lane 15,  $\lambda$  DNA + 5 unit Bst 11071 + 50  $\mu$ M **2**; Lane 16, 1kb DNA ladder.

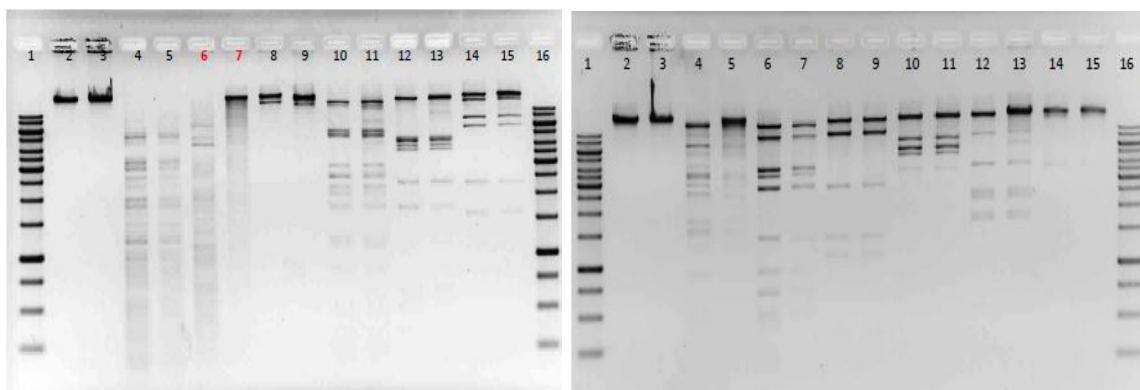


Figure S3.5 RE inhibition by **3**.

Electrophoresis results of incubating  $\lambda$  DNA ( $0.5 \mu\text{g}/\mu\text{L}$ ) with 5 unit of restriction enzyme in the presence or absence of  $50 \mu\text{M}$  **3** for 2 hours at  $37^\circ\text{C}$ .

(a) Lane 1, 1kb DNA ladder; Lane 2,  $\lambda$  DNA alone ( $0.5 \mu\text{g}$ ); Lane 3,  $\lambda$  DNA +  $50 \mu\text{M}$  **3**; Lane 4,  $\lambda$  DNA + 5 unit Tsp 509I (control); Lane 5,  $\lambda$  DNA + 5 unit Tsp 509I +  $50 \mu\text{M}$  **3**; Lane 6,  $\lambda$  DNA + 5 unit Hae III (control); Lane 7,  $\lambda$  DNA + 5 unit Hae III +  $50 \mu\text{M}$  **3**; Lane 8,  $\lambda$  DNA + 5 unit Sal I (control); Lane 9,  $\lambda$  DNA + 5 unit Sal I +  $50 \mu\text{M}$  **3**; Lane 10,  $\lambda$  DNA + 5 unit Pst I (control); Lane 11,  $\lambda$  DNA + 5 unit Pst I +  $50 \mu\text{M}$  **3**; Lane 12,  $\lambda$  DNA + 5 unit Pvu II (control); Lane 13,  $\lambda$  DNA + 5 unit Pvu II +  $50 \mu\text{M}$  **3**; Lane 14,  $\lambda$  DNA + 5 unit Sca I (control); Lane 15,  $\lambda$  DNA + 5 unit Sca I +  $50 \mu\text{M}$  **3**; Lane 16, 1kb DNA ladder.

(b) Lane 1, 1kb DNA ladder; Lane 2,  $\lambda$  DNA alone ( $0.5 \mu\text{g}$ ); Lane 3,  $\lambda$  DNA +  $50 \mu\text{M}$  **3**; Lane 4,  $\lambda$  DNA + 5 unit Ssp I (control); Lane 5,  $\lambda$  DNA + 5 unit Ssp I +  $50 \mu\text{M}$  **3**; Lane 6,  $\lambda$  DNA + 5 unit Ase I (control); Lane 7,  $\lambda$  DNA + 5 unit Ase I +  $50 \mu\text{M}$  **3**; Lane 8,  $\lambda$  DNA + 5 unit Mun I (control); Lane 9,  $\lambda$  DNA + 5 unit Mun I +  $50 \mu\text{M}$  **3**; Lane 10,  $\lambda$  DNA + 5 unit EcoR I (control); Lane 11,  $\lambda$  DNA + 5 unit EcoR I +  $50 \mu\text{M}$  **3**; Lane 12,  $\lambda$  DNA + 5 unit NdeI (control); Lane 13,  $\lambda$  DNA + 5 unit NdeI +  $50 \mu\text{M}$  **3**; Lane 14,  $\lambda$  DNA + 5 unit Bst 11071 (control); Lane 15,  $\lambda$  DNA + 5 unit Bst 11071 +  $50 \mu\text{M}$  **3**; Lane 16, 1kb DNA ladder.

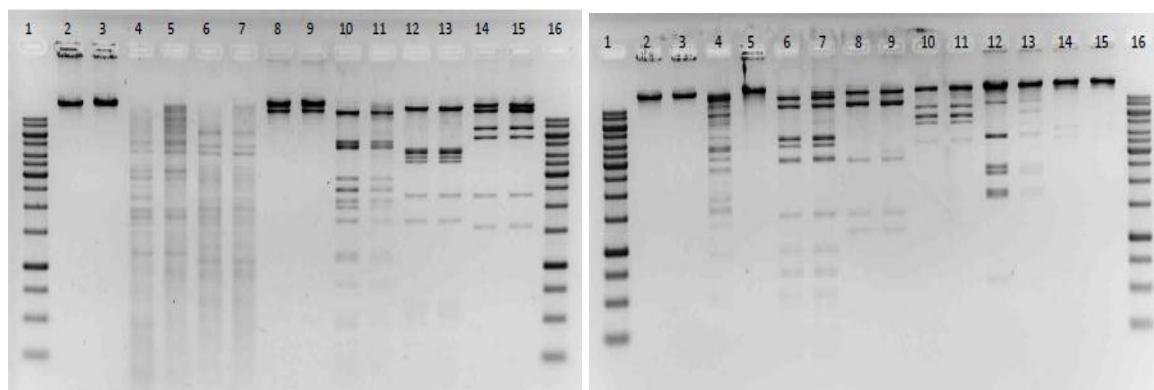


Figure S3.6 RE inhibition by **4**.

Electrophoresis results of incubating  $\lambda$  DNA ( $0.5 \mu\text{g}/\mu\text{L}$ ) with 5 unit of restriction enzyme in the presence or absence of  $50 \mu\text{M}$  **4** for 2 hours at  $37^\circ\text{C}$ .

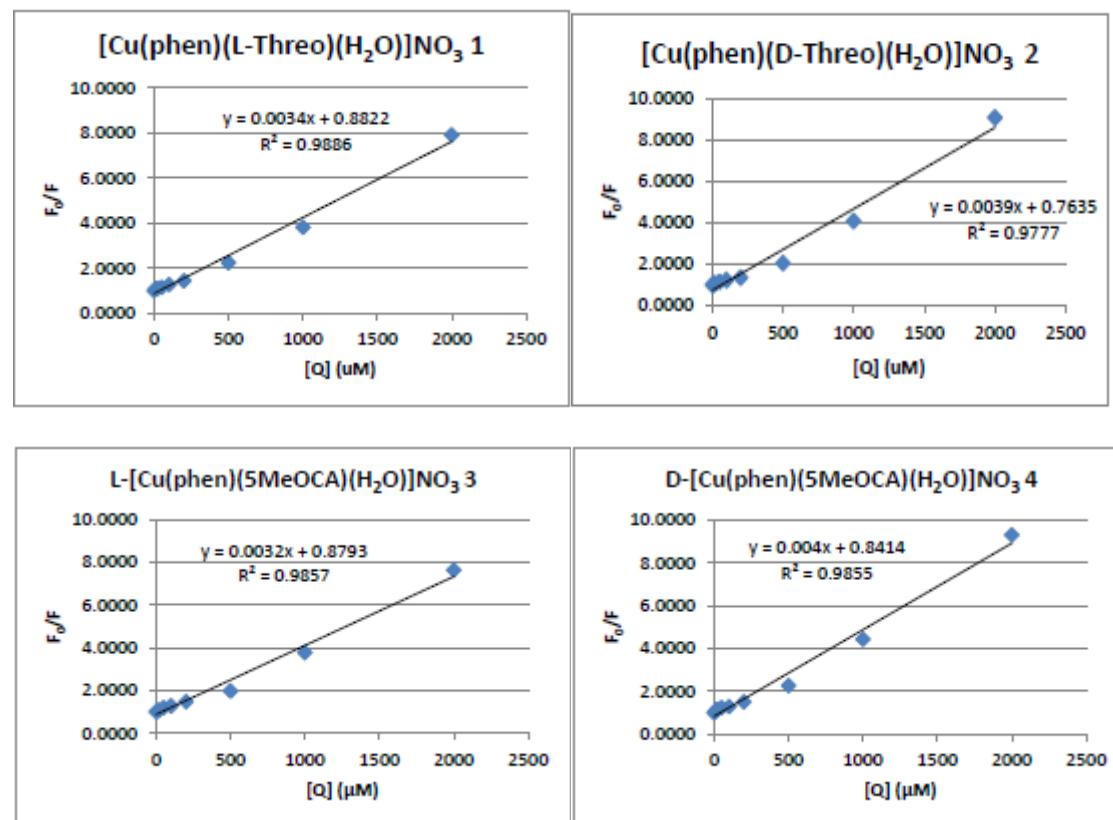
(a) Lane 1, 1kb DNA ladder; Lane 2,  $\lambda$  DNA alone ( $0.5 \mu\text{g}$ ); Lane 3,  $\lambda$  DNA +  $50 \mu\text{M}$  **4**; Lane 4,  $\lambda$  DNA + 5 unit Tsp 509I (control); Lane 5,  $\lambda$  DNA + 5 unit Tsp 509I +  $50 \mu\text{M}$  **4**; Lane 6,  $\lambda$  DNA + 5 unit Hae III (control); Lane 7,  $\lambda$  DNA + 5 unit Hae III +  $50 \mu\text{M}$  **4**; Lane 8,  $\lambda$  DNA + 5 unit Sal I (control); Lane 9,  $\lambda$  DNA + 5 unit Sal I +  $50 \mu\text{M}$  **4**; Lane 10,  $\lambda$  DNA + 5 unit Pst I (control); Lane 11,  $\lambda$  DNA + 5 unit Pst I +  $50 \mu\text{M}$  **4**; Lane 12,  $\lambda$  DNA + 5 unit Pvu II (control); Lane 13,  $\lambda$  DNA + 5 unit Pvu II +  $50 \mu\text{M}$  **4**; Lane 14,  $\lambda$  DNA + 5 unit Sca I (control); Lane 15,  $\lambda$  DNA + 5 unit Sca I +  $50 \mu\text{M}$  **4**; Lane 16, 1kb DNA ladder.

(b) Lane 1, 1kb DNA ladder; Lane 2,  $\lambda$  DNA alone ( $0.5 \mu\text{g}$ ); Lane 3,  $\lambda$  DNA +  $50 \mu\text{M}$  **4**; Lane 4,  $\lambda$  DNA + 5 unit Ssp I (control); Lane 5,  $\lambda$  DNA + 5 unit Ssp I +  $50 \mu\text{M}$  **4**; Lane 6,  $\lambda$  DNA + 5 unit Ase I (control); Lane 7,  $\lambda$  DNA + 5 unit Ase I +  $50 \mu\text{M}$  **4**; Lane 8,  $\lambda$  DNA + 5 unit Mun I (control); Lane 9,  $\lambda$  DNA + 5 unit Mun I +  $50 \mu\text{M}$  **4**; Lane 10,  $\lambda$  DNA + 5 unit EcoR I (control); Lane 11,  $\lambda$  DNA + 5 unit EcoR I +  $50 \mu\text{M}$  **4**; Lane 12,  $\lambda$  DNA + 5 unit NdeI (control); Lane 13,  $\lambda$  DNA + 5 unit NdeI +  $50 \mu\text{M}$  **4**; Lane 14,  $\lambda$  DNA + 5 unit Bst 11071 (control); Lane 15,  $\lambda$  DNA + 5 unit Bst 11071 +  $50 \mu\text{M}$  **4**; Lane 16, 1kb DNA ladder.

(C) FL spectra of BSA in the absence and presence of **1-4** (Figs. S4.1-S4.4 (a), (b), (c))

The experiment to obtain the FL emission spectra of BSA in the absence and presence of increasing concentration of **1-4** [0 (a), 2 (b), 4 (c), 10 (d), 20 (e), 40 (f), 100 (g), 200 (h) and 400 (i)  $\mu\text{M}$ ] were done in triplicate. See **Appendix II**.

(D) Stern-Volmer plots of  $F_0/F$  vs  $[Q]$  for **1-4** (Figs. S5.1-S5.4)



Figs. S5.1-S5.4 Stern-Volmer plots of  $F_0/F$  vs  $[Q]$  for **1-4**

(E) Plots of  $\log(\Delta F/F)$  vs  $\log[Q]$  for **1-4** (Figs. S6.1-S6.4)

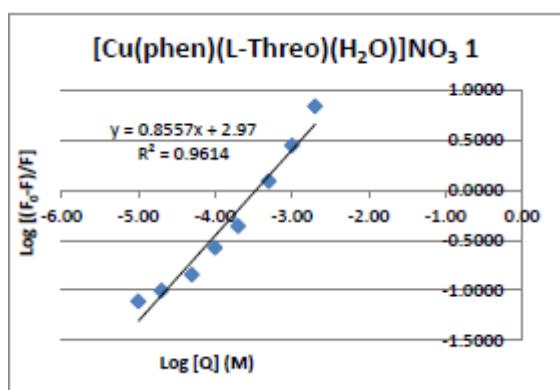


Fig. S6.1

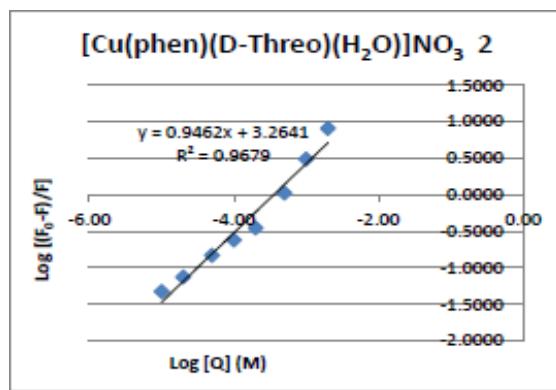
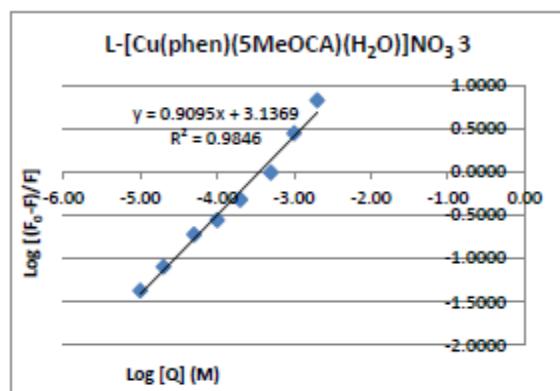


Fig. S6.2



Figs. S6.3

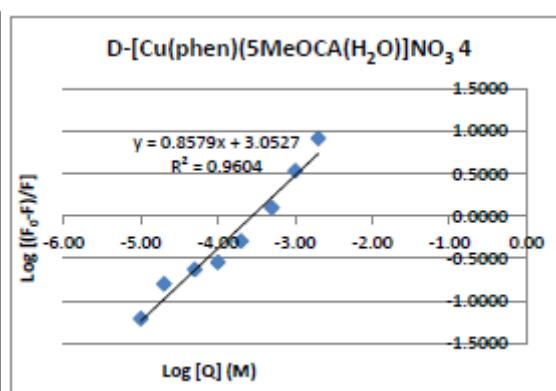


Fig. S6.4

Figs. S6.1-S6.4 Plots of  $\log(\Delta F/F)$  vs  $\log[Q]$  for **1-4**.

(F) UV spectra of BSA in absence and presence of increasing concentration of **1-4** (Figs. S671-S7.4)

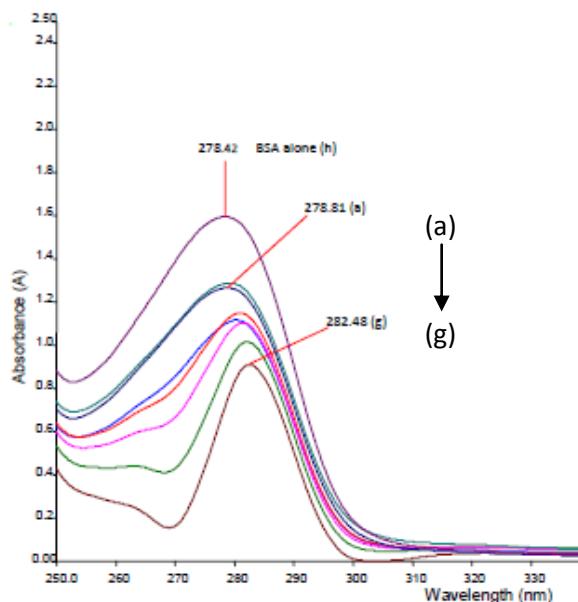


Fig. S7.1 for **1**

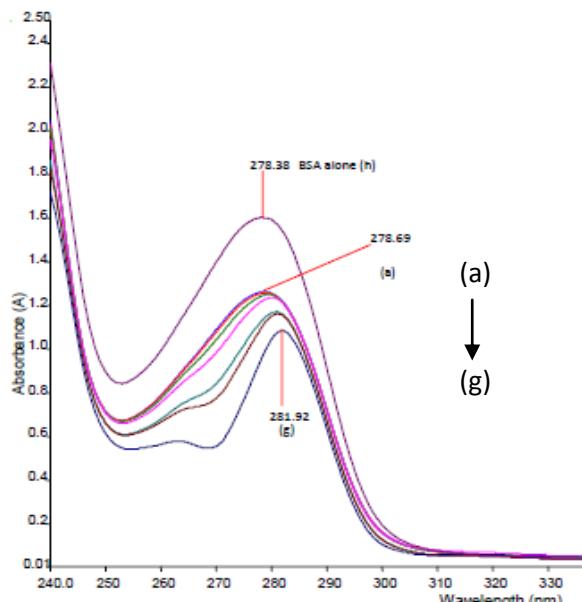


Fig. S7.2 for **2**

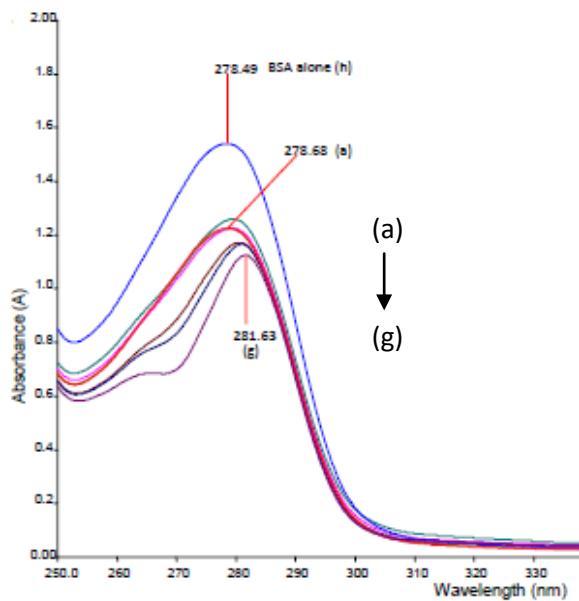


Fig. S7.3 for **3**

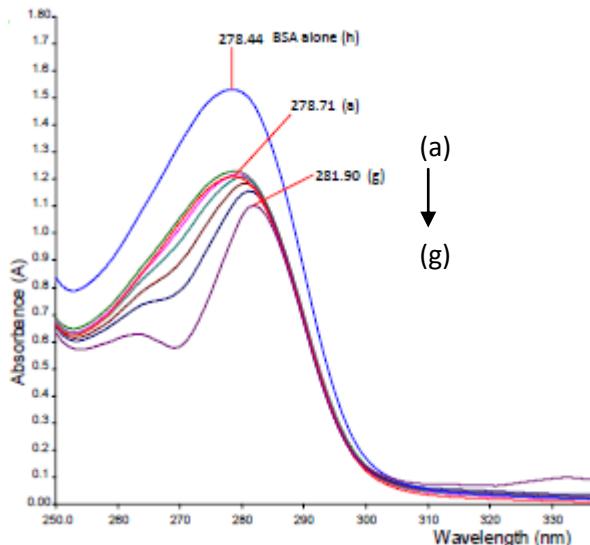


Fig. S7.4 for **4**

Figures S7.1-S7.4. Absorption spectra of BSA in the absence (h) and presence of increasing concentration [2 (a), 4 (b), 10 (c), 20 (d), 30 (e), 40 (f) and 50  $\mu$ M (g)] of copper(II) complexes (1-4). Concentration of BSA alone in the absence of complex was 32.5  $\mu$ M (h).

(G) Synchronous FL spectra of BSA in the absence (*a*) and presence of increasing concentration [2 (*b*), 4 (*c*), 10 (*d*), 20 (*e*), 40 (*f*), 100 (*g*), 200 (*h*) and 400 (*i*)  $\mu$ M] of **1-4** at (a)  $\Delta\lambda = 15$  nm (tyrosine); (b)  $\Delta\lambda = 60$  nm (tryptophan) (Figs. S8.1-S8.4 (a) and (b))

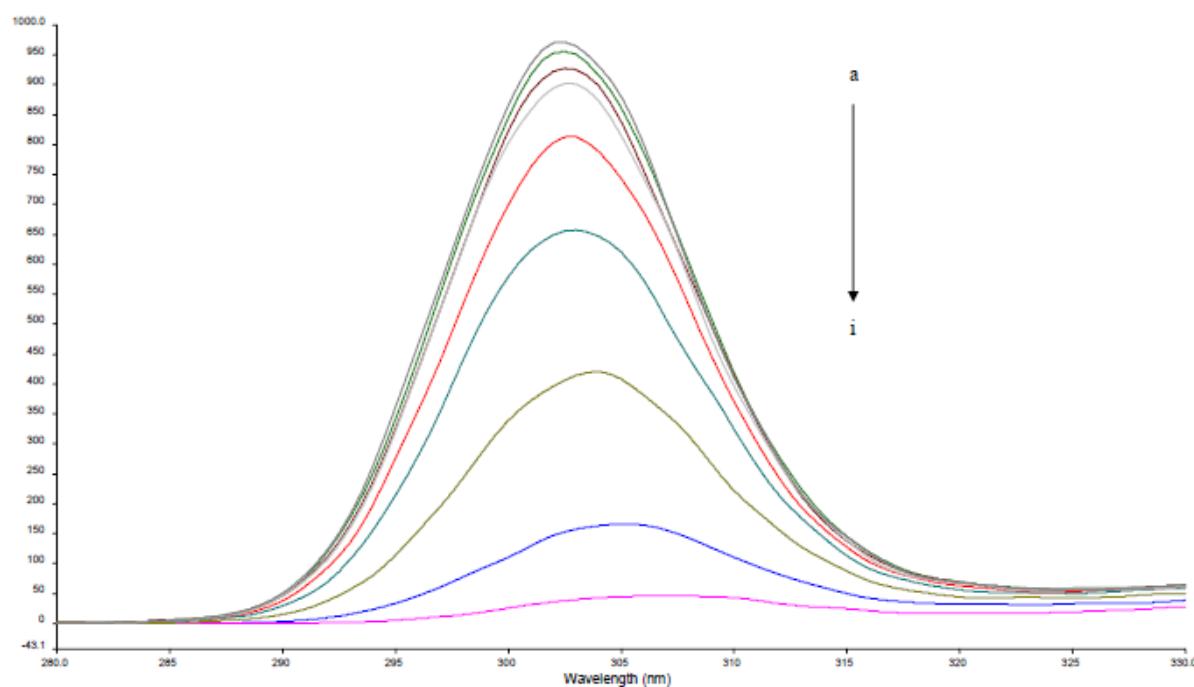


Fig. S8.1(a) for **1**

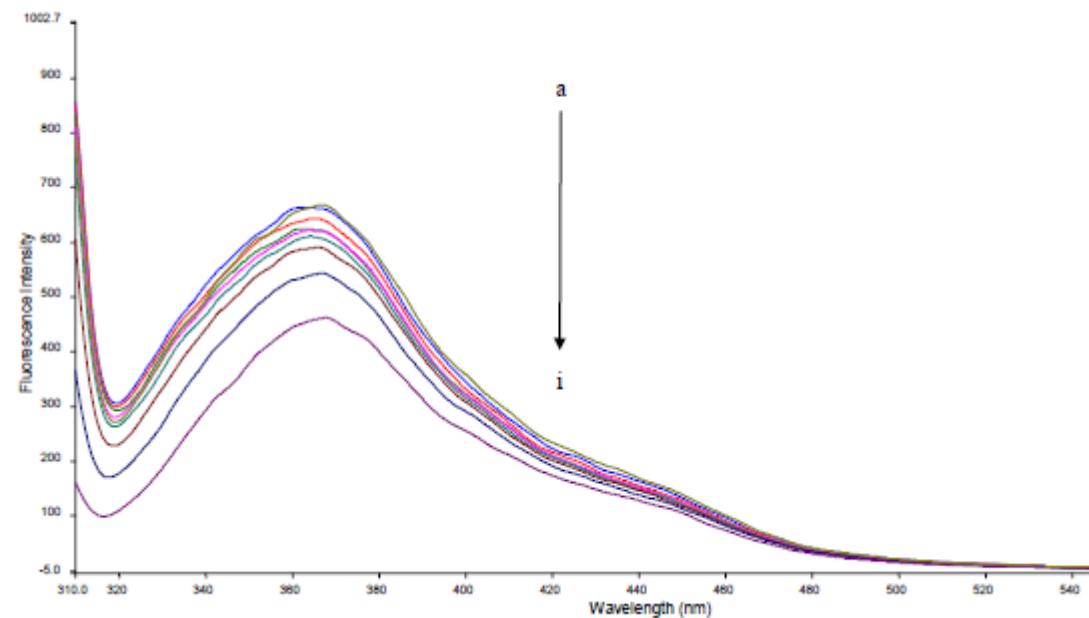


Fig. S8.1(b) for **1**

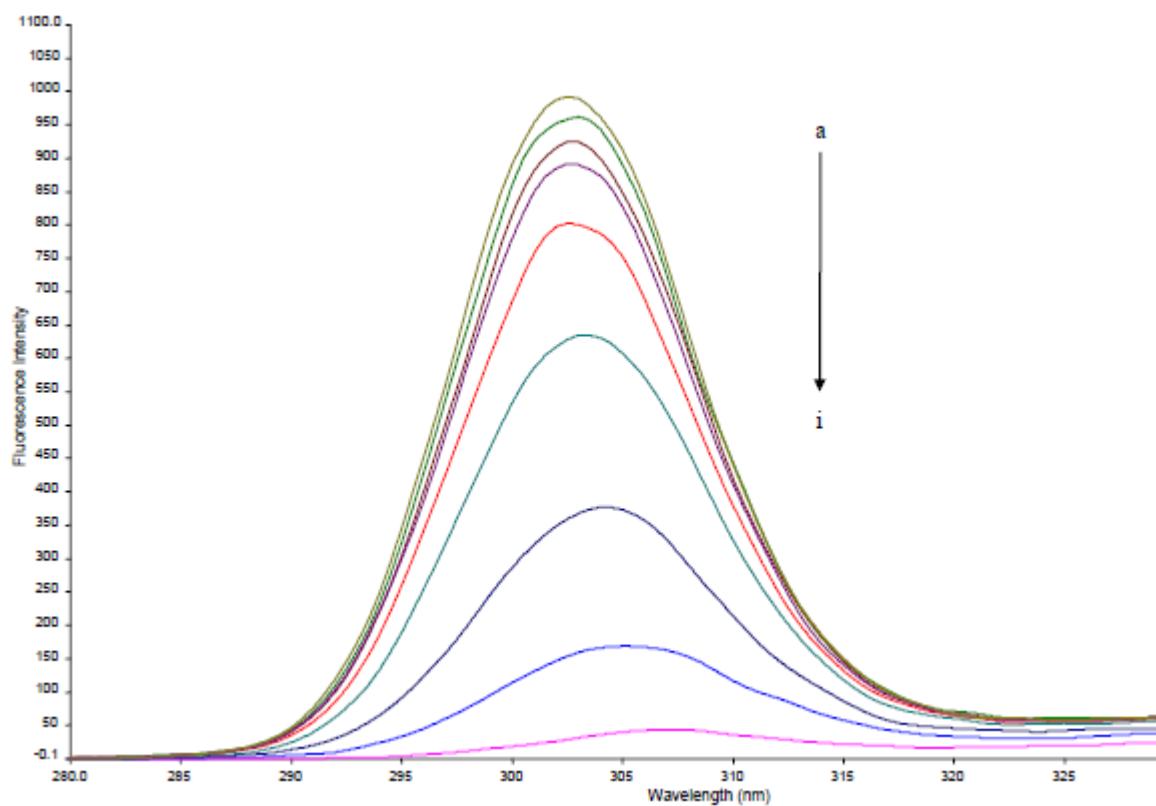


Fig. 8.2(a) for 2

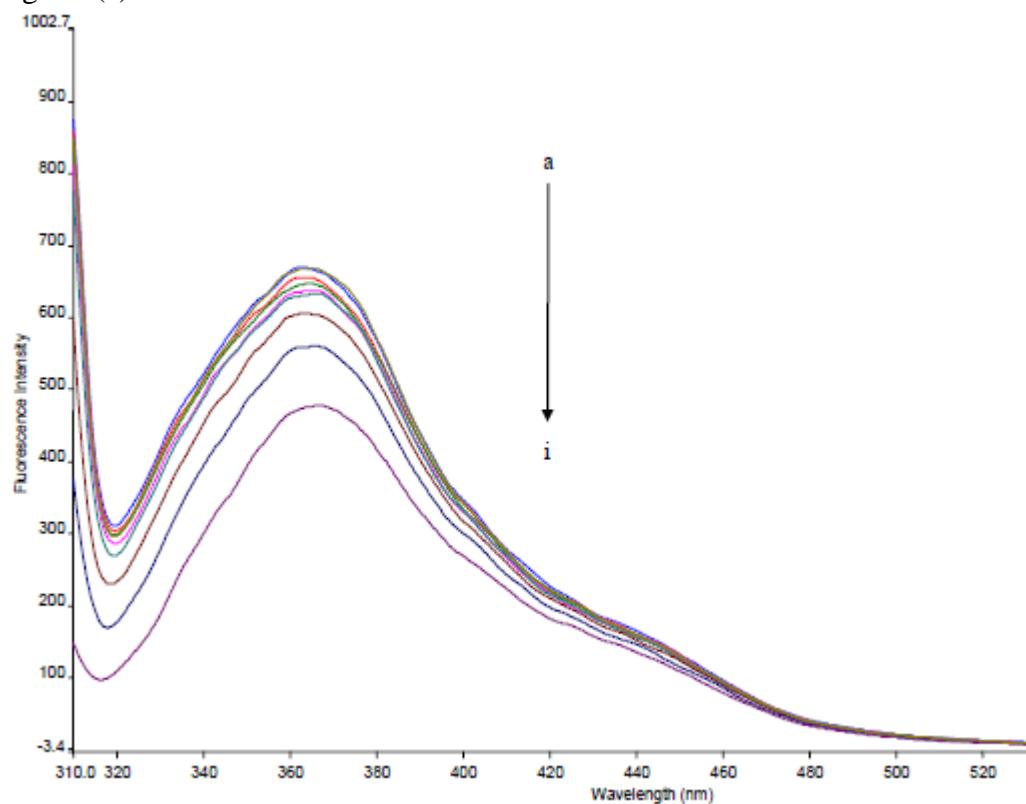


Fig. 8.2(b) for 2

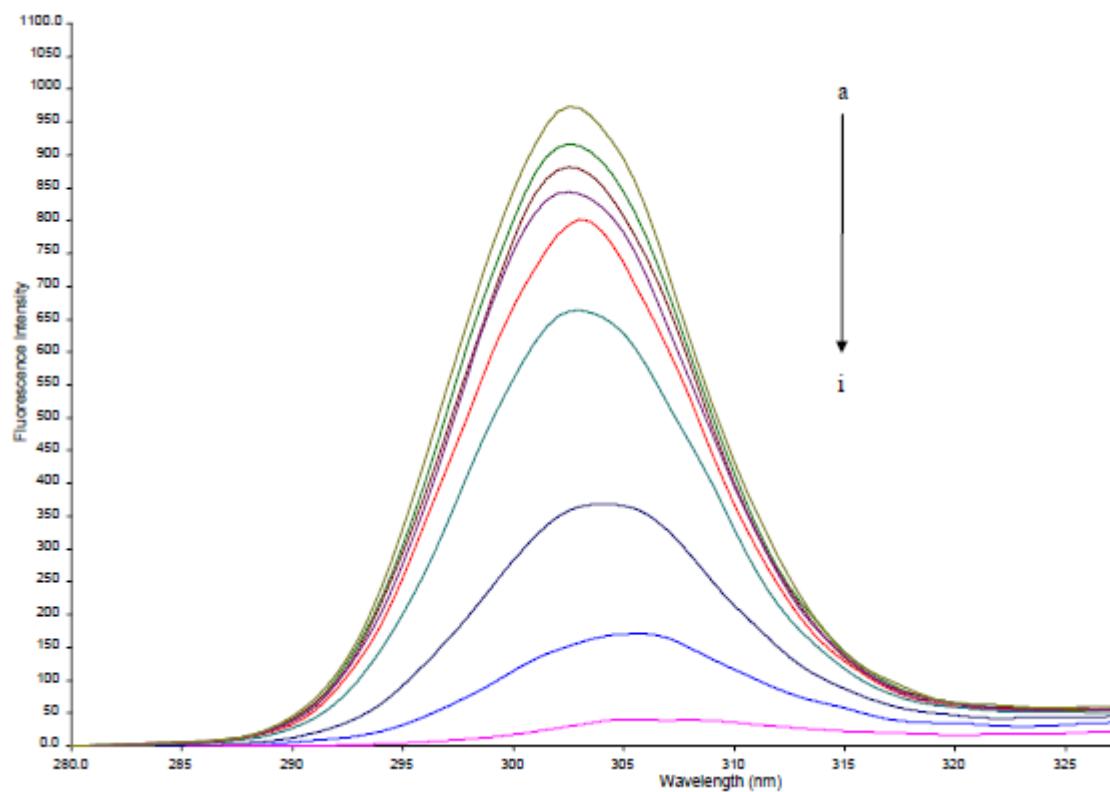


Fig. 8.3(a) for 3

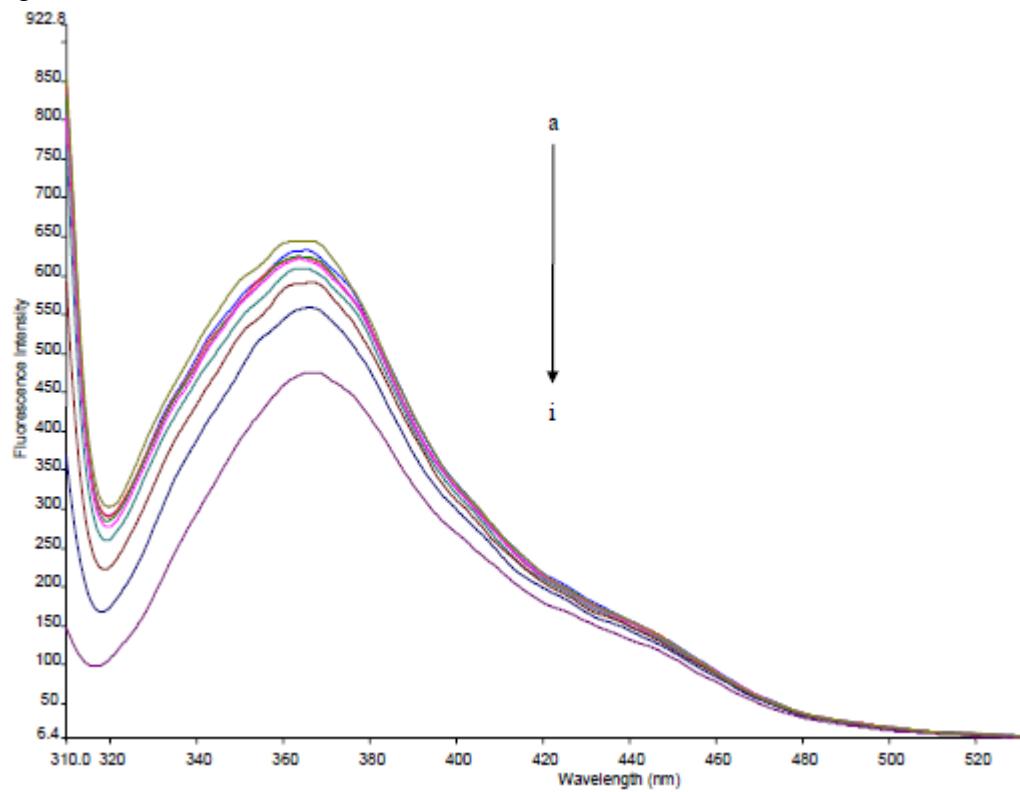


Fig. 8.3(b) for 3

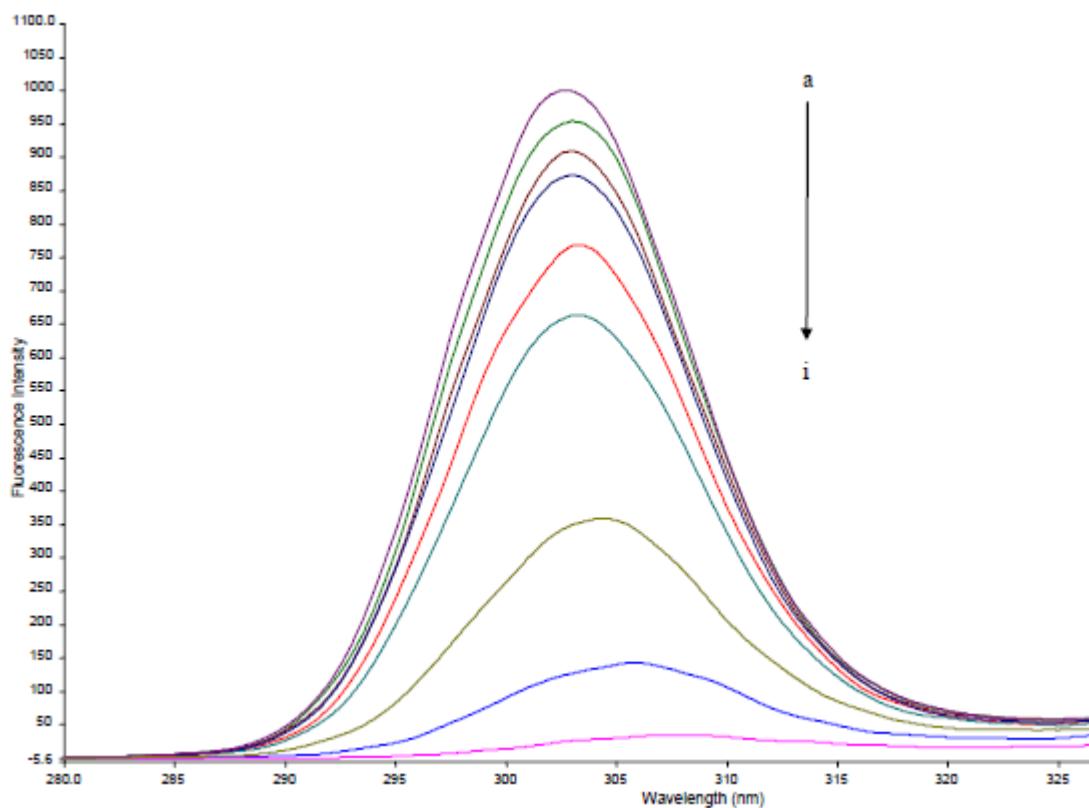


Fig. S8.4(a) for 4

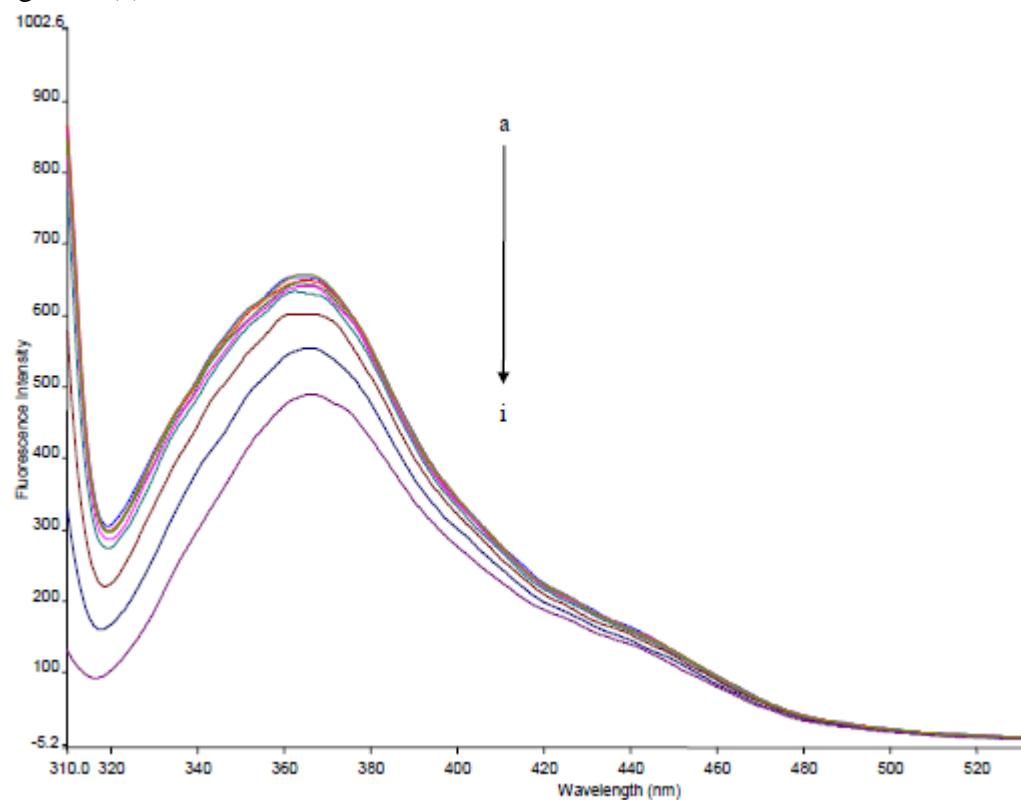


Fig. S8.4(b) for 4

Figs. S8.1-S8.4 Synchronous FL spectra of BSA in the absence (a) and presence of increasing concentration of **1-4** [2 (b), 4 (c), 10 (d), 20 (e), 40 (f), 100 (g), 200 (h) and 400 (i)  $\mu\text{M}$ ] at (a)  $\Delta\lambda = 15 \text{ nm}$  (tyrosine); (b)  $\Delta\lambda = 60 \text{ nm}$  (tryptophan)

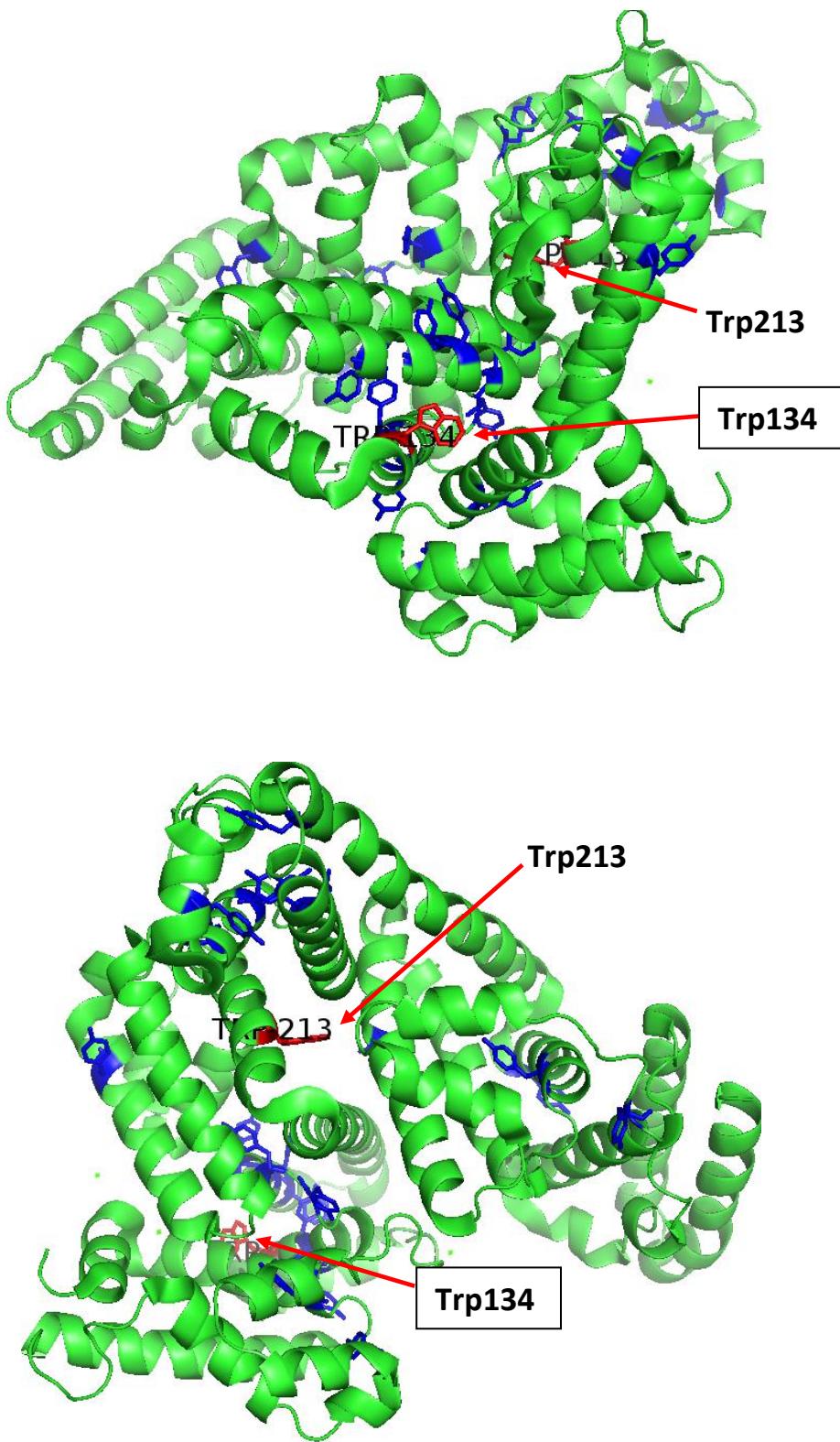


Fig. S9 Comparison of Trp134 and Trp213 binding sites. The **Trp134-binding site** (Top) (Red = Trp134; blue = Tyr residue) has Trp134 with nearby Tyr residues (140, 142, 148, 149, 153). Trp134 is located in the hydrophobic pocket of subdomain IA near the protein surface. **Trp213-binding site** (bottom) contains Trp213 with no nearby Tyr residues but has charged Arg and Asp residues (not shown). Trp213 is in the hydrophobic core of subdomain IIA

**Supplementary Tables S1 and S2**

Crystallographic data and selected bond lengths and angles of **3** and **4**

**Table S1** Crystal data and structure refinement for **3** and **4**

	<b>3</b>	<b>4</b>
Empirical formula	C <sub>17</sub> H <sub>19.25</sub> N <sub>4</sub> O <sub>7.63</sub> Cu	C <sub>17</sub> H <sub>19.25</sub> N <sub>4</sub> O <sub>7.63</sub> Cu
Formula weight	465.15	465.15
Temperature, K	103(2)	100(2)
Wavelength, Å	0.71073	0.71073
Crystal system	Triclinic	Triclinic
Space group	<i>P</i> 1	<i>P</i> 1
Unit cell dimensions	a = 7.6300(4) Å α = 81.2860(10)° b = 10.8699(7) Å β = 84.1770(10)° c = 11.7670(7) Å γ = 75.6260(10)°	a = 7.6383(3) Å α = 81.250(2)° b = 10.8787(4) Å β = 84.085(2) ° c = 11.7750(4) Å γ = 75.541(2) °
Volume, Å <sup>3</sup>	932.38(10)	934.26(6)
Z	2	2
Density (calculated), mg/m <sup>3</sup>	1.657	1.654
Absorption coefficient, mm <sup>-1</sup>	1.226	1.223
F(000)	479	479
Crystal size, mm <sup>3</sup>	0.40 x 0.20 x 0.20	0.40 x 0.38 x 0.20
θ range for data collection	2.45 to 30.56°	2.45 to 26.37°
Index ranges	-7 ≤ h ≤ 9, -13 ≤ k ≤ 13, -14 ≤ l ≤ 14	-9 ≤ h ≤ 9, -13 ≤ k ≤ 13, -14 ≤ l ≤ 14
Reflections collected	11702	25521
Independent reflections	5712 [R(int) = 0.0456]	6572[R(int) = 0.0440]
Absorption correction	Multi-scan	Multi-scan
Max. and min. transmission	0.7916 and 0.6399	0.7920 and 0.6404
Refinement method	Full-matrix least-squares on F <sup>2</sup>	Full-matrix least-squares on F <sup>2</sup>
Data / restraints / parameters	5984 / 3 / 549	6972 / 17 / 557
Goodness-of-fit on F <sub>2</sub>	1.043	1.074
Final R indices [I > 2σ(I)]	R <sub>1</sub> = 0.0272, wR <sub>2</sub> = 0.0703	R <sub>1</sub> = 0.0367, wR <sub>2</sub> = 0.0938
R indices (all data)	R <sub>1</sub> = 0.0290, wR <sub>2</sub> = 0.0714	R <sub>1</sub> = 0.0337, wR <sub>2</sub> = 0.0873
Largest diff. peak and hole, e Å <sup>-3</sup>	0.771 and -0.586	0.717 and -0.689

Tables S2.1 and S2.2 Selected bond lengths and angles for **3** and **4**.

Table S2.1 Selected bond lengths ( $\text{\AA}$ ) and angles ( $^\circ$ ) for **3**

<b>Bond lengths (<math>\text{\AA}</math>)</b>			
C(1) – O(2)	1.222 (5)	C(5) – O(3)	1.414 (4)
C(1) – O(1)	1.270 (4)	C(5) – N(1)	1.473 (4)
C(1) – C(2)	1.529 (4)	Cu(1) – O(1)	1.932 (3)
C(2) – N(1)	1.485 (4)	Cu(1) – N(3)	1.992 (4)
C(2) – C(3)	1.546 (4)	Cu(1) – N(1)	2.002 (3)
C(3) – O(3)	1.422 (4)	Cu(1) – N(2)	2.013 (3)
C(3) – C(4)	1.503 (5)	Cu(1) – O(4)	2.285 (3)

<b>Bond angles (<math>^\circ</math>)</b>			
O(2) – C(1) – O(1)	124.5 (3)	O(1) – Cu(1) – N(3)	92.34 (13)
O(2) – C(1) – C(2)	118.3 (3)	O(1) – Cu(1) – N(1)	85.15 (12)
O(1) – C(1) – C(2)	117.1 (3)	N(3) – Cu(1) – N(1)	177.15 (15)
N(1) – C(2) – C(1)	111.2 (3)	O(1) – Cu(1) – N(2)	169.35 (13)
N(1) – C(2) – C(3)	104.6 (3)	N(3) – Cu(1) – N(2)	82.67 (14)
C(1) – C(2) – C(3)	112.1 (3)	N(1) – Cu(1) – N(2)	99.59 (13)
O(3) – C(3) – C(4)	109.4 (3)	O(1) – Cu(1) – O(4)	88.69 (10)
O(3) – C(3) – C(2)	104.0 (2)	N(3) – Cu(1) – O(4)	95.12 (12)
C(4) – C(3) – C(2)	114.7 (3)	N(1) – Cu(1) – O(4)	86.18 (11)
O(3) – C(5) – N(1)	104.9 (3)	N(2) – Cu(1) – O(4)	101.08 (11)
C(5) – N(1) – C(2)	104.2 (3)	C(5) – O(3) – C(3)	105.3 (2)
C(5) – N(1) – Cu(1)	120.7 (2)	C(1) – O(1) – Cu(1)	114.9 (2)
C(2) – N(1) – Cu(1)	106.9 (2)		

Table S2.2: Selected bond lengths ( $\text{\AA}$ ) and angles ( $^{\circ}$ ) for **4**

Bond lengths ( $\text{\AA}$ )			
C(1A) – O(2A)	1.229 (5)	C(5A) – O(3A)	1.415 (4)
C(1A) – O(1A)	1.272 (5)	C(5A) – N(1A)	1.470 (5)
C(1A) – C(2A)	1.532 (5)	Cu(1A) – O(1A)	1.932 (3)
C(2A) – N(1A)	1.484 (5)	Cu(1A) – N(3A)	1.990 (4)
C(2A) – C(3A)	1.545 (5)	Cu(1A) – N(1A)	2.005 (3)
C(3A) – O(3A)	1.426 (4)	Cu(1A) – N(2A)	2.016 (4)
C(3A) – C(4A)	1.503 (5)	Cu(1A) – O(4A)	2.290 (3)

Bond angles ( $^{\circ}$ )			
O(2A) – C(1A) – O(1A)	124.3 (4)	O(1A) – Cu(1A) – N(3A)	92.36 (14)
O(2A) – C(1A) – C(2A)	118.2 (3)	O(1A) – Cu(1A) – N(1A)	84.97 (14)
O(1A) – C(1A) – C(2A)	117.4 (3)	N(3A) – Cu(1A) – N(1A)	177.04 (17)
N(1A) – C(2A) – C(1A)	110.6 (3)	O(1A) – Cu(1A) – N(2A)	169.38 (14)
N(1A) – C(2A) – C(3A)	104.5 (3)	N(3A) – Cu(1A) – N(2A)	82.66 (15)
C(1A) – C(2A) – C(3A)	111.7 (3)	N(1A) – Cu(1A) – N(2A)	99.75 (14)
O(3A) – C(3A) – C(4A)	109.6 (3)	O(1A) – Cu(1A) – O(4A)	88.64 (11)
O(3A) – C(3A) – C(2A)	104.2 (3)	N(3A) – Cu(1A) – O(4A)	94.96 (12)
C(4A) – C(3A) – C(2A)	114.7 (3)	N(1A) – Cu(1A) – O(4A)	86.29 (12)
O(3A) – C(5A) – N(1A)	105.2 (3)	N(2A) – Cu(1A) – O(4A)	101.10 (12)
C(5A) – N(1A) – C(2A)	104.4 (3)	C(5A) – O(3A) – C(3A)	105.1 (2)
C(5A) – N(1A) – Cu(1A)	120.6 (3)	C(1A) – O(1A) – Cu(1A)	114.9 (3)
C(2A) – N(1A) – Cu(1A)	107.3 (2)		

Table S3 UV-visible spectral data of aqueous solutions of phen and **1-4** (solvent: water-methanol 1:1 v/v)

Compounds	$\lambda_1/\text{nm}$ ( $\epsilon/\text{M}^{-1} \text{cm}^{-1}$ )	$\lambda_2/\text{nm}$ ( $\epsilon/\text{M}^{-1} \text{cm}^{-1}$ )	$\lambda_3/\text{nm}$ ( $\epsilon/\text{M}^{-1} \text{cm}^{-1}$ )
phen	-	227(40,000)	264(30,000)
<b>1</b>	613 (60)	225 (60,000)	273 (50,000)
<b>2</b>	613 (60)	224 (30,000)	273 (30,000)
<b>3</b>	623 (60)	225 (40,000)	273 (40,000)
<b>4</b>	623 (70)	224 (40,000)	273 (30,000)

Concentration of compounds for visible and UV spectra are  $5 \times 10^{-3}$  M and  $3 \times 10^{-5}$  M respectively.

Table S4 Molar conductivity ( $\text{S cm}^2 \text{mol}^{-1}$ ) for **1-4** and other precursor compounds ( $1 \times 10^{-3}$  M)

Compounds	0 h	1 h	24 h
<b>1</b>	50	50	50
<b>2</b>	50	50	50
<b>3</b>	50	50	50
<b>4</b>	50	50	50
$\text{Cu}(\text{NO}_3)_2$	120	120	120
phen	1	1	1
L-threo	1	1	1
D-threo	1	1	1
KCl	1410	1410	1410
(standard Solution)			

Table S5 Restriction enzymes (REs) inhibited by copper(II) compounds

Compounds	<i>Hae</i> III	<i>Ssp</i> I	<i>Ase</i> I	<i>Nde</i> I	<i>Bst</i> 11071
$\text{CuCl}_2$	-	-	-	-	-
$[\text{Cu}(\text{phen})\text{Cl}_2]$	+	+	+	+	+
<b>1</b>	-	+	-	+	-
<b>2</b>	-	+	-	+	+
<b>3</b>	+	-	-	-	-
<b>4</b>	-	-	+	-	-

RE (binding site): *Hae* III (5'-CGGC-3'); *Ssp* I (5'-AATATT-3'); *Ase* I (5'-ATTAAT-3'); *Nde* I (5'-CATATG-3'); *Bst* 11071 (5'-GTATAC-3')