## Supporting Information

## New Chemodosimetric Probe for the Specific Detection of Hg<sup>2+</sup> in Physiological Condition and its Utilisation for Cell **Imaging Studies**

Sukdeb Saha,<sup>b,c</sup> Hridesh Agarwalla,<sup>a,b</sup> Horiom Gupta,<sup>b</sup> Mithu Baidya,<sup>d</sup> Sudip Kumar Ghosh<sup>d</sup>\* and Amitava Das<sup>a,b</sup>\*

<sup>a</sup> CSIR-National Chemical Laboratory, Organic Chemistry Division, Pune: 411008, Maharashtra, India; E-Mail: a.das@ncl.res.in

<sup>b</sup> CSIR-Central Salt & Marine Chemicals Research Institute,Bhavnagar-364002, Gujarat, India

<sup>c</sup> AcSIR-Central Salt & Marine Chemicals Research Institute,Bhavnagar-364002, Gujarat, India <sup>d</sup> Indian Institute Technology, Kharagapur, West Bengal-721302, India

	Contents	Page No
1.	<sup>1</sup> H NMR spectrum of $L_1$	2
2.	Mass spectrum of $L_1$	3
3.	FTIR spectrum of L <sub>1</sub>	4
4.	<sup>1</sup> H NMR spectra of <b>L<sub>2</sub></b> in CD <sub>2</sub> Cl <sub>2</sub>	5
5.	1H NMR spectra of L <sub>2</sub> in DMF-d <sub>7</sub>	6
6.	<sup>13</sup> C NMR spectra of <b>L</b> <sub>2</sub>	7
7.	ESI-MS spectra of L <sub>2</sub>	8
8.	FTIR Spectrum of L <sub>2</sub>	9
9.	ESI-Ms spectrum of $L_2$ in presence of Hg <sup>2+</sup>	10
10.	Crystal data & Refinement parameters for compounds $L_2$	11
11.	Change in <sup>1</sup> H NMR spectra of $L_2$ upon addition of Hg <sup>2+</sup> in CD <sub>3</sub> CN	12
12.	2D-ROESY spectra of $L_2$ in presence of $\beta$ -CD	13
13.	Time dependent spectrophotometry study of $\boldsymbol{L_2}$ in presence of $Hg^{2^+}$	14
14.	Hg <sup>2+</sup> concentration dependent plot of the absorption titration data of	L <sub>2</sub> 15
15.	MTT assay for measuring cytotoxicity of $L_2$ to HeLa cells	16
16.	Recognition of $\mathrm{Hg}^{2^+}$ ions in presence of other metal ions by $L_2$	17
17.	Recognition of $Hg^{2+}$ ions in presence of anions by $L_2$	18
18.	<sup>1</sup> H NMR spectra of $L_2$ in presence of Hg <sup>2+</sup> confirming disappearance –CH <sub>2</sub> groups	e of 19

## <sup>1</sup>H NMR spectra of L<sub>1</sub> in CDCI<sub>3</sub>:



**SI Figure 1:** <sup>1</sup>H NMR spectra of L<sub>1</sub> in CDCl<sub>3</sub>.

#### ESI-MS spectra of L<sub>1</sub>:



SI Figure 2: ESI-MS spectra of L1.

### FTIR spectra of L<sub>1</sub> :



SI Figure 3: FTIR spectra of L1.



**SI Figure 4:** <sup>1</sup>H NMR spectra of  $L_2$  in  $CD_2Cl_2$ .

<sup>1</sup>H NMR spectra of L<sub>2</sub> in DMF-d<sub>7</sub>:



**SI Figure 5:** <sup>1</sup>H NMR spectra of L<sub>2</sub> in DMF-d<sub>7</sub>.



Figure 6: <sup>13</sup>C NMR spectra of L<sub>2</sub> in CDCl<sub>3</sub>.

#### ESI-MS spectra of L<sub>2</sub>:



SI Figure 7: ESI-MS spectra of L<sub>2</sub>.

## FTIR spectra of L<sub>2</sub>:



SI Figure 8: FTIR spectra of L2.





**Figure 9:** ESI-Ms spectra of  $L_2$  in presence of Hg<sup>2+</sup> showing the reappearance of  $L_1$  mass peak.

### Crystal Data and Refinement Parameters for Compound L<sub>2</sub>:

#### Table S1

Identification code	Compound1
Chemical formula	$C_{23}H_{25}BF_2N_2S_2$
Formula weight	442.38
Crystal Colour	orange
Crystal Size (mm)	0.23x0.10 x 0.04
Temperature (K)	150(2)
Crystal System	orthorhombic
Space Group	Pbca
a(Å )	12.572(3)
b(Å )	18.211(5)
<b>C(</b> Å )	19.677(5)
$\alpha(\circ)$	90
β(°)	90
γ (°)	90
Z	8
V(Å <sup>3</sup> )	4505.1(19)
Density (Mg/m <sup>3</sup> )	1.304
Absorption Coefficient(mm <sup>-1</sup> )	0.265
F(000)	1856
Reflections Collected	19526
Independent Reflections	3531
R <sub>(int)</sub>	0.0940
Number of parameters	285
S(Goodness of Fit) on F <sup>2</sup>	1.106
Final R1/wR2 (I>2o(I)	0.0974/ 0.1981
Weighted R1/wR2(all data)	0.1515/ 0.2235
CCDC Number	921458





**Figure 10:** A plot of change in <sup>1</sup>H NMR spectral pattern for the receptor (i)  $L_2$ ; (ii)  $L_2$  with 0.25 equivalents Hg<sup>2+</sup>; (iii)  $L_2$  with 0.5 equivalents Hg<sup>2+</sup> and (iv)  $L_2$  with 1 equivalent Hg<sup>2+</sup> in CD<sub>3</sub>CN medium.



2D-ROESY NMR spectra for L<sub>2</sub> in presence of  $\beta$ -CD in DMF-d<sub>7</sub> solvent:

**Figure 11:** 2D-ROESY NMR showing interaction of Methyl protons ( $H_{21}$ ,  $H_{22}$ ) with aromatic protons and nearby protons ( $H_2$  and  $H_{10}$ ).

Time dependent spectrophotometry study of  $L_2$  in presence of Hg<sup>2+</sup>:



**Figure 12:** Time dependent absorption spectra of  $L_2$  (1.0 x 10<sup>-5</sup> M) in presence of Hg<sup>2+</sup> (2.8 x 10<sup>-5</sup>M) in Acetonitrile/ HEPES buffer medium (3:2, v/v, pH 7.1) over a period of 25 min showing instant conversion of  $L_2$  into  $L_1$ .

 $Hg^{2+}$  Concentration dependent plot of the absorption titration data of L<sub>2</sub> in Acetonitrile/ HEPES buffer medium in presence of  $\beta$ -CD:



Figure 13: Change in absorption value of  $L_2$  at 506 nm with changing the [Hg<sup>2+</sup>].

# MTT Assay for the measuring of the Cytotoxicity of Chemodosimeter $L_2$ to HeLa cells:

Cytotoxicity of L<sub>2</sub> on HeLa cells was determined by conventional MTT assay (J. Natl. Cancer Inst., 1990, 8, 1113-1117). HeLa cells in their exponential growth phase were trypsinised and seeded in 96-well flat-bottom culture plates at a density of 3 x 10<sup>3</sup> cells per well in 100 µl DMEM complete medium (Himedia, India). The cells were allowed to adhere and grow for 24 hr at 37 °C in CO<sub>2</sub> incubator (New Brunswick Scientific, U.S.A.), and then the medium was replaced with 100 µl fresh incomplete medium containing various concentrations of  $L_1$  (0 to 5  $\mu$ M). The assay was performed in quadruplet for each concentration. Cells were then incubated for 6h, after which the culture medium was removed, and 100 µl of 1 mg/ml MTT reagent in PBS was added to each well. Thereafter, it was incubated for 4 hrs; during this period active mitochondria of viable cells reduce MTT to purple formazan. Unreduced MTT were then discarded and DMSO (100 µl) was added into each well to dissolve the formazan precipitate, which was then measured spectrophotometrically using a microplate reader at 570 nm. The cytotoxic effect of each treatment was expressed as percentage of cell viability relative to the untreated control cells. [MTT= (3-(4, 5-Dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide, a yellow tetrazole].



Recognition of Hg<sup>2+</sup> ions in presence of other metal ions by L<sub>2</sub>:

**Figure 14:** Recognition of Hg<sup>2+</sup> ion (1.0 x 10<sup>-4</sup> M) in presence of different metal (M<sup>n+</sup> = 1.0 x 10<sup>-4</sup> M) ions as their perchlorate salts by L<sub>2</sub> (1.1 x 10<sup>-5</sup> M) in acetonitrile/HEPES (3:2, v/v) buffer medium ( $\Delta A$  is the change in absorbances at  $\lambda_{abs}$  490 nm).



Recognition of Hg<sup>2+</sup> ions in presence of other anions by L<sub>2</sub>:

**Figure 15:** Recognition of Hg<sup>2+</sup> ion (1.0 x 10<sup>-4</sup> M) in presence of different anions (X<sup>n-</sup> = 1.0 x 10<sup>-4</sup> M) as their sodium salt by L<sub>2</sub> (1.1 x 10<sup>-5</sup> M) in acetonitrile/ HEPES (3:2, v/v) buffer medium ( $\Delta A$  is the change in absorbances at  $\lambda_{abs}$  490 nm).

<sup>1</sup>H NMR spectra of  $L_2$  in presence and absence of  $Hg^{2+}$  ions related to disappearance of  $-CH_2$  protons of dithiane group:



**Figure 16:** <sup>1</sup>H NMR spectra of  $L_2$  in presence and absence of  $Hg^{2+}$  ions showing disappearance of  $-CH_2$  protons of dithiane group in  $CD_3CN$  medium (Red circles for indicating the  $-CH_2$  protons of dithiane moiety).