

**A functionalized heterobimetallic  $^{99m}\text{Tc}/\text{Re}$  complex as a potential bimodal imaging probe:**

**synthesis, photophysical properties, cytotoxicity and cellular imaging investigations**

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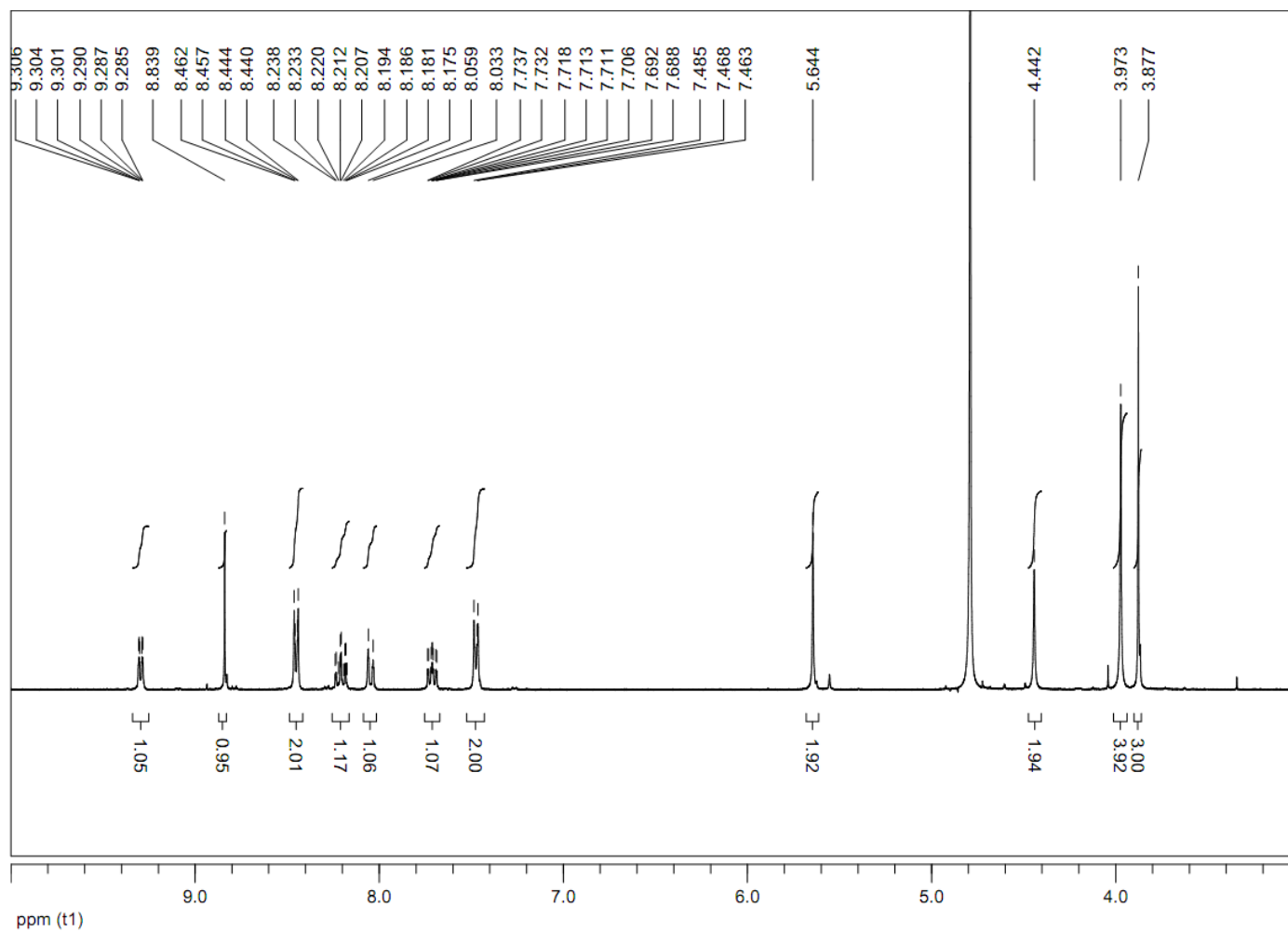
***Electronic Supporting Information***

- I. Selected spectra
- II. Stability assays
- III. Cytotoxicity studies
- IV. Confocal microscopic study

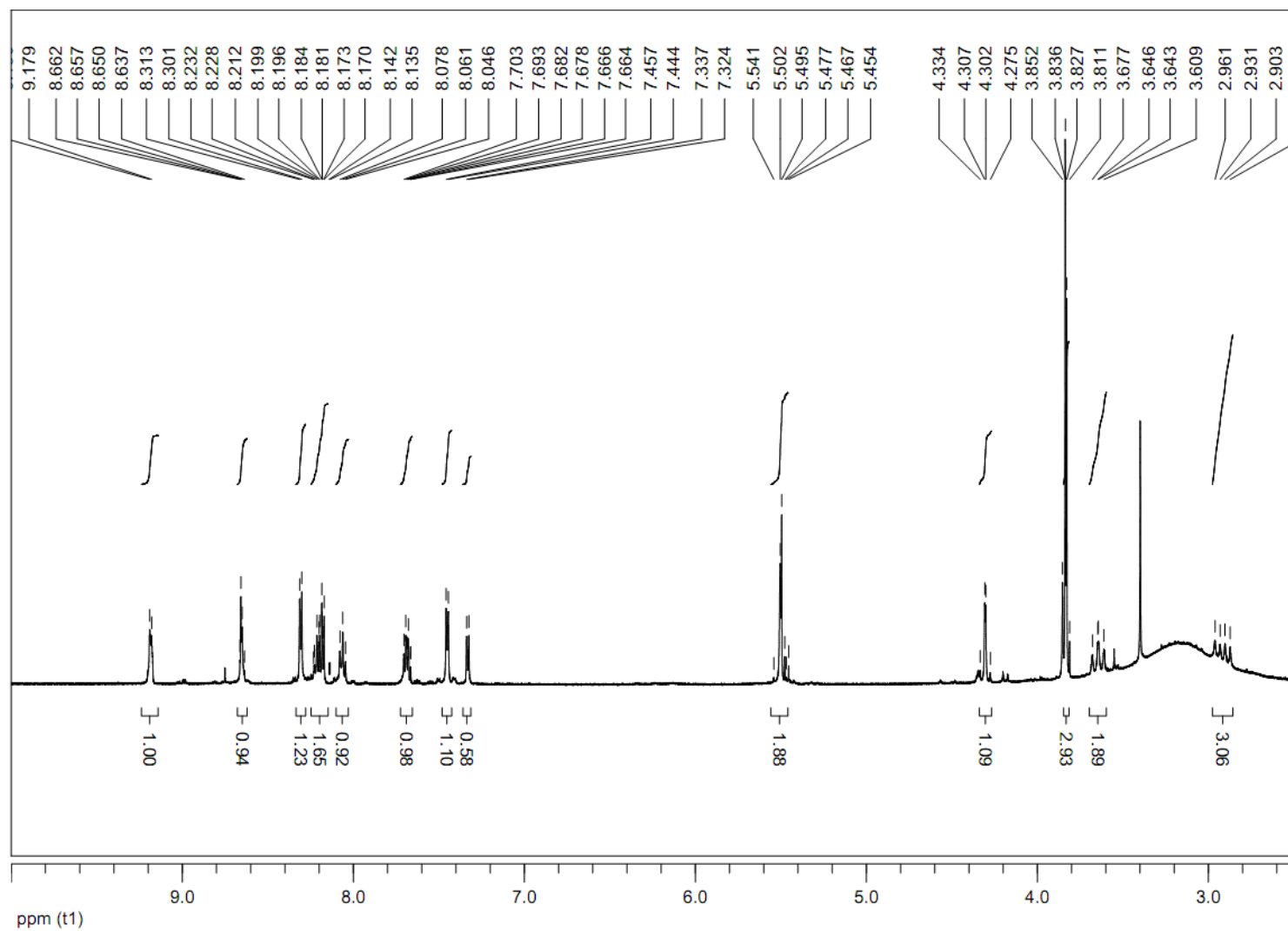
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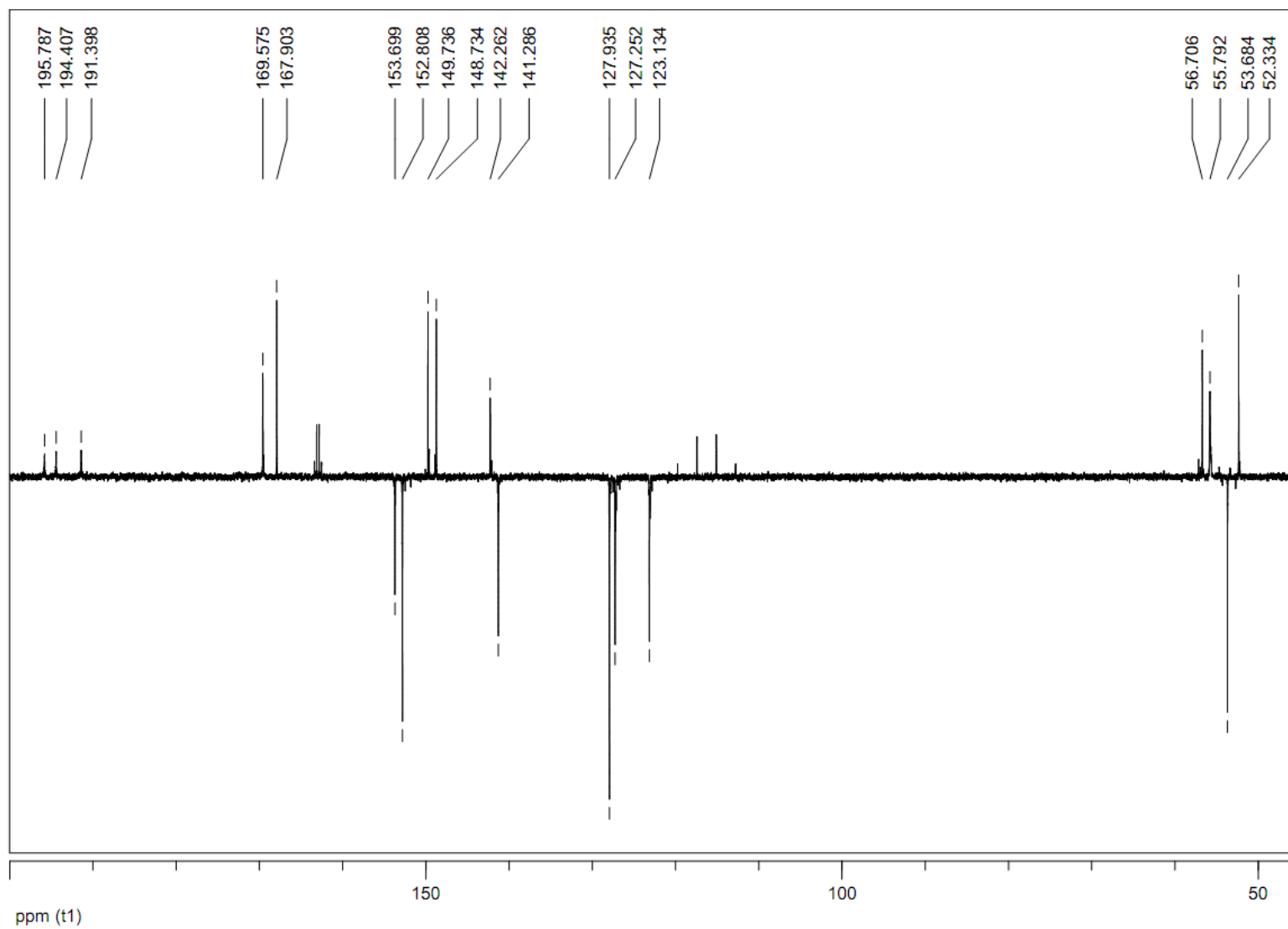
## I. Selected spectra



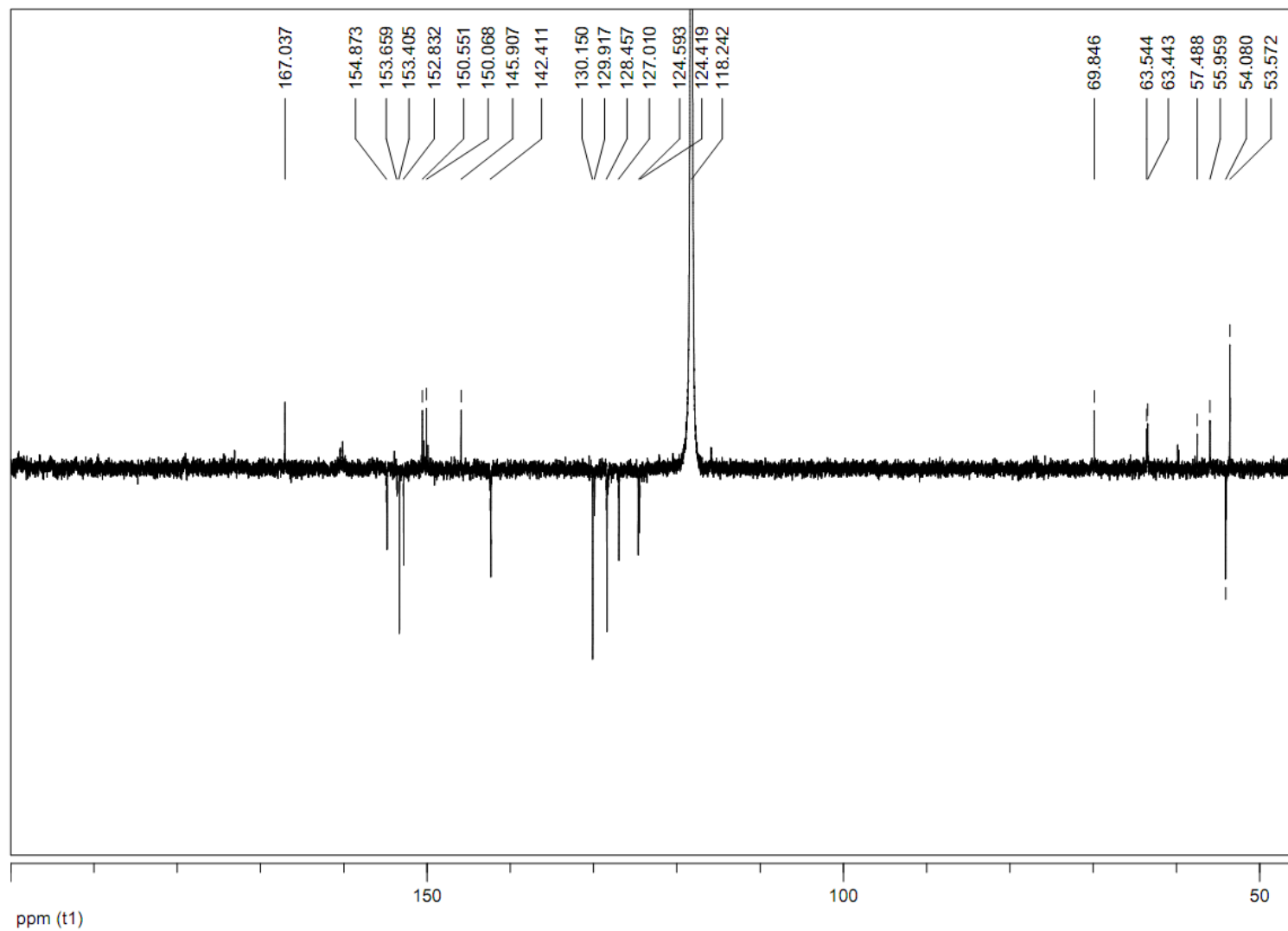
$^1\text{H}$  NMR spectrum of **5** ( $\text{D}_2\text{O}$ , 300 MHz)



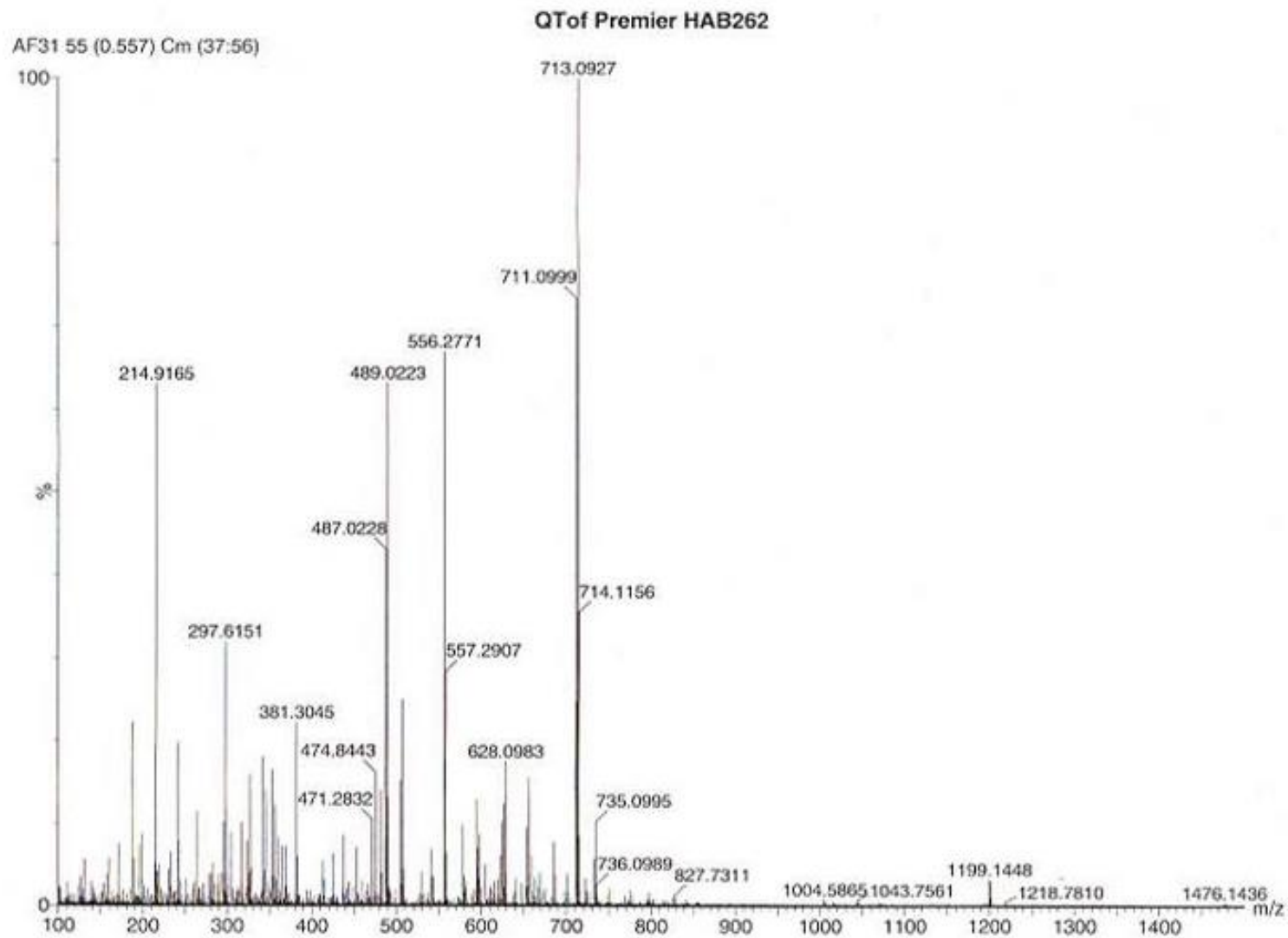
<sup>1</sup>H NMR spectrum of **7** (CD<sub>3</sub>CN, 500 MHz)



$^{13}\text{C}$ -JMOD NMR spectrum of **5** ( $\text{D}_2\text{O}$ , 125 MHz)



$^{13}\text{C}$ -JMOD NMR spectrum of **7** ( $\text{CD}_3\text{CN}$ , 125 MHz)



Mass Spectrum of **5** (ESI<sup>+</sup> mode)

### Elemental Composition Report

#### Multiple Mass Analysis: 2 mass(es) processed

Tolerance = 3.0 mDa / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Odd Electron Ions

3182 formula(e) evaluated with 16 results within limits (all results (up to 1000) for each mass)

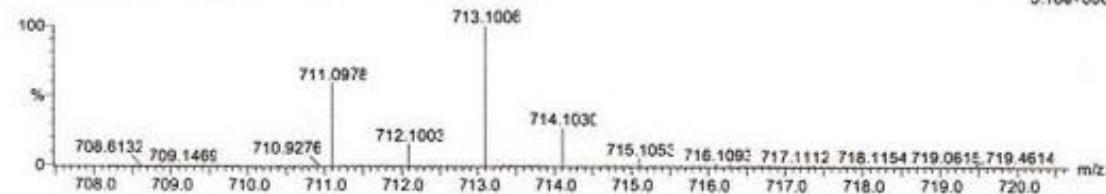
Elements Used:

C: 0-50 H: 0-100 N: 0-6 O: 0-9 185Re: 0-1 187Re: 0-1

AF31

Unit\_11 453 (0.896) AM2 (Av,22000,0.556,28,0.00,LS 10); Cm (441:517)

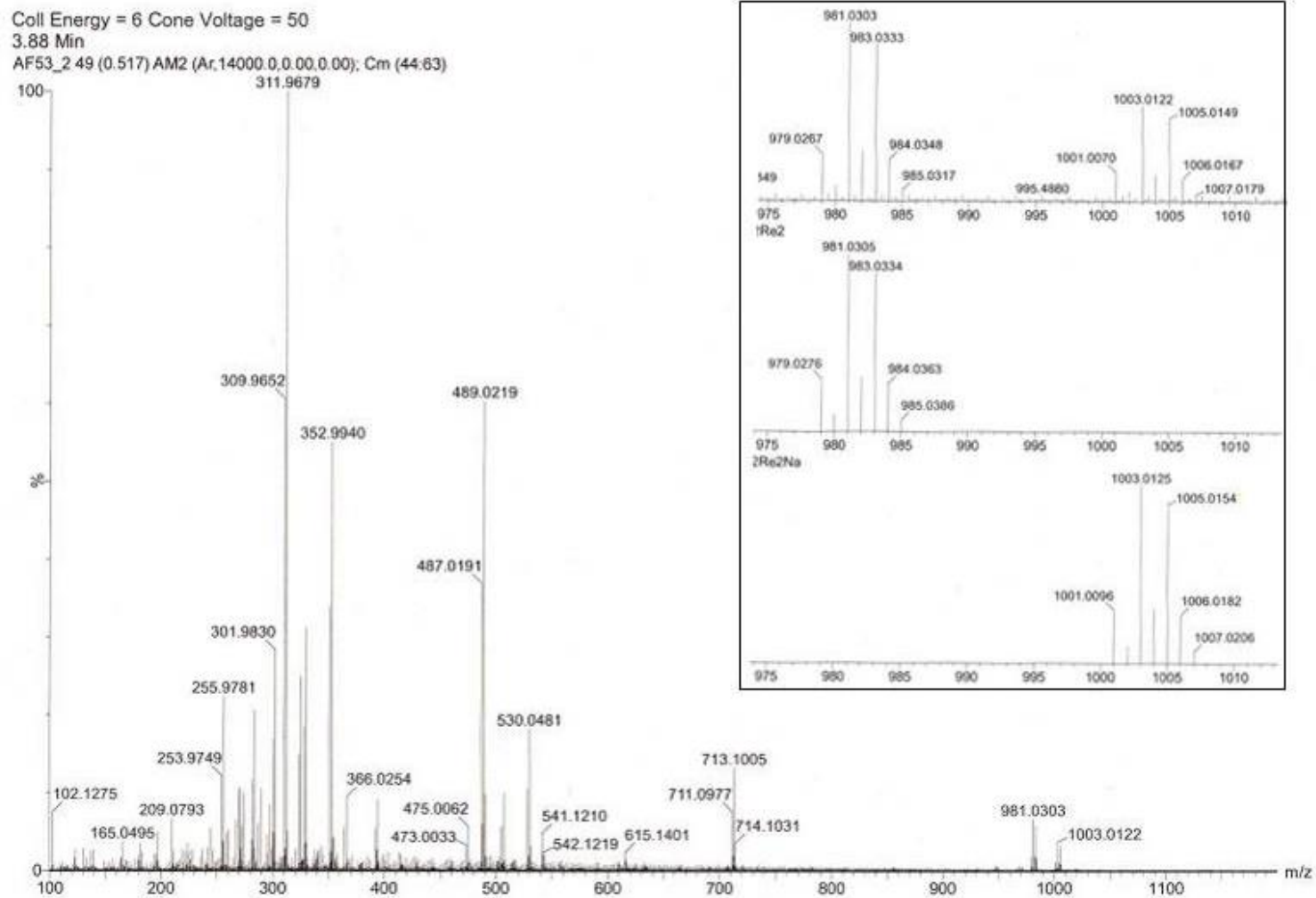
1: TOF MS ES+  
3.18e+006



Minimum: 50.00 -1.5  
 Maximum: 100.00 3.0 10.0 50.0

Mass	RA	Calc. Mass	mDa	PPM	DBE	i-FIT	Norm	Conf(%)	Formula
711.0978	58.35	711.0978	0.0	0.0	16.0	406.2	1.347	26.00	C23 H22 N6 O9 185Re
711.0980	-0.2	-0.3	1.0	406.2	1.444	23.60			C14 H29 N O8 185Re
									187Re
711.0994	-1.6	-2.3	6.0	406.8	2.043	12.97			C15 H25 N5 O4 185Re
									187Re
711.0968	1.0	1.4	44.0	407.2	2.428	8.84			C47 H13 N5 O4
711.0959	1.9	2.7	29.0	407.3	2.454	8.60			C35 H18 N4 O2 185Re
711.0954	2.4	3.4	39.0	407.5	2.698	6.74			C46 H17 N O8
711.0970	0.8	1.1	29.0	407.7	2.884	5.59			C38 H20 O3 187Re
711.1002	-2.4	-3.4	21.0	407.7	2.932	5.33			C27 H20 N6 O6 187Re
711.1000	-2.2	-3.1	33.0	408.6	3.753	2.35			C40 H18 N2 185Re
713.1006	100.00	713.1006	0.0	0.0	16.0	337.6	0.001	99.91	C23 H22 N6 O9 187Re
713.0979	2.7	3.8	14.0	344.8	7.221	0.07			C23 H23 N3 185Re
									187Re
713.0998	0.8	1.1	1.0	347.1	9.498	0.01			C11 H27 N5 O7 185Re
									187Re
713.0987	1.9	2.7	29.0	347.9	10.389	0.00			C35 H18 N4 O2 187Re
713.1003	0.3	0.4	28.0	348.1	10.525	0.00			C36 H20 N2 O3 185Re
713.1012	-0.6	-0.8	43.0	351.2	13.627	0.00			C48 H15 N3 O5
713.1027	-2.1	-2.9	33.0	351.2	13.662	0.00			C40 H18 N2 187Re

High Resolution Mass Spectrum of 5 (ESI+ mode)



Mass Spectrum of **7** (ESI<sup>+</sup> mode).

In the window: experimental MS zoom (up), [M+H]<sup>+</sup> (middle) and [M+Na]<sup>+</sup> (bottom) theoretical isotopic signatures



## Elemental Composition Report

Page 1

### Single Mass Analysis

Tolerance = 2.0 PPM / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Odd and Even Electron Ions

762 formula(e) evaluated with 7 results within limits (all results (up to 1000) for each mass)

Elements Used:

C: 0-50 H: 0-50 N: 0-10 O: 0-15 185Re: 2-2

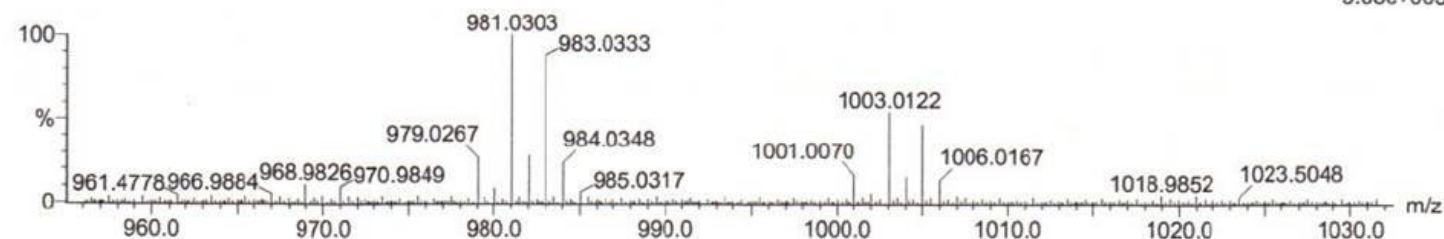
Coll Energy = 6 Cone Voltage = 50

XEVO-G2QTOF#YCA210

3.88 Min

AF53\_2 49 (0.517) AM2 (Ar,14000.0,0.00,0.00); Cm (44:63)

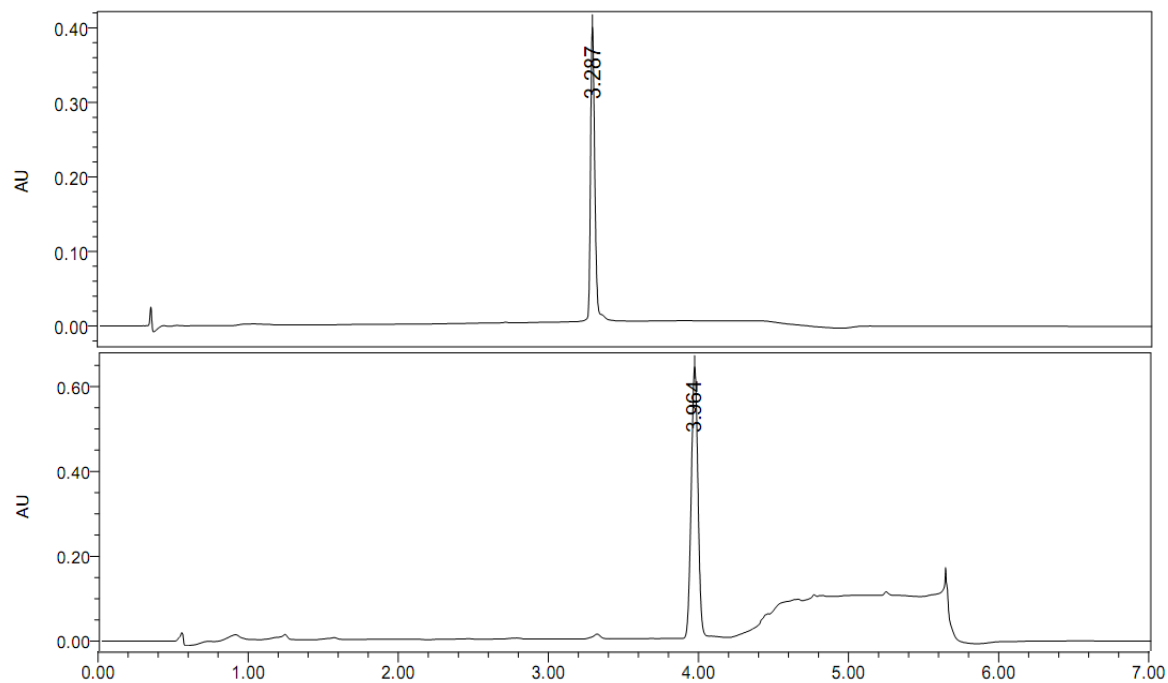
2: TOF MS ES+  
3.08e+005



Minimum: -1.5  
 Maximum: 5.0 2.0 50.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	Norm	Conf (%)	Formula
979.0267	979.0263	0.4	0.4	20.0	140.5	1.022	35.97	C24 H19 N9 O11 185Re2
	979.0250	1.7	1.7	15.0	140.6	1.197	30.22	C23 H23 N5 O15 185Re2
	979.0258	0.9	0.9	32.5	141.4	1.981	13.80	C38 H17 N4 O5 185Re2
	979.0271	-0.4	-0.4	37.5	142.0	2.597	7.45	C39 H13 N8 O 185Re2
	979.0277	-1.0	-1.0	19.5	142.1	2.707	6.68	C26 H21 N6 O12 185Re2
	979.0271	-0.4	-0.4	32.0	142.6	3.186	4.13	C40 H19 N O6 185Re2
	979.0285	-1.8	-1.8	37.0	143.5	4.046	1.75	C41 H15 N5 O2 185Re2

High Resolution Mass Spectrum of 7 (ESI+ mode)



Analytical Chromatograms of **5** (top) and **7** (bottom) after HPLC purification  
(Column: Acquity BEH UPLC C18 column, 1.7  $\mu\text{m}$  2.1x50 mm, 0.3 mL/min,  
Solvent (A/B): 0.1% TFA in water / 0.1% TFA in  $\text{CH}_3\text{CN}$ , Gradient: from A:B 90:10 to 0:100 in 4.5 min,  $\lambda=239$  nm)

## II. Stability assays

### Procedure

#### *Radiocomplex $^{99m}\text{Tc}/\text{Re}$ (8) stability vs. Histidine*

The histidine challenge experiment was performed on purified radiocomplex **8** using a large excess of histidine (1:20 molar ratio). In a vial containing 100  $\mu\text{L}$  of freshly prepared histidine solution (Histidine monohydrate monohydrochlorate PROLABO;  $4.77 \cdot 10^{-3}$  M in water), 20  $\mu\text{L}$  of purified  $^{99m}\text{Tc}/\text{Re}$  complex **8** ( $1.2 \cdot 10^{-3}$  M) were added. The mixture was stirred at  $37^\circ\text{C}$  from 1 to 6h. After this incubation time, 100  $\mu\text{L}$   $\text{CH}_3\text{CN}$  were added, the mixture was centrifuged 5 min at 300 rpm. The supernatant was then analyzed by HPLC, under the previous conditions.

### Results

**Table 1.** Stability of complexes **7** and **8** in buffer solution, and against ligand exchange with histidine

Conditions	Complex <b>7</b> <sup>a</sup>			Complex <b>8</b> <sup>a</sup>		
	0.5 h	2 h	6 h	0.5 h	2 h	6 h
Aqueous <sup>b</sup>	>99%	>99%	>99%	>99%	>99%	>99%
Histidine <sup>c</sup>	<i>n.d.</i>	<i>n.d.</i>	<i>n.d.</i>	99%	95%	88%

a: Percent of complex remaining at the indicated time, b: Tris buffer, pH 7.4 at ambient temperature;  
c: ligand exchange with an excess of histidine at  $37^\circ\text{C}$ , *n.d.*: not determined

### III. Cytotoxicity studies

#### **Material and methods:**

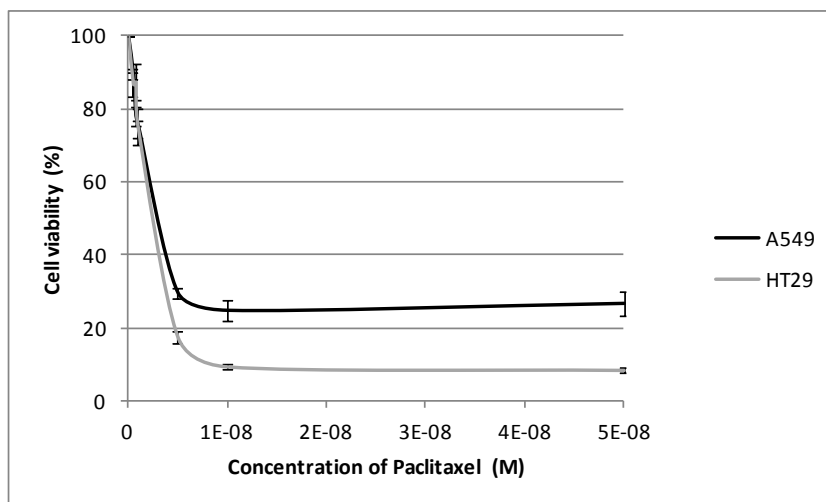
Human lung alveolar (A549), human breast (MCF7) and human colon (HT29) adenocarcinoma cells were obtained from the Developmental Therapeutics Branch of the National Cancer Institute, Bethesda, MD (USA). Cells were grown with RPMI-1640 GlutaMax medium, 10% fetal bovine serum (GIBCO, Grand Island, USA), at 37°C and 5% CO<sub>2</sub> in flasks purchased from Nunc (Denmark). Cell passaging was performed twice a week. Adherent cells cultures were washed once with phosphate-buffer saline (PBS, 10x, pH 7.2, GIBCO) and harvested by stripping of flasks with trypsin (0.25% Trypsin-EDTA, 1X, Phenol Red, GIBCO) after 5 minutes incubation period at 37°C. Cells were counted on Countess Automatic Cells Counter, using trypan blue, before being plated into a 96-well plate (800 MCF7 or HT29 cells / 1,000 A549 cells per well). 24 h later, the medium was removed and replaced by 200 µL dilutions of **7** in RPMI, 1% DMSO.<sup>1</sup> After a 72 h incubation time, the number of surviving cells was estimated by MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide) assay<sup>2</sup>: after removing the medium, MTT reagent (200 µL per well, 0.5 mg/mL in RPMI) was added. After 4h, it was removed and DMSO was added (150 µL per well) to dissolve formazan crystals. Absorbance of the purple solutions was evaluated on a microplate spectrophotometer (Bio-Tek Instruments; measurements at 570 and 630 nm). Wells treated with solvent only were used as a control, with 100% viability. Absorbance of treated wells was compared to this control value, to estimate cell viability. All experiments were performed in triplicate.

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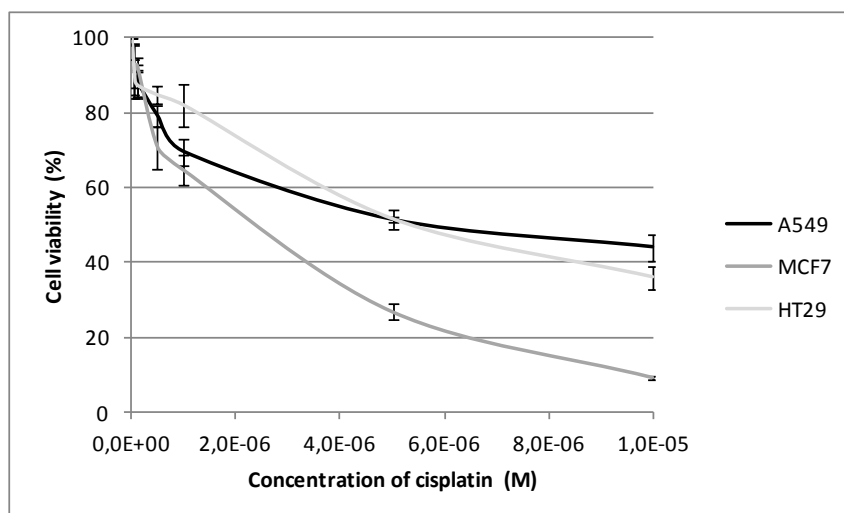
<sup>1</sup> (a) K.K.W. Lo, M.W. Louie, K.S. Sze, J.S.Y. Lau, *Inorg. Chem.*, 2008, **47**, 602-611, (b) M.W. Louie, T.T.H. Fong, K.K.W. Lo, *Inorg. Chem.*, 2011, **50**, 9465-9471.

<sup>2</sup> T. Mosmann, *J. Immunol. Methods*, **1983**, *65*, 55-63.

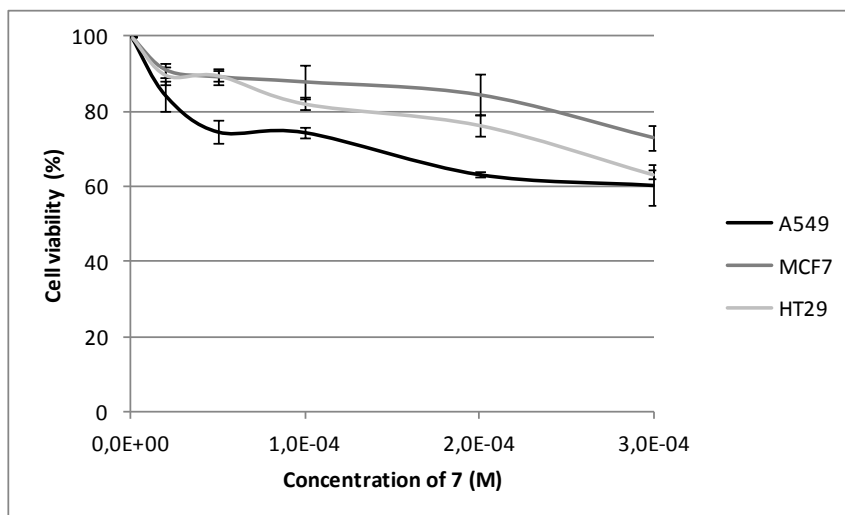
**Cytotoxicity studies of Paclitaxel, Cisplatin and complex 7 on A549, MCF7 and HT29 cells  
(standard deviation between three replicates)**



**Figure 1.** Cytotoxicity positive assay with Paclitaxel (nanomolar  $IC_{50}$ )



**Figure 2.** Cytotoxicity positive assay with Cisplatin (micromolar  $IC_{50}$ )



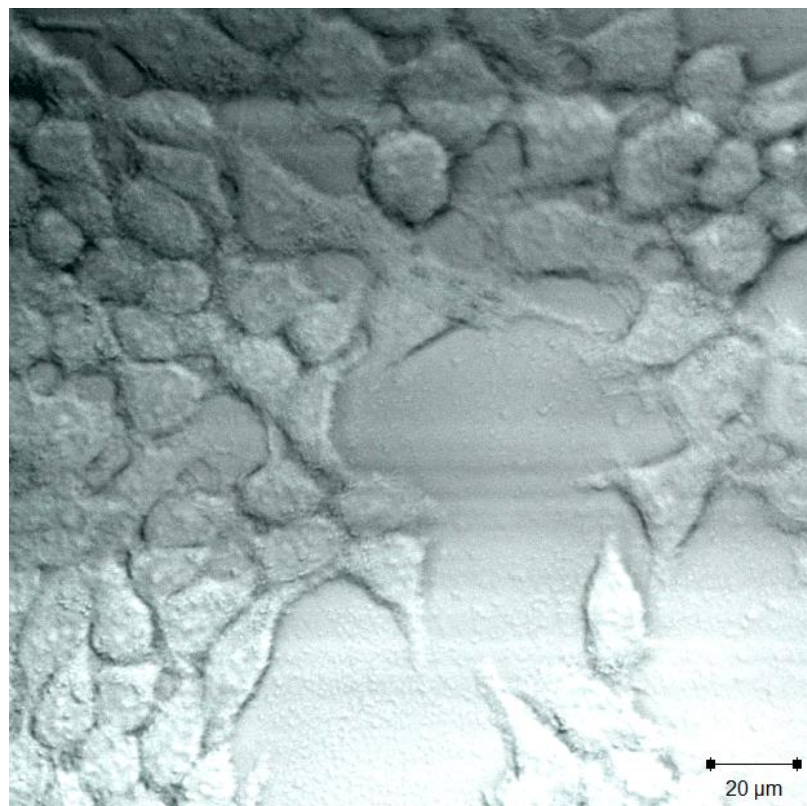
**Figure 3.** Cytotoxicity studies of complex 7 (micromolar  $IC_{50}$ )

## IV. Confocal microscopic study

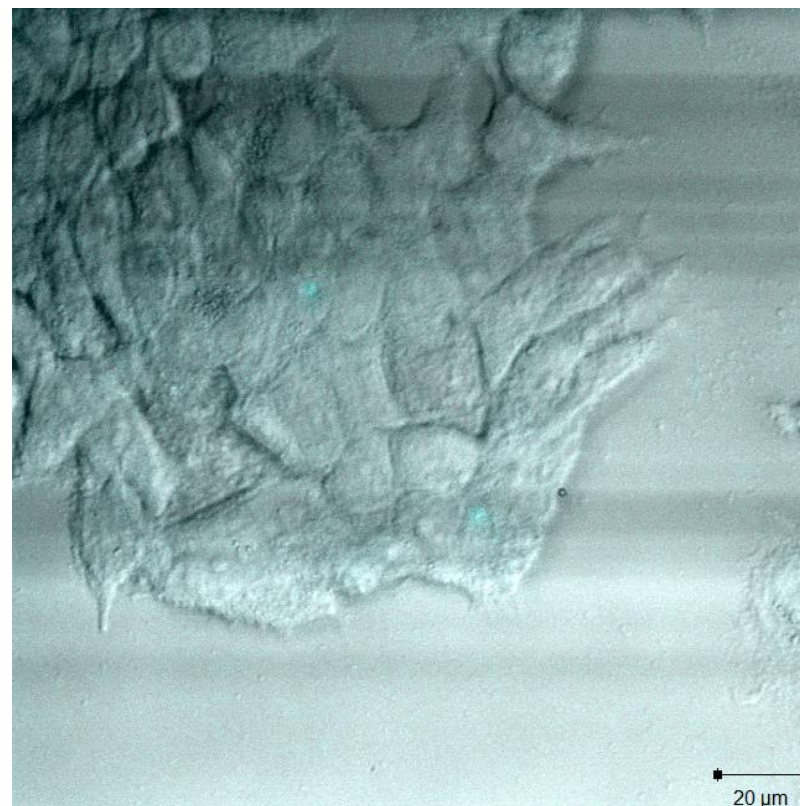
### Material and methods

5,000 cells of A549 were plated onto 96-black well plates and incubated at 37°C, 5% CO<sub>2</sub> for 24 h, in 200 µL RPMI medium. After this incubation time, the medium was removed and replaced by 200 µL solutions of **7** (0, 50, 100 and 200 µM) in RPMI + 1% DMSO medium. The compound was incubated 72h and then removed. Cells were washed twice with PBS and viewed, still in PBS, under a Zeiss LSM 510 confocal microscope (Carl Zeiss AG, Oberkochen, Germany) with LSM Image Browser software, equipped with a diode laser, amongst others. Fluorescence was observed at room temperature with a 25x objective, with an excitation at 405 nm, using the DAPI configuration (emission in blue area; Dye (emission): DAPI; laser (excitation): 405 nm at 25 mW). Wells without compound **7** were used as a control, to ensure that no background fluorescence was recorded.

**Confocal microscopic study of A549 cells after treatment with complex 7 : dose-response relationship**  
*(various dose concentrations,  $\lambda_{ex} = 405\text{ nm}$ , 72h incubation time)*

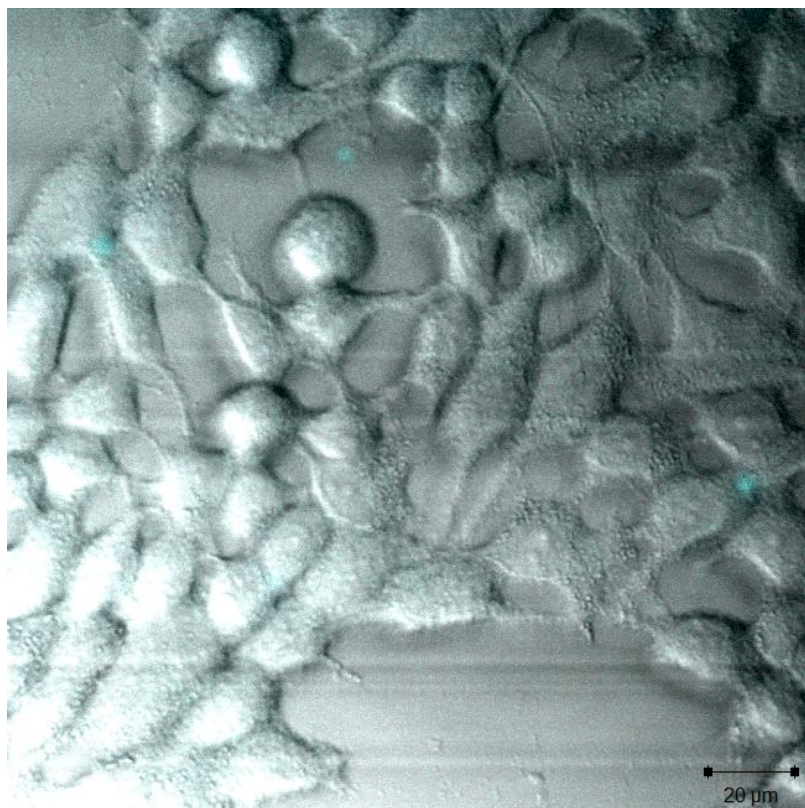


Confocal microscopic image of A549 cells without incubation of complex **7** and two washes with PBS, as a reference.

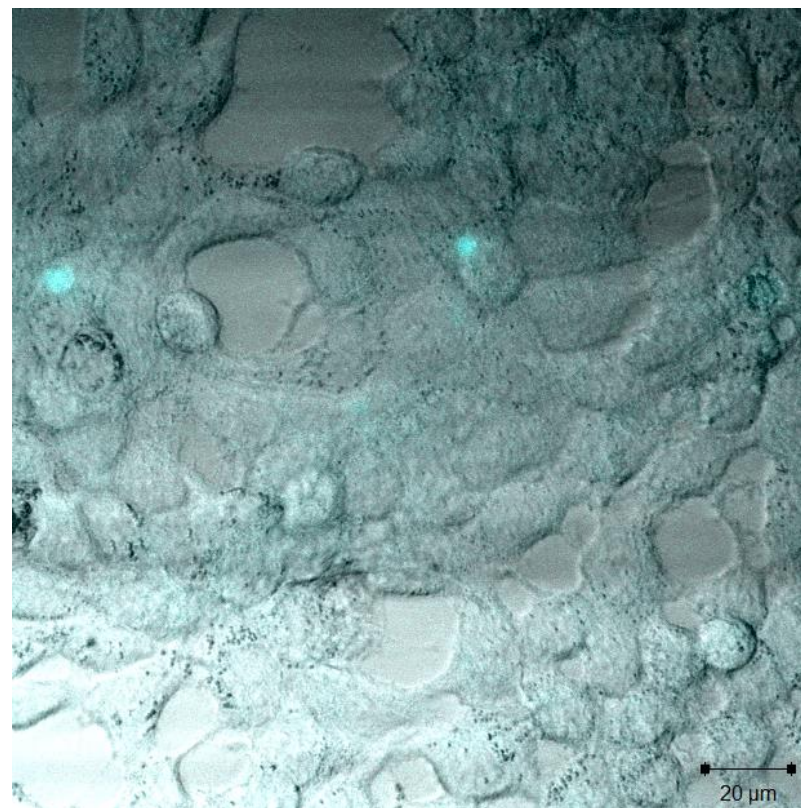


Confocal microscopic image of A549 after a 72h-incubation time with  $5 \times 10^{-5}\text{ M}$  complex **7** and two washes with PBS.

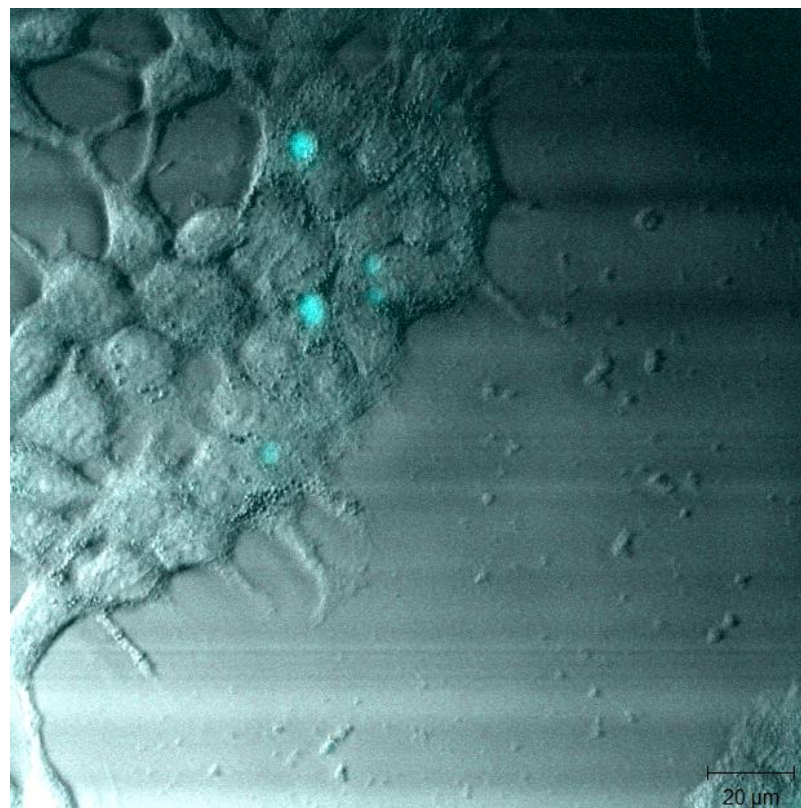




Confocal microscopic image of A549 after a 72h-incubation time with  $1 \times 10^{-4} M$  complex **7** and two washes with PBS.



Confocal microscopy image of A549 after a 72h-incubation time with  $1.5 \times 10^{-4} M$  complex **7** and two washes with PBS.



Confocal microscopy image of A549 after a 72h-incubation time with  $2 \cdot 10^{-4}$  M complex **7** and two washes with PBS.