

***Supporting information for :***

**Harnessing medically-relevant Metals onto water-soluble  
Subphthalocyanines: towards Bimodal Imaging and Theranostics**

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## A. CHEMISTRY

For clarity purposes simple labels are used all along the manuscript text:

- Subphthalocyanine (**SubPc**) with a distal phenoxy pocket are labelled according to the substituent in para position: **SubPc-DOTA (1)**, **SubPc-DTPA (2)**, **SubPc-MDOTA (1-M)**, **SubPc-MDTPA (2-M)**.

### A.1. Chemicals

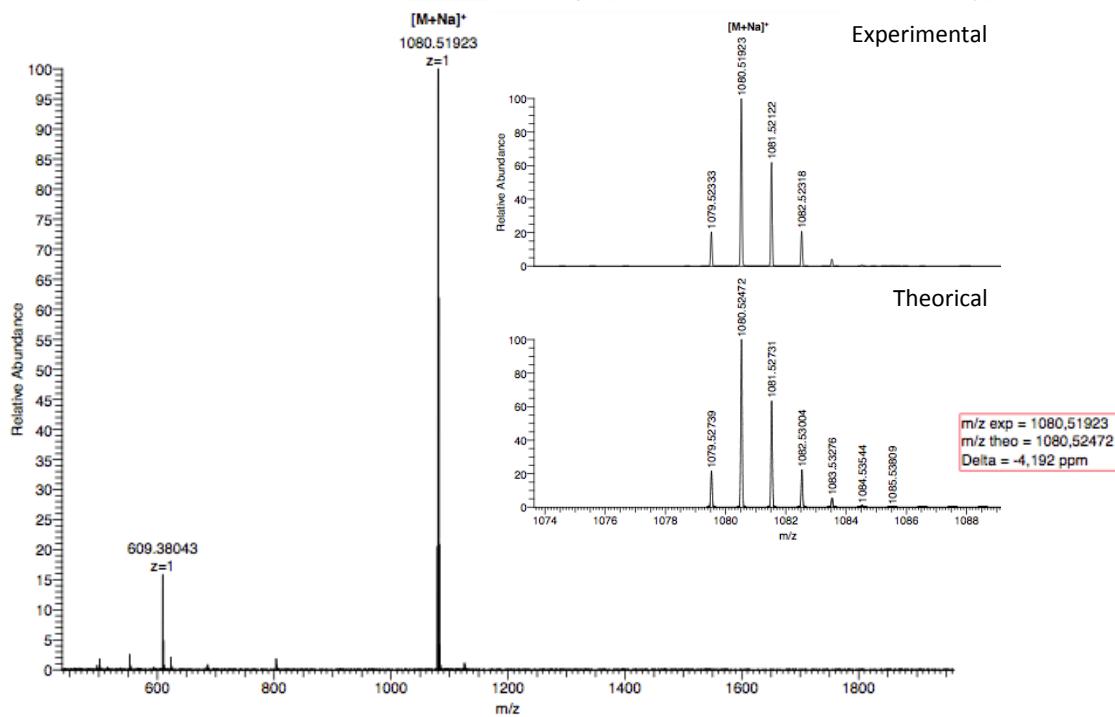
Chemicals used in this study are from various providers: Acros Organics [1,2-dicyanobenzene (98 %, ref. 174012500), 4-nitrophenol (99 %, ref. 157052500), gadolinium (III) nitrate pentahydrate (99.9 %, ref. 199140250), copper (II) nitrate trihydrate (99 %, 207680025)], Sigma Aldrich [boron trichloride (1 M in p-xylene, 345458), 1,2-dipalmitoyl-*sn*-glycero-3-phosphocholine ( $\geq$  99 %, P4329), lutethium (III) chloride hexahydrate (99.9 %, ref. 298131)], Alfa Aesar [palladium 10 % on carbon (A12012), trifluoroacetic acid (99 %, A12198), indium (III) chloride anhydrous (99.9 %, 41977), gallium (III) chloride anhydrous (99.99 %, 11867)], Chematech [2-(4,7,10-tris(2-(tert-butoxy)-2-oxoethyl)-1,4,7,10-tetraazacyclododecan-1-yl)acetic acid ( $\geq$  98 %, C060)] Strem Chemicals [yttrium chloride hexahydrate (99.9 %, ref. 93-3903)]. All chemicals and solvents were used as supplied without further purification.

### A.2. Chromatography

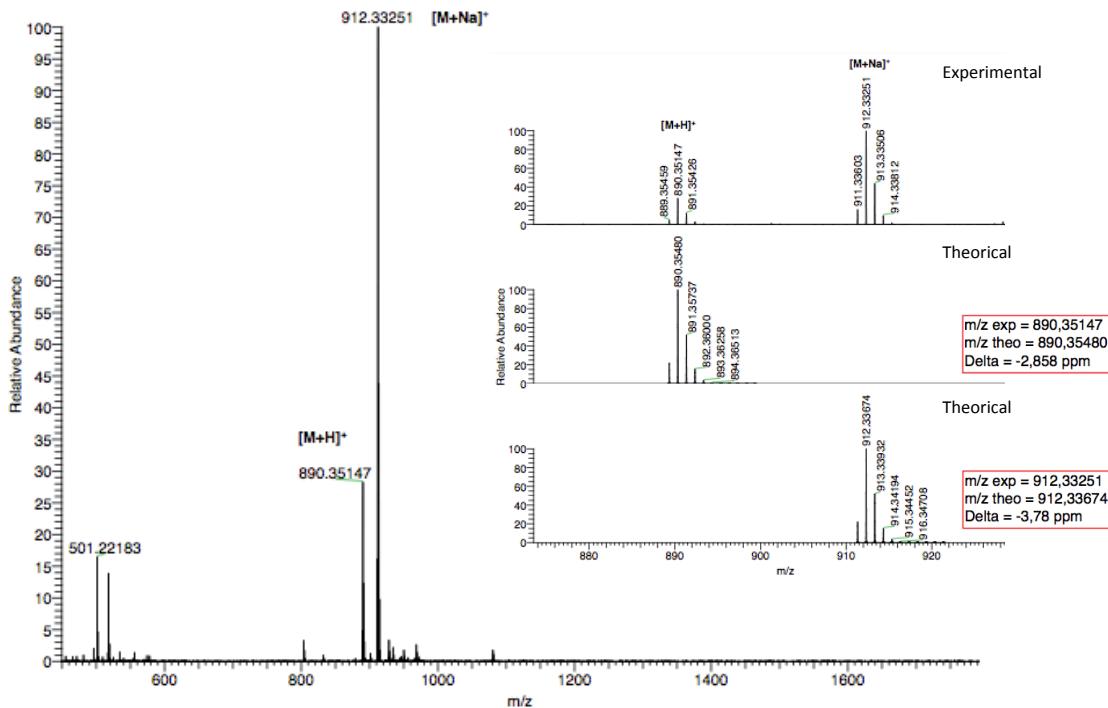
Compounds **3**, **4**, **5** and **6** were purified on column chromatography using silica gel (60A, SDS) and a specific mixture of solvent as described in full text. Compound **3** was purified on column chromatography using alumina gel (60A, SDS) and a specific mixture of solvent as described in full text.

## B. ANALYSIS

### B.1. SubPc-DOTA (1)



*Fig. S1.* HR-MS spectrum of SubPc-DOTAtBu<sub>3</sub> (5)



*Fig. S2.* HR-MS spectrum of SubPc-DOTA (1)

## B.2. SubPc-DTPA (2)

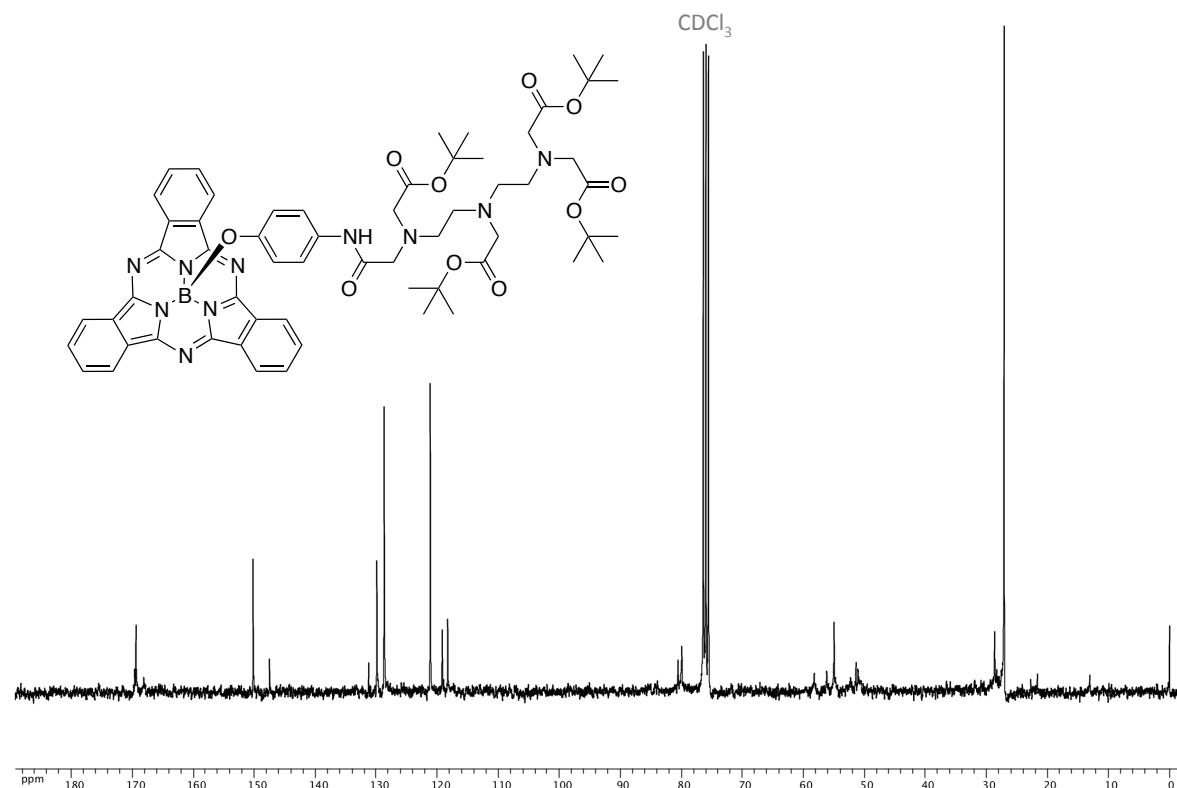


Fig. S3.  $^{13}\text{C}$  NMR spectrum of SubPc-DTPA- $t\text{Bu}_3$  (**6**) ( $\text{CDCl}_3$ , 75 MHz, 300K)

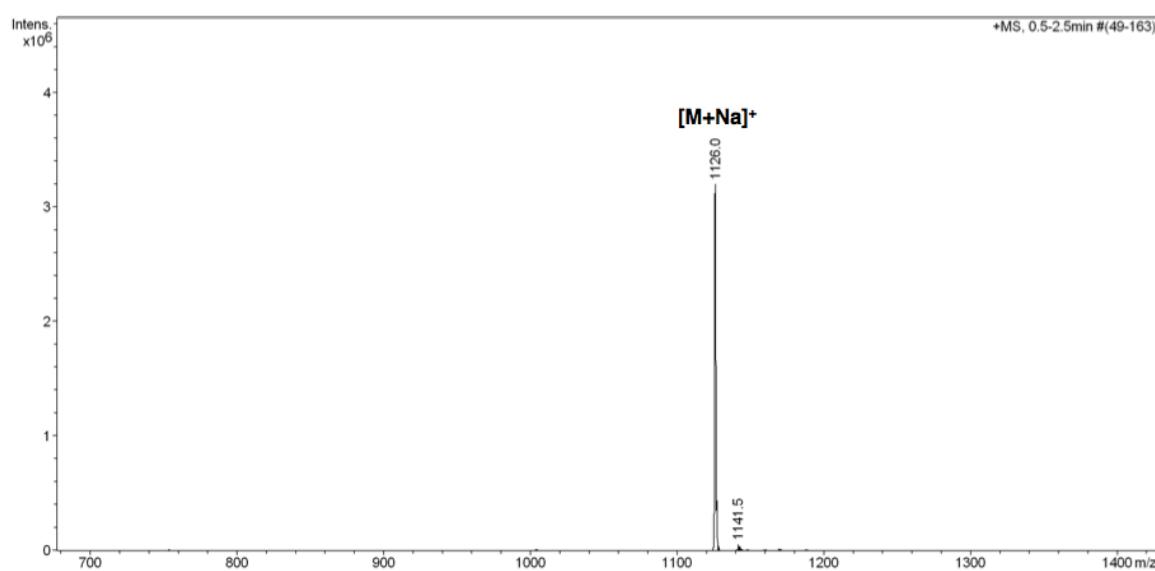
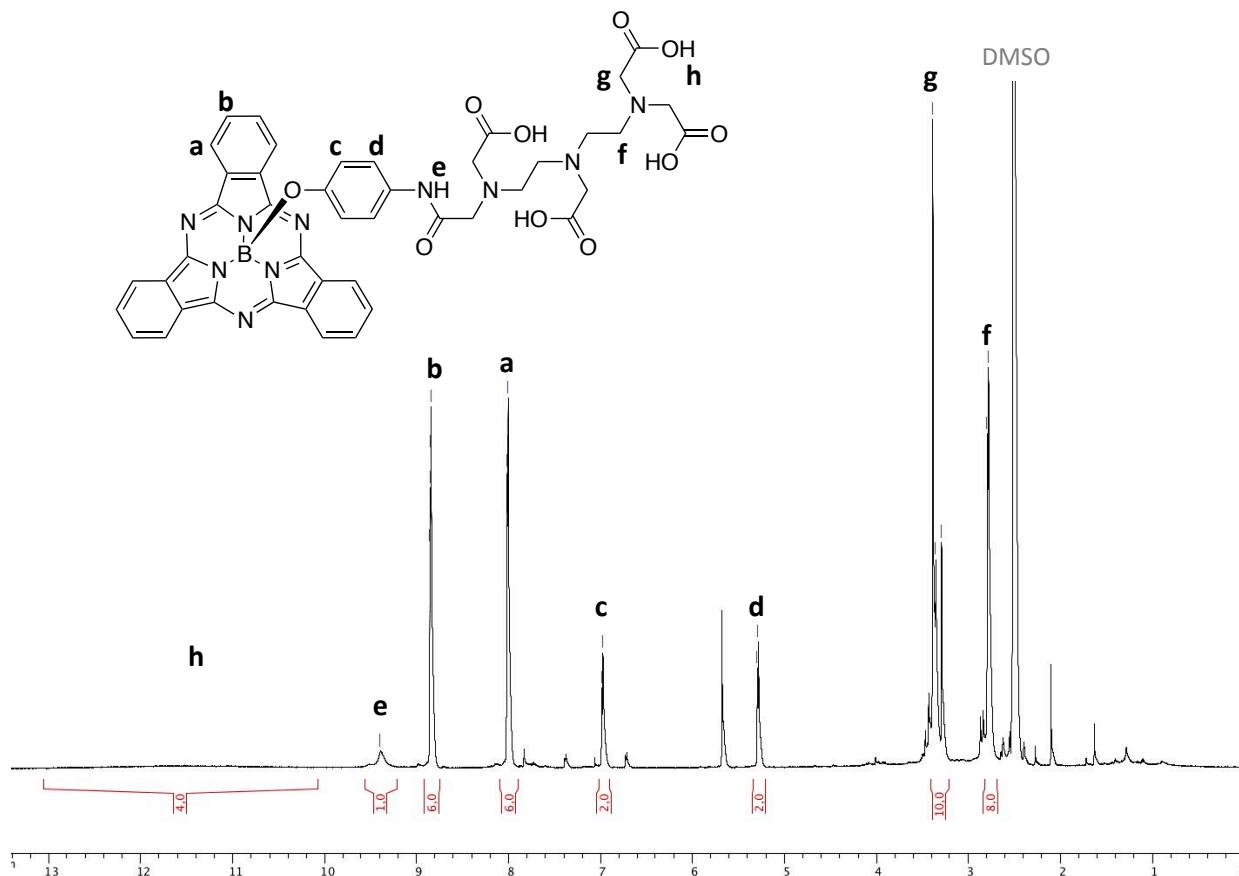
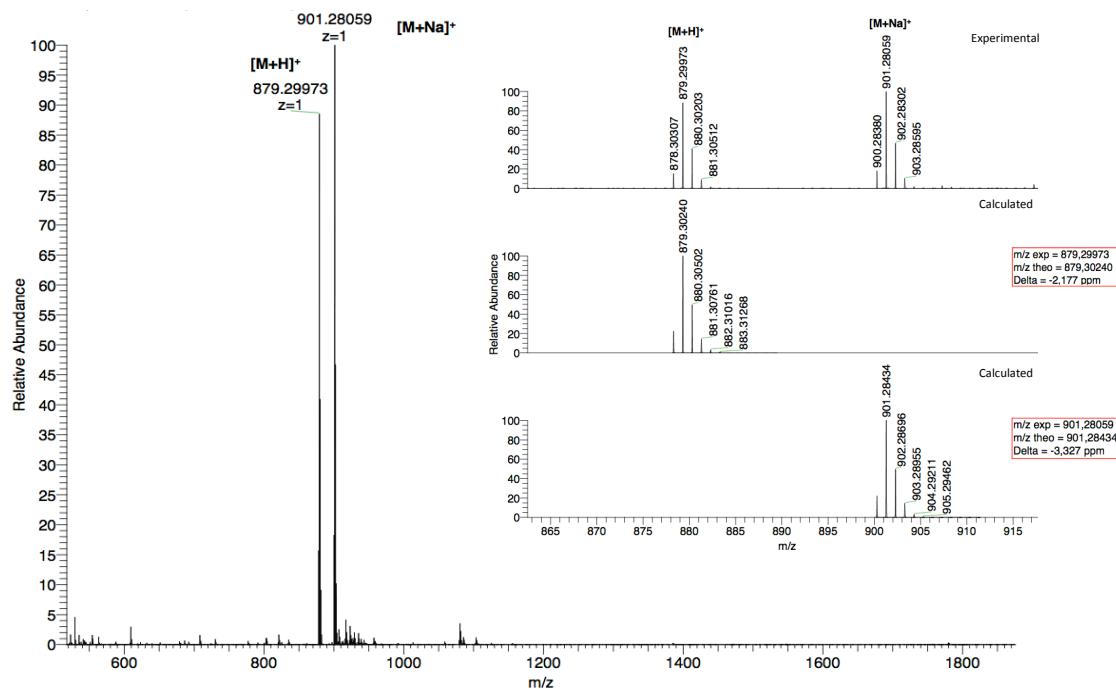


Fig. S4. MS spectrum of SubPc-DTPA- $t\text{Bu}_3$  (**6**)

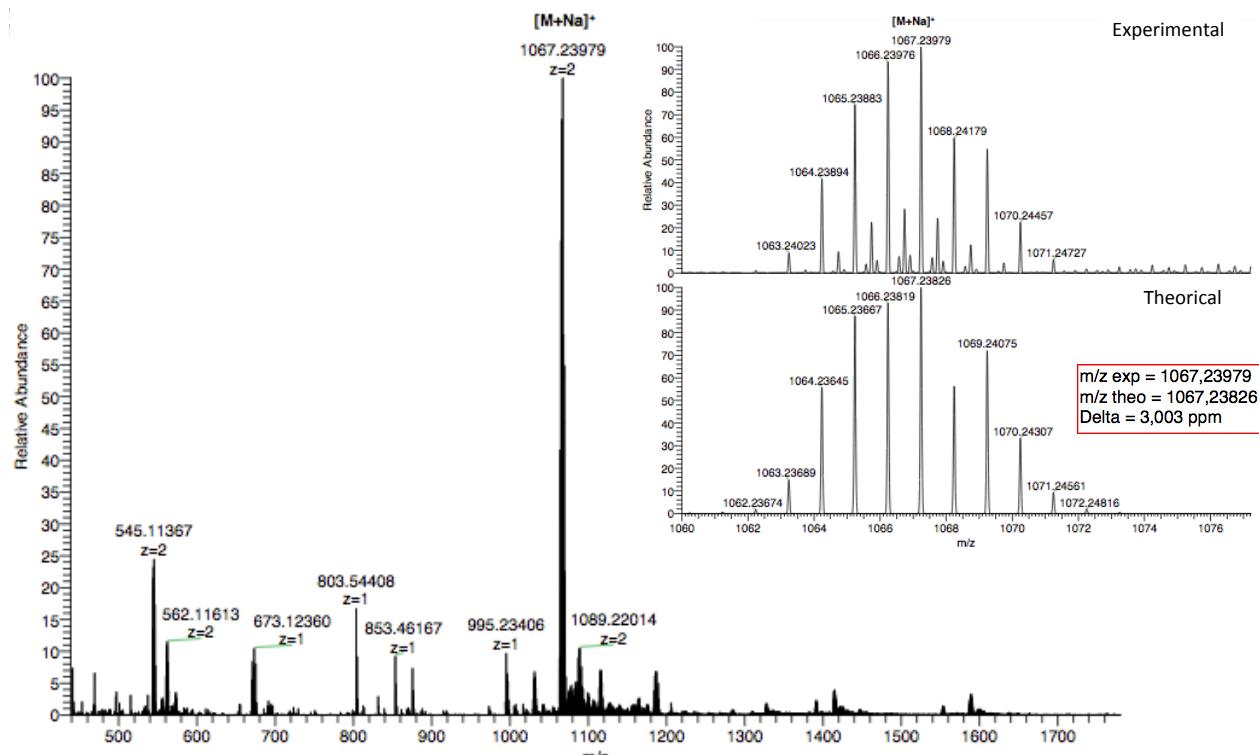


*Fig. S5.*  $^1\text{H}$  NMR spectrum of SubPc-DTPA (2) (DMSO, 600 MHz, 400K)

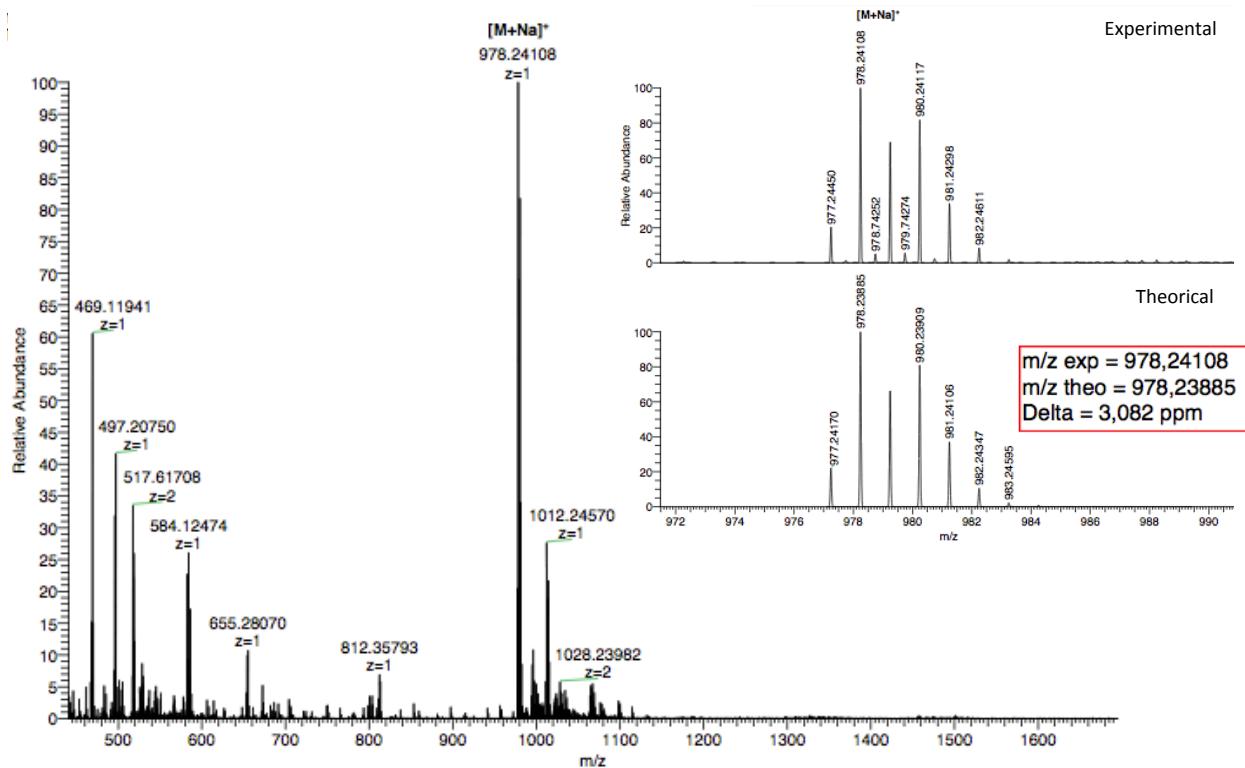


*Fig. S6.* HR-MS spectrum of SubPc-DTPA (2)

### B.3. SubPc-MDOTA (1-M) and SubPc-MDTPA (2-M)



*Fig. S7. HR-MS spectrum of SubPc-GdDOTA (1-Gd)*



*Fig. S8. HR-MS spectrum of SubPc-GaDOTA (1-Ga)*

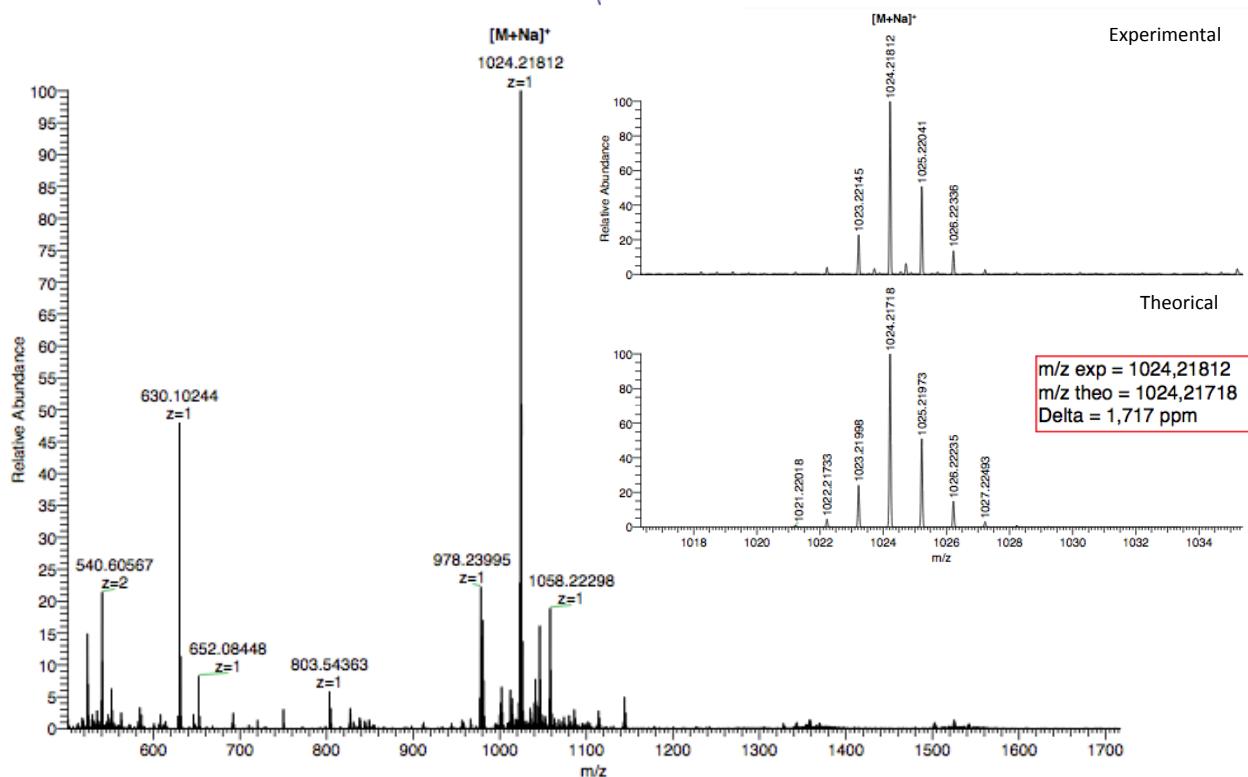


Fig. S9. HR-MS spectrum of SubPc-InDOTA (1-In)

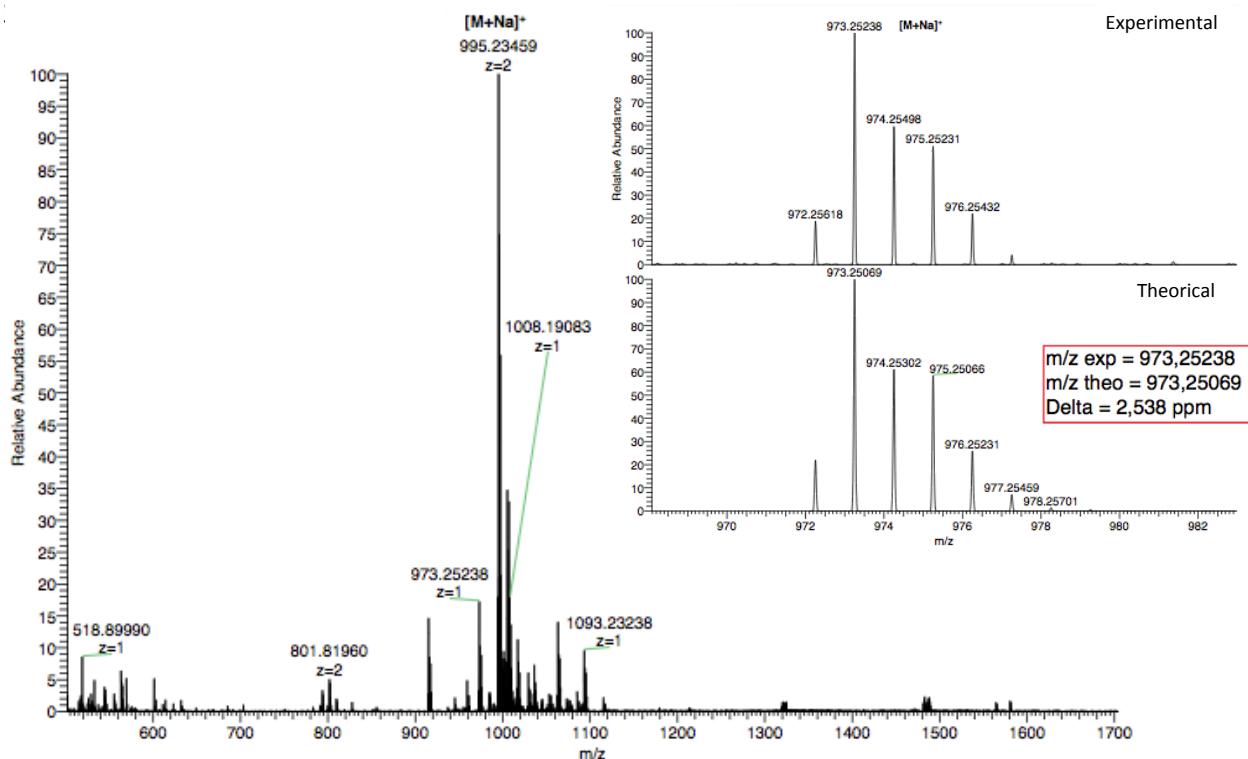
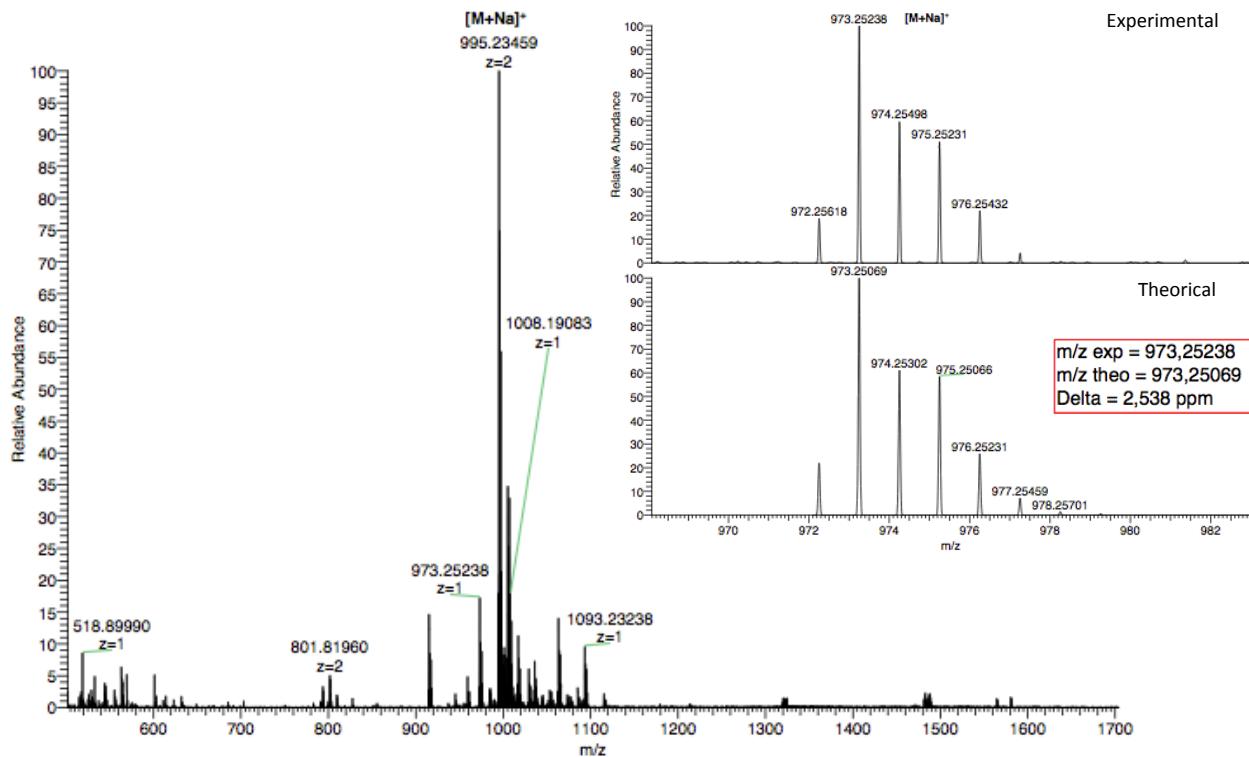
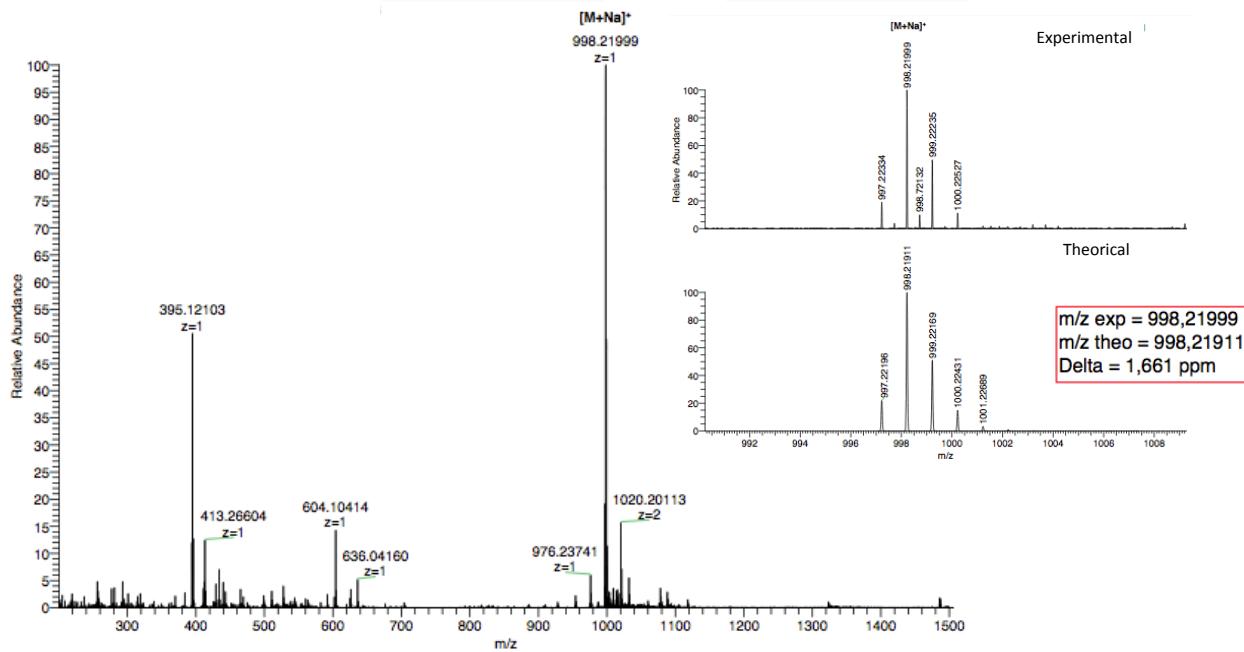


Fig. S10. HR-MS spectrum of SubPc-LuDOTA (1-Lu)

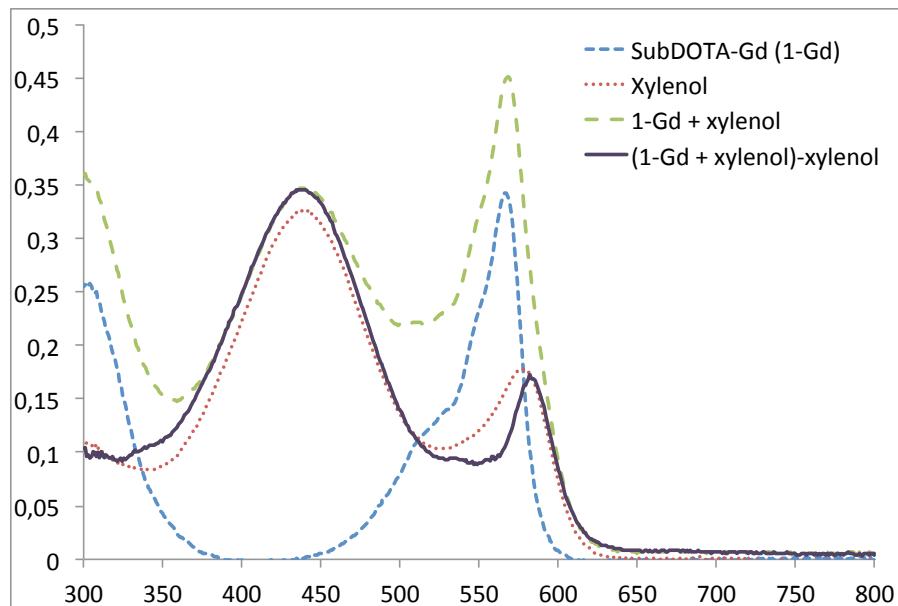


*Fig. S11. HR-MS spectrum of SubPc-LuDOTA (1-Lu)*



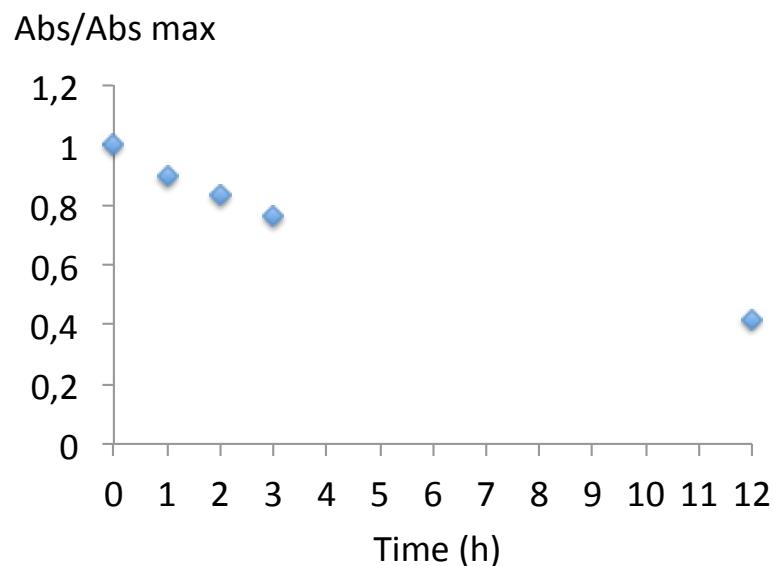
*Fig. S12. HR-MS spectrum of SubPc-LuDOTA (1-Lu)*

## C. DETECTION OF FREE GADOLINIUM



*Fig. S13. Absorption spectra of **1-Gd** in methanol (blue curve), xlenol in water (red curve), mixture of **1-Gd** and xlenol (green curve). The full line curve (black curve) correspond to the subtraction of mixture of **1-Gd** and xlenol curve minus the free xlenol curve.*

#### D. STABILITY STUDIES IN RPMI



*Fig. S14. Absorption of **1-Gd** in RPMI in dark as a function of time*