Electronic Supplementary Information

Synthesis, X-ray structure and *in vitro* cytotoxicity studies of Cu(I/II) complexes of thiosemicarbazone: Special emphasis on their interactions with DNA

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Table S1 DNA binding parameters for the ligands.

Fig. S1 Absorption spectral traces of the complexes 1 (a), 2 (b), 3 (c) and 4 (d) (25 µM each) in 10 mM Tris-HCl buffer (pH

8.0) containing 1% DMF. The spectra were recorded over the period of the biological assays, *i.e.* 72 h, at room temperature.

Fig. S2 ¹H NMR spectra of complex 1 in DMSO-d₆ (a) at time 0 hrs and room temp; (b) at time 24 hrs and room temp; (c) at time 0 hrs and 100 °C.

Fig. S3 ESI-MS of complex 1 in CH₃CN in the range of molecular mass of metal precursor.

Fig. S4 ESI-MS of complex 4 in CH₃OH.

Fig. S5 Cyclic voltammogram of 1 in cathodic region.

Fig. S6 Cyclic voltammogram of 1 in anodic region.

Fig. S7 Cyclic voltammogram of ligand HL³ in anodic region.

Fig. S8 Electronic absorption spectra of HL¹ (a), HL² (b), HL³ (c) and H₂L⁴ (d) (25 μ M each) upon the titration of CT–DNA (0 – 350 μ M) in 10 mM Tris–HCl buffer (pH 8.0) containing 1% DMF. The inset shows the linear fit of [DNA]/($\epsilon_a - \epsilon_f$) vs [DNA] and binding constant (K_b) was calculated using Eq. 1.

Fig. S9 Fluorescence absorption spectra of HL¹ (a), HL² (b), HL³ (c) and H₂L⁴ (d) (0–60 μ M) on the emission intensity of ethidium bromide (2 μ M) bound CT-DNA (50 μ M) at different concentrations in 10 mM Tris–HCl buffer (pH 8.0) containing 1% DMF. Arrow indicates the effect of increasing concentration of complex on the fluorescence emission of CT–DNA bound ethidium bromide. The inset shows the linear fit of F₀/F *vs* [complex] and Stern-Volmer quenching constant (K_{sv}) was calculated using Eq. 2.

Fig. S10 Effect of DMF (1%) and ligands on the chemical-induced cleavage of SC pUC19 DNA. 300 ng SC pUC19 DNA was treated with hydrogen peroxide (0.5 mM) in dark for 1 h at 37 °C in presence of 1 % DMF and various ligands (100 μ M). Lane 1, DNA only; Lane 2, DNA in presence of 1% DMF; Lane 3, DNA + HL¹; Lane 4, DNA + HL²; Lane 5, DNA + HL³; Lane 6, DNA + H₂L⁴.

Fig. S11 Gel diagram depicting cleavage of SC pUC19 DNA by 1–4 in presence of various additives in 50 mM Tris-HCl buffer (pH 8.0) containing 1% DMF. SC pUC19 DNA (300 ng) in the presence of various additives was treated with hydrogen peroxide (0.5mM) in dark for 1 h at 37 °C with 1–4 (100 μ M). The additive concentrations were: sodium azide (0.5 mM), L-histidine (0.5 mM), KI (0.5 mM) and D-mannitol (0.5 mM). Lane 1, DNA + complex; Lane 2, DNA + complex + sodium azide; Lane 3, DNA + complex + L-histidine; Lane 4, DNA + complex + KI; Lane 5, DNA + complex + D-mannitol.

Fig. S12 Effect of DMF (1%) and ligands on the photo-induced cleavage of SC pUC19 DNA. 300 ng SC pUC19 DNA was photo-irradiated in presence of 1% DMF and various ligands (100 μ M) with UVA at 350 nm for 1 h. Lane 1, DNA only; Lane 2, DNA in presence of 1% DMF; Lane 3, DNA + HL¹; Lane 4, DNA + HL²; Lane 5, DNA + HL³; Lane 6, DNA + H₂L⁴.

Fig. S13 Gel diagram depicting cleavage of SC pUC19 DNA by 1–4 in presence of various additives in 50 mM Tris–HCl buffer (pH 8.0) containing 1% DMF. SC pUC19 DNA (300 ng) in the presence of various additives was photo–irradiated at 350 nm for 1 h with 1–4 (100 μ M). The additive concentrations were: sodium azide (0.5 mM), L–histidine (0.5 mM), KI (0.5 mM) and D–mannitol (0.5 mM). Lane 1, DNA + complex; Lane 2, DNA + complex + sodium azide; Lane 3, DNA + complex + L-histidine; Lane 4, DNA + complex + KI; Lane 5, DNA + complex + D-mannitol.

Table S1 DNA binding parameters for the ligands

| Ligands | Binding Constant (K _b) ^a (M ⁻¹) | Stern–Volmer Quenching Constant (K _{SV}) (M ⁻¹) ^b | $K_{app} \left(M^{-1} ight)^{c}$ |
|-----------------|---|---|------------------------------------|
| HL ¹ | 5.50×10^{3} | 5.01×10^{2} | 3.02×10^{5} |
| HL ² | 5.20×10^{3} | 5.80×10^2 | 3.60×10^{5} |
| HL ³ | 1.80×10^{3} | 9.28×10^2 | 6.70×10^{5} |
| H_2L^4 | 7.00×10^{3} | 4.50×10^2 | 2.69×10^{5} |

^aDNA binding constant by UV-vis spectral method. ^bStern-Volmer Quenching constant for CT-DNA-EB complex. ^cthe apparent DNA binding constant.



Fig. S1.



Fig. S2.







Fig. S4.



Fig. S5.



Fig. S6.



Fig. S7.



Fig. S8.



Fig. S9.



Fig. S10.



Fig. S11.



Fig. S12.



Fig. S13.