oxazolidinones

### **Electronic Supplementary Information**

## CO<sub>2</sub> capture and activation by superbase/polyethylene glycol and its subsequent conversion

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**S8** 

#### 1. General experimental methods:

#### Caution

Experiments using compressed gases  $CO_2$  are potentially hazardous and must only be carried out by using the appropriate equipment and under rigorous safety precautions.

#### Materials

Superbases and PEG used in this work were purchased from Alfa Aesar-A Johnson Matthey Company and dried under vacuum at 70  $^{\circ}$ C for one week before use. CO<sub>2</sub> with a purity of 99.999% was commercially available. Amines was refluxed for 10 h with CaH<sub>2</sub> and distilled prior to use. NH<sub>2</sub>PEG150NH<sub>2</sub><sup>1</sup> and aziridines<sup>2</sup> was synthesized according to the reported method.

#### **Experimental methods**

<sup>1</sup>H NMR spectra was recorded at Bruck 400 spectrometer in CDCl<sub>3</sub> and CDCl<sub>3</sub> (7.26 ppm) was used as internal reference, <sup>13</sup>C NMR was recorded at 100.6 MHz in CDCl<sub>3</sub> and CDCl<sub>3</sub> (77.0 ppm) was used as internal reference. GC analyses were performed on Shimadzu GC-2014, equipped with a capillary column (RTX-17, 30 m × 0.25  $\mu$ m) using a flame ionization detector. *In situ* FTIR was collected on a Mettler Toledo React IR ic10, Diamond ATR probe, using ic IR analysis system. The probe is placed in the middle of the absorption mixture, which is constantly stirred by magnetic whisk, and the spectra are collected *in situ* during CO<sub>2</sub> absorption. Infrared (IR) spectra were recorded on a Bruker Tensor27 FT-IR spectrophotometer with KBr pellets.

#### General procedure for CO<sub>2</sub> absorption

In a typical procedure,  $CO_2$  capture was carried out in a 10 mL Schlenk flask. The absorbents were charged into the reactor at room temperature. Then, the air in the flask was replaced by  $CO_2$  and a needle was used for  $CO_2$  bubbling, which was inserted in the bottom of the flask. The absorption reaction was conducted at 40 °C with a  $CO_2$  bubbling rate of 0.1 L/min. The amount of  $CO_2$  absorbed was determined by an Analytical Balance within an accuracy of  $\pm 0.0001$  g every five minutes. Absorption/desorption was determined by several cycles of repeated experiments.

#### General procedure for synthesis of value-added chemicals using captured CO<sub>2</sub>

Taking the systhesis of 1,3-dibutylurea as an example: Firstly, DBU (3 mmol, 0.4567 g) and PEG<sub>150</sub> (3 mmol, 0.45 g) were charged into a glass tube, in which CO<sub>2</sub> was bubbled through a needle. Then, 1-butylamine was added after CO<sub>2</sub> absorption reached equilibrium. The tube was placed into a 25 mL stainless steel autoclave and then the mixture was stirred at predetermined temperature for 5 min to reach the equilibration. When the reaction finished, the reactor was cooled in ice-water and CO<sub>2</sub> was ejected slowly. An aliquot of sample was taken from the resultant mixture and dissolved in dry CH<sub>2</sub>Cl<sub>2</sub> for GC analysis.

# 2. Characterization of the absorption system, amidinium/guanidinium alkylcarbonate salts, aziridines, ureas and oxazolidinones

#### DBU (1,8-diazabicyclo[5.4.0]undec-7-ene):

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  3.23 (t, <sup>3</sup>*J* = 5.6 Hz, 2 H), 3.13-3.18 (m, 4 H), 2.33-2.35 (m, 2 H), 1.72-1.77 (m, 2 H), 1.60 (s, 4 H), 1.52 (s, 2 H); <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>)  $\delta$  161.6, 52.8, 48.3, 44.0, 37.2, 29.7, 28.4, 25.9, 22.3. IR 3420, 2924, 2851, 1616, 1486, 1441, 1366, 1312, 1260, 1233, 1183, 1115, 1060, 991, 955 cm<sup>-1</sup>.

#### **PEG**<sub>150</sub> (triethylene glycol, **MW** = 150 Da):

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  3.71 (t, <sup>3</sup>*J* = 4.8 Hz, 4 H), 3.65 (s, 4 H), 3.59 (t, <sup>3</sup>*J* = 4.8 Hz, 4 H), 3.40 (s, 2 H); <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>)  $\delta$  72.6, 70.3, 61.5. IR 3446, 2865, 2360, 2341, 2136, 1954, 1653, 1457, 1351, 1082, 937, 889, 830, 571, 525 cm<sup>-1</sup>.

#### DBU/PEG<sub>150</sub> + CO<sub>2</sub>:

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  3.56-3.70 (m, 12 H), 3.29-3.34 (m, 6 H), 2.61 (s, 2 H), 1.85-1.90 (m, 2 H), 1.66-1.67 (m, 4 H), 1.59 (s, 2 H); <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>)  $\delta$  164.1, 158.5, 72.6, 70.3, 64.1, 61.3, 53.5, 48.4, 40.6, 34.1, 29.3, 27.6, 24.8, 20.8.



Fig. S1 The <sup>1</sup>H NMR of DBU, PEG<sub>150</sub> and DBU/PEG<sub>150</sub> + CO<sub>2</sub> (CDCl<sub>3</sub>, 400 MHz).

	DBU	DBUPEG <sub>150</sub> CO <sub>2</sub>	PEG <sub>150</sub>
	1.52 (2H)	1.59 (2H)	
<sup>1</sup> H NMR /ppm	1.60 (4H)	1.60-1.67 (4H)	
	1.72-1.77 (2H)	1.85-1.90 (2H)	
	2.33-2.35 (2H)	2.61 (2H)	
	3.13-3.18 (4H)		
	3.23 (2H)	3.29-3.34 (6H)	
		3.56-3.70 (12H)	3.59-3.71 (12H)
	22.3	20.8	
	25.9	24.8	
	28.4	27.6	
	29.7	29.3	
	37.2	34.1	
	44.0	40.6	
	48.3	48.4	
<sup>13</sup> C NMR	52.8	53.5	
/ppm		61.3	61.5
		64.1	
		70.3	70.3
		72.6	72.6
		158.5	
	161.6	164.1	

Table S1 The comparison of <sup>1</sup> H and <sup>13</sup> C NMR chemical shifts of	f DBU, PEG <sub>150</sub> and DBU/PEG <sub>150</sub> + CO
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**Figure S2.** Results of *in situ* FT-IR spectroscopy monitoring CO<sub>2</sub> capture by DBU/PEG<sub>150</sub> system at various times starting from CO<sub>2</sub> bubbling; The spectrum of DBU and PEG<sub>150</sub> was subtracted;  $\blacklozenge$ : CO<sub>2</sub> began to be bubbled at 40 °C;  $\diamondsuit$ : Temperature was gradually increased form 40 °C to 120 °C.



**Figure S3.** The scanning thermogravimetric analysis (TGA) results for DBU/PEG<sub>600</sub>+CO<sub>2</sub> with a 10 °C min<sup>-1</sup> temperature ramping rate to 700 °C.

#### PEG<sub>600</sub> (polyethylene glycol, MW = 600 Da):

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 3.57-3.69 (m, 44H), 2.75 (s, 2H); <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>) δ72.4, 70.4, 70.2, 61.6. IR 3375, 2870, 1636, 1457, 1350, 1297, 1249, 1107, 951, 845, 735, 559 cm<sup>-1</sup>.

#### **DBU/PEG<sub>600</sub> + CO<sub>2</sub>:**

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  3.59-3.70 (m, 53H), 3.29 (t, <sup>3</sup>*J* = 5.6 Hz, 2H), 3.20-3.24 (m, 4H), 2.46 (s, 2H), 1.78-1.84 (m, 2H), 1.65 (s, 4H), 1.57 (s, 2H); <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>)  $\delta$  164.6, 157.7, 77.2, 72.0, 69.8, 69.6, 63.5, 60.6, 53.3, 47.9, 38.4, 32.1, 28.5, 26.5, 23.8, 19.4. IR 3350, 2867, 2361, 2343, 1647, 1613, 1456, 1351, 1315, 1291, 1250, 1110, 993, 953, 885, 837, 731, 694, 546, 526 cm<sup>-1</sup>.

#### TBD (1,5,7-triazabicyclo[4.4.0]dec-5-ene):

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  3.29 (t, <sup>3</sup>*J* = 5.6 Hz, 4H), 3.24 (t, <sup>3</sup>*J* = 6 Hz, 4H), 1.96 (m, 4H); <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>)  $\delta$  151.2, 46.6, 38.0, 20.7. IR 3477, 3418, 3234, 3157, 2967, 2943, 2875, 2797, 2749, 2361, 2343, 1662, 1573, 1477, 1445, 1421, 1378, 1321, 1295, 1202, 1069, 884, 711, 671, 590 cm<sup>-1</sup>.

#### TBD/PEG<sub>600</sub> + CO<sub>2</sub>:

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 3.56-3.68 (m, 62H), 3.21-3.27 (m, 8H), 1.91-1.97 (m, 4H); <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>) δ 158.8, 150.7, 77.2, 72.0, 69.9, 69.7, 64.0, 60.7, 46.2, 37.0, 20.3. IR 3308, 3160, 2869, 2362, 1943, 1667, 1574, 1456, 1377, 1350, 1323, 1291, 1252, 1203, 1107, 951, 844, 745, 730, 714 cm<sup>-1</sup>.

#### DBN (1,5-diazabicyclo[4.3.0]non-5-ene):

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  3.28 (t, <sup>3</sup>*J* = 4.8 Hz, 2H), 3.21 (t, <sup>3</sup>*J* = 6.8 Hz, 2H), 3.13 (t, <sup>3</sup>*J* = 6 Hz, 2H), 2.38 (t, <sup>3</sup>*J* = 7.6 Hz, 2H), 1.83-1.91 (m, 2H), 1.70-1.76 (m, 2H); <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>)  $\delta$ 160.5, 51.2, 43.8, 42.8, 31.3, 20.6, 19.4. IR 3409, 2929, 2845, 1651, 1496, 1422, 1363, 1288, 1232, 1195, 1133, 1051, 961, 903, 737, 677, 574 cm<sup>-1</sup>.

#### DBN/PEG<sub>600</sub> + CO<sub>2</sub>:

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  3.58-3.72 (m, 58H), 3.31 (t, <sup>3</sup>*J* = 5.6 Hz, 2H), 3.26 (t, <sup>3</sup>*J* = 6.4 Hz, 2H), 3.17 (t, <sup>3</sup>*J* = 6 Hz, 2H), 2.45 (t, <sup>3</sup>*J* = 8 Hz, 2H), 1.88-1.95 (m, 2H), 1.74-1.80 (m, 2H); <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>)  $\delta$ 162.8, 157.6, 77.2, 72.0, 69.8, 69.5, 63.4, 60.4, 52.0, 41.9, 38.6, 29.4, 18.5, 18.3. IR 3394, 2870, 2361, 2342, 1681, 1651, 1457, 1350, 1291, 1251, 1107, 952, 837, 747, 730, 698 cm<sup>-1</sup>.

#### TMG(tetramethylguanidine):

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 5.01 (s, 1H), 2.70 (s, 12H); <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>) δ 167.9, 39.3. IR 3420, 3334, 3000, 2942, 2845, 2789, 2360, 2341, 1599, 1494, 1458, 1425, 1409, 1382, 1255, 1198, 1145, 1058, 1010, 892, 779, 734, 562, 544 cm<sup>-1</sup>.

#### TMG/PEG<sub>600</sub> + CO<sub>2</sub>:

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 3.55-3.69 (m, 60H), 2.71 (s, 12H); <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>) δ 163.6, 157.4, 77.2, 72.1, 69.9, 69.7, 63.4, 60.6, 38.9. IR 3393, 2870, 2361, 2343, 1647, 1472, 1457, 1351, 1277, 1251, 1108, 952, 838, 749 cm<sup>-1</sup>.

#### DMICH(N-(1,3-dimethylimidazolidin-2-ylidene)cyclohexanamine):

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ3.38–3.43 (m, 1H), 3.11 (s, 4H), 2.76 (s, 6H), 1.15–1.72 (m, 10H); <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>) δ 155.4, 54.0, 44.9, 36.5, 31.3, 25.8, 25.2.

#### DMICH/PEG<sub>600</sub> + CO<sub>2</sub>:

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  3.56-3.69 (m, 56H), 3.34-3.39 (m, 1H), 3.17 (s, 4H), 2.79 (s, 6H), 1.54-1.73 (m, 5H), 1.13-1.41 (m, 5H); <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>)  $\delta$  157.5, 157.3, 124.4, 77.2, 72.3, 70.2, 70.0, 63.6, 61.1, 54.1, 59.2, 44.7, 36.1, 35.0, 34.9, 25.1, 18.0. IR 3393, 2922, 2867, 2361, 2342, 1653, 1635, 1472, 1456, 1350, 1279, 1256, 1109, 1035, 952, 890, 847, 763, 729, 698 cm<sup>-1</sup>.

#### NH<sub>2</sub>PEG<sub>150</sub>NH<sub>2</sub>:

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  3.47 (s, 4 H), 3.35 (t, <sup>3</sup>*J* = 5.2 Hz, 4 H), 2.71 (t, <sup>3</sup>*J* = 5.2 Hz, 4 H), 1.17 (s, 1 H); <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>)  $\delta$  73.2, 70.0, 41.5. IR 3393, 2869, 1576, 1473, 1375, 1352, 1307, 1114, 817 cm<sup>-1</sup>.

#### NH<sub>2</sub>PEG<sub>150</sub>NH<sub>2</sub>/PEG<sub>150</sub>(molar ratio: 1: 2) + CO<sub>2</sub>:

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  3.72 (t, <sup>3</sup>*J* = 4 Hz, 12H), 3.66 (s, 8H), 3.59-3.62 (m, 12H), 3.54 (t, <sup>3</sup>*J* = 4.8 Hz, 2H), 3.26 (s, 1H), 2.89 (s, 2H); <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>)  $\delta$  162.8, 72.3, 69.8, 68.2, 60.7, 40.9, 39.3. IR 3365, 2870, 2361, 2342, 1576, 1558, 1473, 1457, 1350, 1311, 1117, 1078, 936, 888, 821, 748 cm<sup>-1</sup>.

#### 1,3-Dipropyl urea:

<sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  4.90 (bd, N-H), 3.08 (t, <sup>3</sup>*J* = 6.45 Hz, 4H), 1.41-1.54 (m, 4H), 0.89 (t, <sup>3</sup>*J* = 7.5 Hz, 6H); <sup>13</sup>C NMR (75.5 MHz, CDCl<sub>3</sub>)  $\delta$  158.8, 42.1, 23.4, 11.3.

#### 1,3-Diisopropyl urea:

<sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  3.80-3.87 (m, 2H), 1.14 (d, <sup>3</sup>J = 6.4 Hz, 12H); <sup>13</sup>C NMR (75.5 MHz, CDCl<sub>3</sub>)  $\delta$  157.0, 42.2, 23.5.

#### 1,3-Dibutyl urea:

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  4.77 (s, 2 H), 3.12-3.13 (m, 4H), 1.42-1.49 (m, 4H), 1.28-1.37 (m, 4H), 0.91 (t, <sup>3</sup>J = 7.2 Hz, 6H); <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>)  $\delta$  158.7, 40.2, 32.3, 20.0, 13.7.

#### 1,1,3,3-Tetrabutyl urea:

<sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  2.56 (t, <sup>3</sup>*J* = 7.2 Hz, 8H), 1.36-1.49 (m, 8H), 1.24-1.33 (m, 8H), 0.87 (t, <sup>3</sup>*J* = 7.2 Hz, 12H); <sup>13</sup>C NMR (75.5 MHz, CDCl<sub>3</sub>)  $\delta$  164.3, 49.7, 32.2, 20.5, 13.9.

#### Octahydro-benzoimidazol-2-one:

<sup>1</sup>H NMR (300 MHz, D<sub>2</sub>O) δ 3.12-3.32 (m, 2H), 1.53-1.60 (m, 4H), 1.15-1.35 (m, 4H); <sup>13</sup>C NMR (75.5 MHz, D<sub>2</sub>O) δ 164.1, 53.3, 32.2, 24.2.

#### 1-Ethyl-2-phenylaziridine:

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ 7.18-7.31 (m, 5H), 2.44 (q, <sup>3</sup>*J* = 9.6 Hz, 2H), 2.30 (dd, <sup>3</sup>*J* = 4.4 Hz, <sup>3</sup>*J* = 4.8 Hz, 1H), 1.89 (d, <sup>2</sup>*J* = 4.4 Hz, 1H), 1.65 (d, <sup>2</sup>*J* = 8.8 Hz, 1H), 1.17 (t, <sup>3</sup>*J* = 9.6 Hz, 3H); ESI-MS calcd for C<sub>10</sub>H<sub>13</sub>N 147.10, found 148.31 [M + H]<sup>+</sup>.

#### 1-Propyl-2-phenylaziridine:

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  0.95 (t, <sup>3</sup>*J* = 10.0 Hz, 3H), 1.60-1.67 (m, 3H), 1.89 (d, <sup>2</sup>*J* = 4.0 Hz, 1H), 2.24-2.33 (m, 2H), 2.43-2.51 (m, 1H), 7.18-7.31 (m, 5H); ESI-MS calcd for C<sub>11</sub>H<sub>15</sub>N 161.12, found 162.28 [M + H]<sup>+</sup>.

#### 2-(4-Chlorophenyl)-1-ethylaziridine:

<sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  1.18 (t, <sup>3</sup>*J* = 6.9 Hz, 3H), 1.65 (d, <sup>2</sup>*J* = 6.6 Hz, 1H), 1.83 (d, <sup>2</sup>*J* = 3.3 Hz, 1H), 2.25-2.46 (m, 3H), 7.15-7.23 (m, 4H); ESI-MS calcd for C<sub>10</sub>H<sub>12</sub>NCl 181.66, found 182.13 [M + H]<sup>+</sup>.

#### 2-(4-Methylphenyl)-1-ethylaziridine:

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.19 (t, <sup>3</sup>*J* = 7.2 Hz, 3H), 1.62 (d, <sup>2</sup>*J* = 6.4 Hz, 1H), 1.86 (d, <sup>2</sup>*J* = 3.2 Hz, 1H), 2.26 (dd, <sup>3</sup>*J* = 3.6 Hz, <sup>3</sup>*J* = 3.2 Hz, 1H), 2.31 (s, 3H), 2.37-2.48 (m, 2H), 7.09-7.15 (m, 4H); ESI-MS calcd for C<sub>11</sub>H<sub>15</sub>N 161.24, found 162.20 [M + H]<sup>+</sup>.

#### 3-Ethyl-5-phenyloxazolidin-2-one:

Colorless liquid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  1.17 (t, <sup>3</sup>*J* = 7.2 Hz, 3H), 3.29-3.45 (m, 3H), 3.92 (t, <sup>3</sup>*J* = 8.7 Hz, 1H), 5.48 (t, <sup>3</sup>*J* = 7.8 Hz, 1H), 7.34-7.42 (m, 5H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  12.4, 38.8, 51.5, 74.2, 125.4, 128.6, 128.8, 138.8, 157.5; ESI-MS calcd for C<sub>11</sub>H<sub>13</sub>NO<sub>2</sub> 191.09, found 192.29 (M + H)<sup>+</sup>, 214.38 (M + Na)<sup>+</sup>, 405.01 (2 M + Na)<sup>+</sup>.

#### 3-Ethyl-4-phenyloxazolidin-2-one:

Colorless liquid; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.05 (t, <sup>3</sup>*J* = 7.2 Hz, 3H), 2.79-2.88 (m, 1H), 3.48-3.57 (m, 1H), 4.10 (t, <sup>3</sup>*J* = 8.0 Hz, 1H), 4.62 (t, <sup>3</sup>*J* = 8.8 Hz, 1H), 4.81 (t, <sup>3</sup>*J* = 7.2 Hz, 1H), 7.30 7.44 (m, 5H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  12.1, 36.8, 59.3, 69.7, 126.9, 129.0, 129.2, 137.8, 158.1; ESI-MS calcd for C<sub>11</sub>H<sub>13</sub>NO<sub>2</sub> 191.09, found 192.29 (M + H)<sup>+</sup>, 214.38 (M + Na)<sup>+</sup>, 405.01 (2M + Na)<sup>+</sup>.

#### 3-Propyl-5-phenyloxazolidin-2-one:

Colorless liquid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  0.91 (t, <sup>3</sup>*J*=7.2 Hz, 3H), 1.52-1.61 (m, 2H), 3.18-3.31 (m, 2H) 3.40 (t, <sup>3</sup>*J*=8.0 Hz, 1H), 3.90 (t, <sup>3</sup>*J*=8.8 Hz, 1H), 5.46 (t, <sup>3</sup>*J*=8.0 Hz, 1H), 7.31-7.37 (m, 5H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  10.7, 20.3, 45.5, 51.8, 74.0, 125.2, 128.4, 128.5, 138.7, 157.6; ESI-MS calcd for C<sub>12</sub>H<sub>15</sub>NO<sub>2</sub> 205.11, found 206.30 (M + H)<sup>+</sup>, 228.30 (M + Na)<sup>+</sup>, 433.04 (2M + Na)<sup>+</sup>.

#### 3-Ethyl-5-(4-chlorophenyl)oxazolidin-2-one:

White solid; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.17 (t, <sup>3</sup>*J*=7.3 Hz, 3H), 3.30-3.43 (m, 2H), 3.69-3.76 (m, 1H), 3.92 (t, <sup>3</sup>*J*=8.7 Hz, 1H), 5.44 (t, <sup>3</sup>*J*=8.0 Hz, 1H), 7.27-7.38 (m, 4H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  12.6, 38.9, 51.5, 73.6, 126.9, 129.1, 134.7, 137.4, 157.4; ESI-MS calcd for C<sub>11</sub>H<sub>12</sub>ClNO<sub>2</sub> 225.67, found 451.64 (2M + H)<sup>+</sup>.

#### 3-Ethyl-5-p-tolyloxazolidin-2-one:

White solid; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.18 (t, <sup>3</sup>*J*=7.3 Hz, 3H), 1.62 (d, <sup>3</sup>*J*=6.4 Hz, 1H),1.87 (d, <sup>3</sup>*J*=3.2 Hz, 1H), 2.27 (dd, <sup>3</sup>*J*=6.6 Hz, <sup>2</sup>*J*=3.2 Hz, 1H), 2.31 (s, 3H), 2.36-2.48 (m, 2H), 7.09- 7.15 (m, 4H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  12.6, 21.2, 38.9, 51.6, 74.3, 125.6, 129.5, 135.8, 138.7, 157.7; ESI-MS calcd for C<sub>12</sub>H<sub>15</sub>NO<sub>2</sub> 205.25, found 206.45 (M + H)<sup>+</sup>, 411.15 (2M + H)<sup>+</sup>.

#### 3. Reference

(1) Gansow, O. A.; Kausar, A. R.; Triplett, K. B. J. Heterocyclic Chem. 1981, 18, 297.

(2) Du, Y.; Wu, Y.; Liu, A.-H. and He, L.-N. J. Org. Chem. 2008, 73, 4709.

# 4. The <sup>1</sup>H NMR and <sup>13</sup>C NMR Charts, IR spectra for the absorption system, amidinium/guanidinium alkylcarbonate salts, urea and oxazolidinones













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IR spectrum of PEG<sub>600</sub>:

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IR spectrum of DBU/PEG<sub>600</sub> + CO<sub>2</sub>:

150



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IR spectrum of TBD:











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**IR spectrum of DBN:** 



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IR spectrum of DBN/PEG<sub>600</sub> + CO<sub>2</sub>:



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IR spectrum of TMG:

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IR spectrum of TMG/PEG<sub>600</sub> + CO<sub>2</sub>:







IR spectrum of DMICH/PEG<sub>600</sub> + CO<sub>2</sub>:

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#### IR spectrum of NH<sub>2</sub>PEG<sub>150</sub>NH<sub>2</sub>:



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IR spectrum of NH<sub>2</sub>PEG<sub>150</sub>NH<sub>2</sub>/PEG<sub>150</sub> (molar ratio: 1: 2) + CO<sub>2</sub>:

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1,3-Dipropyl urea:

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<sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)



1,3-Diisopropyl-urea:

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<sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)

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<sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)

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#### Octahydro-benzoimidazol-2-one:



<sup>1</sup>H NMR (D<sub>2</sub>O, 300 MHz)















