Supplementary Information

Facile Aerobic Photooxidation of Methyl Group in the Aromatic Nucleus in the Presence of an Organocatalyst under VIS Irradiation

Norihiro Tada, Kasumi Hattori, Tomoya Nobuta, Tsuyoshi Miura, and Akichika Itoh*

Laboratory of Pharmaceutical Synthetic Chemistry, Faculty of Pharmaceutical Science, Gifu Pharmaceutical University, 1-25-4 Daigaku-nishi, Gifu 501-1196, Japan

E-mail: itoha@gifu-pu.ac.jp

1.	General Information	SI-2
2.	Facile Aerobic Photooxidation of Methyl Group in the Aromatic Nucleus in the	the Presence of an
	Organocatalyst under VIS Irradiation	
2.1	Optimization of the Reaction Conditions (Supplementary data for Table 1)	SI-2
2.2	Optimization of the Reaction Conditions (Supplementary data for Table 2)	SI-4
2.3	Aerobic Photooxidation under Lower Concentration of Oxygen	SI-5
2.4	Time Course of the Aerobic Photooxidation	SI-5
2.5	Measurement of Peroxide by Iodometry	SI-7
2.6	General Procedure	SI-8
Appen	dix: ¹ H and ¹³ C NMR spectra	SI-10

1. General Information.

All dry solvents were obtained from Kanto Kagaku Co., Ltd. Other chemicals used were of reagent grade and were obtained from Aldrich Chemical Co., Tokyo Kasei Kogyo Co., Ltd. and Wako Pure Chemical Industries, Ltd. ¹H NMR and ¹³C NMR spectra were obtained on a JEOL ECA 500 spectrometer or JEOL AL 400 spectrometer or JEOL EX 400 spectrometer (500 or 400 MHz for ¹H NMR and 125 or 100 MHz for ¹³C NMR). Chemical shifts (δ) are reported in parts per million (ppm) downfield from internal Me₄Si. Mass spectra (MS) were obtained on a JEOL JMS-SX102A instrument. Preparative thin-layer chromatography (TLC) was carried out on precoated plates of silica gel (MERCK, silica gel F-254).

2. Facile Aerobic Photooxidation of Methyl Group in the Aromatic Nucleus in the Presence of an Organocatalyst under VIS Irradiation

2.1. Optimization of the Reaction Conditions (Supplementary data for Table 1)

^t Bu 1a (0.3 mmol)	air, fluoresce 2-CI-AQN (K ₂ CO ₃ (0.	(0.1 equiv	.) ′ ^t Bu	$\left[\bigcirc\right]$	R = CC = CC = CF = CF	J₃H 3a I₂OOH 6a			
entry			yield (%) ^a						
entry	solvent	2a	3a	6a	7a	s.m. (1a) ^b			
1	EtOAc	85	0	0	0	0			
2	acetone	20	0	50	26	6			
3	MeCN	0	0	49	21	11			
4	MeOH	0	0	0	30	70			
5	EtOH	0	0	0	28	72			
6	ⁱ PrOH	0	0	18	0	63			
7	ⁱ Pr ₂ O	0	0	trace	trace	92			
8	PhH	42	10	4	13	13			
9	hexane	0	0	trace	trace	78			
10 ^c	EtOAc	0	0	20	6	56			
11 ^d	EtOAc	0	0	0	0	100			

Table S1. Study of Solvent under Air

^a Yields were determined by ¹H-NMR. ^b Since the starting material is somewhat volatile, it is difficult to recover it perfectly even by careful work-up.^c Under N₂. ^d In the dark.

t _{Bu} 1a (0.3 m	$\begin{array}{c} \text{air, fluorescent}\\ \text{catalyst (0.}\\ \text{K}_2\text{CO}_3 (0.0)\end{array} \\ \end{array}$	1 equiv.) 5 equiv.)) → ^t Bu'	R	R = CC = CC = CH = CH	J₃H 3a I₂OOH 6a		
optry			yield (%) ^a					
entry	catalyst	2a	3a	6a	7a	s.m.(1a) ^b		
1	AQN	29	0	14	5	32		
2	2-Me-AQN	54	0	11	3	19		
3	2- ^t Bu-AQN	78	0	7	3	8		
4	AQN-2-CO ₂ H	6	0	7	3	68		
5	2-CI-AQN	85	0	0	0	0		
6	1-CI-AQN	33	0	7	4	49		
7	DCA	0	0	0	0	100		
8	anthracene	0	0	3	0	80		
9	benzophenone	0	0	0	0	96		
10	Rose bengal	0	0	0	0	89		
11	Methylene blue	0	0	0	0	77		

Table S2. Study of Catalyst under Air

^a Yields were determined by ¹H-NMR. ^b Since the starting material is somewhat volatile, it is difficult to recover it perfectly even by careful work-up.

2.2. Optimization of the Reaction Conditions (Supplementary data for Table 2)

'Bu	air, fluoresc 2-CI-AQN base (0	R R = CO = CC	-		
1a (0.3 mmol)	EtOAc ((1 mL), 24 h	- 00	-311 Ja	
entry	base	solubility	yield (%) ^a		
		(g /100g of H_2O at 25 °C)	2a	3a	
1	Li ₂ CO ₃	1.3	92	6	
2	Na ₂ CO ₃	29.4	55	24	
3 ^b	Na ₂ CO ₃	29.4.	91	0	
4 ^c	Na ₂ CO ₃	29.4	85	8	
5	NaHCO ₃	10.3	55	31	
6 ^b	NaHCO ₃	10.3	88(86) ^d	0	
7 ^c	NaHCO ₃	10.3	60	35	
8	K ₂ CO ₃	112.0	100(94) ^d	0	
9		0.8	38	61	
10 ^b		0.8	83	17	
11 ^c		0.8	76	15	
12	Ca(OH) ₂	0.2	33	60	
13 ^b	$Ca(OH)_2$	0.2	94(87) ^d	36	
14 ^c	$Ca(OH)_2$	0.2	53	36	
15 ^e	KOAc	256.0	84	16	
16 ^b		112.0	83	0	

Table S3. Study of Additive (Base) under Air

 a Yields were determined by $^1H\text{-}NMR.\ ^bH_2O$ (75 $\mu L) was used. <math display="inline">^c$ For 72 h. d Isolated yields. e For 15 h.

Table S4. Study of Additive (Acid) under Air

^t Bu 1a (0.3 mn	2-CI-AQN (acid (0.3 equiv	nt lamp (VIS) 0.08 equiv.) (.), H ₂ O (75 μL) mL), 24 h	- 'Bu	R = CO2 = CO2 = CO3 = CH4	₃H 3a		
entry	acid	pKa		yield (%) ^a			
		-	2a	3a	7a		
1	TfSO₃H	<i>H</i> ₀ = -15	52	14	trace		
2 ^b	TfSO₃H	<i>H</i> ₀ = -15	78	22	0		
3	H_2SO_4	<i>H</i> ₀ = -12	75	18	0		
4	p-TsOHH ₂ O	-2.8	76	18	0		
5	HBF_4	-0.4	92	0	0		
6 ^b	TFA	0.23	100 (99) ^c	0	0		
7	CCl ₃ CO ₂ H	0.77	94	0	0		
8	F ₂ CHCO ₂ H	1.24	64	32	0		
9	AcOH	4.74	42	53	0		
10 ^b	AcOH	4.74	83	10	0		
11 ^d	TFA	0.23	81	12	0		

 $^{\rm a}$ Yields were determined by $^{\rm 1}\text{H-NMR}.$ $^{\rm b}$ For 72 h. $^{\rm c}$ Isolated yields. $^{\rm d}$ H_2O was not added.

2.3. Aerobic Photooxidation under Low Concentration of Oxygen

^t Bu 1a (0.3 mmol)		2-CI-AQN (0.08 ec additive	fluorescent lamp (VIS) CI-AQN (0.08 equiv.) additive EtOAc (1 mL)		R	$R = CO_2H$ $= CO_3H$ $= CH_2OOH$ $= CHO$		2a 3a)H 6a 7a
entry		additive	O ₂	time		yield	d (%) ^a	
			(%)	(h)	2a	3a	6a	7a
1	TFA (0.3	equiv.) / H ₂ O (75 μL)) 10	24	93 (93)	0	0	0
2	TFA (0.3	equiv.) / H ₂ O (75 μL)) 5	24	87	5	0	trace
3	TFA (0.3	equiv.) / H ₂ O (75 μL)) 5	30	96 (100)	0	0	0
4	K ₂ C	O ₃ (0.05 equiv.)	10	12	83 (79)	2	7	10
5	K ₂ C	O ₃ (0.05 equiv.)	10	15	100 (88)	0	0	0
6	K ₂ C	O ₃ (0.05 equiv.)	10	18	(95)	0	0	0
7	K ₂ C	O ₃ (0.05 equiv.)	5	18	67	0	13	15
8	K ₂ C	O ₃ (0.05 equiv.)	5	24	99 (95)	0	0	0

Table S5. Aerobic Photooxidation under Low Concentration of O2

^a Yields were determined by ¹H-NMR. Numbers in parentheses were isolated yields.

2.4. Time Course of the Aerobic Photooxidation

Figure 1. Without Additive

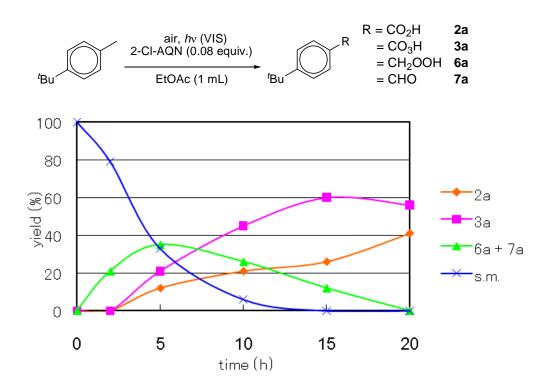


Figure 2. With TFA / H₂O

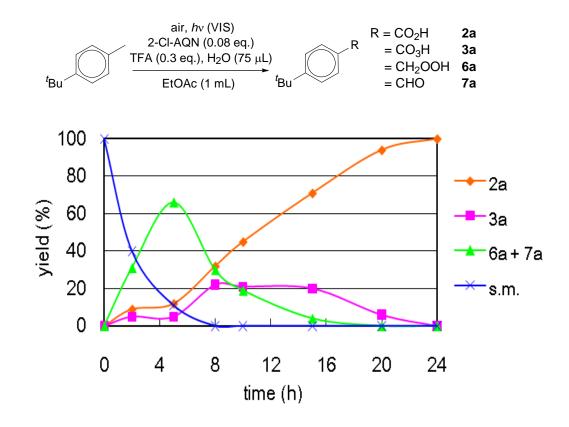
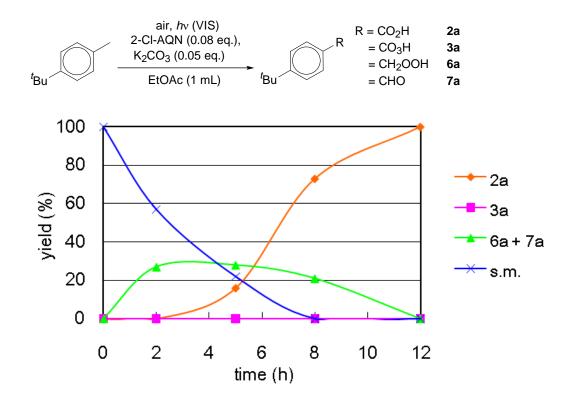


Figure 3. With K₂CO₃



2.5. Measurement of Peroxide by Iodometry

Typical procedure is as follows; 4-*tert*-butyltoluene (0.3 mmol) is oxidized with 2-Cl-AQN (0.08 equiv.), TFA (0.3 equiv.) and H₂O (75 μ L) in EtOAc (1 mL) under irradiation of VIS from fluorescent lamp (22w x 4) in an air atmosphere for 24 h, which usually gives 4-*tert*-butylbenzoic acid quantitatively. The residue is mixed with saturated aq. KI (3 mL), AcOH (1 mL) and *i*-PrOH (10 mL), and warmed at 100°C for 5 min. The result solution is titrated by 0.1 M aq. Na₂S₂O₃, and the volumes of required 0.1 M aq. Na₂S₂O₃ are 5.9 and 5.5 mL for two trials. These results correspond to 0.18 and 0.16 mmol of peroxides respectively after subtracting the required volume of 0.1 M aq. Na₂S₂O₃ for blank experiment.

2.6. General Procedure

Aerobic photooxidation: A solution of substrate (0.3 mmol), 2-Cl-AQN (0.08 equiv) and additive in dry EtOAc (1 mL) in a Pyrex test tube under an air atmosphere is stirred and irradiated externally with four 22W fluorescent lamps for indicated time. The product is extracted with 10% aq. NaHCO₃. Then the aq. solution is acidified with 2N aq. HCl and extracted with Et₂O. The product is purified by PTLC.

4-*tert***-Butylbenzoic acid (2a)**: colorless solid; ¹H NMR (500 MHz, CDCl₃) δ 8.06 (d, J = 8.7 Hz, 2H), 7.49 (d, J = 8.7 Hz, 2H), 1.35 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 172.6, 157.7, 130.2, 126.7, 125.6, 35.3, 31.2; MS m/z 178 (M⁺), 163, 135, 91; CAS Registry Number: 98-73-7

p-Toluic acid (2b): colorless solid; ¹H NMR (500 MHz, CDCl₃) δ 8.00 (d, *J* = 8.6 Hz, 2H), 7.27 (d, *J* = 8.6 Hz, 2H), 2.43 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 172.5, 144.7, 130.4, 129.3, 126.7, 21.9; MS m/z 136 (M⁺), 119, 91; CAS Registry Number: 99-94-5

m-Toluic acid (2c): colorless solid; ¹H NMR (400 MHz, CDCl₃) δ 7.94 (s, 1H), 7.93 (d, *J* = 8.2 Hz, 1H), 7.41 (d, *J* = 7.3 Hz, 1H), 7.38-7.34 (m, 1H), 2.42 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 172.6, 138.3, 134.6, 130.7, 129.2, 128.4, 127.4, 21.2; MS m/z 136 (M⁺), 119, 91; CAS Registry Number: 99-04-7

o-Toluic acid (2d): colorless solid; ¹H NMR (400 MHz, CDCl₃) δ 8.09-8.07 (m, 1H), 7.47-7.43 (m, 1H), 7.30-7.25 (m, 2H), 2.67 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.4, 141.4, 132.9, 131.9, 131.6, 128.3, 125.8, 22.1; MS m/z 136 (M⁺), 118, 91; CAS Registry Number: 118-90-1

p-Methoxybenzoic acid (2e): colorless solid; ¹H NMR (400 MHz, acetone- d_6) δ 7.96 (d, J = 9.2 Hz, 2H), 7.0 (d, J = 9.2 Hz, 2H), 3.85 (s, 3H); ¹³C NMR (100 MHz, acetone- d_6) δ 167.5, 164.4, 132.5, 123.7, 114.5, 55.8; MS m/z 152 (M⁺), 135, 107; CAS Registry Number: 100-09-4

p-Bromobenzoic acid (2f): colorless solid; ¹H NMR (500 MHz, acetone- d_6) δ 7.93 (d, J = 8.6 Hz, 2H), 7.67 (d, J = 8.6 Hz, 2H); ¹³C NMR (100 MHz, acetone- d_6) δ 166.9, 132.6, 132.3, 130.7, 128.1; MS m/z 200 (M⁺), 183, 155; CAS Registry Number: 586-76-5

p-Chlorobenzoic acid (2g): colorless solid; ¹H NMR (500 MHz, acetone- d_6) δ 8.01 (d, J = 8.6 Hz, 2H), 7.53 (d, J = 8.6 Hz, 2H); ¹³C NMR (100 MHz, acetone- d_6) δ 166.7, 139.5, 132.1, 130.3, 129.5; MS m/z 156 (M⁺), 139, 111; CAS Registry Number: 74-11-3

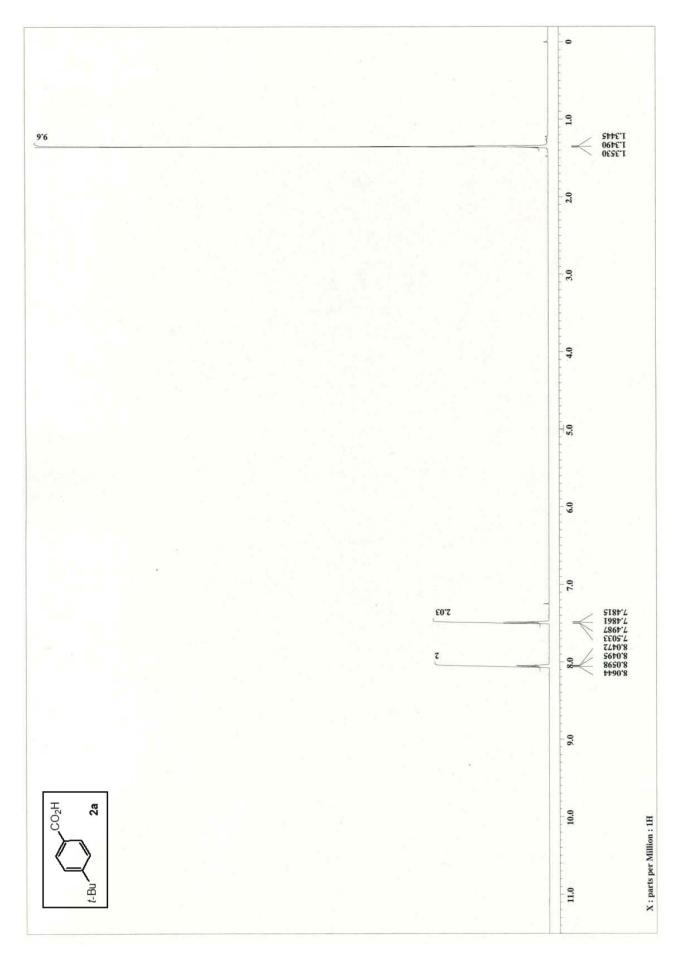
Benzoic acid (2h): colorless solid; ¹H NMR (500 MHz, CDCl₃) δ 8.13 (d, *J* = 7.4 Hz, 2H), 7.62 (t, *J* = 7.4 Hz, 1H), 7.48 (t, *J* = 7.4 Hz, 2H); ¹³C NMR (125 MHz, acetone-*d*₆) δ 172.6, 133.9, 130.3, 129.4, 128.6; MS m/z 122 (M⁺), 105, 77; CAS Registry Number: 65-85-0

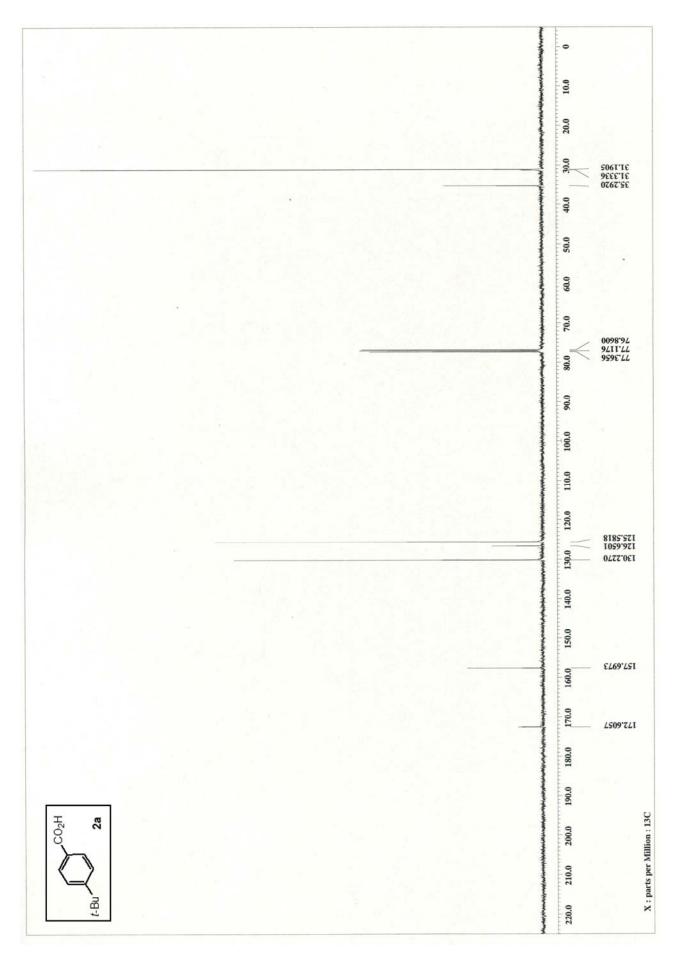
p-Phenylbenzoic acid (2i): colorless solid; ¹H NMR (500 MHz, acetone- d_6) δ 8.10 (d, J = 8.6 Hz, 2H), 7.78 (d, J = 8.6 Hz, 2H), 7.71 (d, J = 7.5 Hz, 2H), 7.48 (t, J = 7.5 Hz, 2H), 7.40 (t, J = 7.5 Hz, 1H); ¹³C SI-8

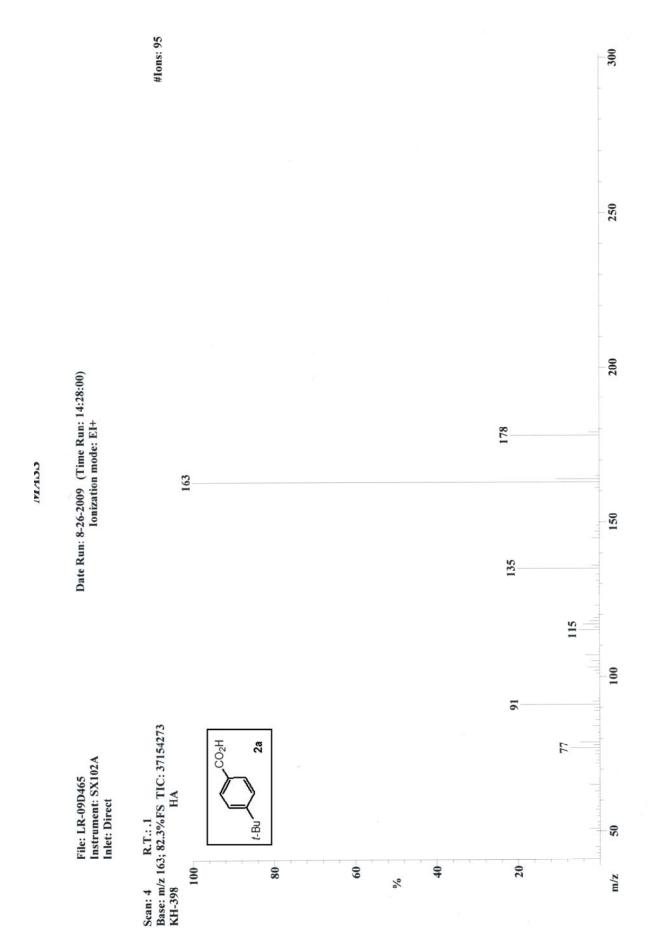
NMR (100 MHz, acetone-*d*₆) δ 167.5, 146.1, 140.7, 131.0, 130.4, 129.9, 129.0, 128.0, 127.7; MS m/z 198 (M⁺), 181, 152; CAS Registry Number: 92-92-2

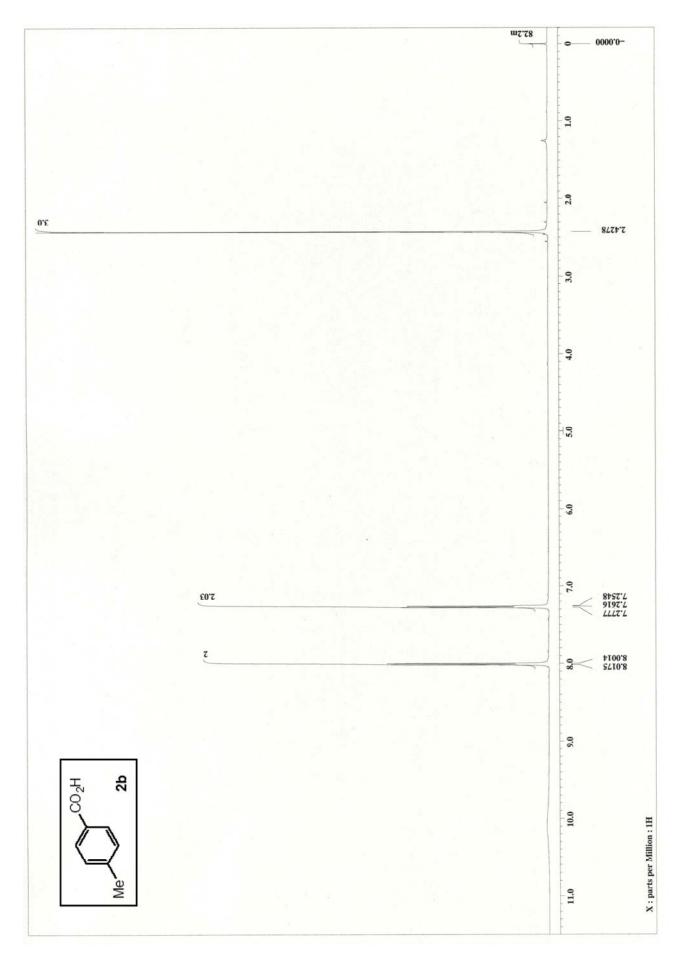
p-Benzoylbenzoic acid (2j): colorless solid; ¹H NMR (500 MHz, acetone- d_6) δ 8.19 (d, J = 8.6 Hz, 2H), 7.86 (d, J = 8.6 Hz, 2H), 7.79 (d, J = 7.4 Hz, 2H), 7.68 (t, J = 7.4 Hz, 1H), 7.56 (t, J = 7.4 Hz, 2H); ¹³C NMR (125 MHz, acetone- d_6) δ 195.3, 166.3, 141.4, 137.2, 133.8, 132.9, 129.9, 129.7, 128.6; MS m/z 226 (M⁺), 181, 149, 105; CAS Registry Number: 611-95-0

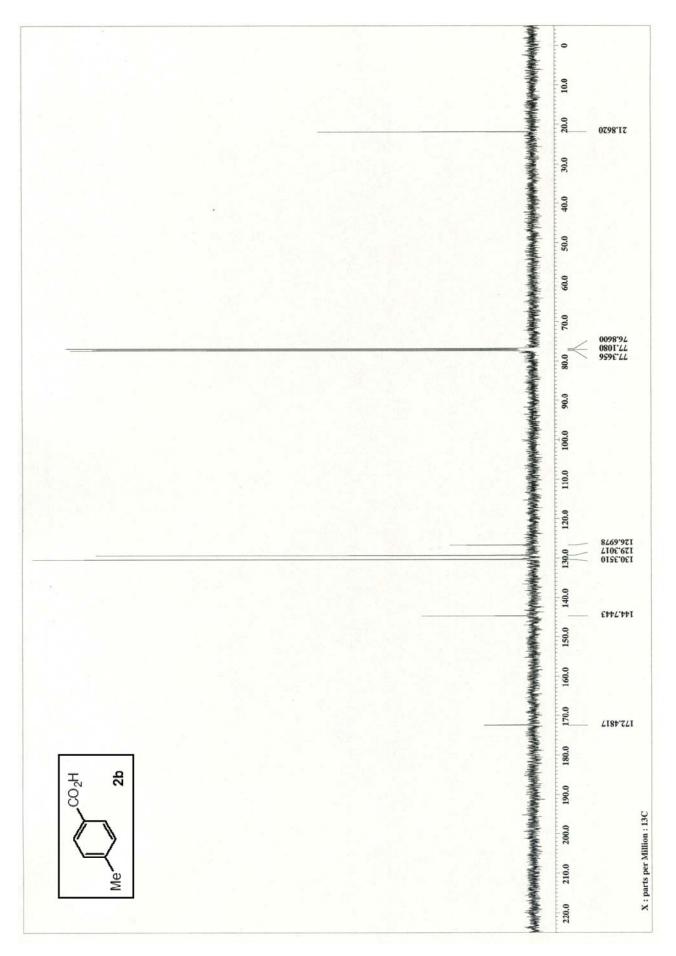
4,4'-Oxybis-benzoic acid (**2k**): white powder; ¹H NMR (500 MHz, DMSO- d_6) δ 7.96 (d, J = 9.2 Hz, 2H), 7.11 (d, J = 9.2 Hz, 2H); ¹³C NMR (125 MHz, DMSO- d_6) δ 167.2, 160.0, 132.3, 126.9, 119.2; MS m/z 258 (M⁺), 241; CAS Registry Number: 2215-89-6.



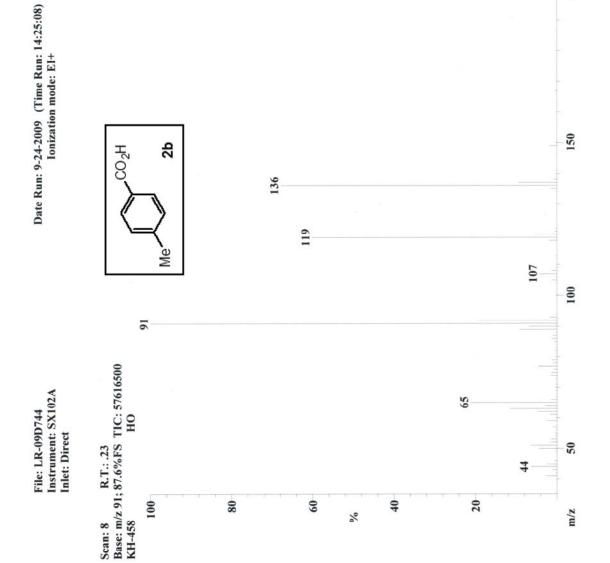












CCEM

SI-15

250

200

