

Entrainer-intensified vacuum reactive distillation process for the separation of 5-hydroxymethylfurfural from the dehydration of carbohydrates catalyzed by metal salt/ionic liquid

Zuojun Wei,^{*,a} Yingxin Liu,^b Dilantha Thushara^a and Qilong Ren^a

^a Institute of Pharmaceutical Engineering, Department of Chemical and Biological Engineering, Zhejiang University, Hangzhou 310027, P.R. China

^b College of Pharmaceutical Science, State Key Laboratory Breeding Base of Green Chemistry Synthesis Technology, Zhejiang University of Technology, Hangzhou 310032, P.R. China

* Corresponding author, E-mail address: weizuojun@zju.edu.cn, phone: 86-13588810769

Contents:

1. General information
2. ¹H NMR and ¹³C NMR spectra of the synthesized [OMIM]Cl
3. ¹H NMR and ¹³C NMR spectra of the synthesized [OMIM]Cl after the dehydration reaction
4. ¹H NMR and ¹³C NMR spectra of the obtained 5-HMF
5. HPLC chromatograms for the analysis of reaction bulks

1. General information

NMR spectra were recorded on a Rarian PLUS-400 spectrometer in CDCl_3 . ^1H and ^{13}C NMR chemical shifts (δ) are given in ppm relative to TMS. ^1H and ^{13}C positive chemical shifts (δ) in ppm are downfield from tetramethylsilane (CDCl_3 : $\delta_{\text{C}} = 77.000$ ppm; residual CHCl_3 in CDCl_3 : $\delta_{\text{H}} = 7.26$ ppm).

Carbohydrates were analyzed by HPLC (Detector: Waters 410 Differential Refractometer; Aminex® HPX-87H column 9 m, 300×7.8 mm), using $5 \text{ mmol}\cdot\text{l}^{-1}$ H_2SO_4 in ultrapure water as mobile phase at a flow rate of $0.6 \text{ ml}\cdot\text{min}^{-1}$ and a column temperature of $60 \text{ }^\circ\text{C}$. 5-HMF was analyzed by HPLC (UV Detector: Waters 2487; wavelength: 284 nm; Sunfire™ C18 column 5 m; 250×4.6 mm), using 70% methanol in ultrapure water as mobile phase at a flow rate of $1 \text{ ml}\cdot\text{min}^{-1}$ and a column temperature of $35 \text{ }^\circ\text{C}$.

2. ^1H NMR and ^{13}C NMR spectra of the synthesized [OMIM]Cl

The ^1H NMR (400 MHz, CDCl_3 , 29 °C) spectrum data for the synthesized IL [OMIM]Cl: $\delta=10.44$ (s, 1H_a), 7.77 (s, 1H_b), 7.54 (s, 1H_c), 4.32 (t, $J = 7.4$ Hz, 2H_d), 4.13 (s, 3H_e), 2.00–1.84 (m, 2H_f), 1.530-1.088 (dd, $J = 22.5, 7.4$ Hz, 10H_g), 0.87 (t, $J = 6.8$ Hz, 3H_h).

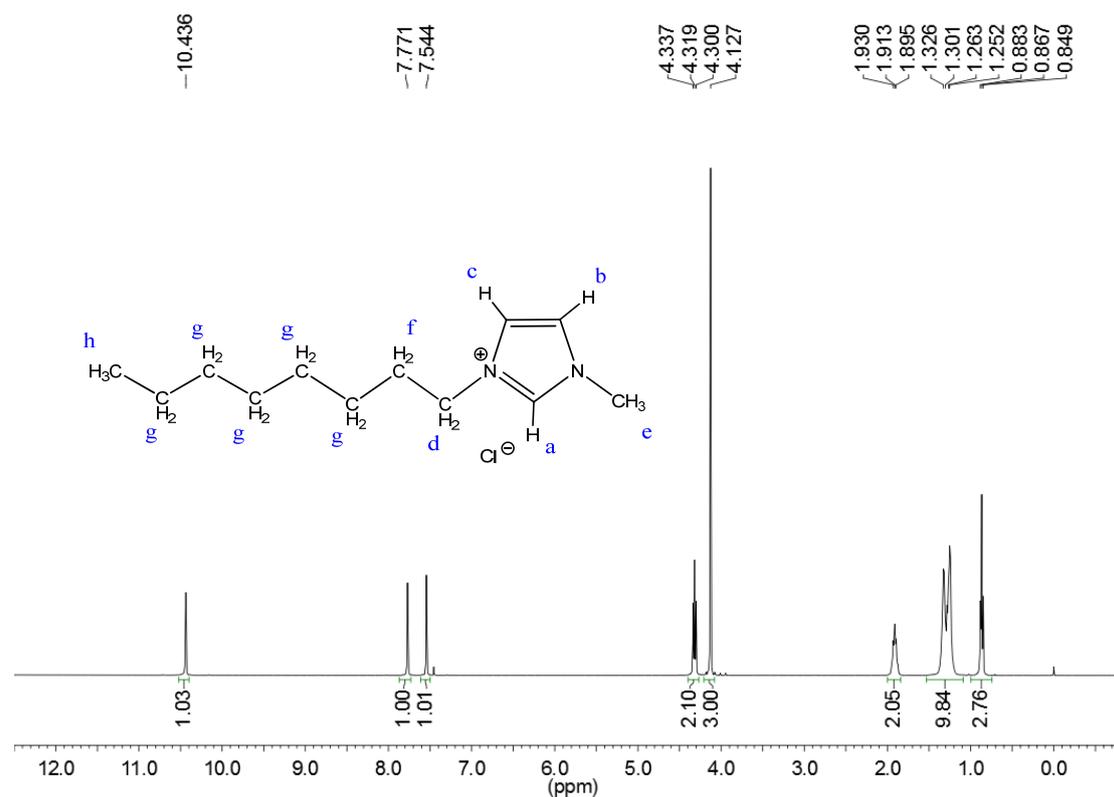


Figure 1s ^1H NMR spectrum of the synthesized [OMIM]Cl

The ^{13}C NMR (101 MHz, CDCl_3 , 29 °C) spectrum data for the synthesized IL [OMIM]Cl: $\delta_{\text{C}}=137.03$ (1C_{a}), 123.37 (1C_{b}), 121.50 (1C_{c}), 49.65 (1C_{d}), 36.21 (1C_{e}), 31.30 (1C_{f}), 29.99 (1C_{g}), 28.66 (1C_{h}), 28.60 (1C_{i}), 25.92 (1C_{j}), 22.22 (1C_{k}), 13.74 (1C_{l}).

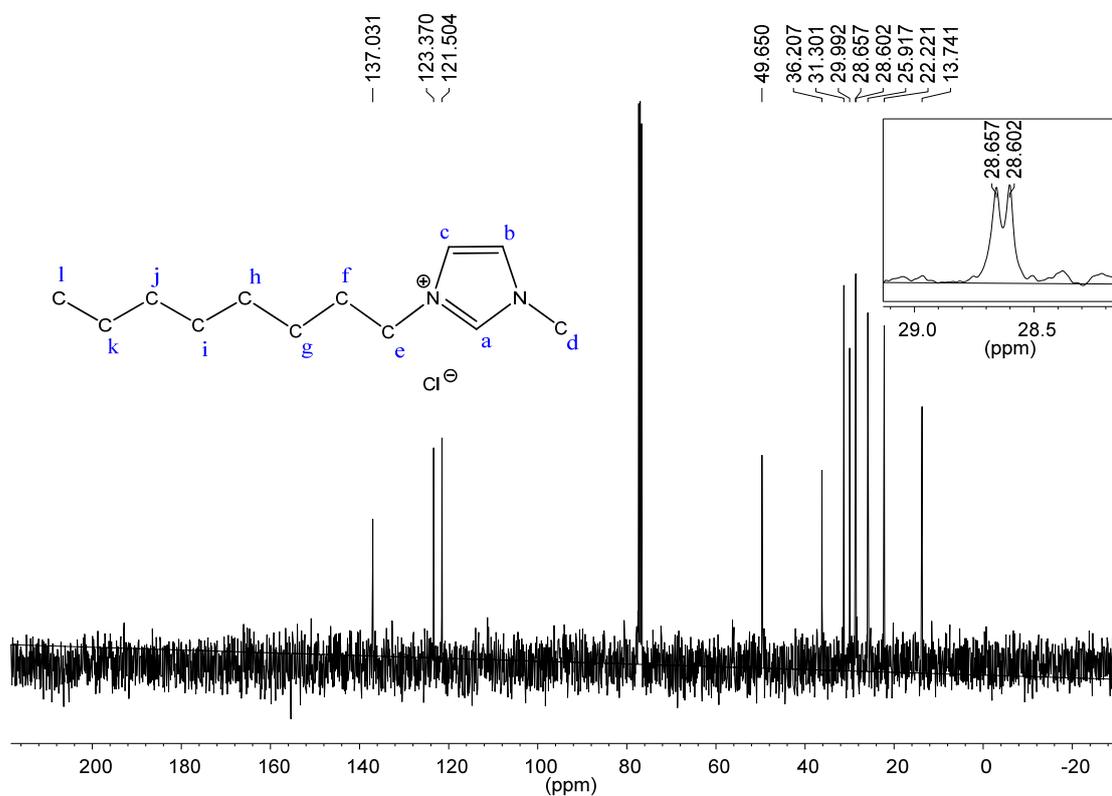


Figure 2s ^{13}C NMR spectrum of the synthesized [OMIM]Cl

3. ^1H NMR and ^{13}C NMR spectra of the synthesized [OMIM]Cl after the dehydration reaction

The ^1H NMR (400 MHz, CDCl_3 , 29 °C) spectrum data for the synthesized IL [OMIM]Cl after five times reused for the dehydration of fructose: $\delta=10.31$ (s, 1H_a), 7.71 (s, 1H_b), 7.49 (s, 1H_c), 4.31 (t, J = 7.4 Hz, 2H_d), 4.11 (s, 3H_e), 2.00–1.84 (m, 2H_f), 1.43–1.17 (dd, 10H_g), 0.87 (t, J = 6.8 Hz, 3H_h).

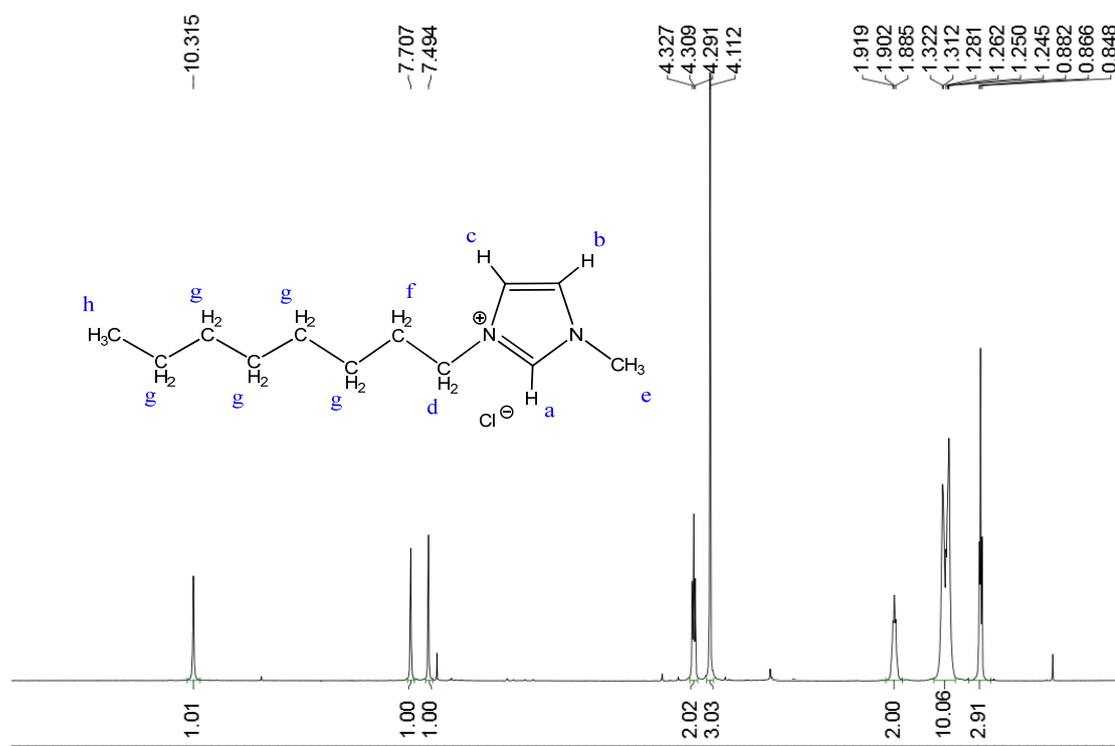


Figure 3s ^1H NMR spectrum of the synthesized [OMIM]Cl after five times reused for the dehydration of fructose

The ^{13}C NMR (101 MHz, CDCl_3 , 29 °C) spectrum data for the synthesized IL [OMIM]Cl after five times reused for the dehydration of fructose: δ_{C} =137.11 (1C_a), 123.43 (1C_b), 121.55 (1C_c), 49.78 (1C_d), 36.36 (1C_e), 31.44 (1C_f), 30.10 (1C_g), 28.79 (1C_h), 28.73 (1C_i), 26.05 (1C_j), 22.35 (1C_k), 13.87 (1C_l).

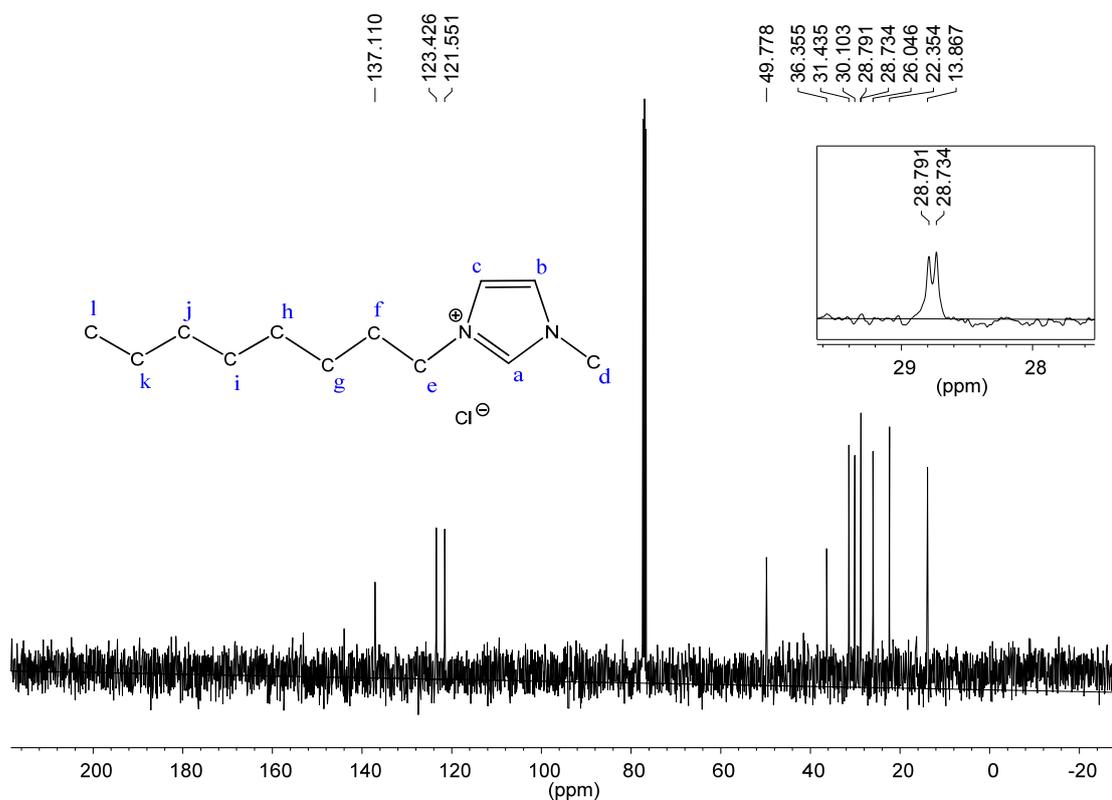


Figure 4s ^{13}C NMR spectrum of the synthesized [OMIM]Cl after five times reused for the dehydration of fructose

4. ^1H NMR and ^{13}C NMR spectra of the obtained 5-HMF

The ^1H NMR (400 MHz, CDCl_3 , 29 °C) spectrum data for obtained 5-HMF: $\delta=9.56$ (s, 1H_a), 7.21 (d, $J = 3.5$ Hz, 1H_b), 6.51 (d, $J = 3.5$ Hz, 1H_c), 4.71 (s, 2H_d), 2.73 (s, 1H_e).

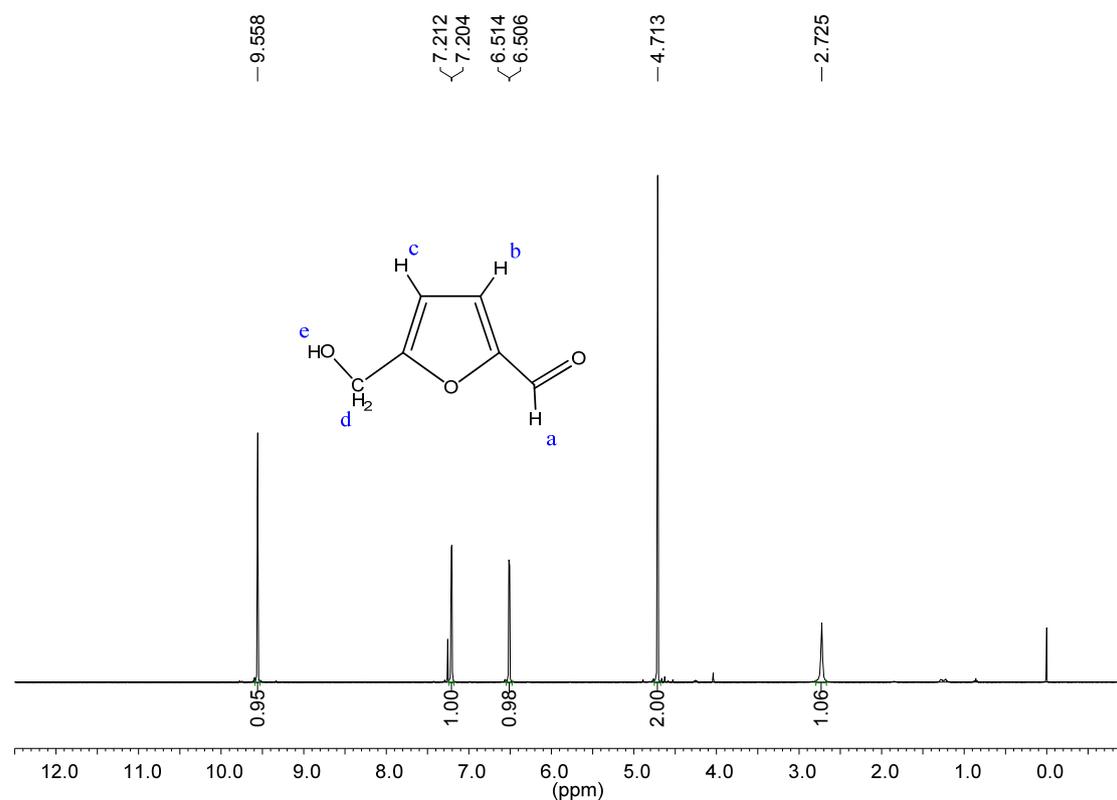


Figure 5s ^1H NMR spectrum of the obtained 5-HMF

The ^{13}C NMR (101 MHz, CDCl_3 , 29 °C) spectrum data for the obtained 5-HMF: $\delta_{\text{C}}=177.40$ (1C_a), 160.42 (1C_b), 152.12 (1C_c), 122.68 (1C_d), 109.90 (1C_e), 57.62 (1C_f).

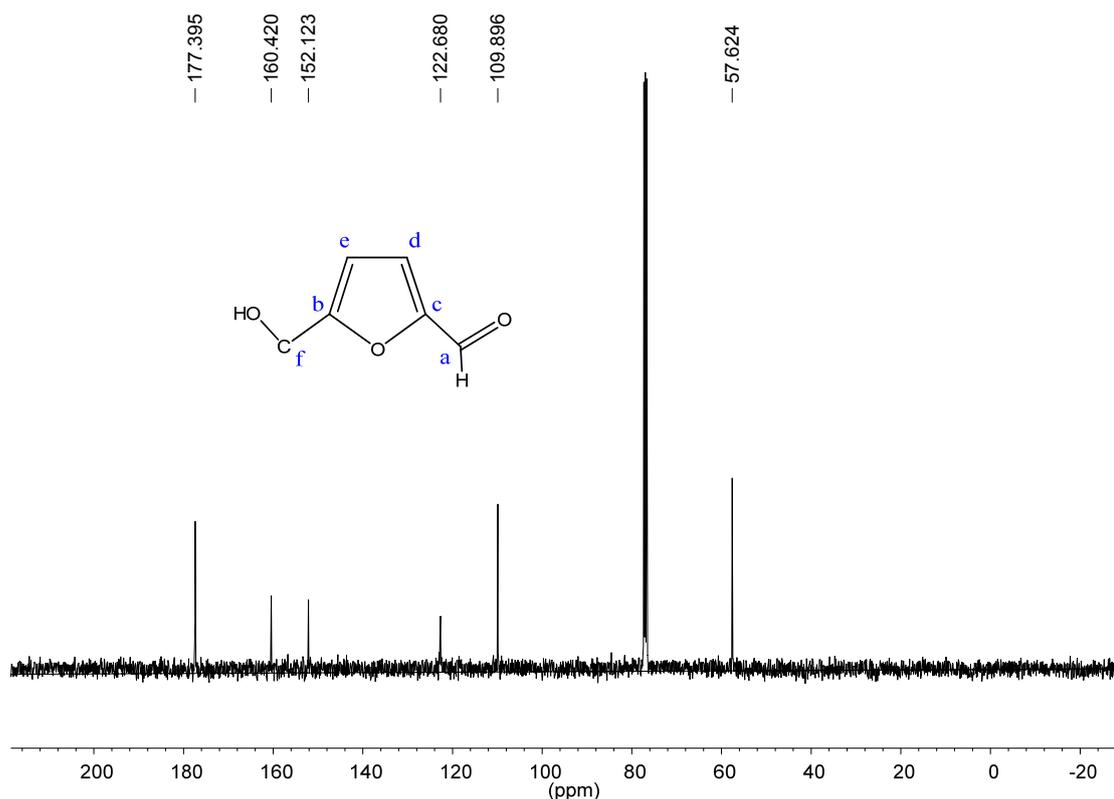


Figure 6s ^{13}C NMR spectrum of the obtained 5-HMF

5. HPLC chromatograms for the analysis of reaction bulks

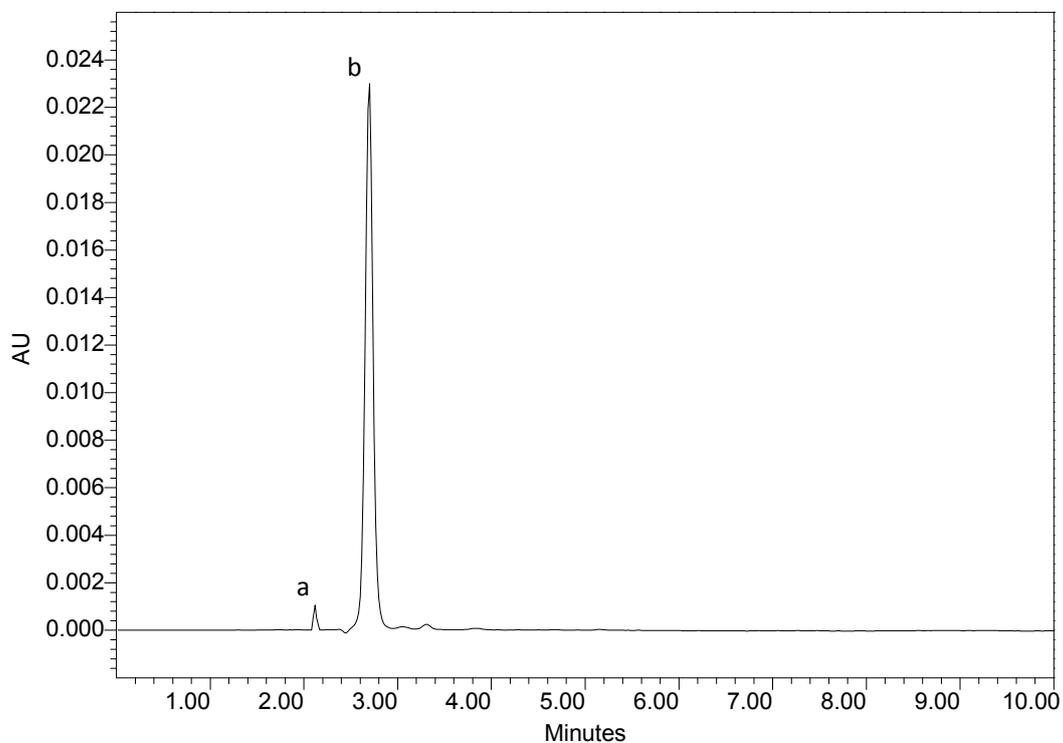


Figure 7s Typical HPLC chromatogram for the analysis of 5-HMF from reaction bulk (peak a belongs to [OMIM]Cl, with r.t. 2.05 min, peak b belongs to 5-HMF, with r.t 2.72 min)

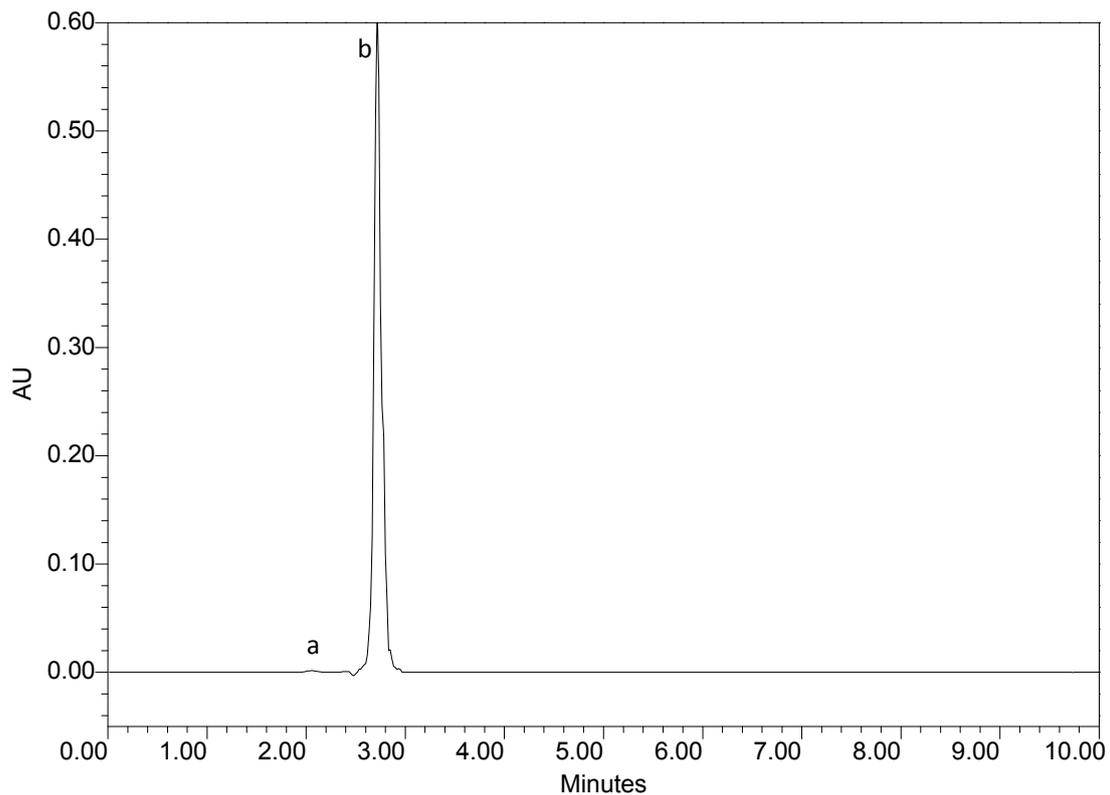


Figure 8s Typical HPLC chromatogram for the analysis of 5-HMF from recovery flask (peak a belongs to [OMIM]Cl, with r.t. 2.07 min, peak b belongs to 5-HMF, with r.t 2.73 min)

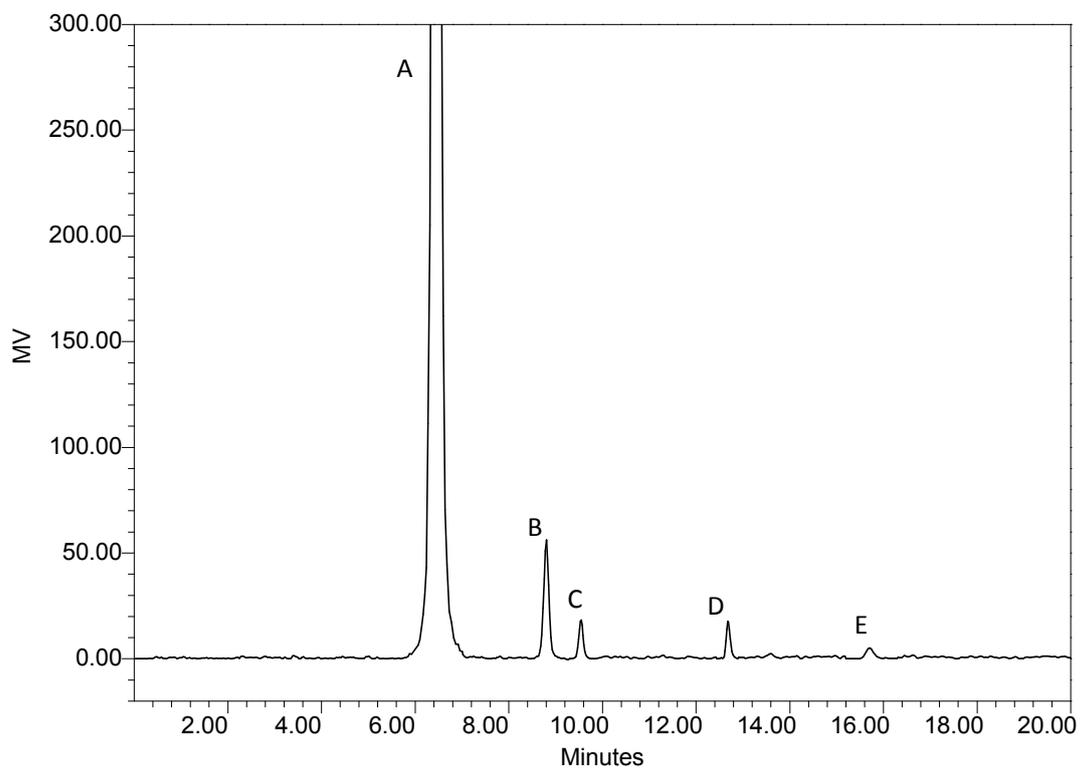


Figure 9s Typical HPLC chromatogram for the analysis of fructose and glucose (peak A belongs to [OMIM]Cl, with r.t. 6.45 min; peak B belongs to glucose, with r.t 8.80 min, peak C belongs to fructose, with r.t. 9.55 min; peak D and E are two unknown byproducts with r.t. 12.7 min and 15.7 min, respectively.)