Supporting Information

L-Proline supported on ionic liquid-modified magnetic nanoparticles as a highly efficient and reusable

organocatalyst for direct asymmetric aldol reaction in water

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Experimental

General Procedure for the Direct Asymmetric Aldol Reaction

The selected catalyst (10 mol%, based on L-proline content in the catalyst), ketone (2.5 mmol), aldehyde (0.25 mmol) and deionized water (2 mL) were added in a 10 mL round-bottom flask in turn. The mixture was allowed to react at room temperature for 12 h. The reaction was monitored constantly by TLC. After completion of the reaction, the magnetic catalyst was separated by a magnet near the bottle. The reaction solution was removed from the reaction vessel by decantation while the external magnet held the magnetic catalyst inside the bottle. The magnetic catalyst was then washed with ethyl acetate, separated by magnetic decantation as described above, dried under vacuum overnight at room temperature for the recycle experiment. Ethyl acetate $(3 \times 10 \text{ mL})$ was used to extract the aldols from the reaction solution. The combined organic layers were washed with brine and died with Na₂SO₄. After the evaporation of ethyl acetate, the residue was purified by column chromatography on silica gel (Acros, 40–60 μ m, 60 Å, eluent *n*-hexane/ethyl acetate = 3/1 (V/V)) to afford the desired aldol products. Enantiomeric excess of the corresponding aldol products was determined by HPLC analysis with a UV-vis detector using the Daicel chiralpak AD-H, OB-H or AD column. All the aldol products have been identified by comparison with those corresponding authentic samples in detailed HPLC analysis. NMR spectra for the aldol products **1-6** are in agreement with those reported in references.^[1] The syn and anti diastereomers of the aldols **1-6** were readily distinguished in ¹H NMR spectroscopy by the diagnostic chemical shifts of -CHOH- proton.

(2R,10S)-2-(Hydroxy-(2-nitrophenyl)methyl)cyclohexan-1-one (1).

Enantiomeric excess (85% ee) was determined by HPLC with a Chiralpak AD column (2-propanol/*n*-hexane =15: 85 (V/V) eluent) UV 254 nm, flow rate 1.0 mL/min, major enantiomer $t_R = 13.4$ min and minor enantiomer $t_R = 14.2$ min (see Fig. S1 and S2).



Fig. S1 HLPC of the authentic sample 1



Fig. S2 HLPC of the as-obtained product 1

The aldol product **1** has been identified by ¹H NMR spectrum, which is in agreement with the reported data.^[1a] The syn and anti diastereomers of the aldol **1** were distinguished in ¹H NMR spectroscopy by the diagnostic chemical shifts of -CHOH– proton according to the corresponding chemical shifts published in the reference.^[1b] (see Fig. S3)



(2R,10S)-2-(Hydroxy-(4-nitrophenyl)methyl)cyclohexan-1-one (2).

Enantiomeric excess (75% ee) was determined by HPLC with a Chiralpak AD column (2-propanol/*n*-hexane =15: 85 (V/V) eluent) UV 254 nm, flow rate 1.0 mL/min, major enantiomer $t_R = 16.2$ min and minor enantiomer $t_R = 21.4$ min (see Fig. S4 and S5).



Fig. S4 HLPC of the authentic sample 2





The aldol product **2** has been identified by ¹H NMR spectrum, which is in agreement with the reported data. ^[1a, 1c] The syn and anti diastereomers of the aldol **2** were distinguished in ¹H NMR spectroscopy by the diagnostic chemical shifts of -CHOH- proton according to the corresponding chemical shifts published in the reference.^[1b] (see Fig. S6)



Fig. S6 ¹H NMR of the as-obtained product 2 (inset: Chemical shifts of -CHOH- proton in

sample 2 (anti/syn) = 76: 24)

(2R,10S)-2-(Hydroxy-(2-chlorophenyl)methyl)cyclohexa -n-1-one (3).

Enantiomeric excess (89% ee) was determined by HPLC with a Chiralpak AD column (2-propanol/*n*-hexane =10: 90 (V/V) eluent) UV 220 nm, flow rate 1.0 mL/min, major enantiomer $t_R = 9.9$ min and minor enantiomer $t_R = 11.1$ min (see Fig. S7 and S8).



Fig. S7 HLPC of the authentic sample 3



Fig. S8 HLPC of the as-obtained product 3

The aldol product **3** has been identified by ¹H NMR spectrum, which is in agreement with the reported data. ^[1a, 1c] The syn and anti diastereomers of the aldol **3** were distinguished in ¹H NMR spectroscopy by the diagnostic chemical shifts of -CHOH– proton according to the corresponding chemical shifts published in the reference. ^[1a, 1c] (see Fig. S9)



(2R,10S)-2-(Hydroxy-(4-bromophenyl)methyl)cyclohexan-1-one (4).

Enantiomeric excess (89% ee) was determined by HPLC with a Chiralpak AD column (2-propanol/*n*-hexane =10: 90 (V/V) eluent) UV 220 nm, flow rate 1.0 mL/min, major enantiomer $t_R = 16.1$ min and minor enantiomer $t_R = 13.2$ min (see Fig. S10 and S11).



Fig. S10 HLPC of the authentic sample 4



Fig. S11 HLPC of the as-obtained product 4

The aldol product **4** has been identified by ¹H NMR spectrum, which is in agreement with the reported data. ^[1a, 1c] The syn and anti diastereomers of the aldol **4** were distinguished in ¹H NMR spectroscopy by the diagnostic chemical shifts of -CHOH– proton according to the corresponding chemical shifts published in the reference.^[1b] (see Fig. S12)



sample **4** (anti/syn) = 89: 11)

(2R,10S)-2-(Hydroxy-(2-naphthyl)methyl)cyclohexan-1-one (5).

The aldol product **5** has been identified by comparison of the HPLC retention times with reported values.^[1d] Enantiomeric excess (45% ee) was determined by HPLC with a Chiralpak AD column (2-propanol/*n*-hexane = 2: 98 (V/V) eluent) UV 220 nm, flow rate 1.0 mL/min, major enantiomer $t_R = 26.2$ min and minor enantiomer $t_R = 29.6$ min (see Fig. S13).



Fig. S13 HLPC of the as-obtained product 5

The aldol product **5** has been identified by ¹H NMR spectrum, which is in agreement with the reported data. ^[1c, 1d] The syn and anti diastereomers of the aldol **5** were distinguished in ¹H NMR spectroscopy by the diagnostic chemical shifts of -CHOH– proton according to the corresponding chemical shifts published in the reference. ^[1c] (see Fig. S14)



sample **5** (anti/syn) = 98: 2)

(2R,10S)-2-(Hydroxy-(4-nitrophenyl)methyl)cyclopentanone-1-one (6).

Enantiomeric excess (86% ee) was determined by HPLC with a Chiralpak AD column (2-propanol/*n*-hexane = 10: 90 (V/V) eluent) UV 220 nm, flow rate 1.0 mL/min, major enantiomer $t_R = 12.9$ min and minor enantiomer $t_R = 16.7$ min (see Fig. S15 and S16).



Fig. S15 HLPC of the authentic sample 6



Fig. S16 HLPC of the as-obtained sample 6

The aldol product **6** has been identified by ¹H NMR spectrum, which is in agreement with the reported data.^[1e] The syn and anti diastereomers of the aldol **6** were distinguished in ¹H NMR spectroscopy by the diagnostic chemical shifts of -CHOH– proton according to the corresponding chemical shifts published in the reference.^[1b] (see Fig. S17)



Fig. S17 ¹H NMR of the as-obtained product **6** (inset: Chemical shifts of –CHOH– proton in

sample **6** (anti/syn) =33: 67)

(4R)-Hydroxy-4-(2-nitrophenyl)-butan-2-one (7).

Enantiomeric excess (77% ee) was determined by HPLC with a Chiralpak OB-H column (2-propanol/*n*-hexane = 15: 85 (V/V) eluent) UV 254 nm, flow rate 1.0 mL/min, major enantiomer $t_R = 8.4$ min and minor enantiomer $t_R = 7.3$ min (see Fig. S18 and S19).



The aldol product 7 has been identified by 1 H NMR and 13 C NMR spectra (see Fig. S20 and S21).

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Fig. S21 ¹³C NMR of the as-obtained product 7

(4R)-Hydroxy-4-(4-nitrophenyl)-butan-2-one (8).

Enantiomeric excess (70% ee) was determined by HPLC with a Chiralpak OB-H column (2-propanol/*n*-hexane = 15: 85 (V/V) eluent) UV 254 nm, flow rate 1.0 mL/min, major enantiomer $t_R = 14.7$ min and minor enantiomer $t_R = 16.9$ min (see Fig. S22 and S23).



The aldol product **8** has been identified by 1 H NMR and 13 C NMR spectra (see Fig. S24 and S25).



Fig. S25 ¹³C NMR of the as-obtained product 8

(4R)-(2-Chlorophenyl)-4-hydroxy-2-butanone (9).

Enantiomeric excess (77% ee) was determined by HPLC with a Chiralpak AD column

(2-propanol/n-hexane = 7.5: 92.5 (V/V) eluent) UV 254 nm, flow rate 0.8 mL/min, major enantiomer $t_R = 10.9$ min and minor enantiomer $t_R = 12.3$ min (see Fig. S26 and S27).



Fig. S27 HLPC of the as-obtained sample 9

The aldol product **9** has been identified by ¹H NMR and ¹³C NMR spectra (see Fig. S28 and S29).



Fig. S28 ¹H NMR of the as-obtained product **9**



Fig. S29¹³C NMR of the as-obtained product **9**

(4R)-(4-Bromophenyl)-4-hydroxy-2-butanone (10).

Enantiomeric excess (77% ee) was determined by HPLC with a Chiralpak AD column (2-propanol/*n*-hexane = 7.5: 92.5 (V/V) eluent) UV 254 nm, flow rate 0.8 mL/min, major enantiomer $t_R = 15.2$ min and minor enantiomer $t_R = 16.1$ min (see Fig. S30 and S31).



Fig. S31 HLPC of the as-obtained sample 10

The aldol product **10** has been identified by 1 H NMR and 13 C NMR spectra (see Fig. S32 and S33).



Fig. S32 ¹H NMR of the as-obtained product **10**



Fig. S33 ¹³C NMR of the as-obtained product 10

(4R)-(4-Acetamidophenyl)-4-hydroxy-2-butanone (11).

Enantiomeric excess (83% ee) was determined by HPLC with a Chiralpak AD column

(2-propanol/*n*-hexane = 10: 90 (V/V) eluent) UV 254 nm, flow rate 0.8 mL/min, major enantiomer $t_R = 50.0$ min and minor enantiomer $t_R = 55.7$ min (see Fig. S34 and S35).





The aldol product **11** has been identified by ¹H NMR and ¹³C NMR spectra (see Fig. S36 and S37).



Fig. S36 ¹H NMR of the as-obtained product **11**



Fig. S37 ¹³C NMR of the as-obtained product **11**

(4R)-Hydroxy-4-(2-naphthyl)-2-butanone (12).

Enantiomeric excess (83% ee) was determined by HPLC with a Chiralpak AD column (2-propanol/*n*-hexane = 7.5: 92.5 (V/V) eluent) UV 254 nm, flow rate 0.8 mL/min, major enantiomer $t_R = 20.9$ min and minor enantiomer $t_R = 25.2$ min (see Fig. S38 and S39).



Fig. S39 HLPC of the as-obtained sample 12

The aldol product **12** has been identified by ¹H NMR and ¹³C NMR spectra (see Fig. S40 and S41).



Fig. S40 ¹H NMR of the as-obtained product **12**



Fig. S41 ¹³C NMR of the as-obtained product **12**

Characterization of Samples

TG-DTG curves



catalyst 2 (B) and pristine L -proline (C).

References

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