

Supplementary Information

**Non Catalytic Synthesis of Chromogen I and III from *N*-acetyl-D-glucosamine in
High-temperature Water**

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List of Supplementary Information:

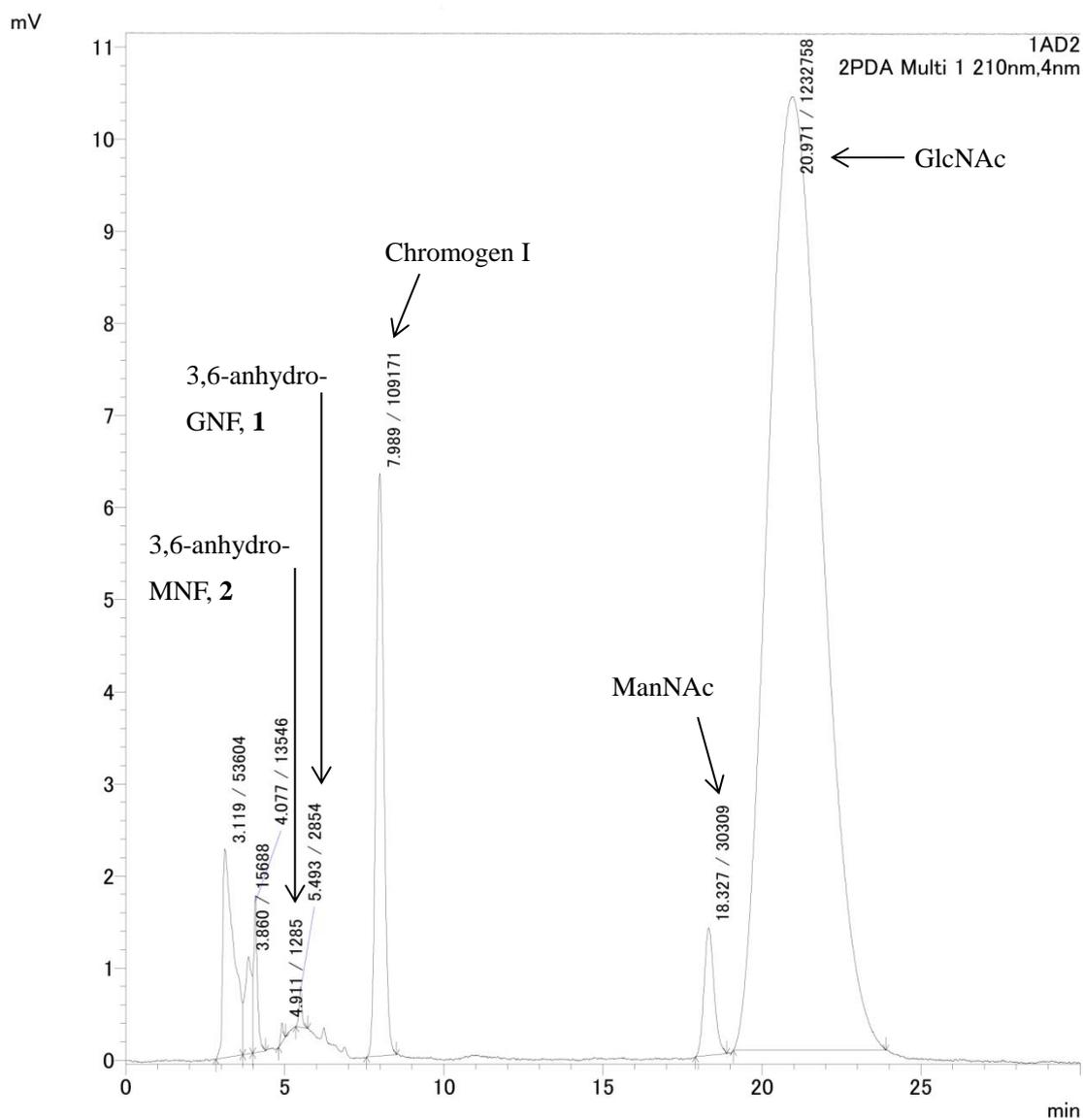
S1. HPLC chromatograph examples

S2. Characterization of Chromogen I, 3,6-anhydro-GNF, **1**, 3,6-anhydro-MNF, **2**, and

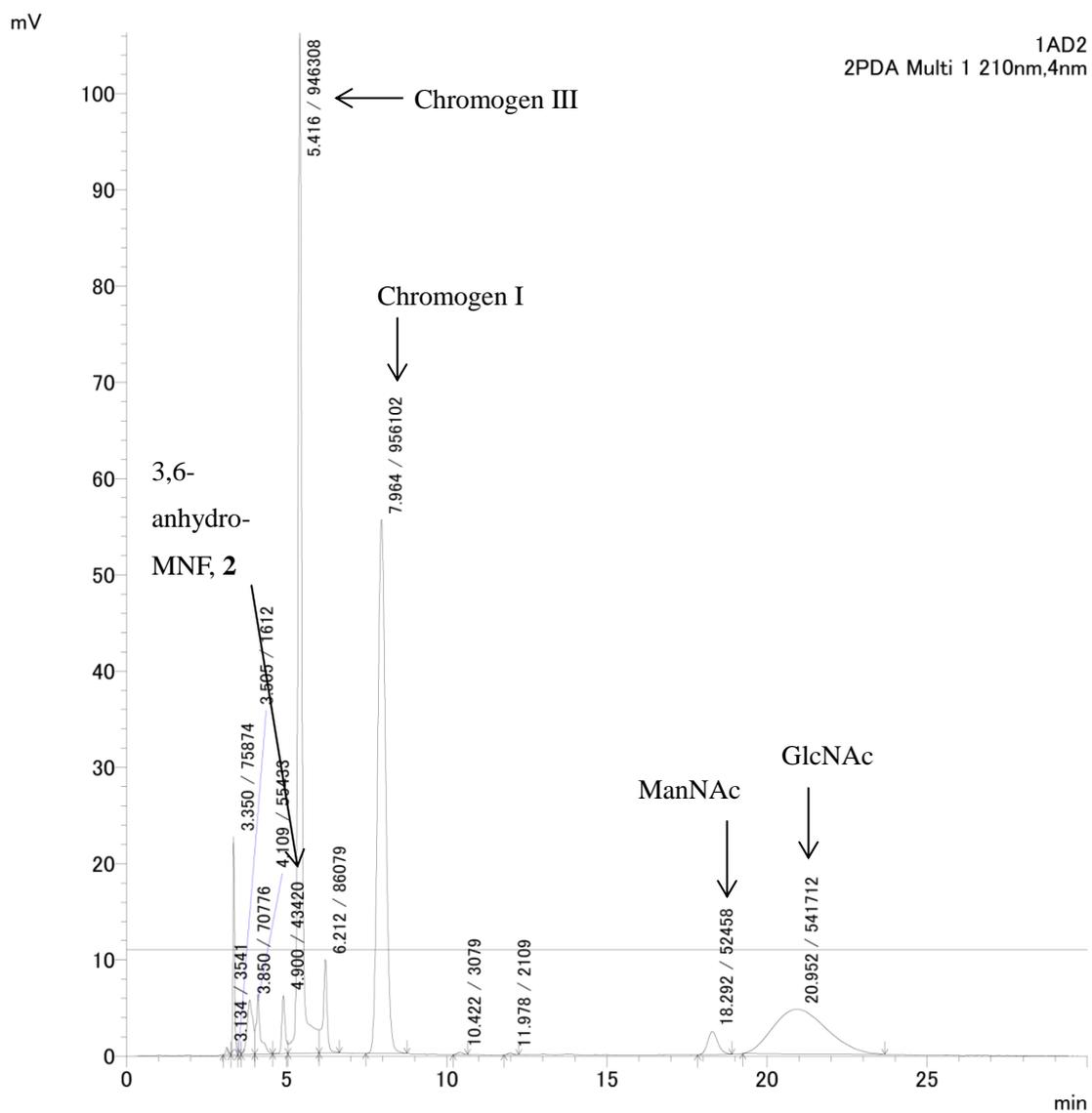
Chromogen III

S1. HPLC chromatograph examples

(a) 180°C, 25 MPa, reaction time 17 sec



(b) 180°C, 25 MPa, reaction time 33 sec



When Chromogen III was high concentration as the product, the peaks of Chromogen III and 3,6-anhydro-GMF, **1** overlapped. To analyze these compounds separately, we conducted the following HPLC method too. However, GlcNAc and ManNAc were not separated and Chromogen I showed two peaks in this method

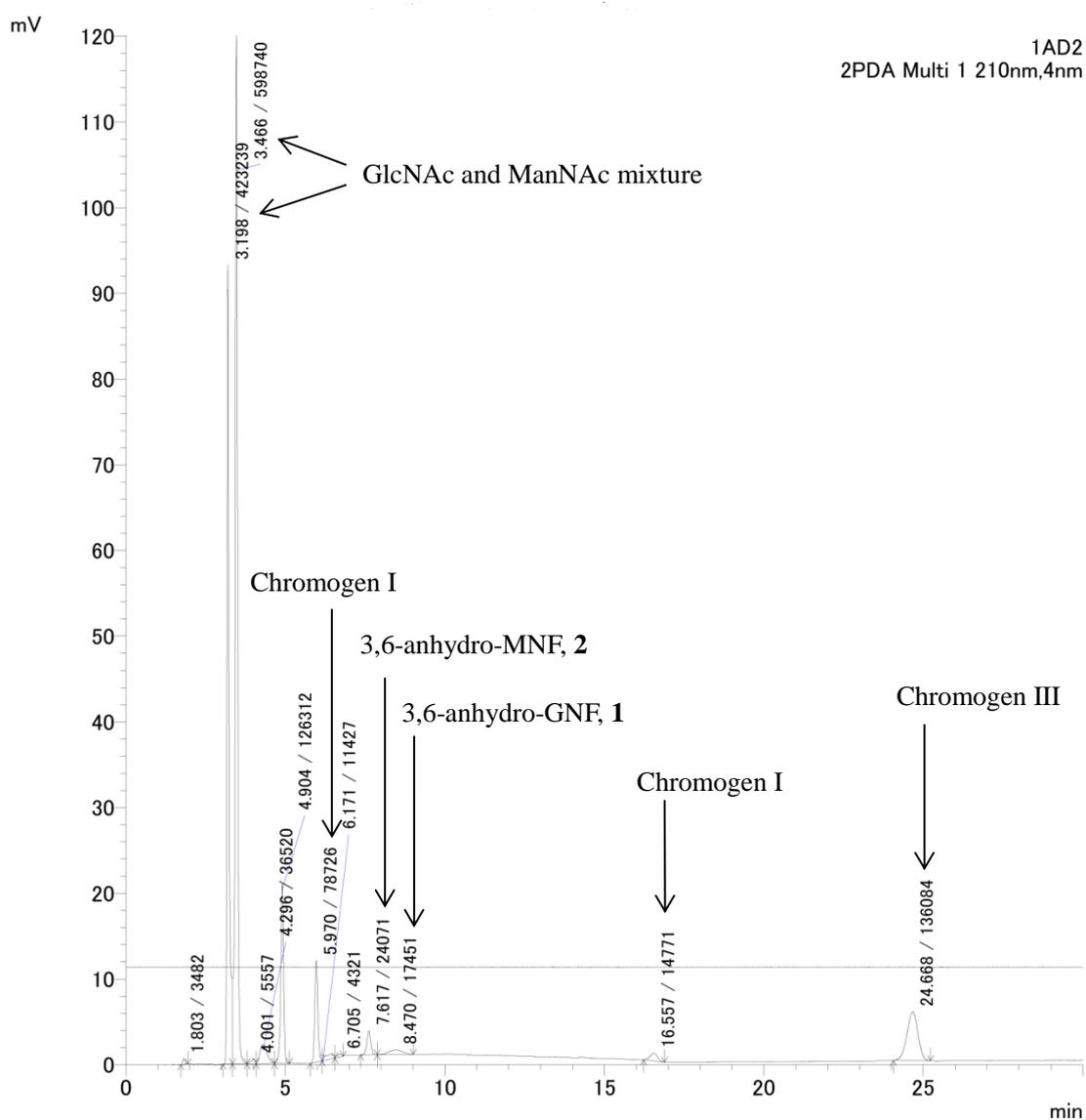
Column: Unison US-C18 (4.6 × 250mm, Imtakt)

Mobile phase: water

Flow rate: 1.0 mL min⁻¹

Column oven temperature: 40°C

(c) 180°C, 25 MPa, reaction time 19 sec



S2. Characterization of Chromogen I, 3,6-anhydro-GNF, **1**, 3,6-anhydro-MNF, **2**, and Chromogen III

Chromogen I ; HRESIMS: m/z 226.06893 $[M+Na]^+$ (calcd for $C_8H_{13}N_1Na_1O_5$, 226.06914).

1H NMR (D_2O , 500 MHz) α -anomer: δ 6.16 (1H, H-3), 6.04 (1H, H-1), 5.06 (1H, H-4),

3.83–3.57 (3H, H-5, H-6b, H-6a), 2.13 (s, 3H, CH_3CONH-); β -anomer: δ 6.21 (1H, H-3),

5.99 (1H, H-1), 4.83 (1H, H-4), 3.83–3.57 (3H, H-5, H-6b, H-6a), 2.13 (s, 3H, CH_3CONH-);

^{13}C NMR (D_2O , 125 MHz) α -anomer: δ 176.1 (CH_3CONH-), 137.0 (C-2), 112.0 (C-3),

102.2 (C-1), 87.5 (C-4), 76.2 (C-5), 65.2 (C-6), 25.4 (CH_3CONH-); β -anomer: δ 176.1

(CH_3CONH-), 136.5 (C-2), 112.7 (C-3), 102.0 (C-1), 87.2 (C-4), 76.5 (C-5), 65.1 (C-6), 25.4

(CH_3CONH-).

3,6-anhydro-GNF, **1** ; HRESIMS: m/z 226.07013 $[M+Na]^+$ (calcd for $C_8H_{13}N_1Na_1O_5$,

226.06914). 1H NMR (D_2O , 500 MHz) α -anomer: δ 5.62 (d, 1H, $J_{1,2} = 5.0$ Hz, H-1), 4.76 (t,

1H, $J_{3,4} = 5.0$, $J_{4,5} = 5.0$ Hz, H-4), 4.66 (t, 1H, $J_{2,3} = 5.0$, $J_{3,4} = 5.0$ Hz, H-3), 4.33–4.29 (1H,

H-5), 4.27 (t, 1H, $J_{1,2} = 5.0$, $J_{2,3} = 5.0$ Hz, H-2), 4.00 (dd, 1H, $J_{5,6b} = 6.5$, $J_{6a,6b} = 8.5$ Hz,

H-6b), 3.66 (t, 1H, $J_{5,6a} = 8.5$, $J_{6a,6b} = 8.5$ Hz, H-6a), 2.06 (s, 3H, CH_3CONH-); β -anomer: δ

5.44 (1H, H-1), 4.79 (t, 1H, $J_{3,4} = 5.0$, $J_{4,5} = 5.0$ Hz, H-4), 4.52 (d, 1H, $J_{3,4} = 5.0$ Hz, H-3),

4.33–4.29 (1H, H-5), 4.18 (1H, H-2), 3.96 (t, 1H, $J_{5,6b} = 8.0$, $J_{6a,6b} = 8.0$ Hz, H-6b), 3.88 (t,

1H, $J_{5,6a} = 8.0$, $J_{6a,6b} = 8.0$ Hz, H-6a), 2.02 (s, 3H, CH_3CONH-); ^{13}C NMR (D_2O , 125 MHz)

α -anomer: δ 177.1 (CH_3CONH-), 100.4 (C-1), 88.5 (C-3), 81.7 (C-4), 73.0 (C-5, C-6), 61.5

(C-2), 24.56 (CH₃CONH-); β-anomer: δ 176.8 (CH₃CONH-), 105.2 (C-1), 88.6 (C-3), 85.6 (C-4), 73.8 (C-6), 73.5 (C-5), 64.7 (C-2), 24.60 (CH₃CONH-).

3,6-anhydro-MNF, **2** ; HRESIMS: *m/z* 226.06938 [M+Na]⁺ (calcd for C₈H₁₃N₁Na₁O₅, 226.06914). ¹H NMR (D₂O, 500 MHz) α-anomer: δ 5.53 (d, 1H, *J*_{1,2} = 5.5 Hz, H-1), 4.70 (t, 1H, *J*_{3,4} = 5.5, *J*_{4,5} = 5.5 Hz, H-4), 4.65–4.62 (1H, H-3), 4.42–4.38 (1H, H-5), 4.35 (t, 1H, *J*_{1,2} = 5.5, *J*_{2,3} = 5.5 Hz, H-2), 3.96–3.89 (2H, H-6b, H-6a), 2.07 (s, 3H, CH₃CONH-); β-anomer: d 5.31 (d, 1H, *J*_{1,2} = 6.0 Hz, H-1), 4.81 (t, 1H, *J*_{3,4} = 4.6, *J*_{4,5} = 4.6 Hz, H-4), 4.65–4.62 (1H, H-3), 4.42–4.38 (1H, H-5), 4.25 (t, 1H, *J*_{1,2} = 6.0, *J*_{2,3} = 6.0 Hz, H-2), 4.02 (dd, 1H, *J*_{5,6b} = 6.7, *J*_{6a,6b} = 8.4 Hz, H-6b), 3.55 (t, 1H, *J*_{5,6a} = 8.4, *J*_{6a,6b} = 8.4 Hz, H-6a), 2.05 (s, 3H, CH₃CONH-); ¹³C NMR (D₂O, 125 MHz) α-anomer: δ 177.0 (CH₃CONH-), 98.3 (C-1), 84.7 (C-4), 83.0 (C-3), 73.95 (C-5), 73.5 (C-6), 57.5 (C-2), 24.4 (CH₃CONH-); β-anomer: δ 177.2 (CH₃CONH-), 103.7 (C-1), 83.5 (C-4), 82.7 (C-3), 74.3 (C-5), 73.90 (C-6), 61.8 (C-2), 24.5 (CH₃CONH-).

Chromogen III ; HRESIMS: *m/z* 393.12810 [2M+Na]⁺ (calcd for C₁₆H₂₂N₂Na₁O₈, 393.12739). ¹H NMR (D₂O, 270 MHz) δ 7.68 (1H, H-1), 6.25 (1H, H-3), 4.59 (1H, H-5), 3.69–3.67 (2H, H-6b, H-6a), 2.00 (s, 3H, CH₃CONH-).