

Supporting Information

Efficient Visible Light-Mediated Cross-Dehydrogenative Coupling Reactions of Tertiary Amines Catalyzed by a Polymer-Immobilized Iridium-Based Photocatalyst

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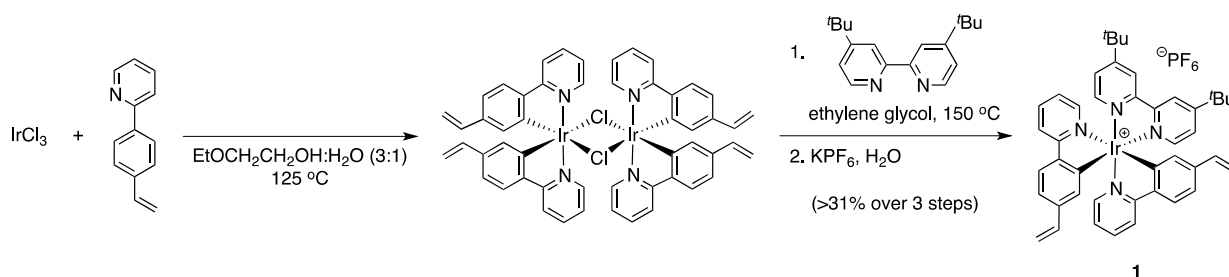
Contents

Part I:	Preparation of the Polymer-Supported Iridium Photocatalyst.....	S-2
Part II:	Substrate Scope for the CDC Reaction of <i>N</i> -Aryl Tetrahydroisoquinolines with P-H Nucleophiles	S-4
Part II:	NMR Spectra of 6a-n	S-10
Part III:	References	S-54

General Information: ^1H and ^{13}C NMR spectra were recorded on a JEOL ECX-400 in CDCl_3 . Chemical shifts were reported in parts per million (ppm) from tetramethylsilane using the solvent resonance as the internal standard (chloroform: δ 7.26 ppm) for ^1H NMR and (deuteriochloroform: δ 77.0 ppm) for ^{13}C NMR. ^{31}P NMR spectra were referenced to external H_3PO_4 (δ 0 ppm). ^{19}F NMR spectra were referenced to external benzotrifluoride (δ -63.7 ppm). IR spectra were measured on a JASCO FT/IR-610 spectrometer. High-resolution mass spectrometry was carried out using a JEOL JMS-T100TD (ESI). Preparative thin-layer chromatography (PTLC) was carried out using Wakogel B-5F from Wako Pure Chemical Industries, Ltd.

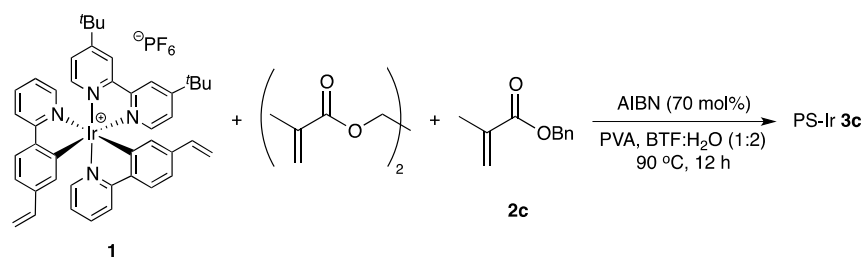
Reagents: Unless stated otherwise, commercial reagents were used as received. 2-(4-vinylphenyl)pyridine,¹ *N*-aryl tetrahydroisoquinolines **4a-e**,² and secondary phosphine oxides **5f-k**³ were prepared according to known literature procedures.

Part I: Preparation of the Polymer-Supported Iridium Photocatalyst



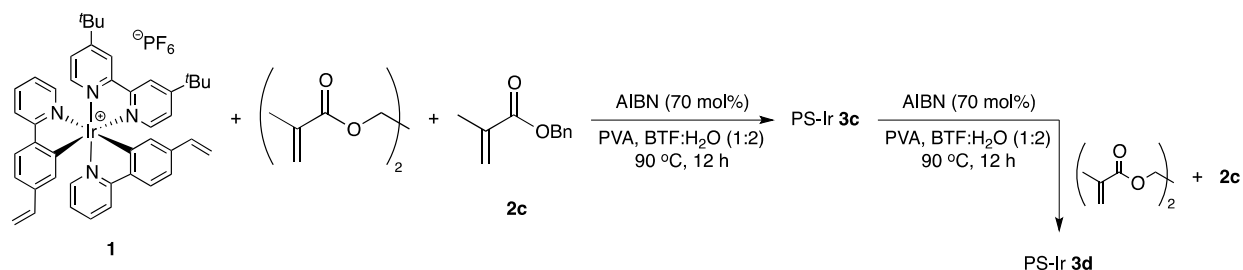
[Ir(vppy)₂Cl]₂. In a 50 mL round-bottom flask, IrCl_3 (0.2470 g, 0.8273 mmol) and 2-(4-vinylphenyl)pyridine (0.5738 g, 3.166 mmol) was dissolved in a solvent mixture of 2-ethoxyethanol (19 mL) and water (6.4 mL). The reaction vessel was fitted with a reflux condenser, flushed with argon gas, and placed in an oil bath set at 125 °C for 16 hours. The reaction mixture was allowed to cool to room temperature, filtered, and washed with H_2O , EtOH, and hexane. The yellow solid was allowed to dry under light vacuum and this crude product (403.6 mg, 0.3425 mmol, <41%) was used directly for the next procedure.

$[\text{Ir}(\text{vppy})_2(\text{dtbbpy})]\text{PF}_6$ (1**).** In a 30 mL round-bottom flask was added $[\text{Ir}(\text{vppy})_2\text{Cl}]_2$ (0.4036 g, 0.3425 mmol), 4,4'-di-*tert*-butyl-2,2'-bipyridine (0.2023 g, 0.7537 mmol), and ethylene glycol (17 mL). The reaction flask was fitted with a reflux condenser, flushed with argon gas, and heated to 150 °C for 16 hours. The reaction mixture was allowed to cool to room temperature and then was poured into a KPF_6 solution (1.261 g in 170 mL of water). The resulting precipitate was filtered and then was purified by column chromatography (basic Al_2O_3 , MeCN) to provide a bright yellow gel. The resulting gel was re-dissolved in a minimal amount of acetone, and then was precipitated in Et_2O to provide a bright yellow solid (0.2530 g, 0.2619 mmol, <76%). This crude solid was used directly for the suspension polymerization reaction.



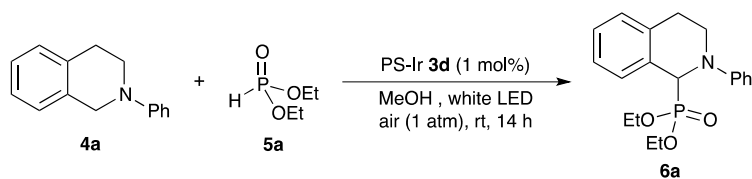
PS-Ir (3c). In a 10 mL test tube was added the crude Ir-based monomer **1** (0.0400 g, 0.0414 mmol), ethylene glycol dimethacrylate (24 μ L, 0.13 mmol), benzyl methacrylate (**2c**) (400 μ L, 2.35 mmol), azobisisobutyronitrile (AIBN) (0.0048 g, 0.0210 mmol), polyvinyl alcohol (0.0048 g), benzotrifluoride (BTF) (0.4 mL), and water (0.8 mL). The reaction vessel was degassed and was allowed to stir at 90 °C for 12 hours under an atmosphere of argon. The resulting solid was filtered and washed successively with water, MeOH, and DCM. The light orange cross-linked polymer was then lightly grounded with a mortar and pestle to give a bright yellow solid. The yellow polymer was washed with DCM and dried under high vacuum to provide **3c** (448.5 mg).

The iridium content of **3c** was determined using the following procedure: In a test tube containing 14.2 mg of **3c** was added conc. H_2SO_4 (1 mL) and then was heated to 200 °C for 1 hour in a heating block. Then conc. HNO_3 was added dropwise until the black solution turned into a light transparent yellow solution. After being heated for an additional 1 hour at 200 °C, the acidic solution was allowed to cool to room temperature. Then, 1 mL of aqua regia was added and then the solution was diluted to 50 mL with de-ionized using a volumetric flask. ICP analysis of the acid digested polymer **3c** was performed and the Ir content was determined using a calibration curve (2.85291 ppm Ir, 0.0523 mmol/g, expected Ir loading: 0.0851 mmol/g).



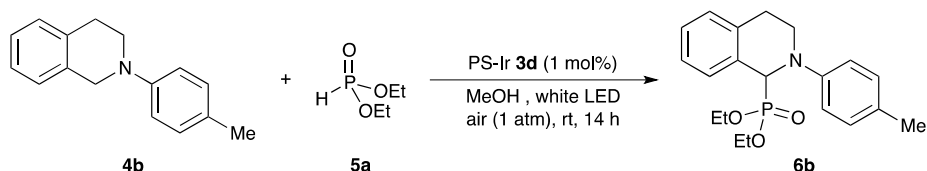
PS-Ir (3d). In a 50 mL round-bottom was added the crude Ir-based monomer **1** (0.3706 g, 0.3836 mmol), ethylene glycol dimethacrylate (235 μ L, 1.246 mmol), benzyl methacrylate (**2c**) (3.8 mL, 22 mmol), azobisisobutyronitrile (AIBN) (0.0441 g, 0.2686 mmol), polyvinyl alcohol (0.0441 g), benzotrifluoride (BTF) (3.8 mL), and water (7.6 mL). The reaction vessel was degassed and was allowed to stir at 90 °C for 12 hours under an atmosphere of argon. The resulting solid was filtered and washed successively with water, MeOH, and DCM. The light orange cross-linked polymer was then lightly grounded with a mortar and pestle to give a bright yellow solid. The yellow polymer was washed with DCM and dried under high vacuum at 90 °C to provide PS-Ir **3c** (4.381 g). The dried yellow solid was then transferred to a 50 mL round-bottom flask and the above procedure was repeated to provide the double-coated cross-linked polymer **3d** (8.4226 g, 0.0255 mmol/g, expected Ir loading: 0.0437 mmol/g).

Part II: Substrate Scope for the CDC Reaction of *N*-Aryl Tetrahydroisoquinolines with P-H Nucleophiles

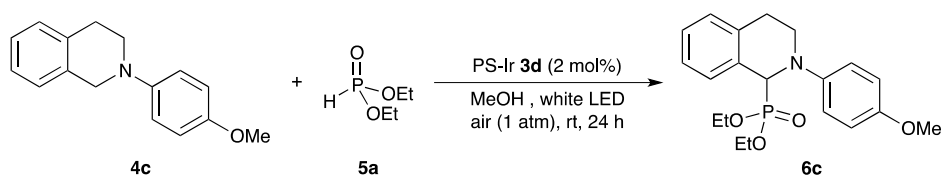


In a 2-necked test-tube was added PS-Ir **3d** (0.1996 g, 0.0255 mmol/g, 0.005 mmol, 1 mol%) and *N*-aryl tetrahydroisoquinoline **4a** (0.1046 g, 0.500 mmol). The reaction vessel was dried under high vacuum, and then re-pressurized with dry air. Then dialkyl phosphite **5a** (65 μ L, 0.5046 mmol) and MeOH (1.6 mL) was added and the reaction mixture was allowed to stir for 14 h under a white LED lamp (Toshiba E-CORE LDA7N/2). Next, the heterogeneous Ir catalyst **3d** was filtered and washed with MeOH. The filtrate was concentrated under reduced pressure, and the crude product was purified by preparative TLC (1:1 EtOAc:hexane) to afford the desired α -amino phosphonate **6a** (0.1499 g, 0.4340 mmol, 87%) as a white solid.

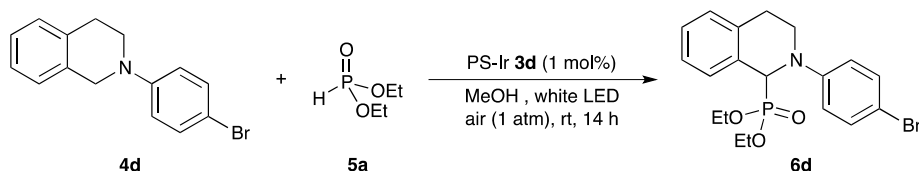
Diethyl (2-phenyl-1,2,3,4-tetrahydroisoquinolin-1-yl)phosphonate (6a) (Table 2, entry 1). ^1H NMR (CDCl_3 , 400 MHz) δ 7.38-7.37 (m, 1H), 7.26-7.13 (m, 5H), 6.97 (d, 2H, $J = 8.3$ Hz), 6.78 (t, 1H, $J = 7.4$ Hz), 5.19 (d, 1H, $J = 20.2$ Hz), 4.11-3.86 (m, 5H), 3.65-3.59 (m, 1H), 3.10-2.98 (m, 2H), 1.24 (t, 3H, $J = 6.9$ Hz), 1.13 (t, 3H, $J = 6.8$ Hz); ^{13}C NMR (CDCl_3 , 100 MHz) δ 149.3 (d, $J_{\text{C-P}} = 5.7$ Hz), 136.3 (d, $J_{\text{C-P}} = 5.7$ Hz), 130.6, 129.0, 128.6 (d, $J_{\text{C-P}} = 2.9$ Hz), 128.0 (d, $J_{\text{C-P}} = 4.8$ Hz), 127.3 (d, $J_{\text{C-P}} = 3.8$ Hz), 125.8 (d, $J_{\text{C-P}} = 2.9$ Hz), 118.4, 114.7, 63.2 (d, $J_{\text{C-P}} = 7.6$ Hz), 62.2 (d, $J_{\text{C-P}} = 7.6$ Hz), 58.7 (d, $J_{\text{C-P}} = 160.2$ Hz), 43.4, 26.7; ^{31}P NMR (CDCl_3 , 160 MHz) δ 23.3. This is a known compound and the spectral data are identical to those reported in the literature.⁴



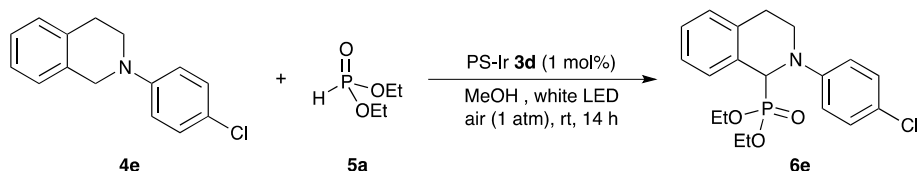
Diethyl (2-(*p*-tolyl)-1,2,3,4-tetrahydroisoquinolin-1-yl)phosphonate (6b) (Table 2, entry 2). Following the above general procedure with **4b** (111.7 mg, 0.500 mmol) and **5a** (65 μ L, 0.505 mmol) for 14 h. The crude reaction mixture was purified by preparative TLC (1:1 EtOAc:hexane) to provide **6b** (0.1736 g, 0.483 mmol, 97%) as a white solid. ^1H NMR (CDCl_3 , 400 MHz) δ 7.17-7.10 (m, 3H), 7.03 (d, 2H, $J = 8.3$ Hz), 6.87 (d, 2H, $J = 8.2$ Hz), 5.11 (d, 1H, $J = 20.6$ Hz), 4.16-3.86 (m, 5H), 3.60-3.56 (m, 1H), 2.98-2.97 (m, 2H), 2.24 (s, 3H), 1.24 (t, 3H, $J = 6.9$ Hz), 1.16 (t, 3H, $J = 6.9$ Hz); ^{13}C NMR (CDCl_3 , 100 MHz) δ 147.4, 136.4 (d, $J_{\text{C-P}} = 5.7$ Hz), 130.6, 129.6, 128.8 (d, $J_{\text{C-P}} = 1.9$ Hz), 128.1, 127.9, 127.3 (d, $J_{\text{C-P}} = 3.8$ Hz), 125.7 (d, $J_{\text{C-P}} = 2.9$ Hz), 115.3, 63.3 (d, $J_{\text{C-P}} = 6.7$ Hz), 62.2 (d, $J_{\text{C-P}} = 7.6$ Hz), 59.0 (d, $J_{\text{C-P}} = 157.3$ Hz), 43.8, 26.4, 20.2, 16.4 (d, $J_{\text{C-P}} = 5.7$ Hz), 16.3 (d, $J_{\text{C-P}} = 5.7$ Hz); ^{31}P NMR (CDCl_3 , 160 MHz) δ 22.9. This is a known compound and the spectral data are identical to those reported in the literature.⁵



Diethyl (2-(4-methoxyphenyl)-1,2,3,4-tetrahydroisoquinolin-1-yl)phosphonate (6c) (Table 2, entry 3). Following the above general procedure with **4c** (119.7 mg, 0.500 mmol), **5a** (65 μ L, 0.505 mmol), and heterogeneous catalyst **3d** (0.3992 g, 0.0255 mmol/g, 0.010 mmol, 2 mol%) for 24 h. The crude reaction mixture was purified by preparative TLC (1:1 EtOAc:hexane) to provide **6c** (0.1641 g, 0.437 mmol, 87%) as a white solid. ^1H NMR (CDCl_3 , 400 MHz) δ 7.38-7.36 (m, 1H), 7.17-7.10 (m, 3H), 6.90 (d, 2H, $J = 8.7$ Hz), 6.79 (d, 2H, $J = 8.7$ Hz), 5.00 (d, 1H, $J = 21.5$ Hz), 4.13-3.87 (m, 5H), 3.72 (s, 3H), 3.53-3.49 (m, 1H), 2.91-2.93 (m, 2H), 1.23 (t, 3H, $J = 7.3$ Hz), 1.14 (t, 3H, $J = 7.3$ Hz); ^{13}C NMR (CDCl_3 , 100 MHz) δ 153.1, 144.2, 136.4 (d, $J_{\text{C-P}} = 5.7$ Hz), 130.5, 128.9 (d, $J_{\text{C-P}} = 2.9$ Hz), 128.1 (d, $J_{\text{C-P}} = 4.8$ Hz), 127.2 (d, $J_{\text{C-P}} = 2.9$ Hz), 125.8 (d, $J_{\text{C-P}} = 2.9$ Hz), 117.6, 114.5, 63.3 (d, $J_{\text{C-P}} = 6.7$ Hz), 62.2 (d, $J_{\text{C-P}} = 7.6$ Hz), 59.4 (d, $J_{\text{C-P}} = 157.2$ Hz), 55.6, 44.6, 26.1, 16.4 (d, $J_{\text{C-P}} = 5.7$ Hz), 16.3 (d, $J_{\text{C-P}} = 5.7$ Hz); ^{31}P NMR (CDCl_3 , 160 MHz) δ 22.8. This is a known compound and the spectral data are identical to those reported in the literature.⁴

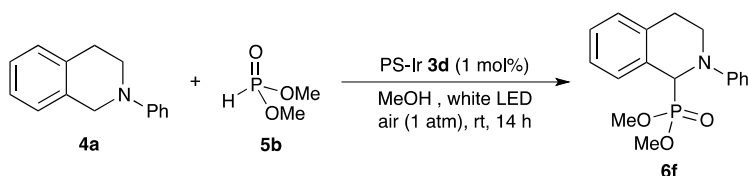


Diethyl (2-(4-bromophenyl)-1,2,3,4-tetrahydroisoquinolin-1-yl)phosphonate (6d) (Table 2, entry 4). Following the above general procedure with **4d** (144.1 mg, 0.500 mmol) and **5a** (65 μ L, 0.505 mmol) for 14 h. The crude reaction mixture was purified by preparative TLC (1:1 EtOAc:hexane) to provide **6d** (0.0827 g, 0.1949 mmol, 39%) as a white solid. ^1H NMR (CDCl_3 , 400 MHz) δ 7.34-7.29 (m, 3H), 7.21-7.13 (m, 3H), 6.84-6.81 (m, 2H), 5.08 (d, 1H, $J = 19.3$ Hz), 4.08-3.81 (m, 5H), 3.54-3.49 (m, 1H), 3.15-3.11 (m, 1H), 2.98-2.93 (m, 1H), 1.21 (t, 3H, $J = 7.3$ Hz), 1.19 (t, 3H, $J = 7.3$ Hz); ^{13}C NMR (CDCl_3 , 100 MHz) δ 148.3 (d, $J_{\text{C-P}} = 4.8$ Hz), 136.3, 131.7, 130.4, 128.6 (d, $J_{\text{C-P}} = 2.9$ Hz), 128.1 (d, $J_{\text{C-P}} = 4.8$ Hz), 127.6 (d, $J_{\text{C-P}} = 2.9$ Hz), 126.0 (d, $J_{\text{C-P}} = 2.9$ Hz), 116.1, 110.3, 63.2 (d, $J_{\text{C-P}} = 7.6$ Hz), 62.4 (d, $J_{\text{C-P}} = 7.6$ Hz), 58.7 (d, $J_{\text{C-P}} = 158.3$ Hz), 43.6, 26.9, 16.4 (d, $J_{\text{C-P}} = 5.7$ Hz), 16.3 (d, $J_{\text{C-P}} = 3.7$ Hz); ^{31}P NMR (CDCl_3 , 160 MHz) δ 22.4. This is a known compound and the spectral data are identical to those reported in the literature.⁶



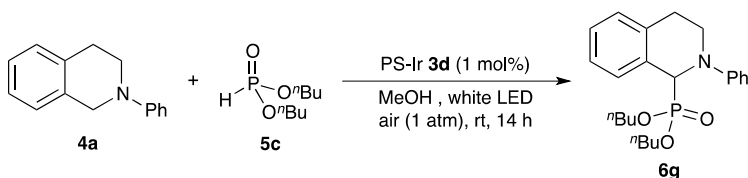
Diethyl (2-(4-chlorophenyl)-1,2,3,4-tetrahydroisoquinolin-1-yl)phosphonate (6e) (Table 2, entry 5). Following the above general procedure with **4e** (121.9 mg, 0.500 mmol) and **5a** (65 μ L, 0.505 mmol) for 14 h. The crude reaction mixture was purified by preparative TLC (1:1 EtOAc:hexane) to provide **6e** (0.0889 g, 0.2341 mmol, 47%) as a white solid. ^1H NMR (CDCl_3 , 400 MHz) δ 7.35-7.33 (m, 1H), 7.21-7.13 (m, 5H), 6.88-6.86 (m, 2H), 5.09 (d, 1H, $J = 19.2$ Hz),

4.10-3.81 (m, 5H), 3.55-3.49 (m, 1H), 3.14-3.10 (m, 1H), 2.98-2.93 (m, 1H), 1.21 (t, 3H, $J = 7.3$ Hz), 1.12 (t, 3H, $J = 6.9$ Hz); ^{13}C NMR (CDCl_3 , 100 MHz) δ 147.9 (d, $J_{\text{C-P}} = 4.8$ Hz), 136.2 (d, $J_{\text{C-P}} = 5.7$ Hz), 130.4, 128.6 (d, $J_{\text{C-P}} = 2.9$ Hz), 128.1 (d, $J_{\text{C-P}} = 4.8$ Hz), 127.6 (d, $J_{\text{C-P}} = 3.8$ Hz), 126.0 (d, $J_{\text{C-P}} = 2.9$ Hz), 123.1, 115.7, 63.2 (d, $J_{\text{C-P}} = 6.7$ Hz), 62.4 (d, $J_{\text{C-P}} = 7.6$ Hz), 58.8 (d, $J_{\text{C-P}} = 160.2$ Hz), 43.7, 26.9, 16.4 (d, $J_{\text{C-P}} = 4.8$ Hz), 16.3 (d, $J_{\text{C-P}} = 5.7$ Hz); ^{31}P NMR (CDCl_3 , 160 MHz) δ 22.5. This is a known compound and the spectral data are identical to those reported in the literature.⁵



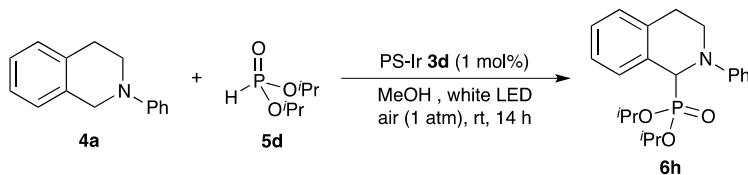
Dimethyl (2-phenyl-1,2,3,4-tetrahydroisoquinolin-1-yl)phosphonate (6f) (Table 2, entry 6).

Following the above general procedure with **4a** (104.6 mg, 0.500 mmol) and **5b** (46 μL , 0.502 mmol) for 14 h. The crude reaction mixture was purified by preparative TLC (1:1 EtOAc:hexane) to provide **6f** (0.1511 g, 0.4762 mmol, 95%) as a white solid. ^1H NMR (CDCl_3 , 400 MHz) δ 7.28-7.26 (m, 1H), 7.19-7.06 (m, 5H), 6.89 (d, 2H, $J = 8.7$ Hz), 6.72 (t, 1H, $J = 7.3$ Hz), 5.12 (d, 1H, $J = 19.7$ Hz), 3.96-3.90 (m, 1H), 3.58-3.54 (m, 7H), 3.02-2.87 (m, 2H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 149.2 (d, $J_{\text{C-P}} = 6.7$ Hz), 136.3 (d, $J_{\text{C-P}} = 4.8$ Hz), 130.4, 129.2, 128.8 (d, $J_{\text{C-P}} = 1.9$ Hz), 127.9 (d, $J_{\text{C-P}} = 4.8$ Hz), 127.5 (d, $J_{\text{C-P}} = 3.8$ Hz), 126.0 (d, $J_{\text{C-P}} = 2.9$ Hz), 118.6, 114.7, 58.7 (d, $J_{\text{C-P}} = 158.3$ Hz), 53.9 (d, $J_{\text{C-P}} = 1.9$ Hz), 53.8 (d, $J_{\text{C-P}} = 1.9$ Hz), 43.5, 26.6; ^{31}P NMR (CDCl_3 , 160 MHz) δ 25.0. This is a known compound and the spectral data are identical to those reported in the literature.⁴



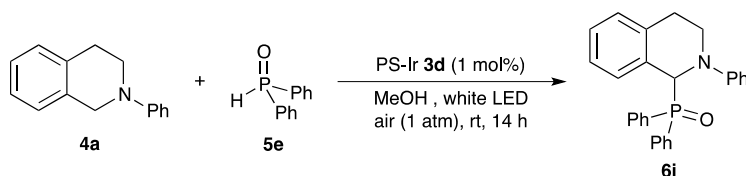
Dibutyl (2-phenyl-1,2,3,4-tetrahydroisoquinolin-1-yl)phosphonate (6g) (Table 2, entry 7).

Following the above general procedure with **4a** (104.6 mg, 0.500 mmol) and **5c** (98 μL , 0.502 mmol) for 14 h. The crude reaction mixture was purified by preparative TLC (1:1 EtOAc:hexane) to provide **6g** (0.1766 g, 0.4399 mmol, 88%) as a white solid. ^1H NMR (CDCl_3 , 400 MHz) δ 7.39-7.37 (m, 1H), 7.26-7.13 (m, 5H), 6.97 (d, 2H, $J = 8.2$ Hz), 6.78 (t, 1H, $J = 7.3$ Hz), 5.20 (d, 1H, $J = 19.7$ Hz), 4.06-3.76 (m, 5H), 3.66-3.60 (m, 1H), 3.15-2.97 (m, 2H), 1.60-1.51 (m, 2H), 1.49-1.42 (m, 2H), 1.38-1.20 (m, 4H), 0.88 (t, 3H, $J = 7.3$ Hz), 0.82 (t, 3H, $J = 7.3$ Hz); ^{13}C NMR (CDCl_3 , 100 MHz) δ 149.3, 136.4, 130.7, 129.0, 128.7 (d, $J_{\text{C-P}} = 1.9$ Hz), 128.1 (d, $J_{\text{C-P}} = 4.8$ Hz), 127.3 (d, $J_{\text{C-P}} = 2.9$ Hz), 125.8 (d, $J_{\text{C-P}} = 2.9$ Hz), 118.3, 114.7, 66.8 (d, $J_{\text{C-P}} = 7.6$ Hz), 65.9 (d, $J_{\text{C-P}} = 7.6$ Hz), 58.7 (d, $J_{\text{C-P}} = 157.3$ Hz), 43.4, 32.5 (d, $J_{\text{C-P}} = 5.7$ Hz), 32.4, 26.8, 18.6 (d, $J_{\text{C-P}} = 6.7$ Hz), 13.5 (d, $J_{\text{C-P}} = 4.8$ Hz); ^{31}P NMR (CDCl_3 , 160 MHz) δ 23.0. This is a known compound and the spectral data are identical to those reported in the literature.⁶

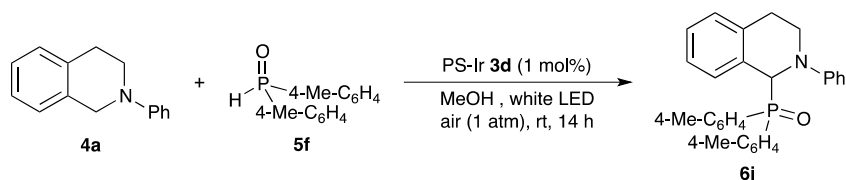


Diisopropyl (2-phenyl-1,2,3,4-tetrahydroisoquinolin-1-yl)phosphonate (6h) (Table 2, entry 8).

Following the above general procedure with **4a** (104.6 mg, 0.500 mmol) and **5d** (84 μ L, 0.504 mmol) for 14 h. The crude reaction mixture was purified by preparative TLC (1:1 EtOAc:hexane) to provide **6h** (0.1260 g, 0.3374 mmol, 67%) as a white solid. ^1H NMR (CDCl_3 , 400 MHz) δ 7.43-7.41 (m, 1H), 7.27-7.13 (m, 5H), 6.97 (d, 2H, $J = 8.2$ Hz), 6.78 (t, 1H, $J = 7.3$ Hz), 5.16 (d, 1H, $J = 21.1$ Hz), 4.68-4.60 (m, 2H), 4.10-4.03 (m, 1H), 3.76-3.64 (m, 1H), 3.07-2.96 (m, 2H), 1.31 (t, 6H, $J = 6.9$ Hz), 1.18 (d, 3H, $J = 6.0$ Hz), 0.97 (d, 3H, $J = 6.4$ Hz); ^{13}C NMR (CDCl_3 , 100 MHz) δ 149.5 (d, $J_{\text{C-P}} = 6.7$ Hz), 136.4 (d, $J_{\text{C-P}} = 5.7$ Hz), 130.9, 128.9, 128.6 (d, $J_{\text{C-P}} = 2.9$ Hz), 128.4 (d, $J_{\text{C-P}} = 4.8$ Hz), 127.2 (d, $J_{\text{C-P}} = 2.9$ Hz), 125.6 (d, $J_{\text{C-P}} = 2.9$ Hz), 118.2, 115.0, 72.1 (d, $J_{\text{C-P}} = 7.6$ Hz), 70.8 (d, $J_{\text{C-P}} = 8.6$ Hz), 58.7 (d, $J_{\text{C-P}} = 159.2$ Hz), 43.4, 26.5, 24.5 (d, $J_{\text{C-P}} = 2.9$ Hz), 24.1 (d, $J_{\text{C-P}} = 3.8$ Hz), 23.7 (d, $J_{\text{C-P}} = 4.8$ Hz), 23.3 (d, $J_{\text{C-P}} = 5.7$ Hz); ^{31}P NMR (CDCl_3 , 160 MHz) δ 21.5. This is a known compound and the spectral data are identical to those reported in the literature.⁶

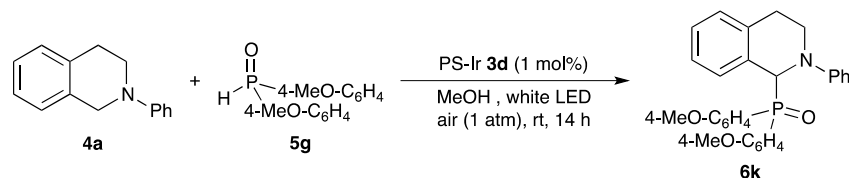
**Diphenyl (2-phenyl-1,2,3,4-tetrahydroisoquinolin-1-yl)phosphine oxide (6i) (Table 2, entry 9).**

Following the above general procedure with **4a** (104.6 mg, 0.500 mmol) and **5e** (101.1 mg, 0.500 mmol) for 14 h. The crude reaction mixture was purified by preparative TLC (1:49 MeOH: CH_2Cl_2) to provide **6i** (0.1874 g, 0.4577 mmol, 92%) as a white solid. ^1H NMR (CDCl_3 , 400 MHz) δ 7.83-7.67 (m, 4H), 7.53-7.29 (m, 6H), 7.14-7.05 (m, 4H), 6.92 (t, 1H, $J = 7.4$ Hz), 6.80-6.74 (m, 3H), 6.64 (d, 1H, $J = 7.8$ Hz), 5.56 (d, 1H, $J = 10.5$ Hz), 4.06-3.99 (m, 1H), 3.59-3.53 (m, 1H), 2.85-2.78 (m, 1H), 2.69-2.64 (m, 1H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 149.9, 136.8, 132.6, 132.1 (d, $J_{\text{C-P}} = 7.6$ Hz), 131.8 (d, $J_{\text{C-P}} = 2.9$ Hz), 131.7 (d, $J_{\text{C-P}} = 6.7$ Hz), 131.6 (d, $J_{\text{C-P}} = 4.8$ Hz), 130.8, 129.8, 129.1 (d, $J_{\text{C-P}} = 1.9$ Hz), 129.0, 128.4, 128.3 (d, $J_{\text{C-P}} = 5.7$ Hz), 128.1, 127.7 (d, $J_{\text{C-P}} = 2.9$ Hz), 127.3 (d, $J_{\text{C-P}} = 2.9$ Hz), 125.4 (d, $J_{\text{C-P}} = 2.9$ Hz), 119.5, 116.7, 61.9 (d, $J_{\text{C-P}} = 79.1$ Hz), 45.1, 25.5; ^{31}P NMR (CDCl_3 , 160 MHz) δ 31.4. This is a known compound and the spectral data are identical to those reported in the literature.⁵

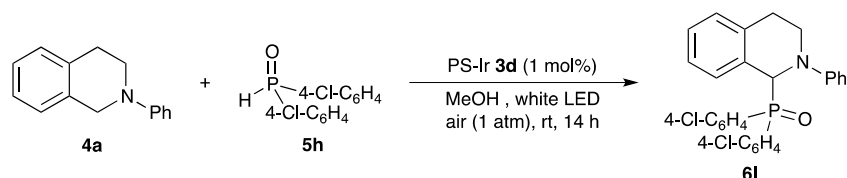
**(2-Phenyl-1,2,3,4-tetrahydroisoquinolin-1-yl)di-*p*-tolylphosphine oxide (6j) (Table 2, entry 10).**

Following the above general procedure with **4a** (104.6 mg, 0.500 mmol) and **5f** (115.1 mg, 0.500 mmol) for 14 h. The crude reaction mixture was purified by preparative TLC (1:49 MeOH: CH_2Cl_2) to provide **6j** (0.1860 g, 0.4251 mmol, 85%) as a white solid. ^1H NMR (CDCl_3 , 400 MHz) δ 7.59-7.43 (m, 4H), 7.16-7.14 (m, 2H), 7.06-7.00 (m, 6H), 6.73-6.61 (m, 4H), 5.44 (d, 1H, $J = 11.0$ Hz), 3.95-3.88 (m, 1H), 3.50-3.47 (m, 1H), 2.75-2.71 (m, 1H), 2.58-2.53 (m, 1H), 2.30 (s, 3H), 2.23 (s, 3H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 149.9 (d, $J_{\text{C-P}} = 8.6$ Hz), 142.2 (d, $J_{\text{C-P}} = 2.9$ Hz), 141.9 (d, $J_{\text{C-P}}$

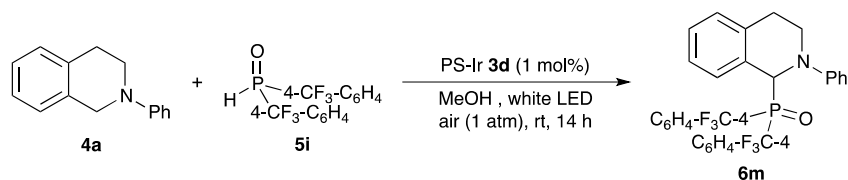
= 2.9 Hz); 136.7, 132.1 (d, J_{C-P} = 8.6 Hz), 131.6 (d, J_{C-P} = 9.5 Hz), 130.1, 129.6, 129.11, 129.08 (d, J_{C-P} = 1.9 Hz), 128.97 (d, J_{C-P} = 1.9 Hz), 128.8, 128.6 (d, J_{C-P} = 5.7 Hz), 127.8 (d, J_{C-P} = 2.9 Hz), 127.6, 127.2 (d, J_{C-P} = 2.9 Hz), 125.4 (d, J_{C-P} = 2.9 Hz), 119.2, 116.5, 61.9 (d, J_{C-P} = 79.1 Hz), 44.8, 25.5, 21.5 (d, J_{C-P} = 8.6 Hz); ^{31}P NMR (CDCl_3 , 160 MHz) δ 31.8. This is a known compound and the spectral data are identical to those reported in the literature.⁵



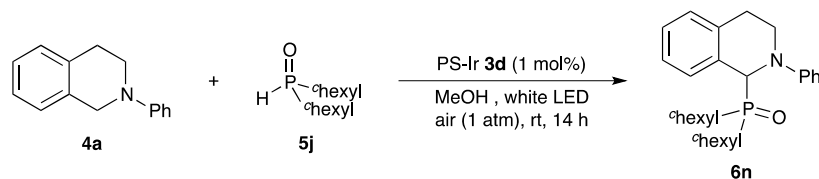
Bis(4-methoxyphenyl)(2-phenyl-1,2,3,4-tetrahydroisoquinolin-1-yl)phosphine oxide (6k) (Table 2, entry 11). Following the above general procedure with **4a** (104.6 mg, 0.500 mmol) and **5g** (131.1 mg, 0.500 mmol) for 14 h. The crude reaction mixture was purified by preparative TLC (1:49 MeOH: CH_2Cl_2) to provide **6k** (0.2092 g, 0.4456 mmol, 89%) as a white solid. ^1H NMR (CDCl_3 , 400 MHz) δ 7.70-7.65 (m, 2H), 7.58-7.53 (m, 2H), 7.14-7.10 (m, 3H), 7.05-7.03 (m, 1H), 6.98-6.91 (m, 3H), 6.83-6.72 (m, 6H), 5.48 (d, 1H, J = 11.5 Hz), 3.98-3.89 (m, 1H), 3.80 (s, 3H), 3.74 (s, 3H), 3.60-3.52 (m, 1H), 2.84-2.78 (m, 1H), 2.63-2.58 (m, 1H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 162.3 (d, J_{C-P} = 2.9 Hz), 162.1 (d, J_{C-P} = 2.9 Hz), 149.9 (d, J_{C-P} = 7.6 Hz), 136.7, 134.0 (d, J_{C-P} = 9.5 Hz), 133.4 (d, J_{C-P} = 9.5 Hz), 130.2, 129.0, 127.9 (d, J_{C-P} = 2.9 Hz), 127.2 (d, J_{C-P} = 2.9 Hz), 125.4 (d, J_{C-P} = 2.9 Hz), 124.1, 123.1, 122.6, 121.7, 119.2, 116.3, 113.9 (d, J_{C-P} = 11.4 Hz), 62.3 (d, J_{C-P} = 80.1 Hz), 55.2, 55.1, 44.6, 25.6; ^{31}P NMR (CDCl_3 , 160 MHz) δ 31.6; IR (KBr) cm^{-1} 3423 (m), 3023 (m), 2966 (m), 2840 (m), 1570 (s), 1502 (s), 1297 (s), 1255 (s), 1024 (s); ESI-HRMS (m/z) calcd. for $\text{C}_{29}\text{H}_{29}\text{N}_1\text{O}_3\text{P}_1$ [(M+H) $^+$]: 470.18850, found: 470.18622.



Bis(4-chlorophenyl)(2-phenyl-1,2,3,4-tetrahydroisoquinolin-1-yl)phosphine oxide (6l) (Table 2, entry 12). Following the above general procedure with **4a** (104.6 mg, 0.500 mmol) and **5h** (135.5 mg, 0.500 mmol) for 14 h. The crude reaction mixture was purified by preparative TLC (1:49 MeOH: CH_2Cl_2) to provide **6l** (0.2080 g, 0.4348 mmol, 87%) as a white solid. ^1H NMR (CDCl_3 , 400 MHz) δ 7.70 (t, 2H, J = 8.2 Hz), 7.57 (t, 2H, J = 8.2 Hz), 7.43-7.40 (m, 2H), 7.31-7.28 (m, 2H), 7.17-7.06 (m, 3H), 6.98 (t, 1H, J = 7.8 Hz), 6.82-6.71 (m, 4H), 5.50 (d, 1H, J = 11.0 Hz), 4.03-3.91 (m, 1H), 3.56-3.51 (m, 1H), 2.84-2.77 (m, 1H), 2.66-2.61 (m, 1H); ^{13}C NMR (CDCl_3 , 100 MHz) δ 149.8 (d, J_{C-P} = 8.6 Hz), 138.7 (d, J_{C-P} = 3.8 Hz), 138.4 (d, J_{C-P} = 3.8 Hz), 136.7, 133.4 (d, J_{C-P} = 9.5 Hz), 132.9 (d, J_{C-P} = 9.5 Hz), 130.9, 130.1, 129.9, 129.3 (d, J_{C-P} = 2.9 Hz), 129.2, 128.9, 128.7 (d, J_{C-P} = 9.5 Hz), 128.6, 127.6 (d, J_{C-P} = 2.9 Hz), 125.7 (d, J_{C-P} = 2.9 Hz), 120.1, 117.2, 61.9 (d, J_{C-P} = 80.1 Hz), 45.4, 25.4; ^{31}P NMR (CDCl_3 , 160 MHz) δ 30.5. This is a known compound and the spectral data are identical to those reported in the literature.⁵

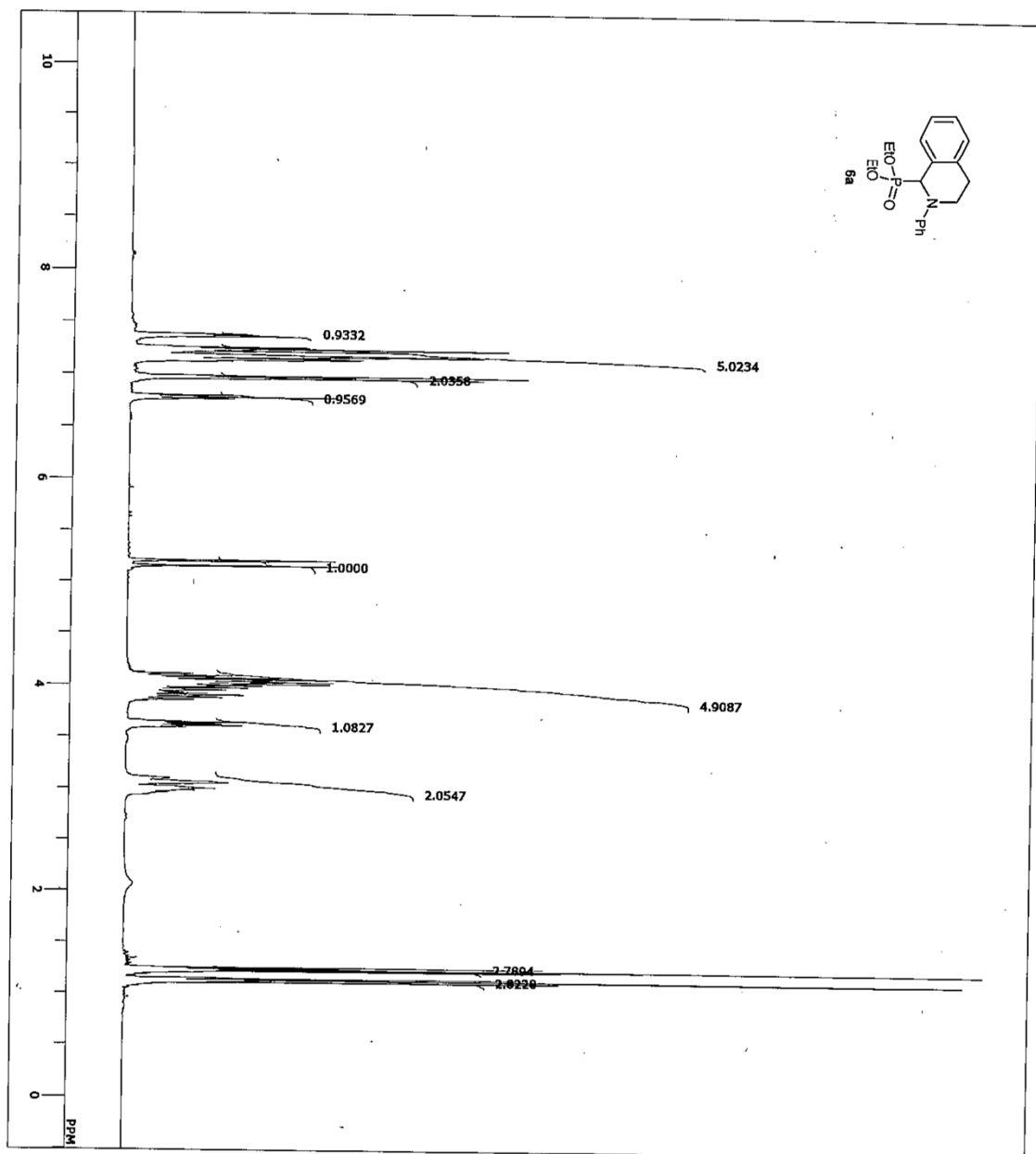


(2-Phenyl-1,2,3,4-tetrahydroisoquinolin-1-yl)bis(4-(trifluoromethyl)phenyl)phosphine oxide (6m) (Table 2, entry 13). Following the above general procedure with **4a** (104.6 mg, 0.500 mmol) and **5i** (169.1 mg, 0.500 mmol) for 14 h. The crude reaction mixture was purified by preparative TLC (1:49 MeOH:CH₂Cl₂) to provide **6m** (0.1658 g, 0.3040 mmol, 61%) as a white solid. ¹H NMR (CDCl₃, 400 MHz) δ 7.92 (t, 2H, *J* = 9.6 Hz), 7.81 (t, 2H, *J* = 9.2 Hz), 7.71 (d, 2H, *J* = 6.4 Hz), 7.59 (d, 2H, *J* = 7.8 Hz), 7.20-7.09 (m, 4H), 6.98 (t, 1H, *J* = 7.8 Hz), 6.84-6.77 (m, 3H), 6.64 (d, 1H, *J* = 7.8 Hz), 5.59 (d, 1H, *J* = 10.1 Hz), 4.05-3.95 (m, 1H), 3.58-3.53 (m, 1H), 2.85-2.77 (m, 1H), 2.71-2.66 (m, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ 149.8, 149.7, 136.88, 136.84, 136.5, 136.2, 135.6, 135.3, 134.16, 134.14, 133.85, 133.83, 133.53, 133.50, 132.5, 132.4, 132.1, 132.0, 129.58, 129.56, 129.3, 128.7, 127.90, 127.88, 127.42, 127.38, 125.81, 125.79, 125.52, 125.48, 125.45, 125.41, 125.38, 125.30, 125.26, 125.23, 125.19, 125.15, 125.11, 125.07, 125.04, 124.8, 122.1, 120.7, 117.7, 62.1, 61.3, 45.9, 25.4; ³¹P NMR (CDCl₃, 160 MHz) δ 29.5; ¹⁹F NMR (CDCl₃, 376 MHz) δ -64.2, -64.3; IR (KBr) cm⁻¹ 3434 (m), 3061 (m), 2940 (m), 1598 (m), 1506 (m), 1327 (s), 1172 (s), 1065 (s), 1018 (m); ESI-HRMS (*m/z*) calcd. for C₂₉H₂₃N₁O₁F₆P₁ [(M+H)⁺]: 546.14214, found: 546.14106.

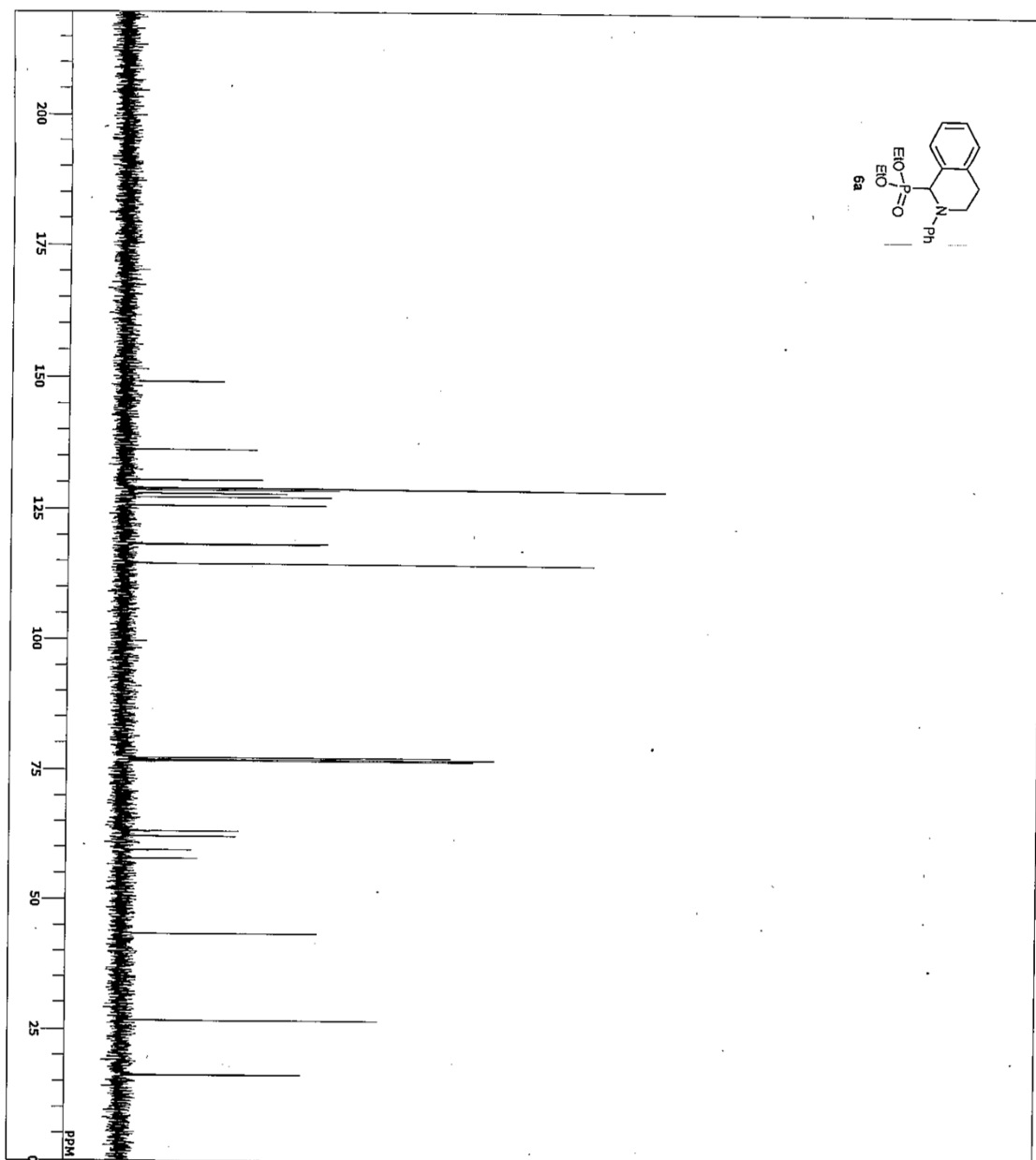


Dicyclohexyl(2-phenyl-1,2,3,4-tetrahydroisoquinolin-1-yl)phosphine oxide (6n) (Table 2, entry 14). Following the above general procedure with **4a** (104.6 mg, 0.500 mmol) and **5j** (107.1 mg, 0.500 mmol) for 14 h. The crude reaction mixture was purified by preparative TLC (1:49 MeOH:CH₂Cl₂) to provide **6n** (0.1616 g, 0.3833 mmol, 77%) as a white solid. ¹H NMR (CDCl₃, 400 MHz) δ 7.41-7.40 (m, 1H), 7.20-7.12 (m, 4H), 7.05-7.03 (m, 1H), 6.89 (d, 2H, *J* = 8.2 Hz), 6.82 (t, 1H, *J* = 7.3 Hz), 5.05 (d, 1H, *J* = 13.8 Hz), 4.12-4.05 (m, 1H), 3.73-3.69 (m, 1H), 2.76-2.74 (m, 2H), 2.08-1.91 (m, 4H), 1.85-1.07 (m, 18H); ¹³C NMR (CDCl₃, 100 MHz) δ 150.4 (d, *J*_{C-P} = 9.5 Hz), 136.2 (d, *J*_{C-P} = 3.8 Hz), 131.7, 129.4 (d, *J*_{C-P} = 1.9 Hz), 129.2, 127.5 (d, *J*_{C-P} = 3.8 Hz), 127.0 (d, *J*_{C-P} = 2.9 Hz), 125.9 (d, *J*_{C-P} = 1.9 Hz), 120.3, 118.3, 57.5 (d, *J*_{C-P} = 1.9 Hz), 56.8 (d, *J*_{C-P} = 2.9 Hz), 46.9, 37.8, 37.4, 36.8, 27.0 (d, *J*_{C-P} = 7.6 Hz), 26.9 (d, *J*_{C-P} = 3.8 Hz), 26.8 (d, *J*_{C-P} = 2.9 Hz), 26.2 (d, *J*_{C-P} = 3.8 Hz), 26.1, 26.0, 25.9 (d, *J*_{C-P} = 3.8 Hz), 25.0; ³¹P NMR (CDCl₃, 160 MHz) δ 52.9; IR (KBr) cm⁻¹ 3426 (m), 3056 (w), 3032 (w), 2926 (s), 2852 (s), 1595 (s), 1505 (s), 1449 (m), 1312 (m), 1151 (s); ESI-HRMS (*m/z*) calcd. for C₂₇H₃₇N₁O₁P₁ [(M+H)⁺]: 422.26128, found: 422.25985.

Part III: NMR Spectra of **6a-n**

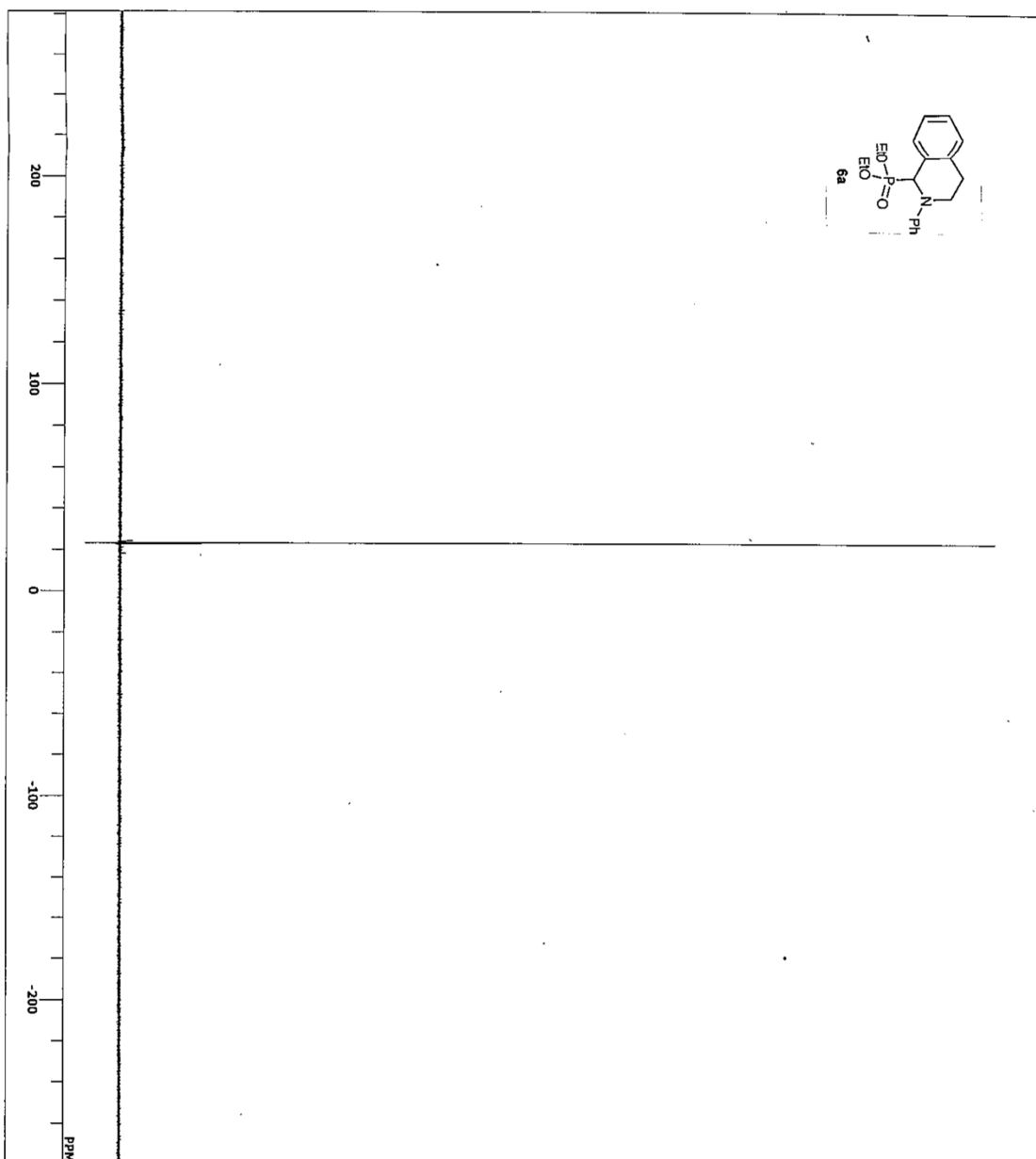
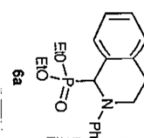


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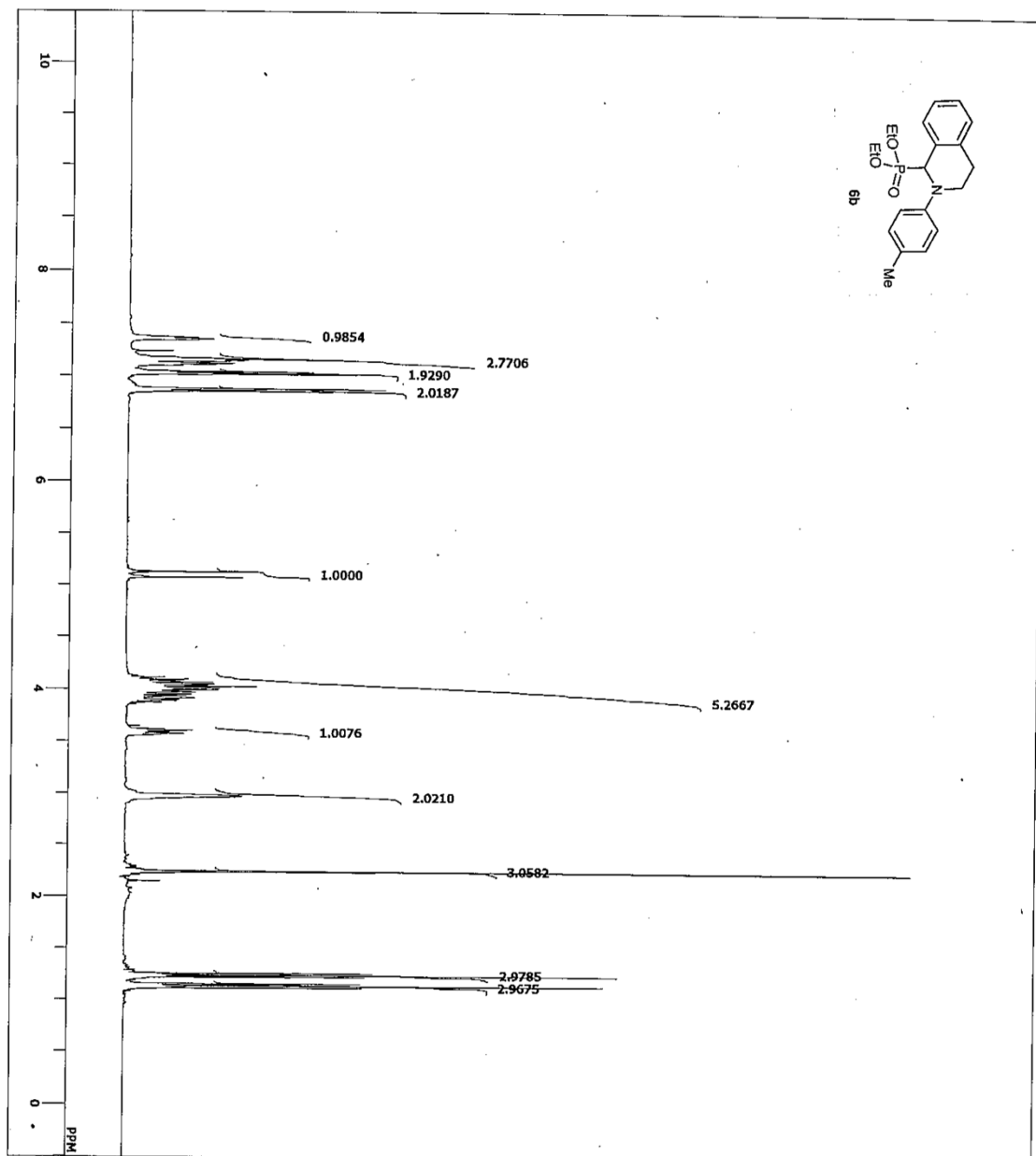


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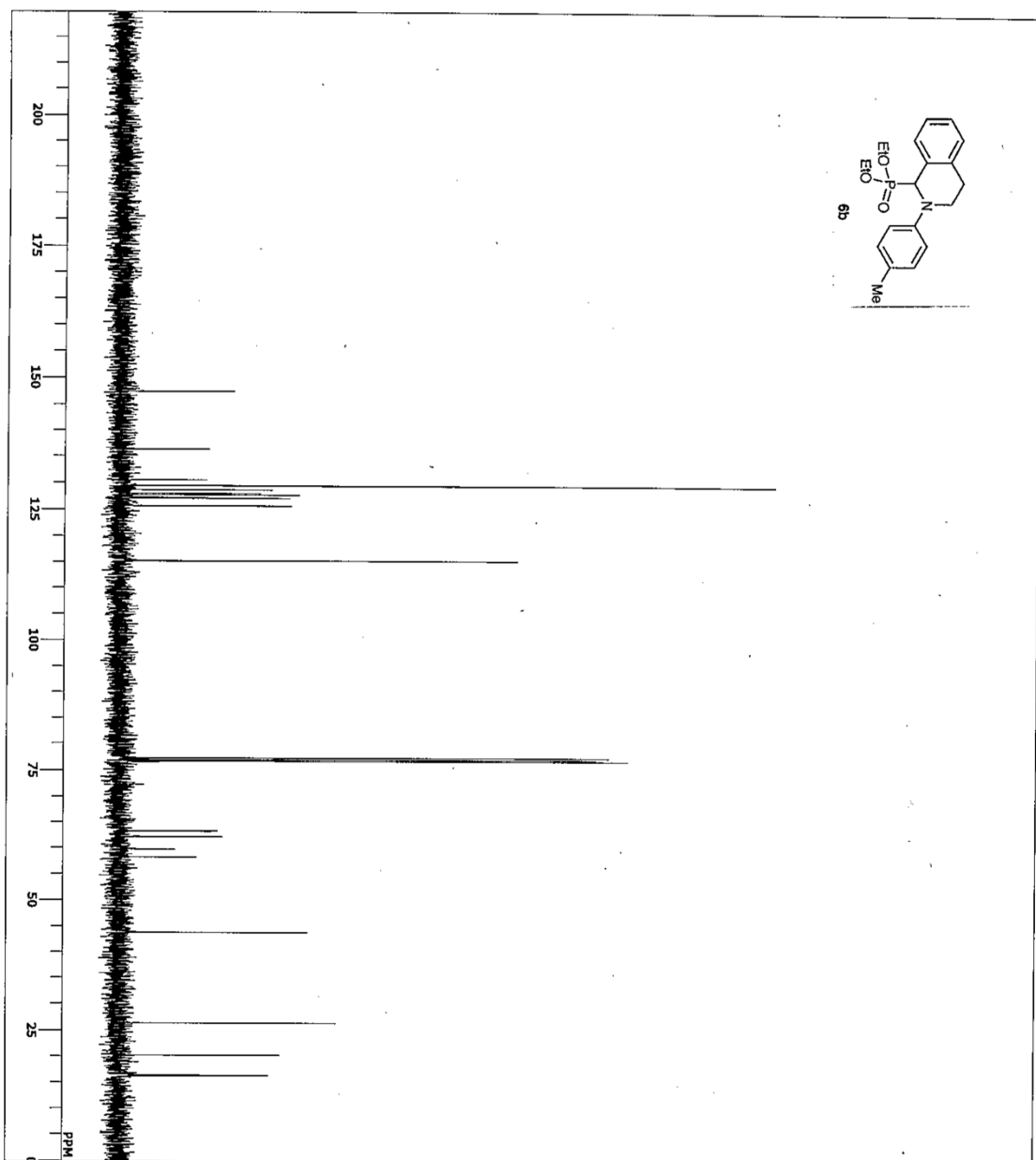
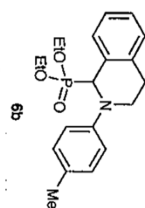
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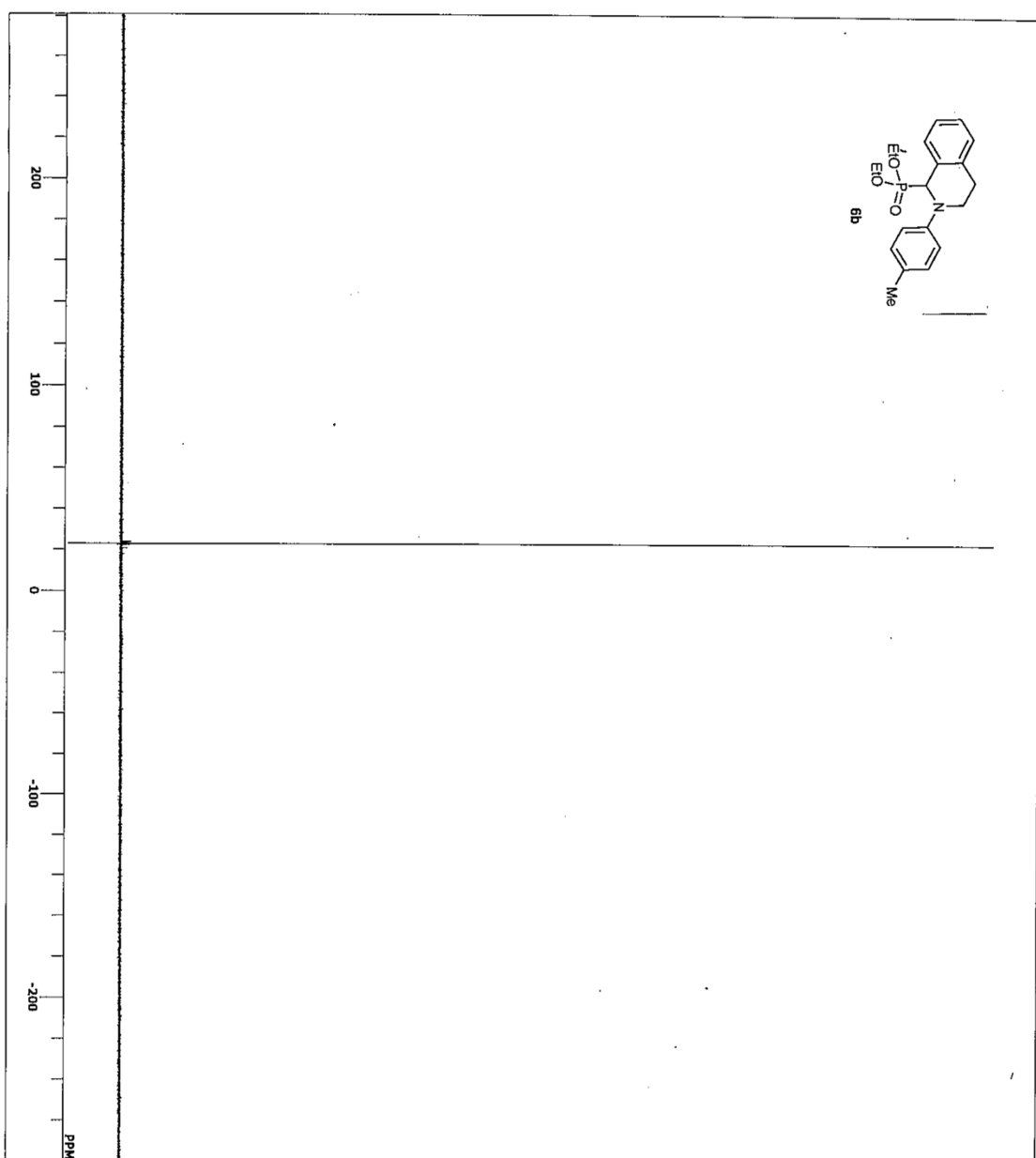
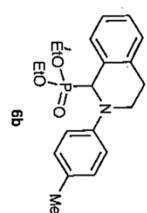


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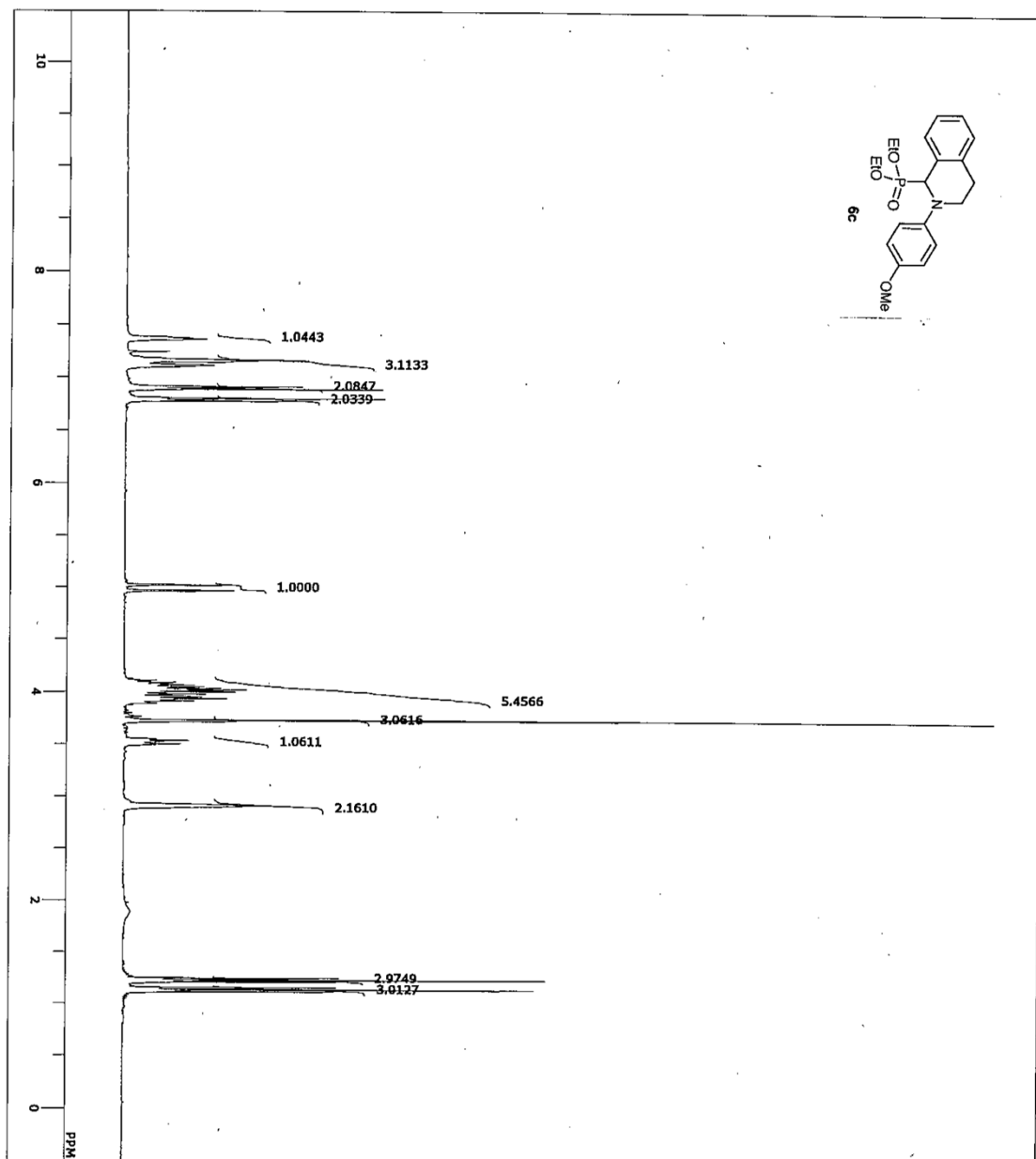
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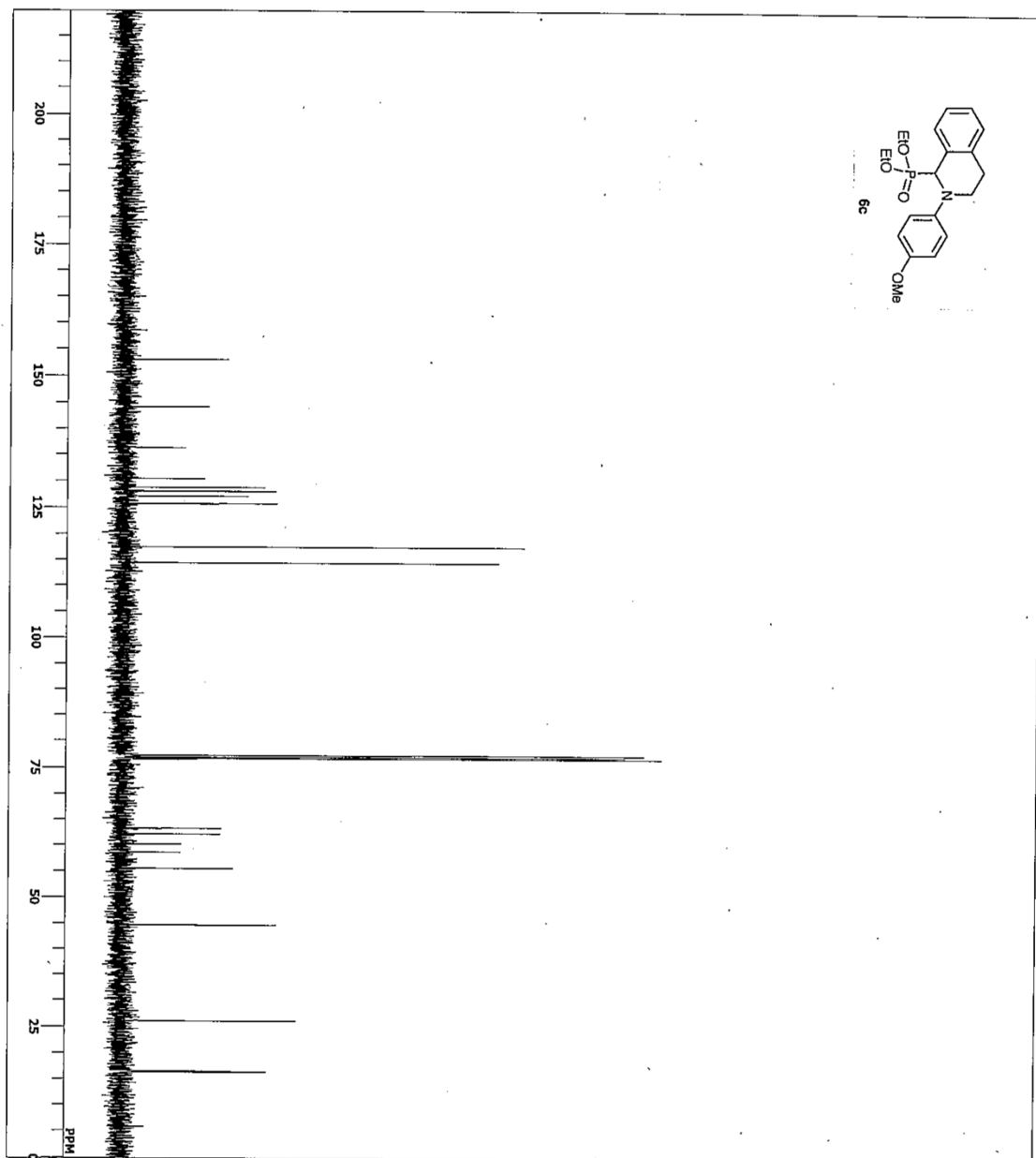


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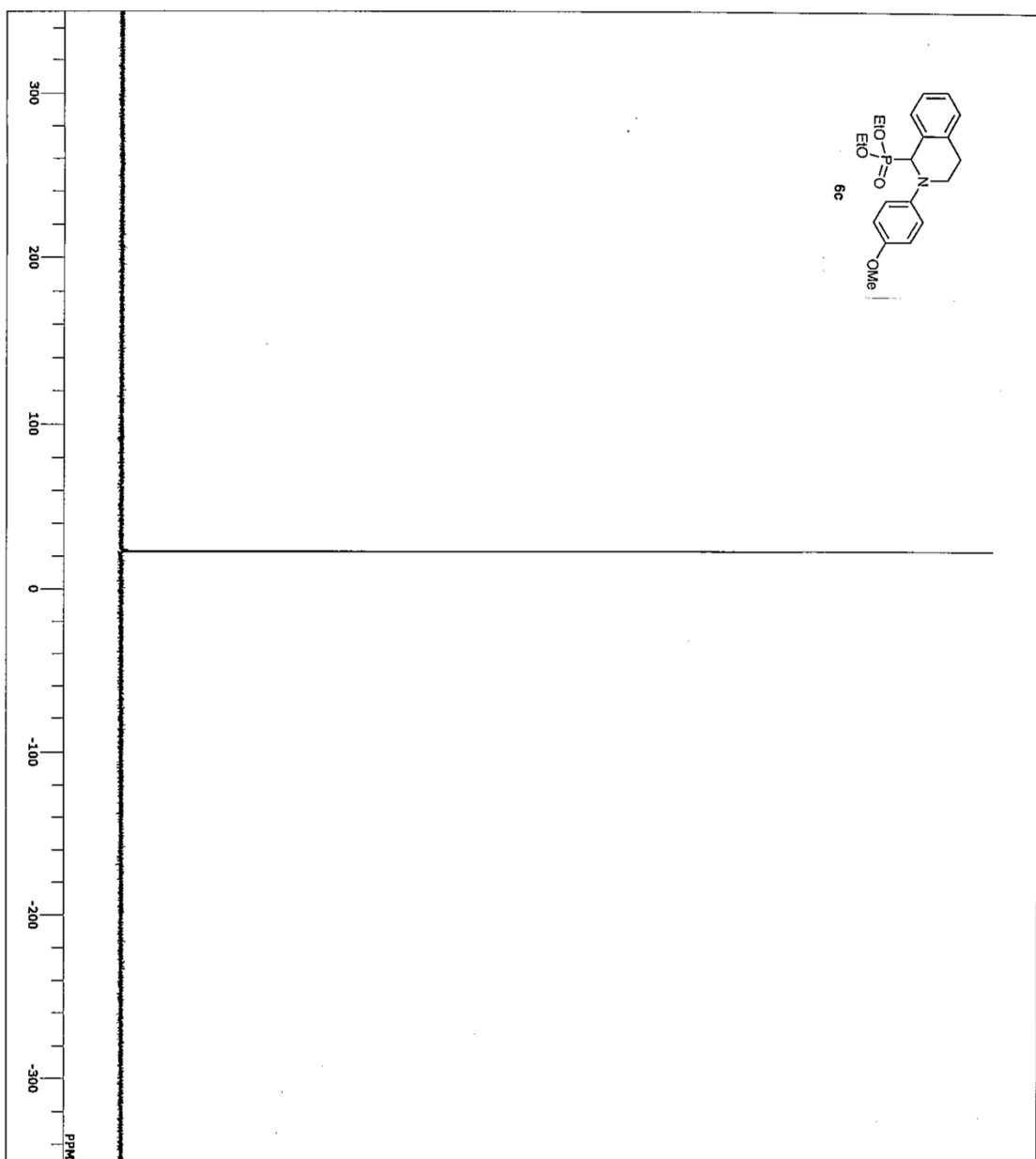
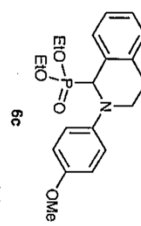
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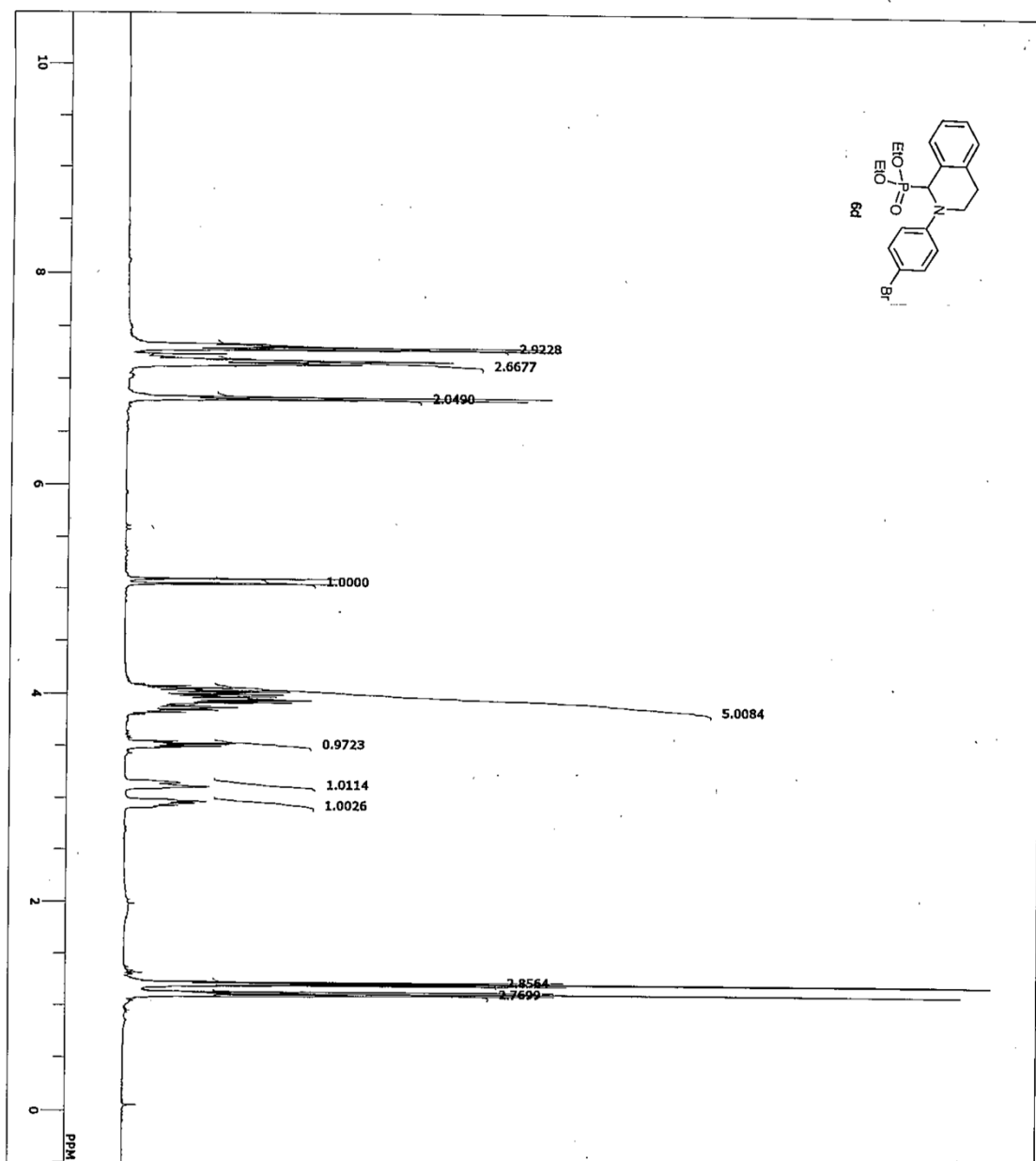
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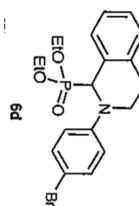
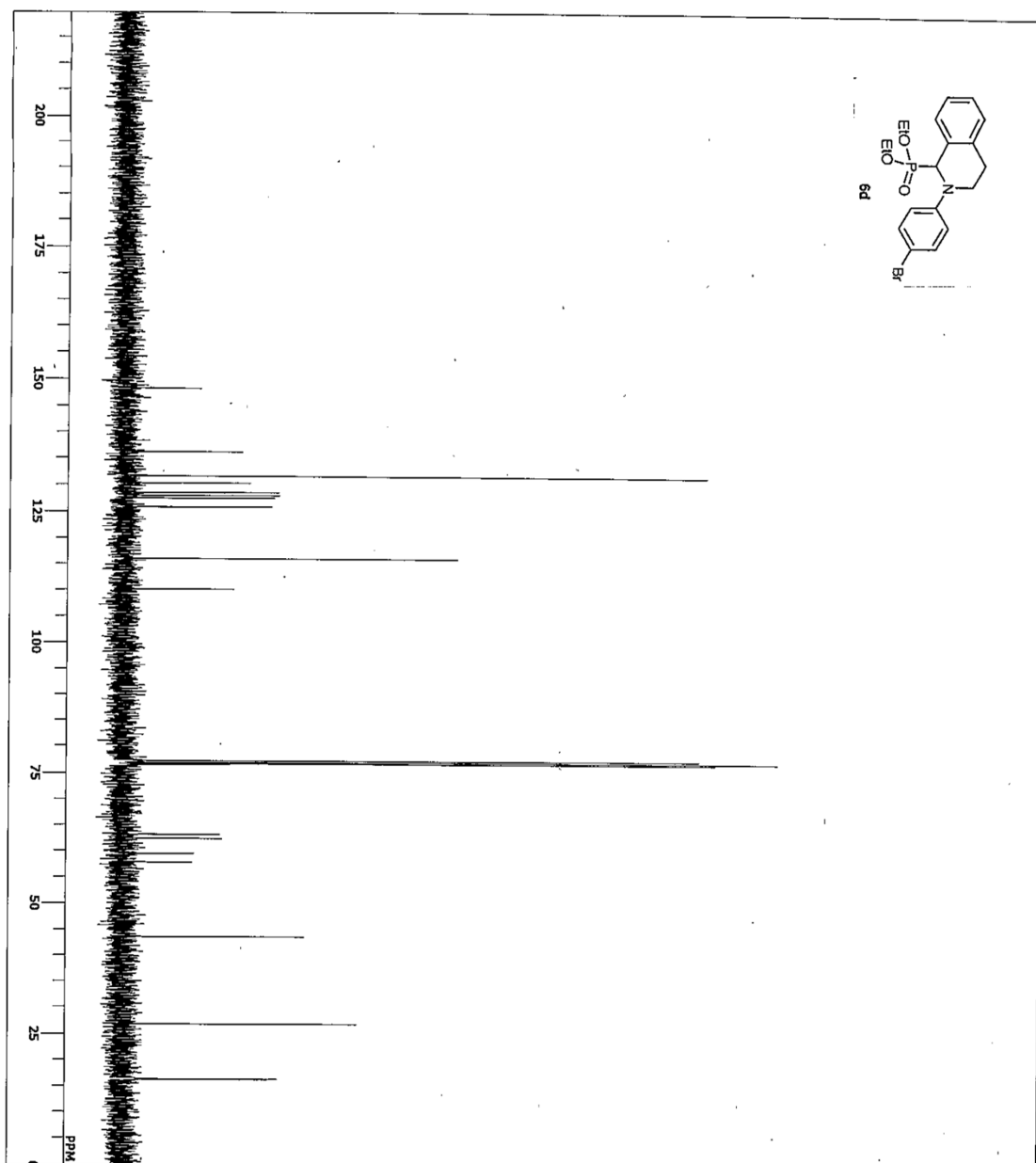
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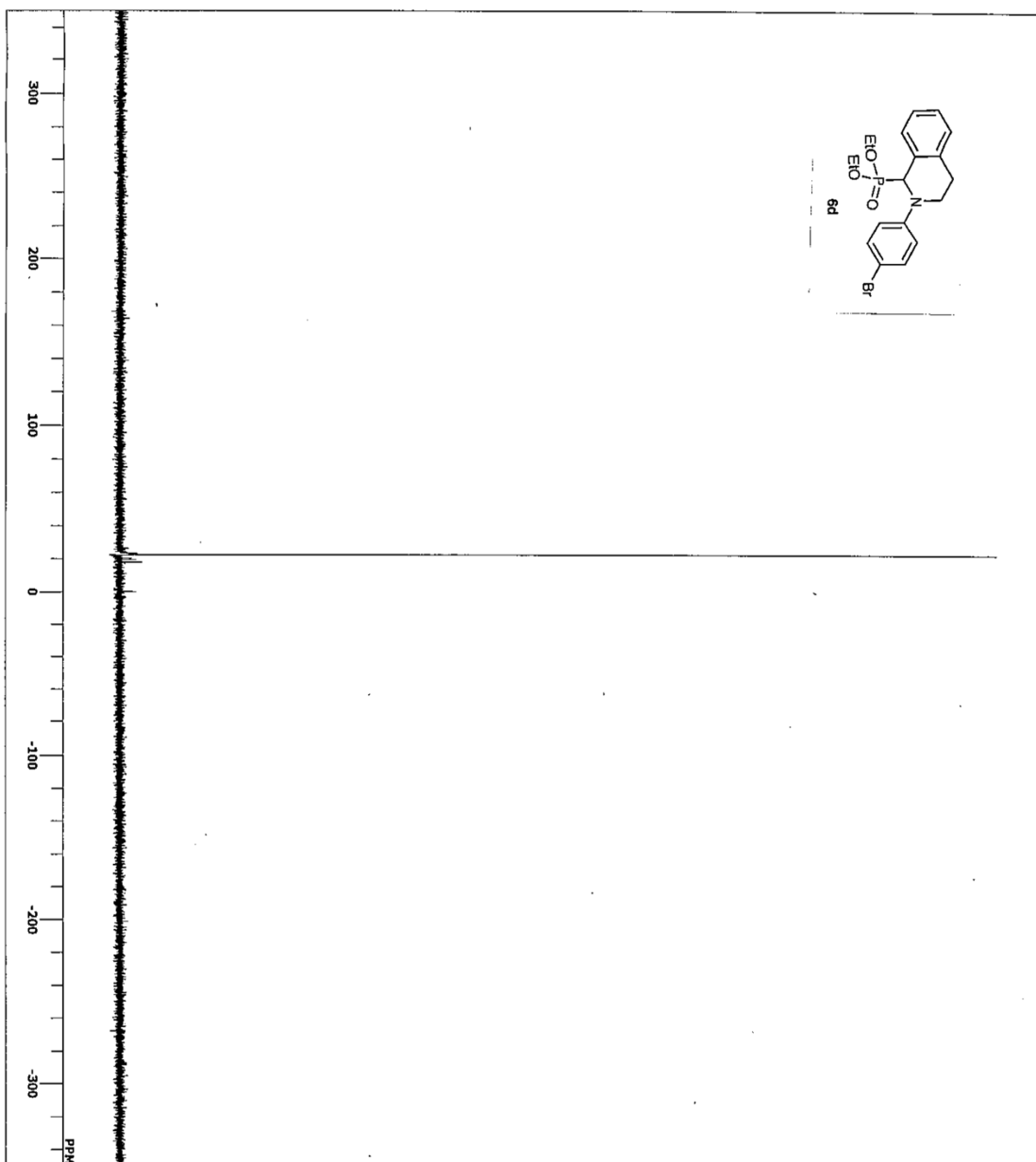
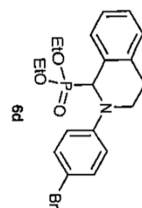
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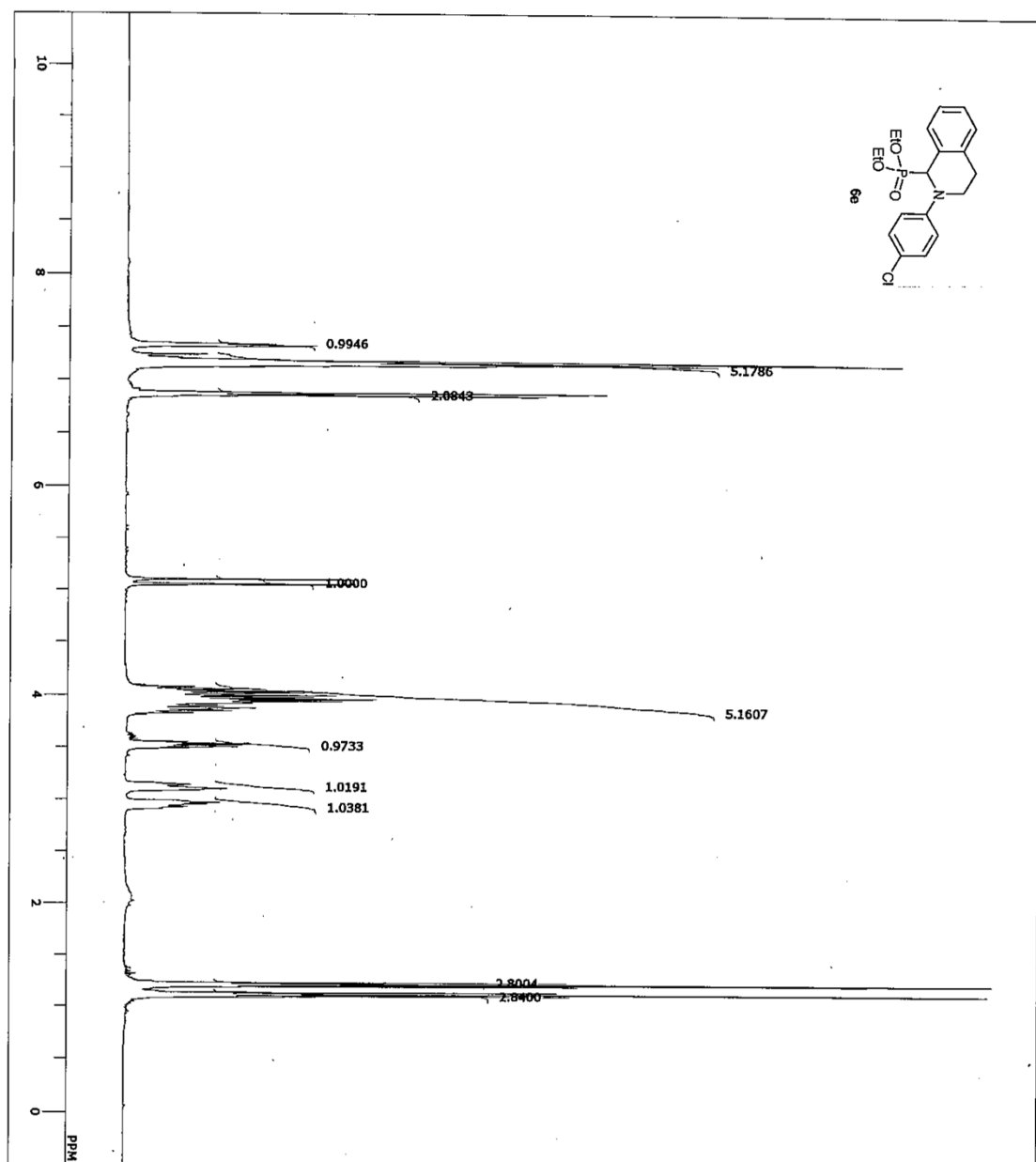
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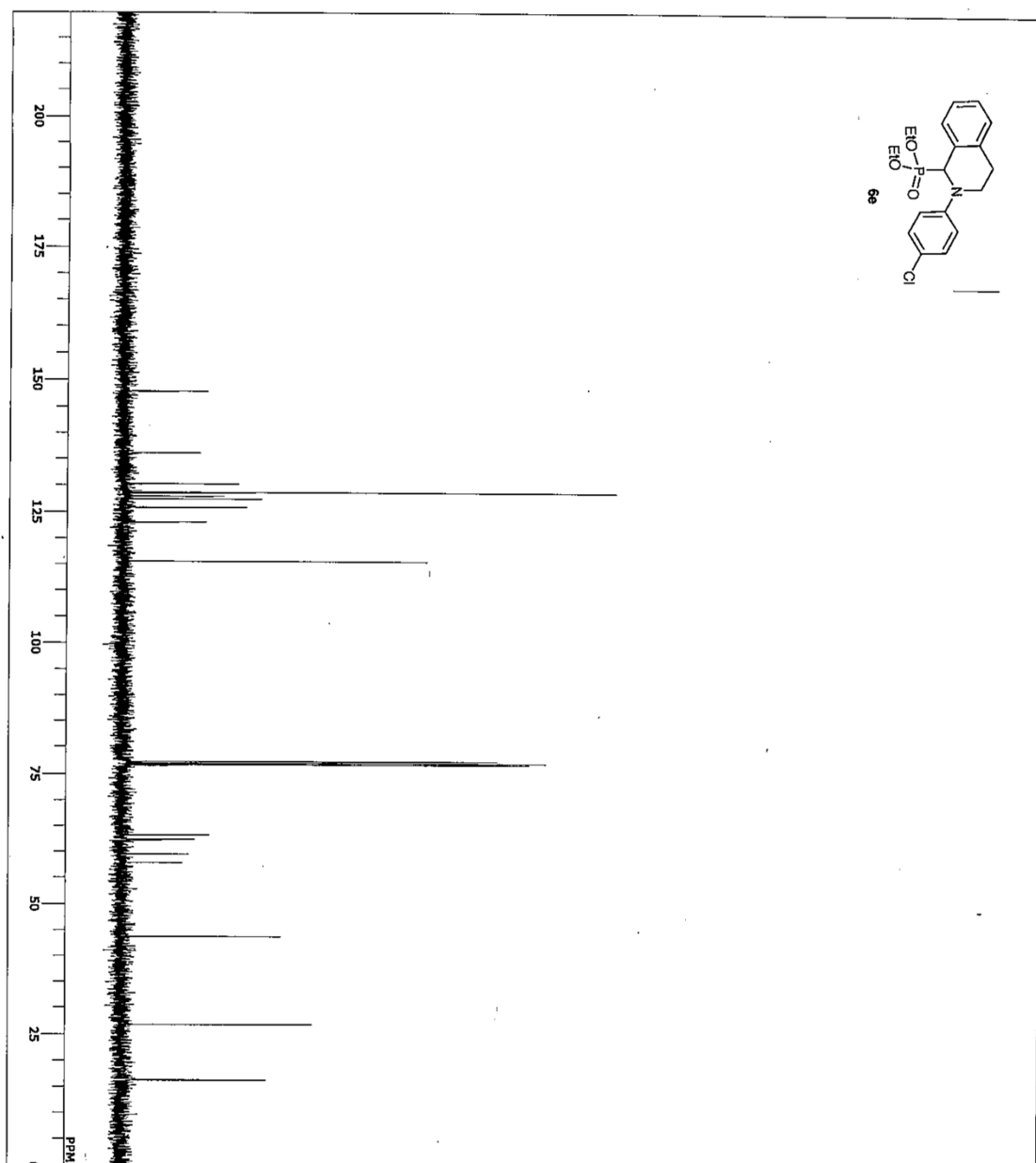
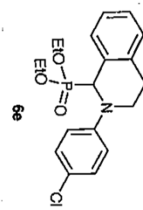
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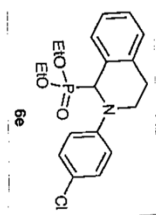


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 EXMUD single_pulse.ac2
 DBFRO 399.76 MHz
 OBSF 400.14 MHz
 OBEN 7.24 Hz
 POINT 16384
 FREQ 7503.00 Hz
 SCANS 8
 ACQTM 2.1837 sec
 PD 2.0000 sec
 PW1 6.50 usec
 IRNUC 1H
 CTMP 408.4 c
 CLVNT CDCL3
 EXREF 7.24 ppm
 BF 0.42 Hz
 RGAIN 32

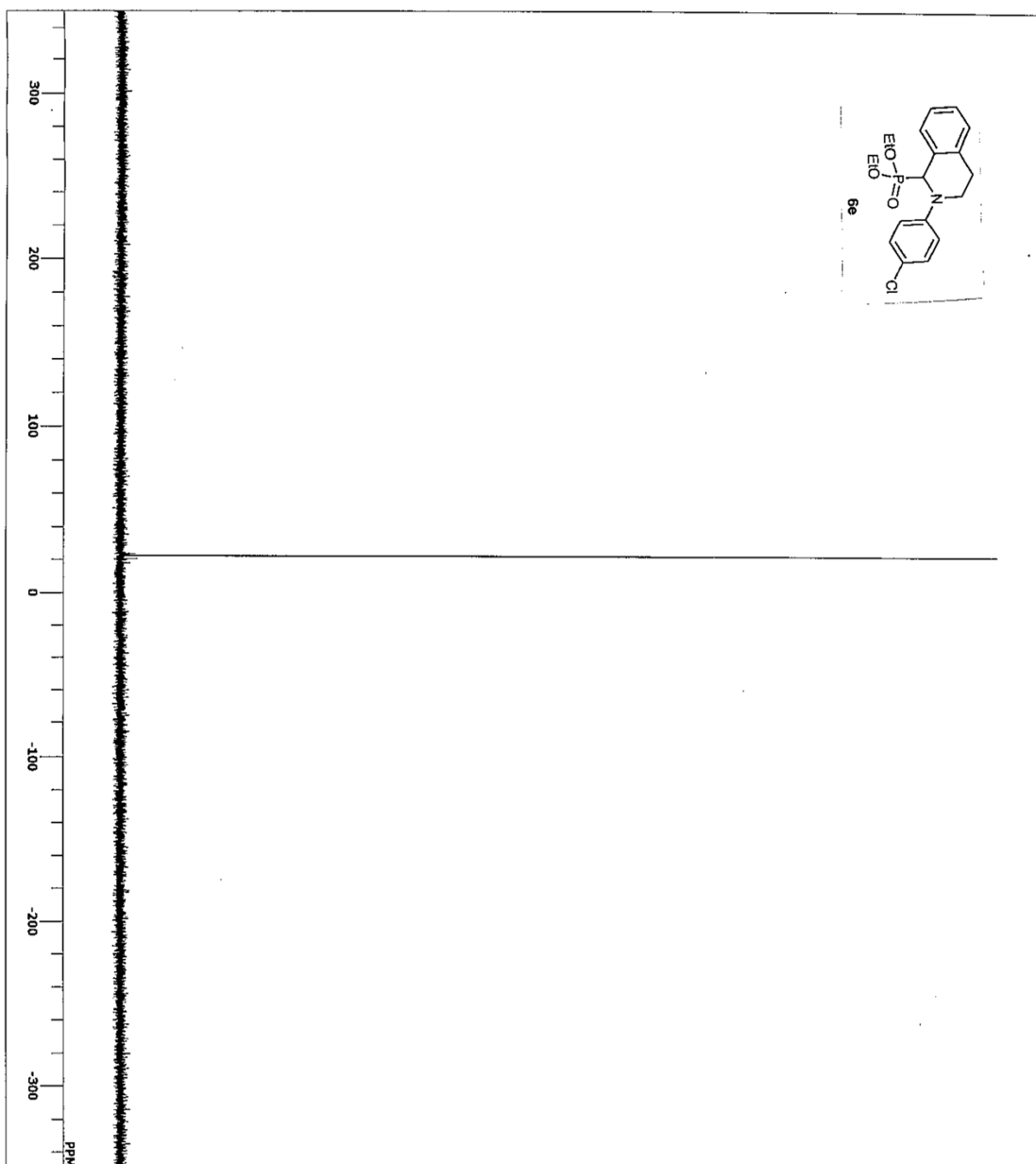


D:\Documents and Settings\Kobayashi\My Desktop\WV-16

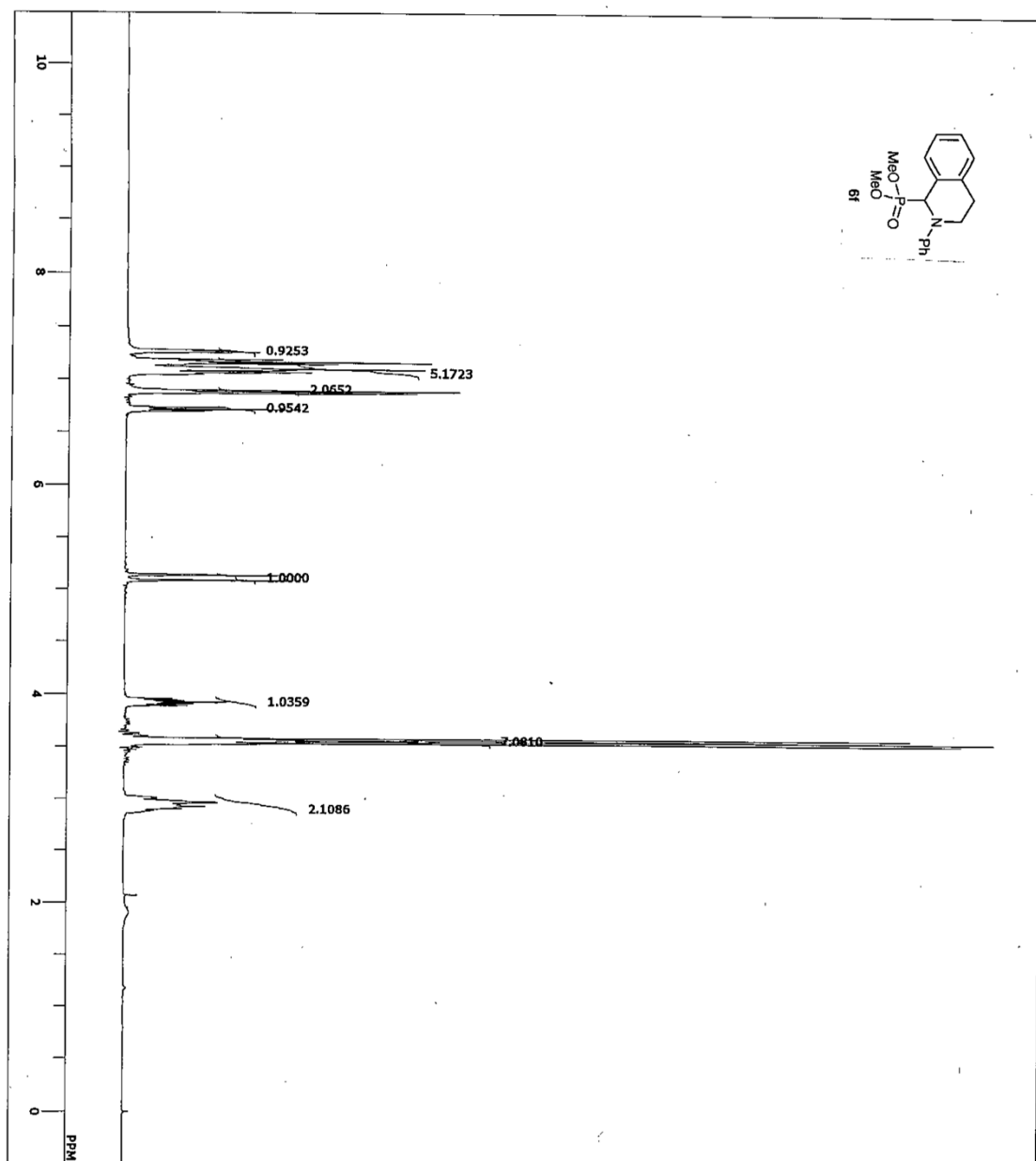
DFILE COMNT 24-07-2013 10:15:03
 DATIN 13C
 ORNUC single_pulse_dec
 EXMOC 100.53 MHz
 ORPRQ 5.35 KHz
 ORSET 5.85 Hz
 ORFTN 32768
 POINT 31607.03 Hz
 FREQ 31607.03 Hz
 SCANS 1.0433 sec
 ACQTM 2.0000 sec
 PD 3.50 usec
 PWL 1H 408.4 c
 TRNUC CDCL3
 CTENP 77.00 ppm
 SILNT 0.42 Hz
 EXREF 54
 BF 54
 RGAIN 54



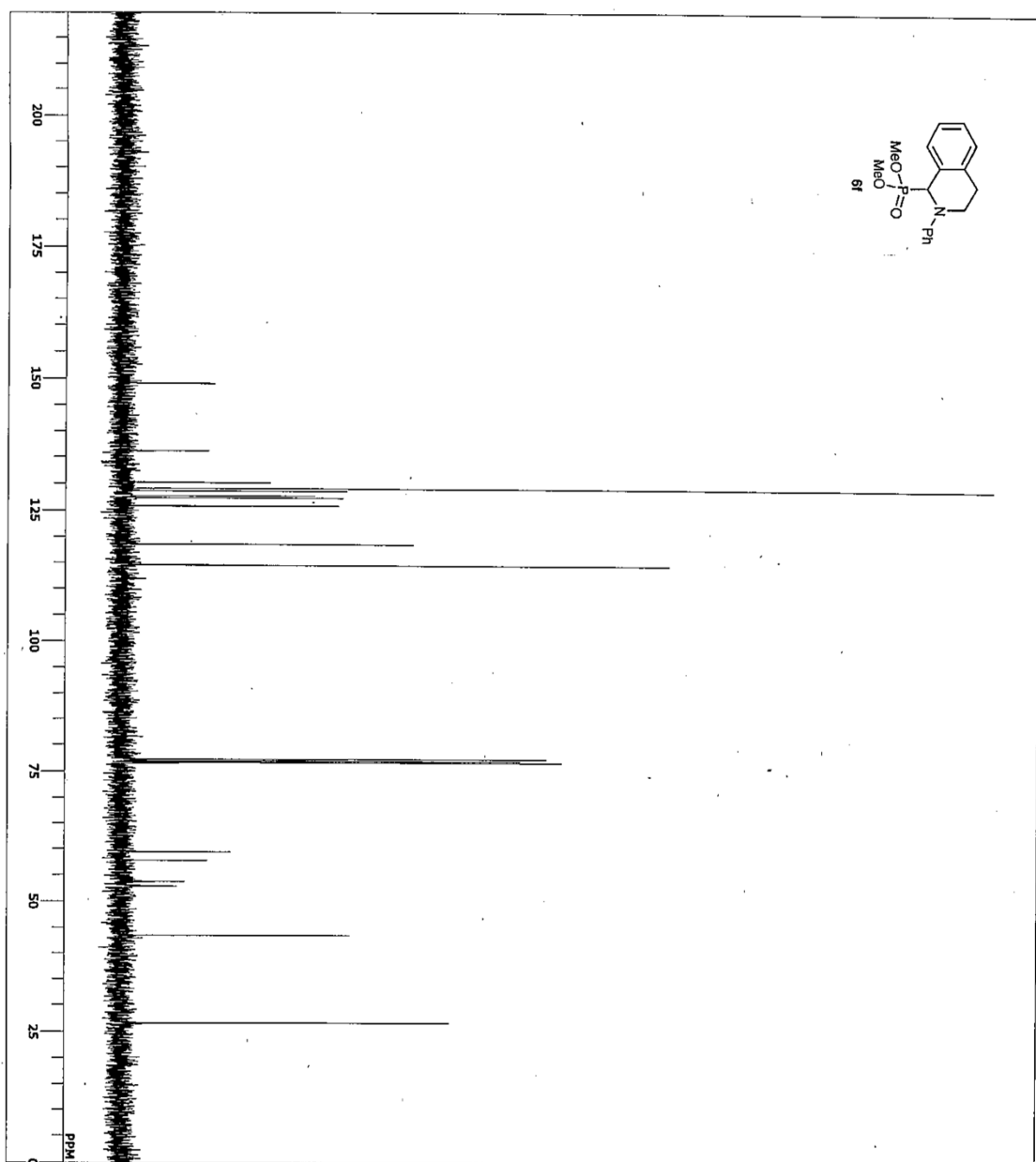
6e



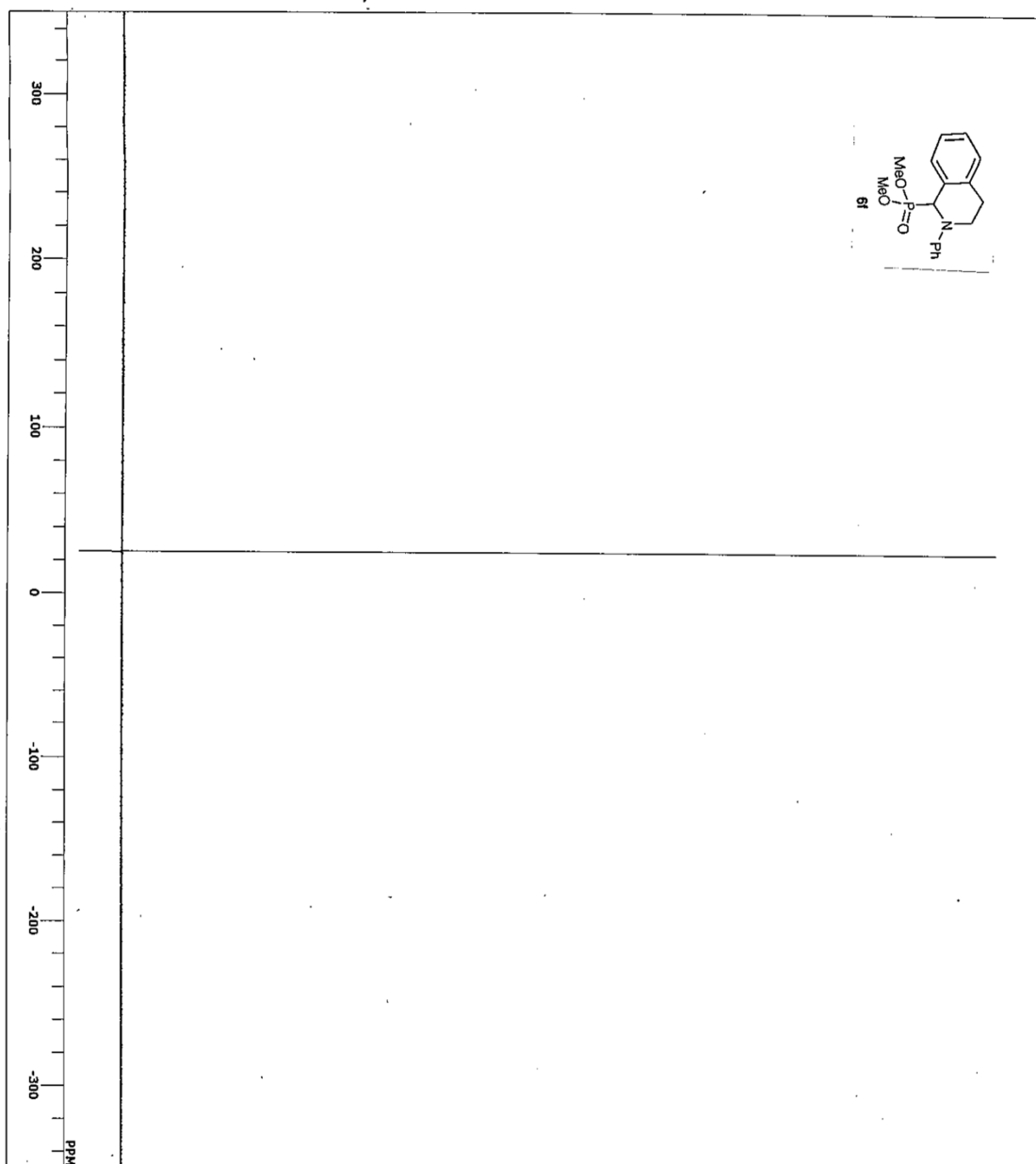
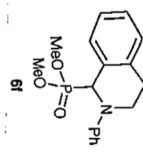
D:\Documents and Settings\Kobayashi\lab\Desktop\WV-16
 24-07-2013 10:17:25
 31P
 single, pulse, dec
 161.83 MHz
 4.69 kHz
 3.09 Hz
 32768
 142045.45 Hz
 3
 0.2307 sec
 2.0000 sec
 5.80 usec
 1H 408.5 c
 CDCl3
 0.00 ppm
 0.42 Hz
 56



D:\Documents and Settings\Kobayashi\lab\Desktop\pww-16
 DFILE 23-07-2013 14:26:29
 COMNT 1H
 DATIM 1H
 EXHUC single_pulse.ex2
 EXHUC 359.78 MHz
 OBSFQ 413 MHz
 OBSFQ 413 MHz
 POINT 14584
 FREQU 7503.00 Hz
 SCANS 8
 ACQTM 2.1837 sec
 PD 2.0000 sec
 PW1 6.50 usec
 TRNUC 1H
 CTMPC 408.6 c
 SILVNT CDCl₃
 EXREF 0.00 ppm
 BF 0.42 Hz
 RGAIN 28

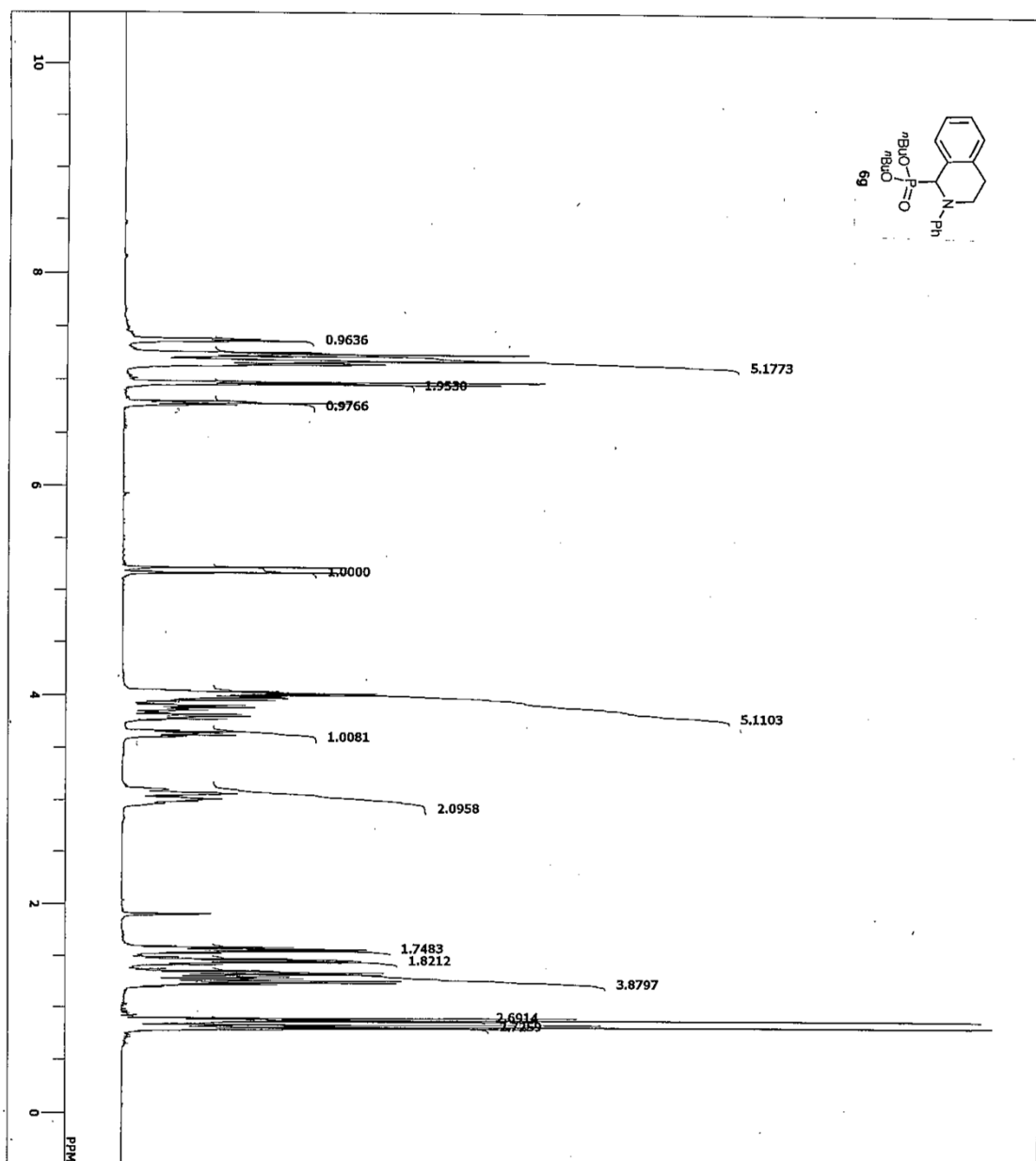


D:\Documents and Settings\Kobayashi\lab\Desktop\WV-16
 DFILE
 COMNT 23-07-2013 14:30:59
 DATM 13C
 OBNUC single_pulse_dec
 EXMOD 100.53 MHz
 OBSFQ 5.35 kHz
 OBSL 5.86 Hz
 OBFN 32758
 PD 31407.03 Hz
 FREQ 1.033 sec
 SCNS 2.0000 sec
 ACQTM 3.50 usec
 PD 77.00 ppm
 PUL 54
 IRNUC 1H 408.6 c
 CTMP CDCL3
 SLVT 0.42 Hz
 EXREF 54
 RGAIN 54

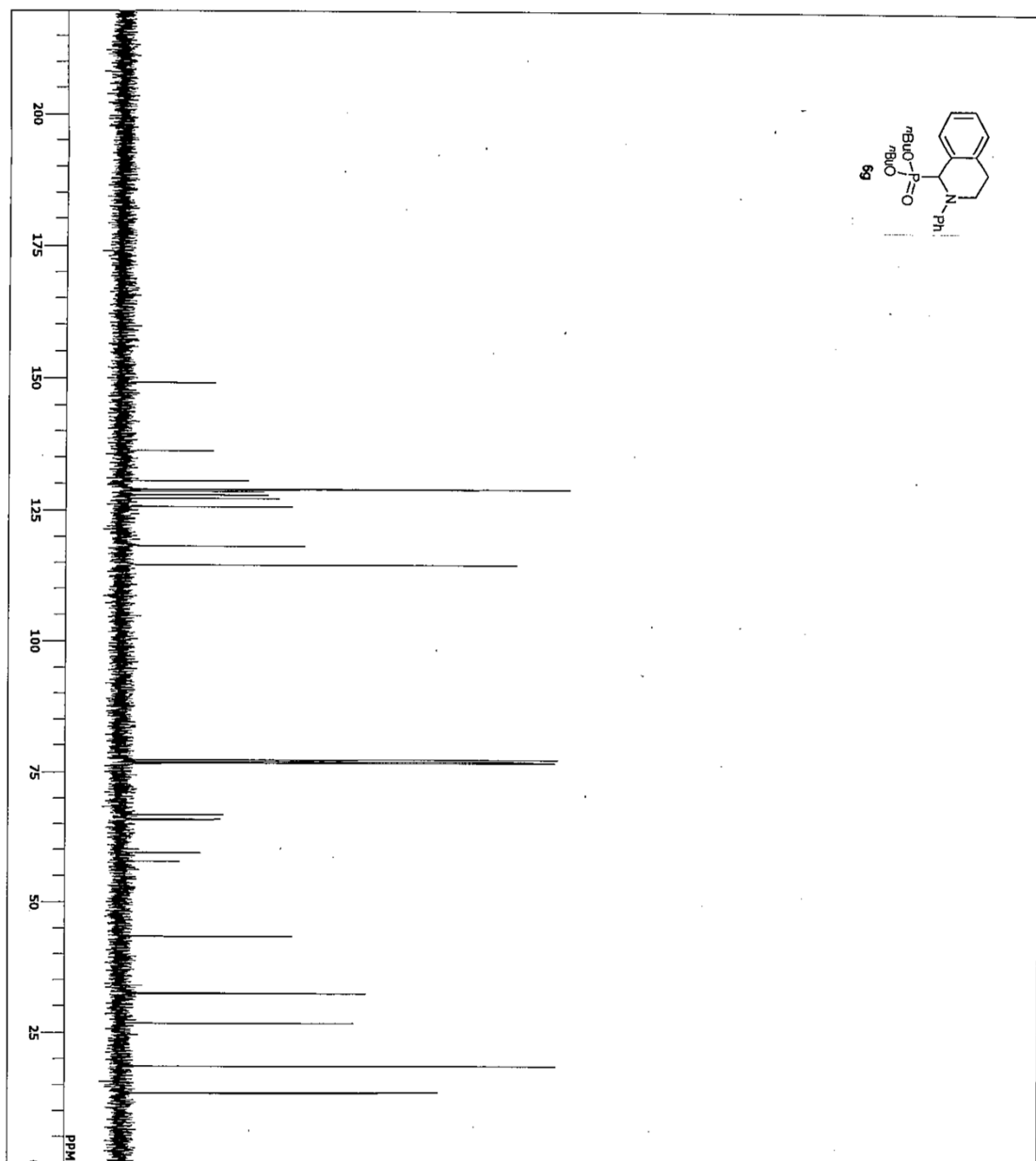
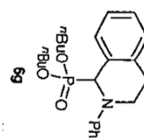


D:\Documents and Settings\Kobayashi\My Desktop\WY-16

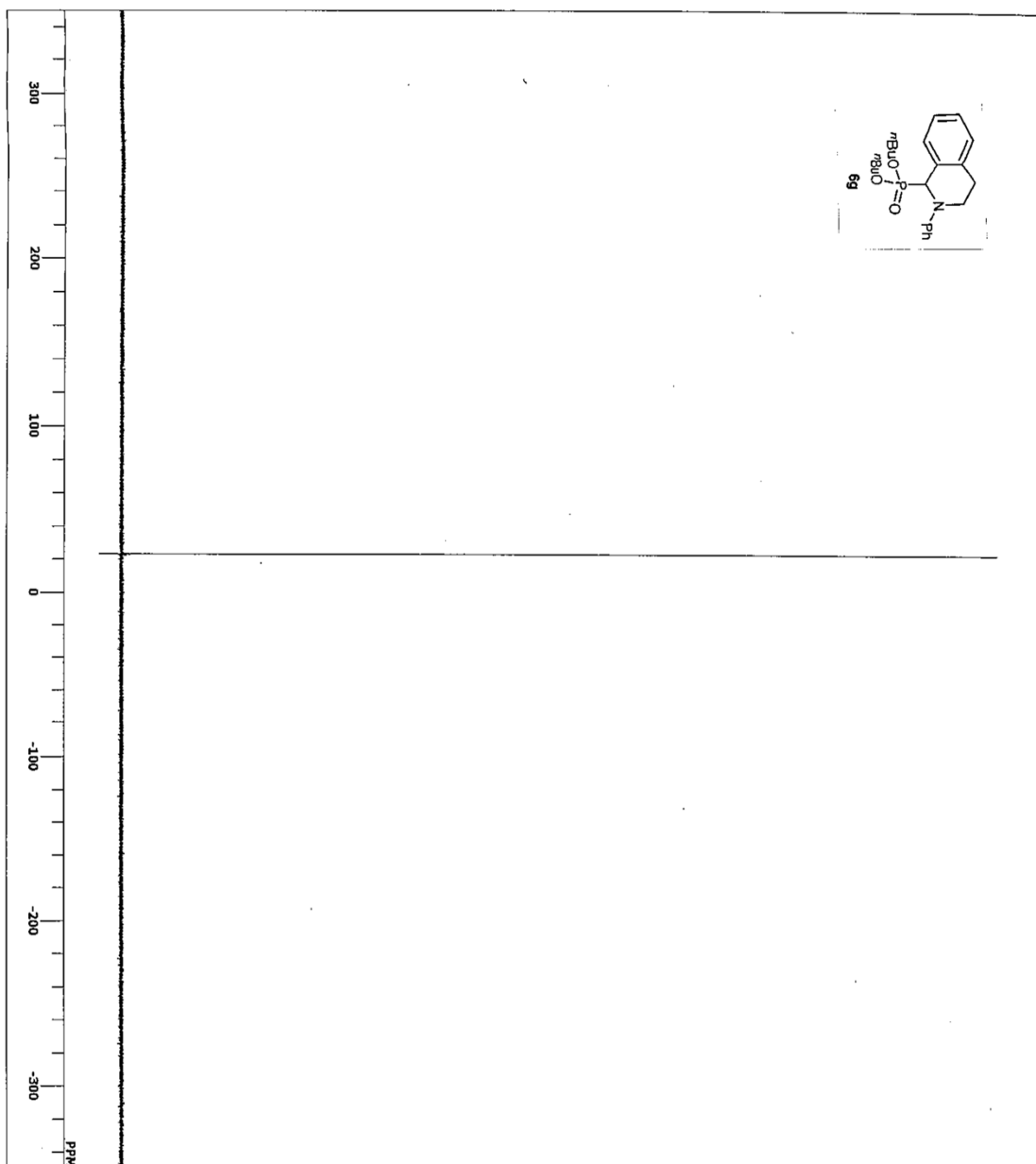
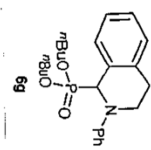
DFILE	COMNT	23-07-2013 14:33:40
DATIM	31P	
OBNUC	single_pulse_dec	
EXMOD	161.83 MHz	
OBFRQ	4.69 KHz	
OBSET	3.09 Hz	
OBFTN	32768	
POINT	142045.45 Hz	
FREQ	9	
SCANS	0.2307 sec	
ACQTM	2.0000 sec	
PD	5.80 usec	
INUC	1H 408.6 c	
CTEMP	CDCl3	
SIVNT	0.00 ppm	
EXREF	0.42 Hz	
BF	54	
RGAIN		



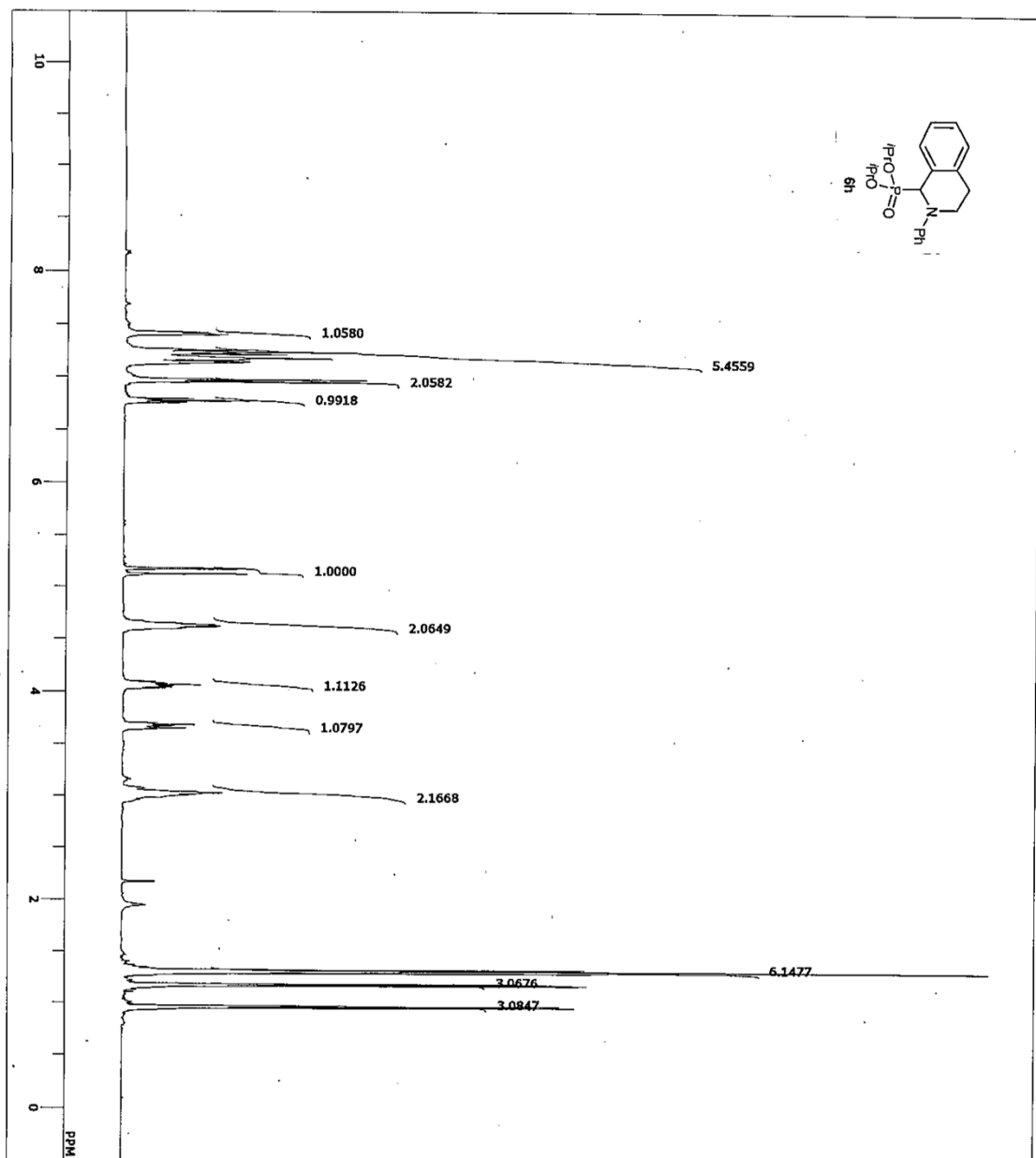
D:\Documents and Settings\Kobayashi\lab\Desktop\WV-16
 25-07-2013 16:34:31
 1H
 single, pulse-ex2
 399.78 MHz
 4.12 Hz
 7.24 Hz
 14.58 Hz
 7503.00 Hz
 8
 2.1837 sec
 2.0000 sec
 6.50 usec
 408.7 °C
 CDCl₃
 7.24 ppm
 0.42 Hz
 Z8
 RGAIN



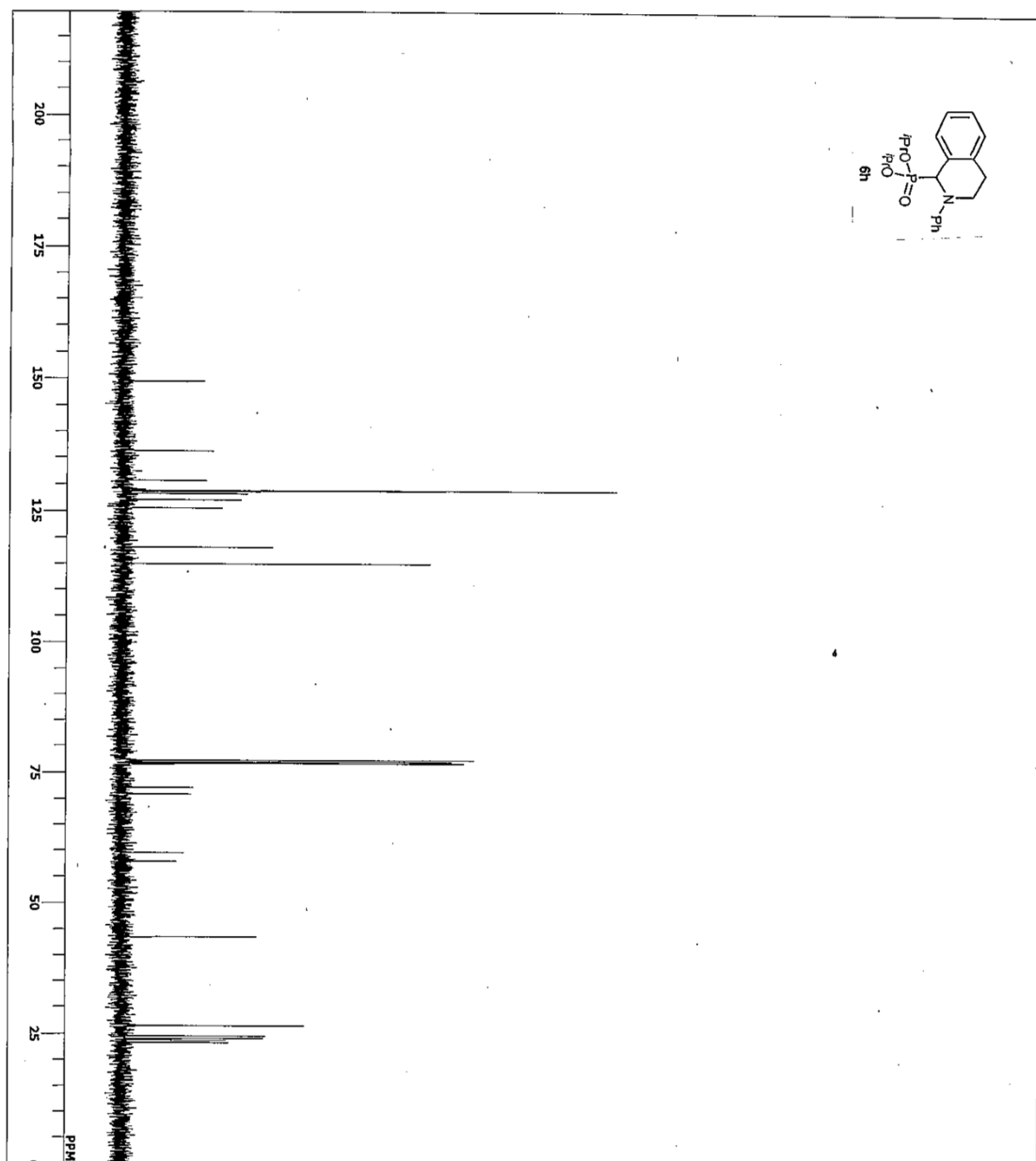
D:\Documents and Settings\Kobayashi\Desktop\WV-16
 DFILE 25-07-2013 16:40:34
 CONNT 13C
 DATIN
 OBNUC
 EXMOD Single_pulse_dec
 OBRQ 100.53 MHz
 OBRF 500 MHz
 OBRN 500 MHz
 POINT 32768
 FREQ 31407.03 Hz
 SCANS 56
 ACQTM 1.0433 sec
 PD 2.0000 sec
 PW1 3.50 usec
 TRNUC 1H
 CTEMP 408.7 c
 SLVNT CDCl3
 EXREF 77.00 ppm
 BF 0.42 Hz
 RGAIN 60



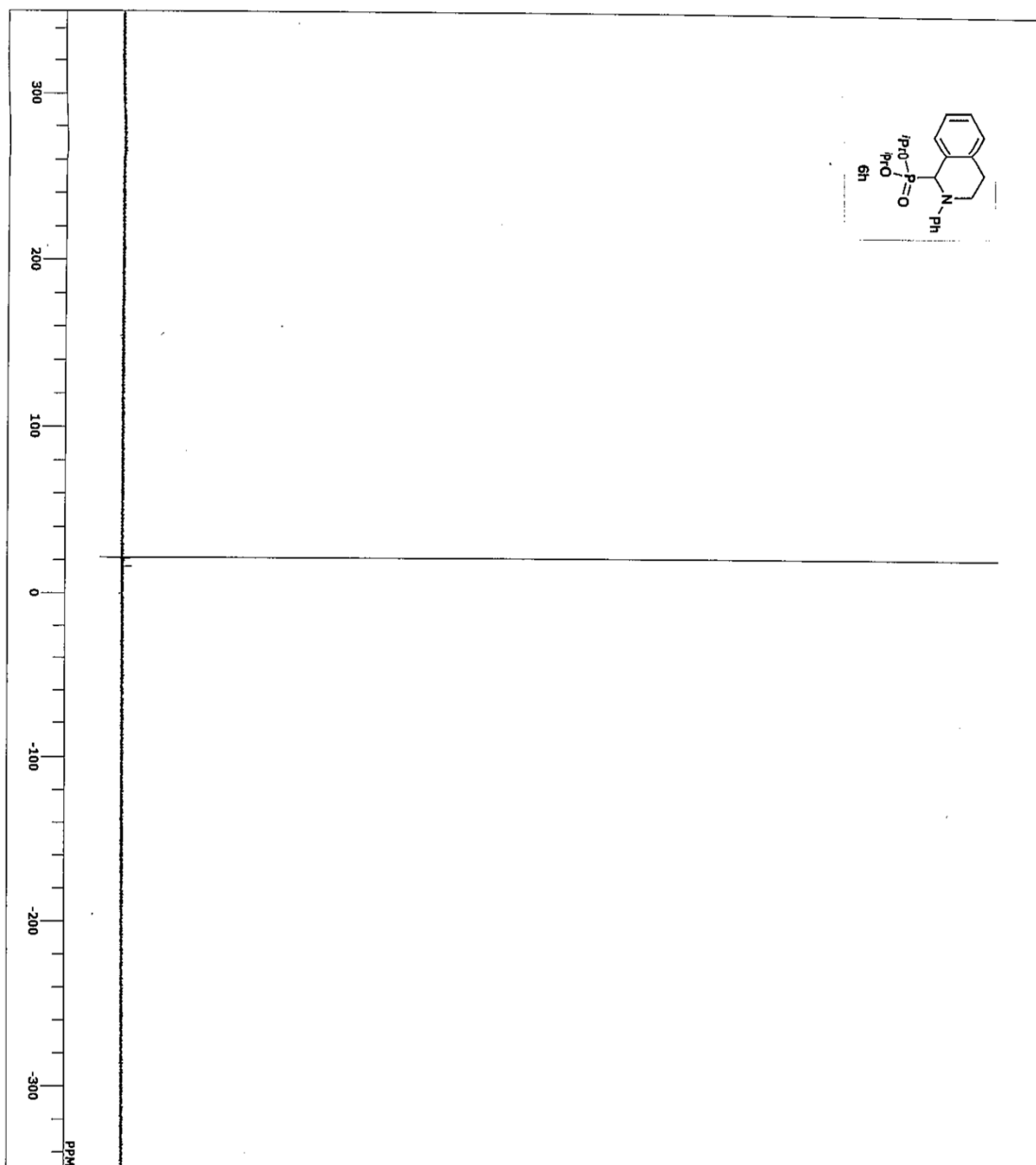
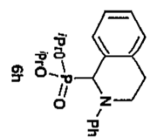
D:\Documents and Settings\Kobayashi\lab\Desktop\WV-16
 25-07-2013 16:43:04
 31P
 single, pulse, dec
 161.83 MHz
 4.69 KHz
 3.09 Hz
 32768
 142045.45 Hz
 5
 0.2307 sec
 2.0000 sec
 5.80 usec
 1H 408.7 c
 CDCl3
 0.00 ppm
 0.42 Hz
 54
 RGAIN



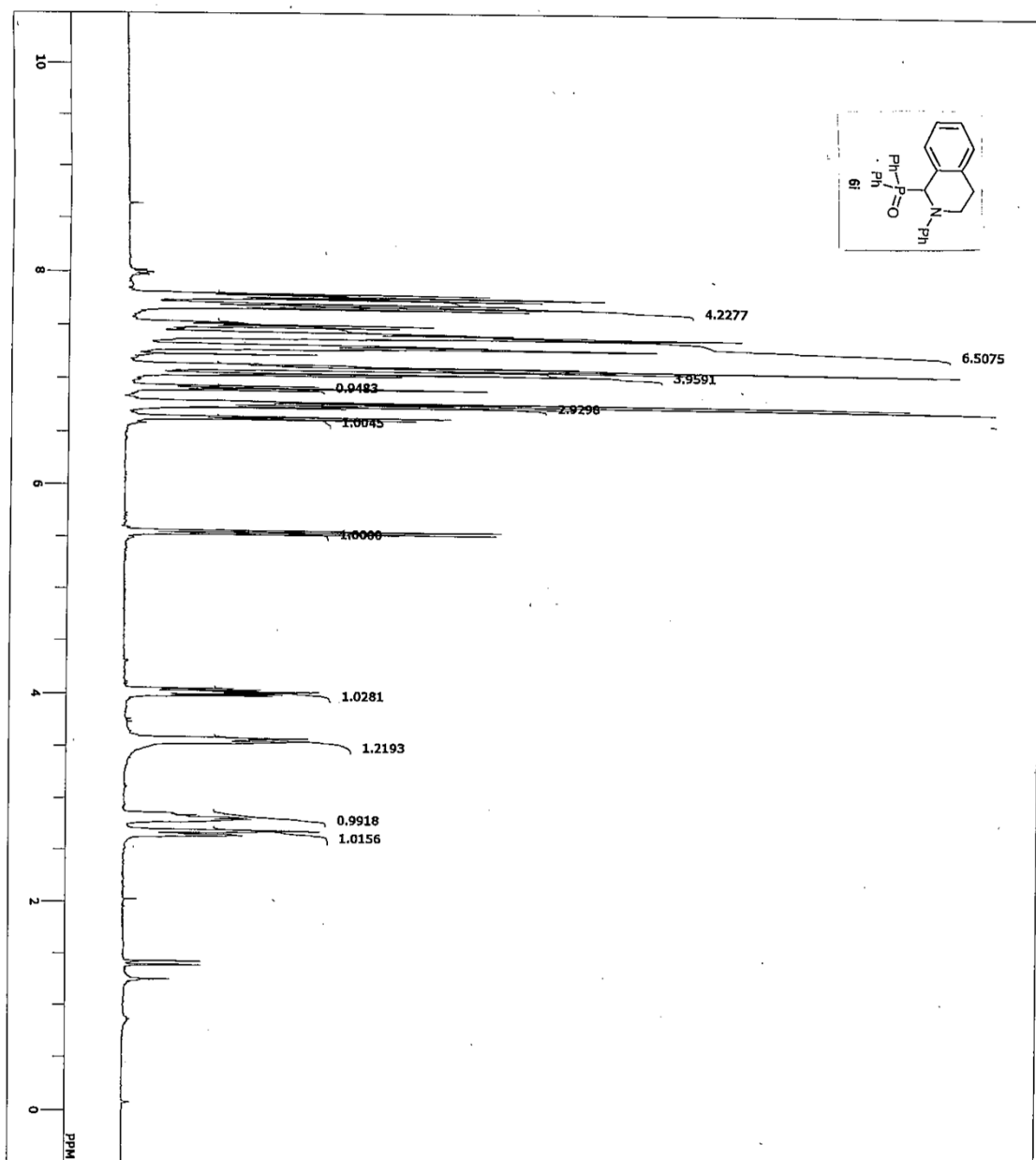
D:\Documents and Settings\Kobayashi\Desktop\WV-16
 D:\Documents and Settings\Kobayashi\Desktop\WV-16
 23-07-2013 14:37:04
 1H
 single_pulse.ex2
 399.78 MHz
 4.19 KHz
 7.29 Hz
 16384
 7503.00 Hz
 8
 2.1637 sec
 2.0000 sec
 6.50 usec
 1H 408.6 c
 CDCl3
 7.24 ppm
 0.42 Hz
 28
 RGAIN



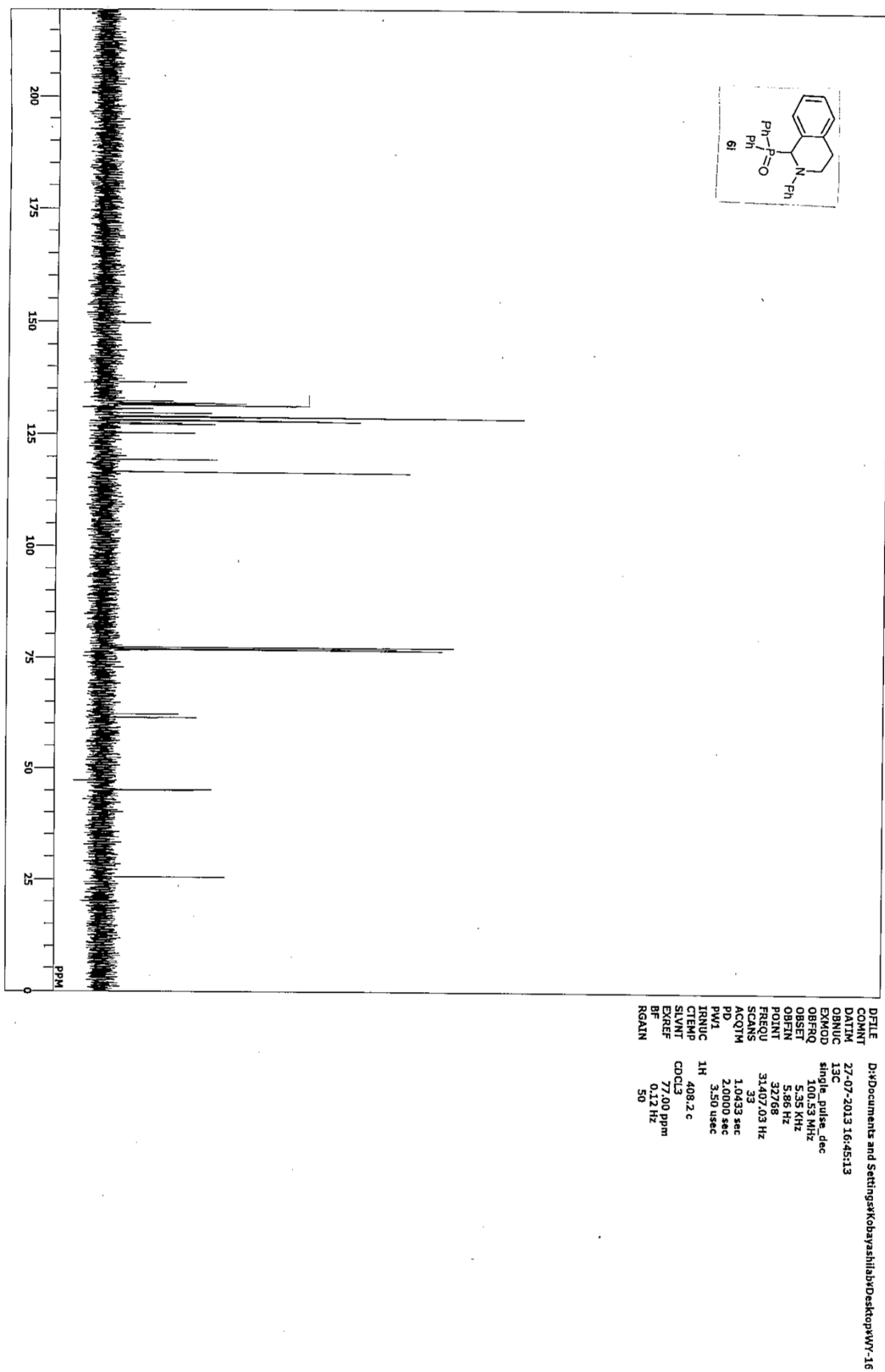
D:\Documents and Settings\Kobayashi\lab\Desktop\WV-16
 DFILE
 CONNT 23-07-2013 14:42:33
 DATE 13C
 DNAME Single_pulse_dec
 EXPRD 100.53 MHz
 OBSFQ 5.08 Hz
 OBSFZ 32758
 POINT 31407.03 Hz
 FREQ 45
 SCANS 1.0433 sec
 ACQTM 2.0000 sec
 PD 3.50 usec
 PUL 1H
 IRNUC 408.6 c
 CTEMP CDCL3
 SLVNT 77.00 ppm
 EXREF 0.42 Hz
 BF 60
 RGAIN

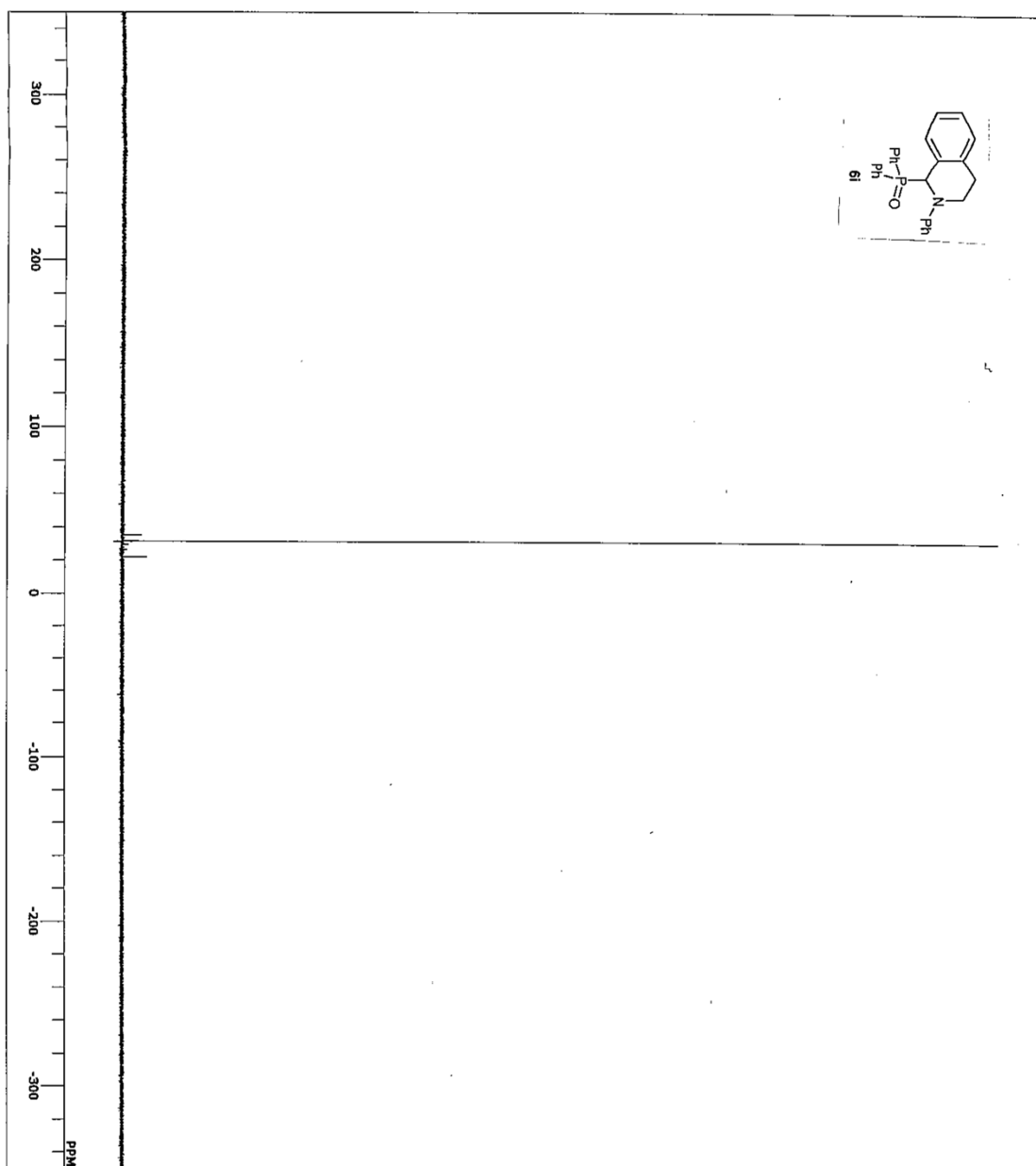
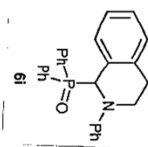


FILE D:\Documents and Settings\Kobayashi\My Desktop\WV-16
 COUNT 23-07-2013 14:45:39
 DATUM 31P
 OBNUC single, pulse, dec
 EXMOD 161.83 MHz
 OBFREQ 4.69 KHz
 OBFET 3.09 Hz
 POINT 32768
 FREQU 142045.45 Hz
 SCANS 11
 ACQTM 0.2307 sec
 PD 2.0000 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 408.6 c
 STVNT CDCL3
 EXREF 0.00 ppm
 BF 0.42 Hz
 RGAIN 54

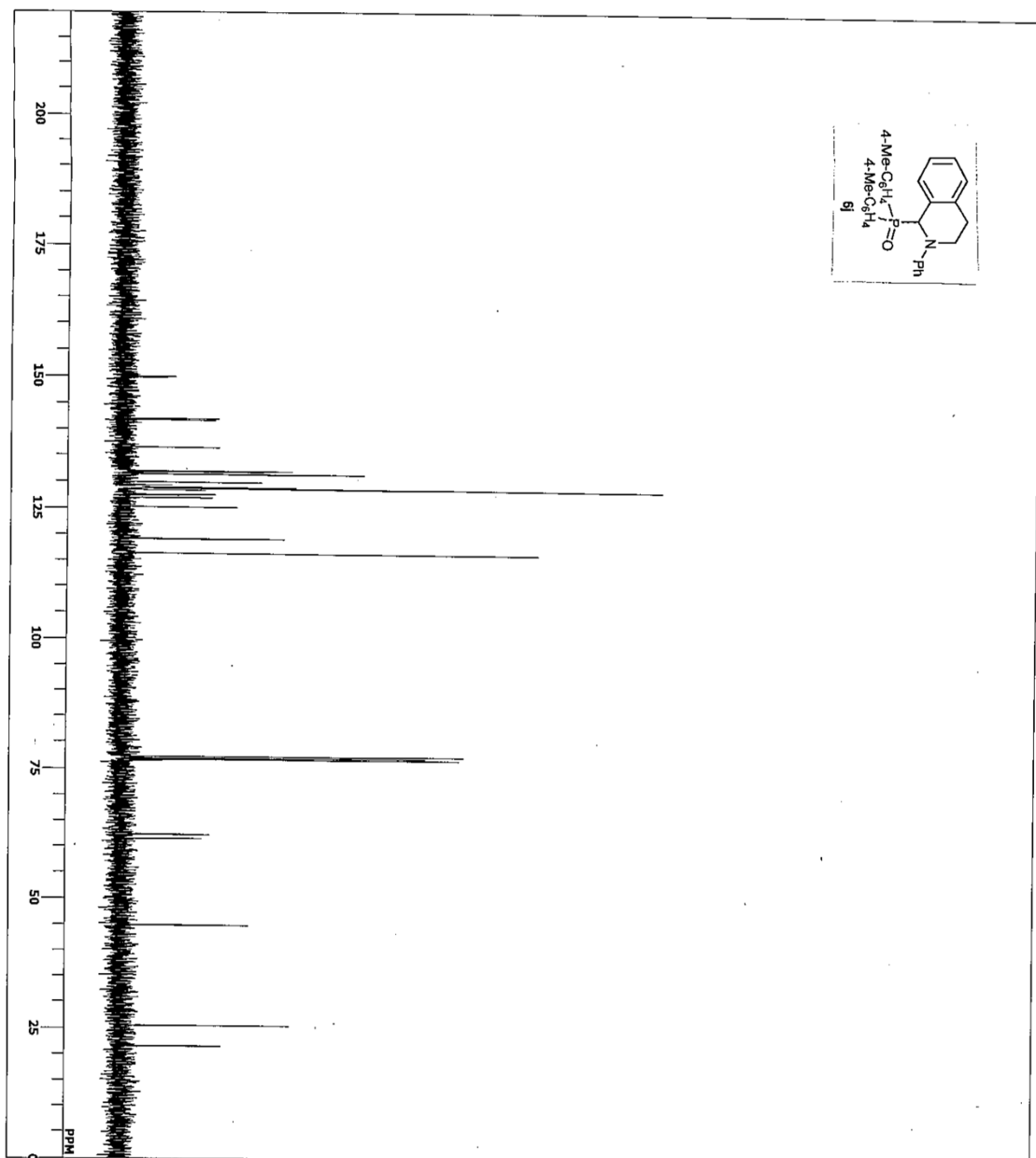
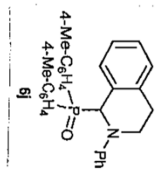


D:\Documents and Settings\Kobayashi\Desktop\WY-16
 DEFILE
 COMMENT 27-07-2013 16:41:07
 DATUM
 OBNUC 1H
 EXMOD single, pulse, ex2
 OBFRQ 399.75 MHz
 OBSET 4.19 KHz
 OBFIN 7.29 Hz
 POINT 16384
 FREQU 7503.00 Hz
 SCANS 8
 ACQTM 2.1837 sec
 PD 2.0000 sec
 PUL 6.50 usec
 INRUC 1H
 STEMP 408.2 c
 CDCL3
 EXREF 7.24 ppm
 B1 0.12 Hz
 RGAIN 28

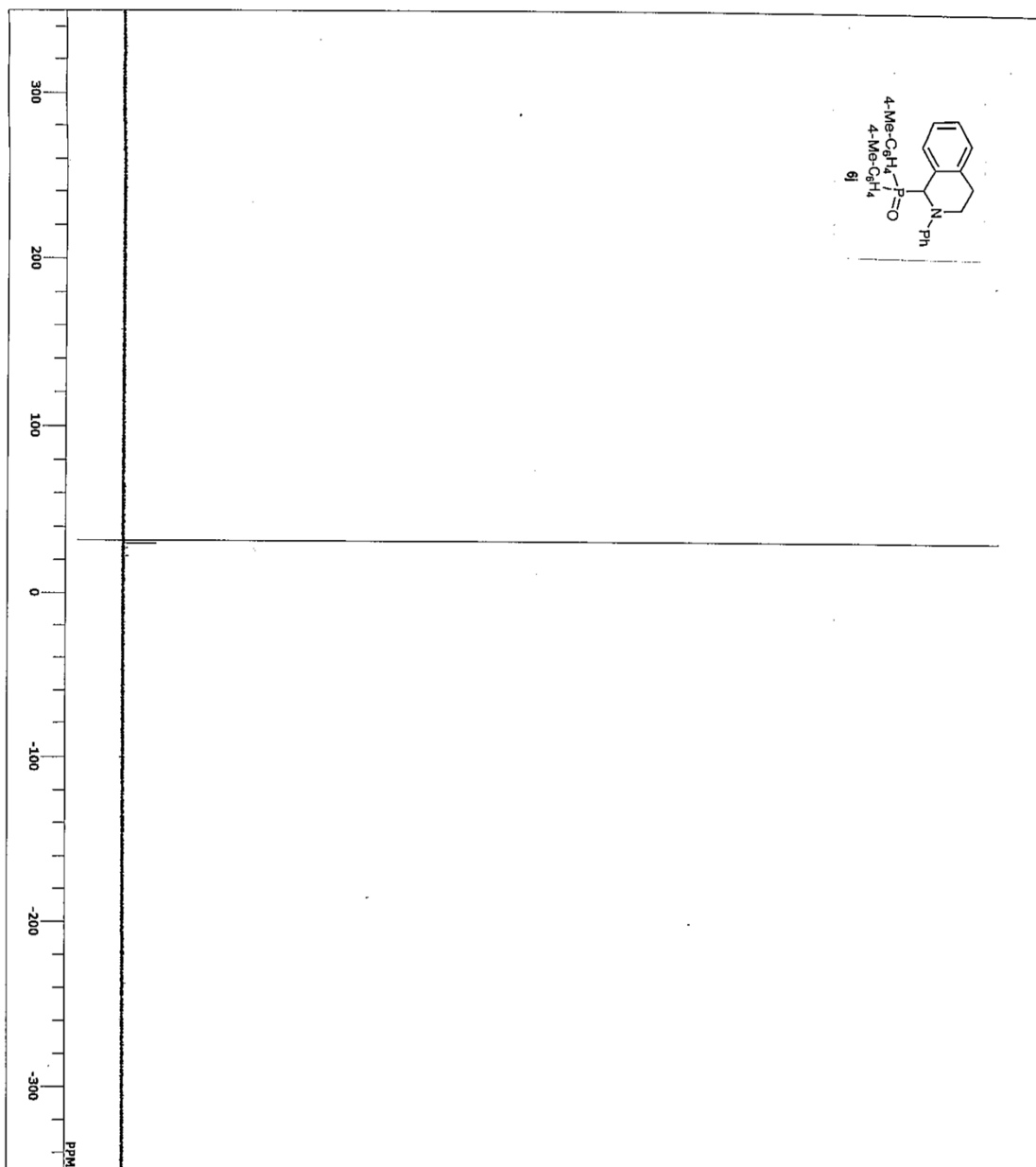
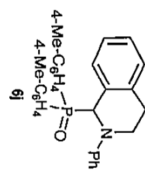




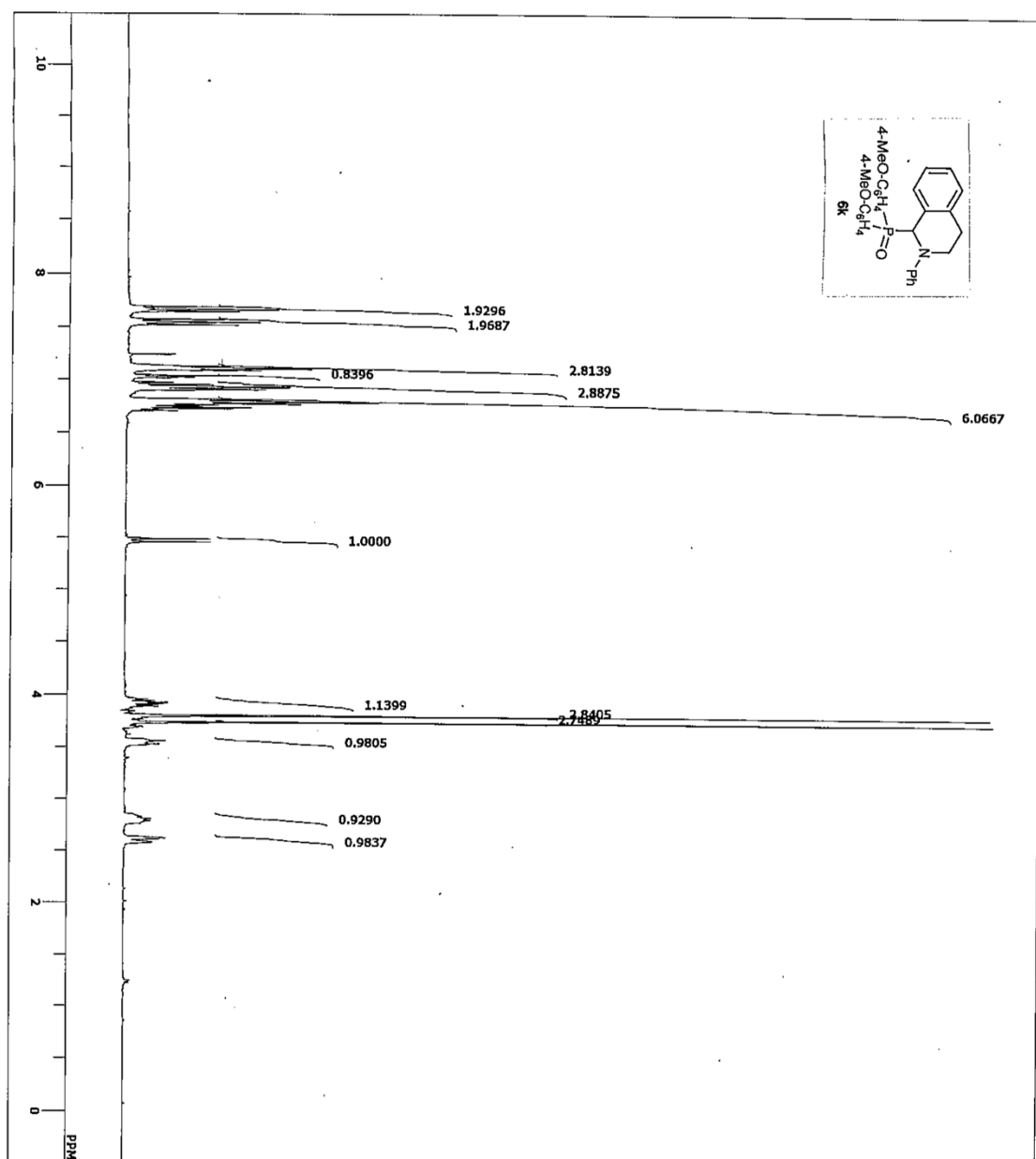
D:\Documents and Settings\Kobayashi\lab\Desktop\WV-16
 FILE 27-07-2013 16:48:18
 COUNT 31P
 DATE 27-07-2013 16:48:18
 OBNUC single, pulse, dac
 EXMOD 161.33 MHz
 OBFREQ 4.69 kHz
 OBFSET 3.09 Hz
 OBFEN 32768
 POINT 142045.45 Hz
 FREQU 11
 SCANS 11
 ACQTM 0.2307 sec
 PD 2.0000 sec
 PW1 5.80 usec
 IRNUC 1H 408.2 c
 CTEMP 0.000 ppm
 SOLVT CDCL3
 SKREF 0.42 Hz
 RGAIN 50



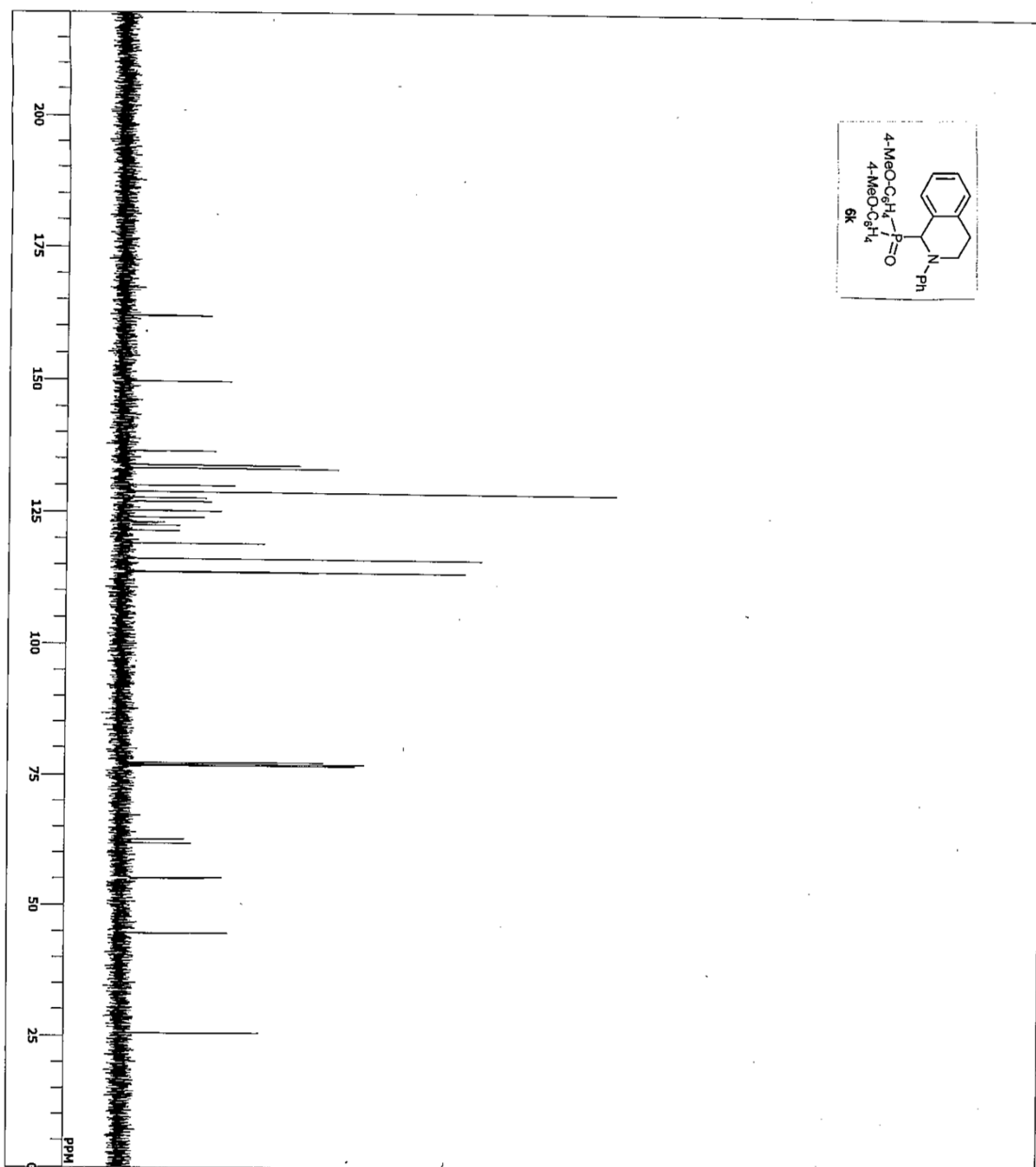
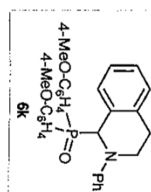
D:\Documents and Settings\Kobayashi\lab\Desktop\WY-16
 DF16 27-07-2013 16:57:35
 COMMENT 13C
 DATE 27-07-2013 16:57:35
 EXMOD single_pulse_dec
 OBNUC 13C
 OFREQ 100.63 MHz
 OBSET 5.35 KHz
 OFIN 5.86 Hz
 POINT 32768
 FREQ 31407.03 Hz
 SCANS 38
 ACQIM 1.0433 sec
 PD 2.0000 sec
 PW1 3.50 usec
 TRUNC 1H 408.2 c
 CHUP C13
 SUMP 77.00 ppm
 EXREF 0.12 Hz
 BF 58
 RGAIN 58



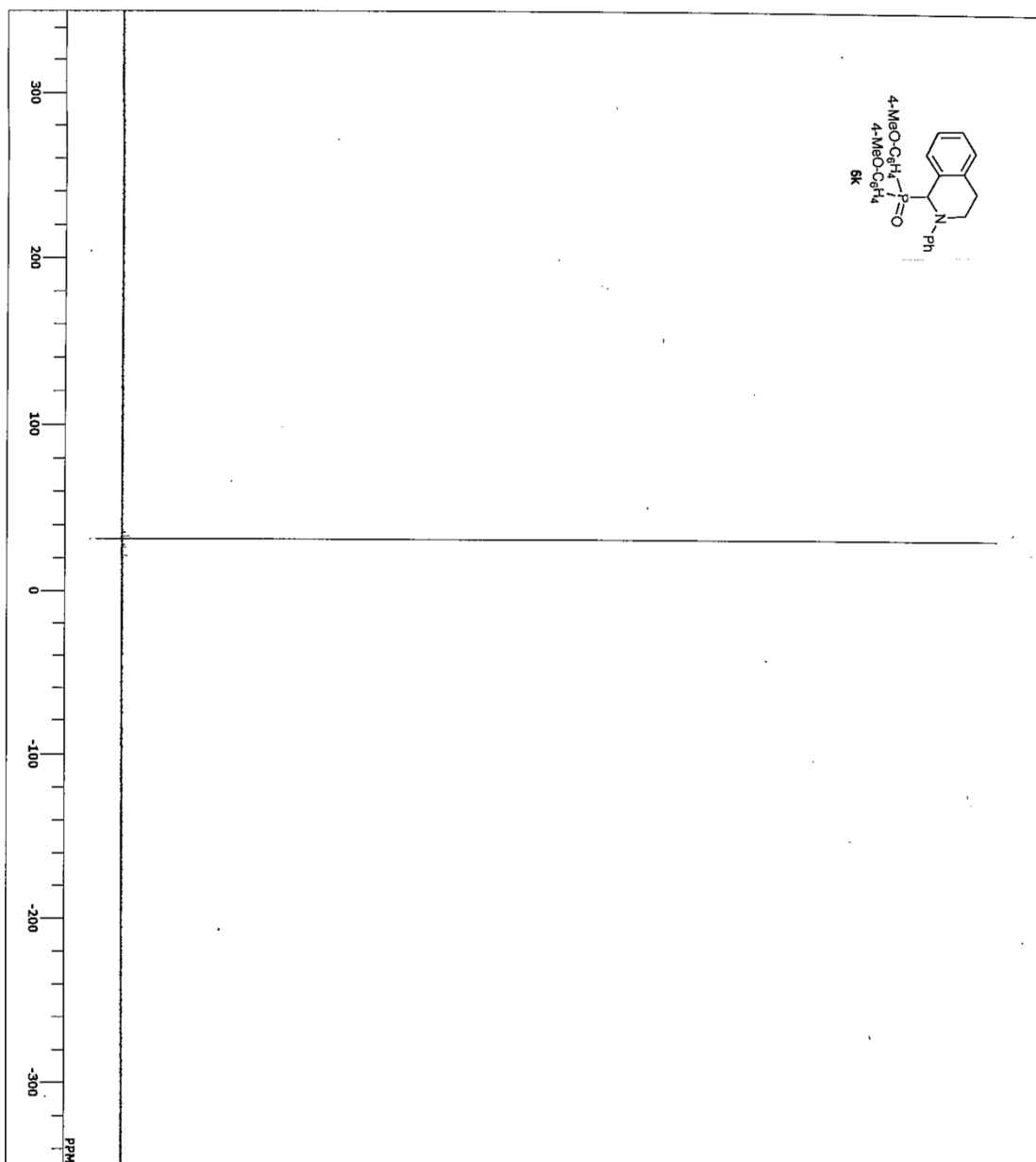
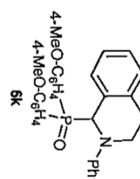
D:\Documents and Settings\Kobayashi\lab\Oasktop\WV-16
 CONT 27-07-2013 17:00:55
 DATIM 31P
 OBNUC single_pulse_dec
 EXMOD 161.83 MHz
 OBFRQ 4.69 kHz
 OBSET 3.09 Hz
 OBFIN 32768
 POINT 142045.45 Hz
 FREQU 6
 SCANS 0.2307 sec
 ACQTM 2.0000 sec
 PD 5.80 usec
 PW1 1H 408.2 c
 TRNUC CDCL3
 GTEMP 0.00 ppm
 SLENT 0.02 Hz
 EXREF 54
 RGAIN



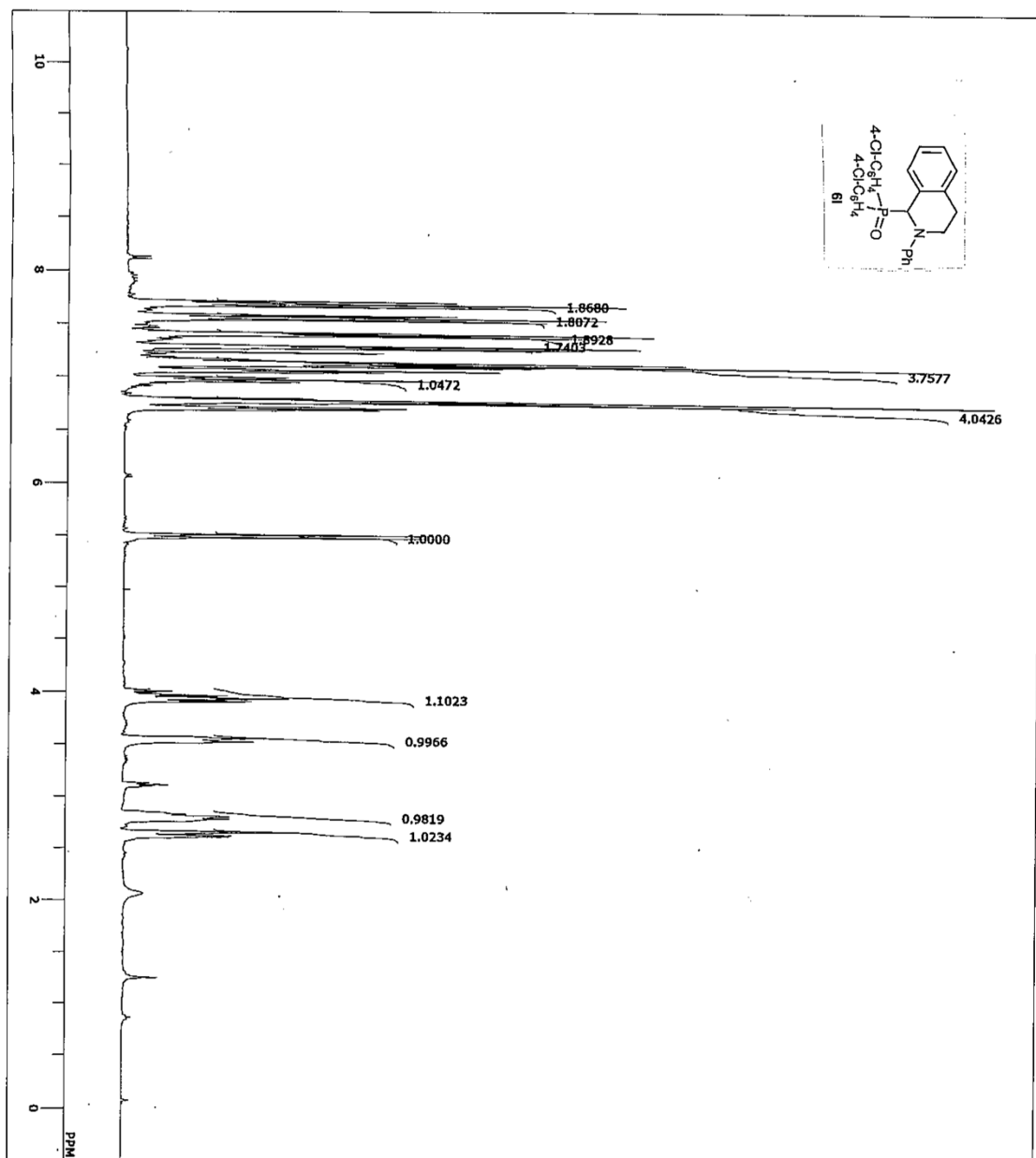
D:\Documents and Settings\Kobayashi\My Desktop\WY-16
 D:\Documents and Settings\Kobayashi\My Desktop\WY-16
 27-07-2013 17:05:08
 1H
 single_pulse.ac2
 399.78 MHz
 4.19 KHz
 7.29 Hz
 16384
 7503.00 Hz
 8
 2.1837 sec
 2.0000 sec
 6.50 usec
 1H 408.2 c
 CDCl3
 7.24 ppm
 0.2 Hz
 28



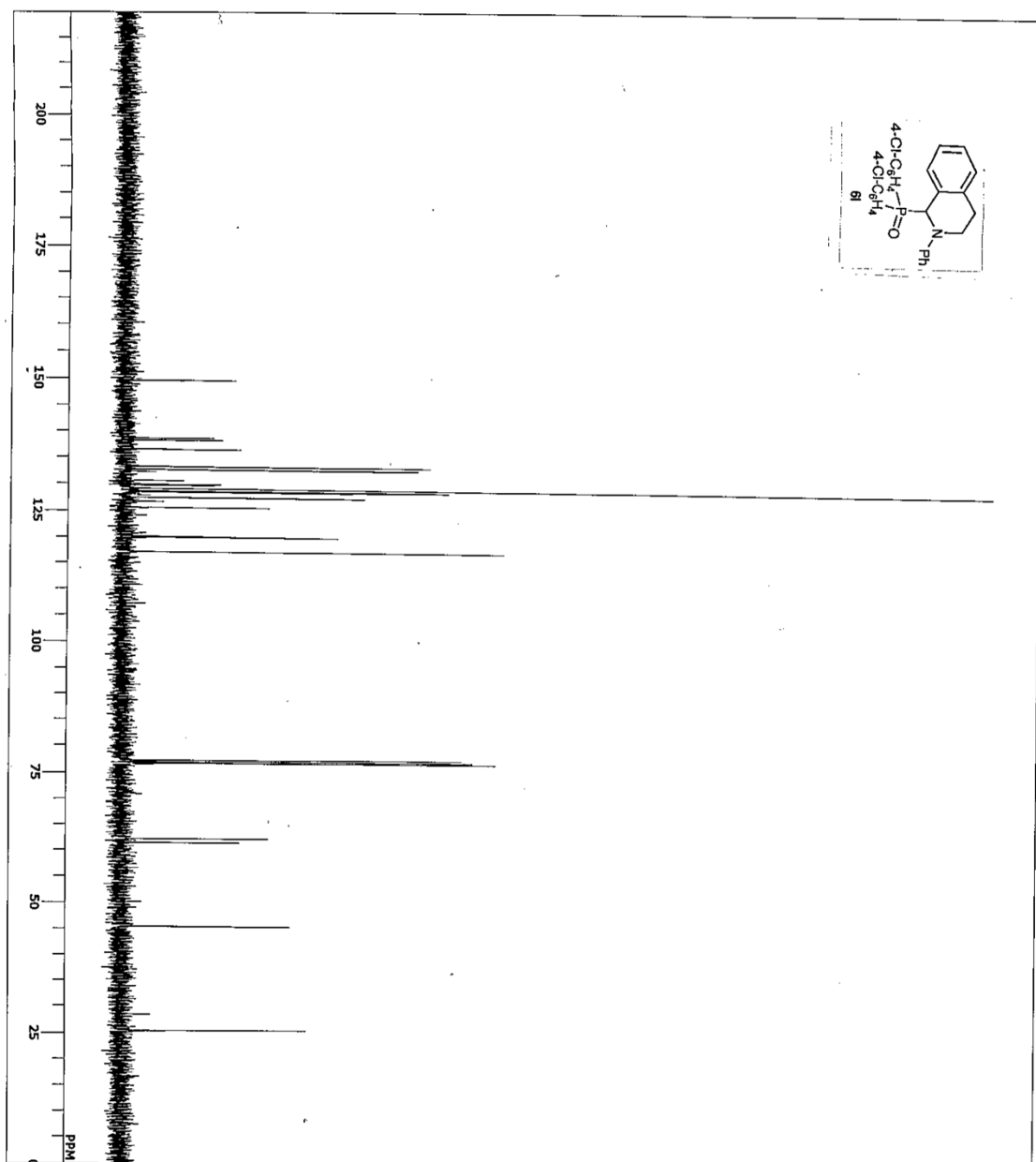
DFILE D:\Documents and Settings\Kobayashi\My Desktop\WY-16
 COMMENT 27-07-2013 17:09:58
 DATIM 13C
 OBNUC single_pulsa_dec
 EXMOD 100.53 MHz
 OBFREQ 5.35 KHz
 OBSET 5.86 Hz
 OBFIN 32768
 POINT 31407.03 Hz
 FREQU 36
 SCANS 1.0433 sec
 ACQTM 2.0000 sec
 PD 3.50 usec
 PWT 1H 408.2 c
 INRUC
 TRIMP
 STIMP 77.00 ppm
 SUNIT
 EXREF 0.12 Hz
 BF 56
 RGAIN



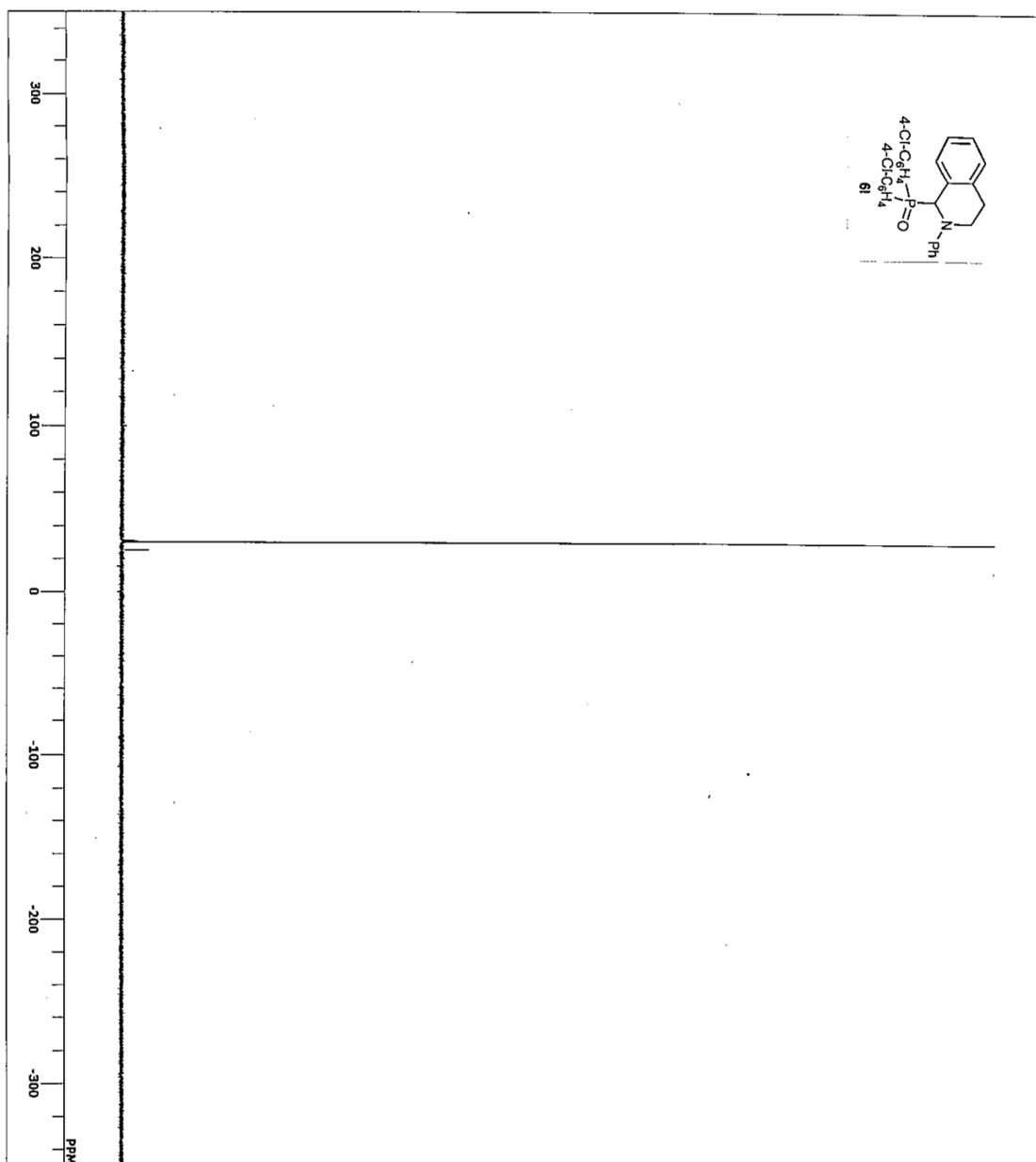
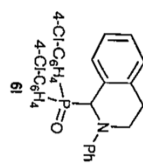
D:\Documents and Settings\Kobayashi\My Desktop\WV-16
 COUNT 31P
 DATE 27-07-2013 17:13:04
 ORNUC single, pulse, dec
 EXMOD 161.83 MHz
 OFREQ 4.69 kHz
 OBSET 3.09 Hz
 OBENT 32768
 POINT 142045.45 Hz
 FREQU 10
 SCANS 10
 ACQTM 0.2307 sec
 PD 2.0000 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 408.2 c
 SLEW CDCL3
 EXREF 0.00 ppm
 BR 0.42 Hz
 RGAIN S4



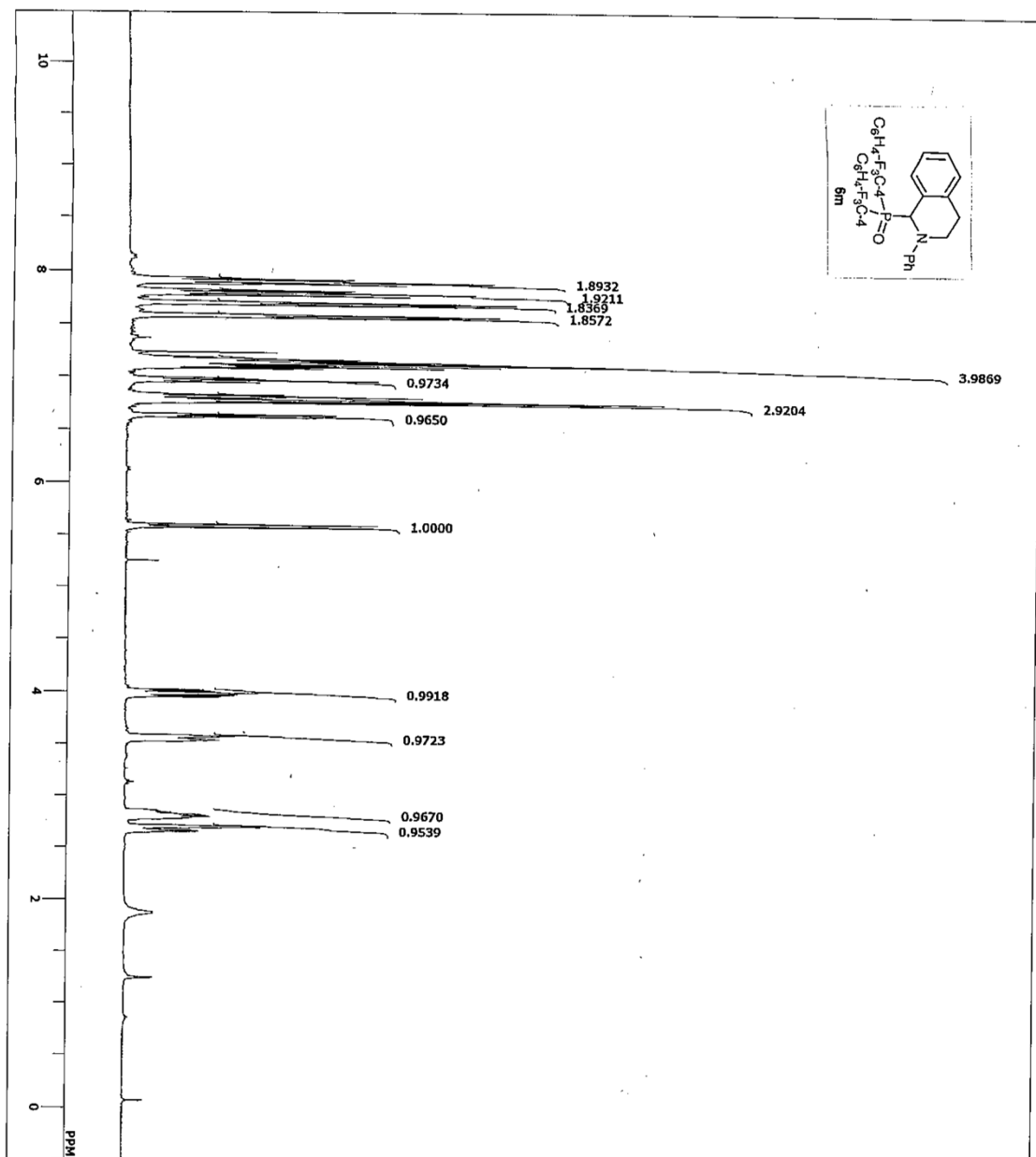
D:\Documents and Settings\kobayashilab\Desktop\WY-1619-1H.2
 FILE 14-08-2013 14:44:26
 COMNT 1H
 DATIM 1H
 OBNUC single_pulse.ex2
 EXMOD 399.76 MHz
 OBFRQ 4.19 KHz
 OBSET 7.29 Hz
 POINT 16384
 PREQU 7503.00 Hz
 SCANS 8
 SFOQIM 2.1837 sec
 PUL1 2.0000 sec
 IRNUC 6.50 usec
 CTMP 1H 400.8 c
 SLVT CDCL3
 EXREF 7.24 ppm
 BF 0.12 Hz
 RGAIN 28



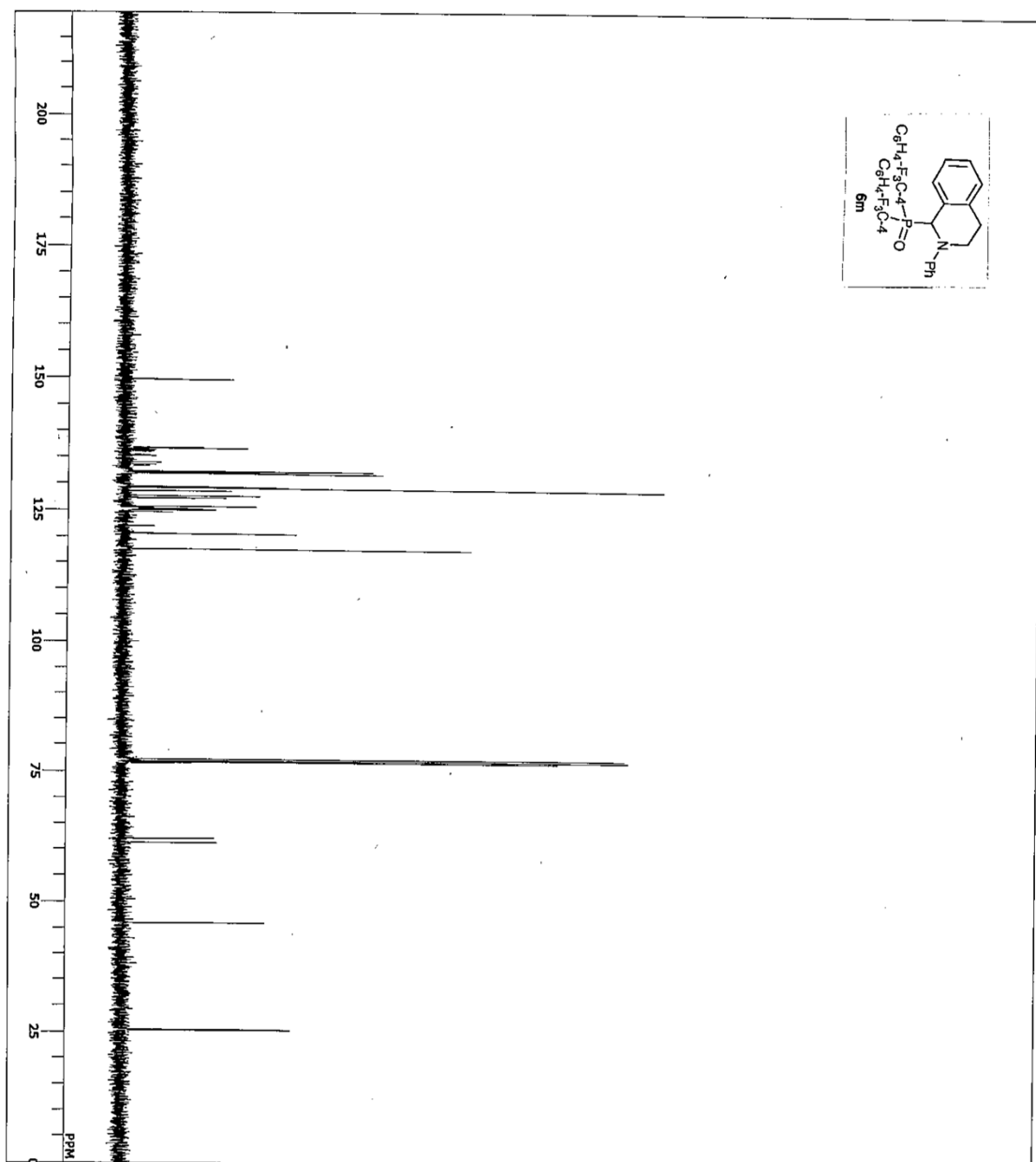
D:\Documents and Settings\Kobayashi\lab\Desktop\WY-16
 DFILE
 COMNT 14-08-2013 14:50:35
 DATIM 13C
 OBNUC single_pulse_dec
 EXMOD 100.53 MHz
 OBFREQ 5.35 KHz
 OBFEN 5.86 Hz
 POINT 32768
 FREQ 31407.03 Hz
 SCANS 71
 ACQTM 1.0633 sec
 PUL 2.0000 sec
 PM1 3.50 usec
 IRNUC 1H 401.2 c
 CTMP C13
 SLVT 77.00 ppm
 EXREF 0.12 Hz
 BF 56
 RGAIN



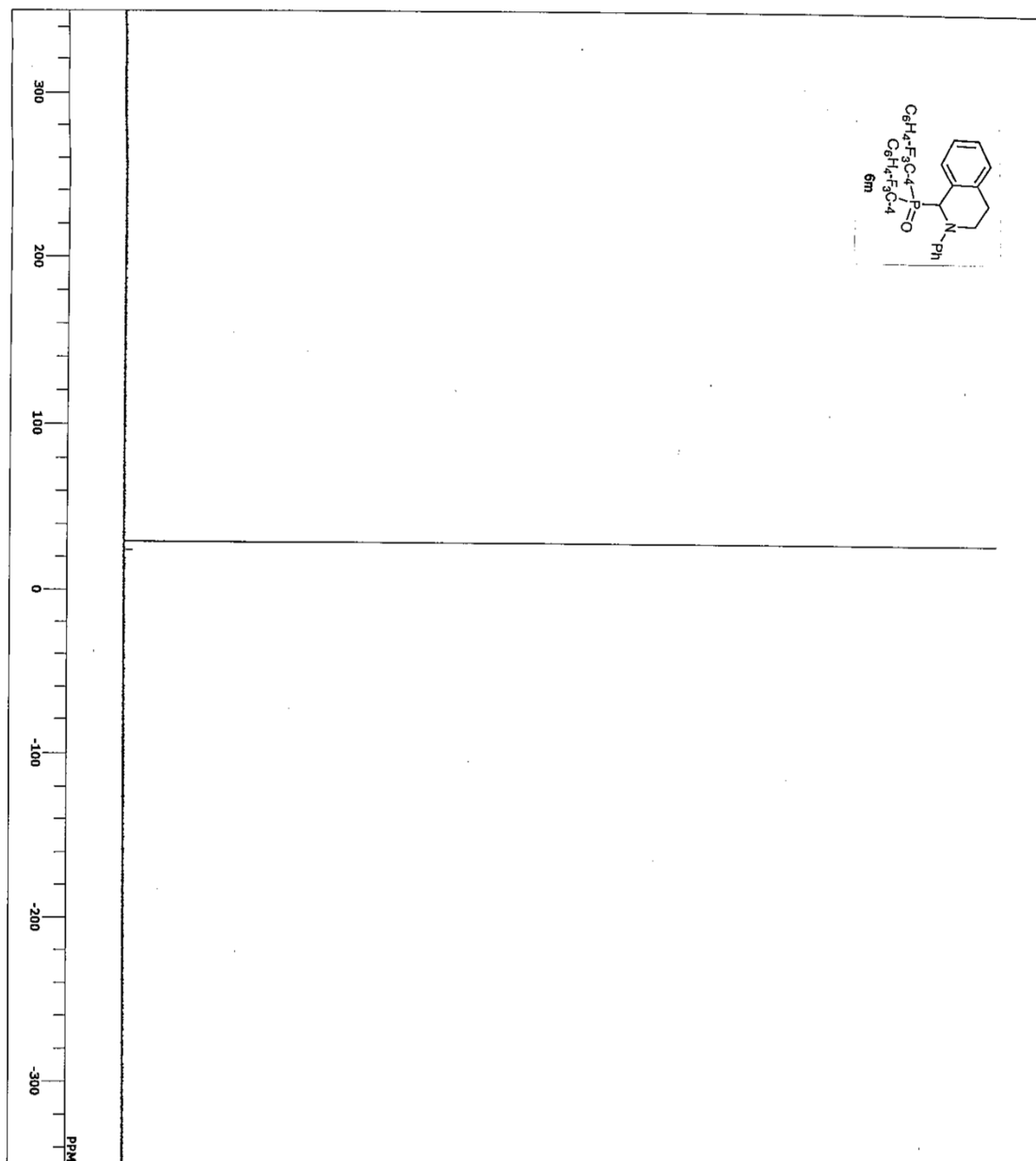
FILE D:\Documents and Settings\Kobayashi\lab\Desktop\WVY-16
 COUNT 31P
 DATE 14-08-2013 14:53:41
 ORNUC 31P
 EXMOD single, pulse, dac
 OBSFQ 161.83 MHz
 OBSRT 4.69 kHz
 OBSIN 3.09 Hz
 POINT 32768
 FREQU 142045.45 Hz
 SCANS 11
 ACQTM PD 0.2307 sec
 PD 2.0000 sec
 PW1 5.80 usec
 IRNUC 1H
 CTEMP 401.5 c
 SOLVT CDCl3
 EXREF 0.000 ppm
 B1 0.42 Hz
 RGAIN 54



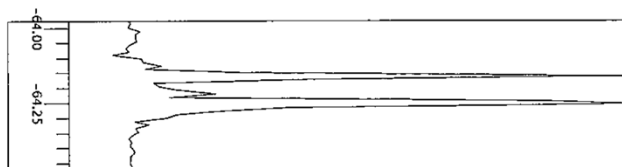
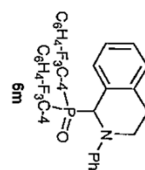
DFTL D:\Documents and Settings\skolayashlab\Desktop\WY-16
 COMNT 09-08-2013 09:41:16
 DATIM 1H
 OBNUC single_pulse.ac2
 EXMOD 399.78 MHz
 OBFREQ 4.19 KHz
 OBFIN 7.29 Hz
 POINT 16384
 FREQU 7503.00 Hz
 SCANS 8
 ACQTM 2.1837 sec
 PD 2.0000 sec
 PUL 6.50 usec
 IRNUC 1H 400.8 c
 CTEMP CDCL3
 SLENT 7.24 ppm
 SREF 0.12 Hz
 RGAIN 32



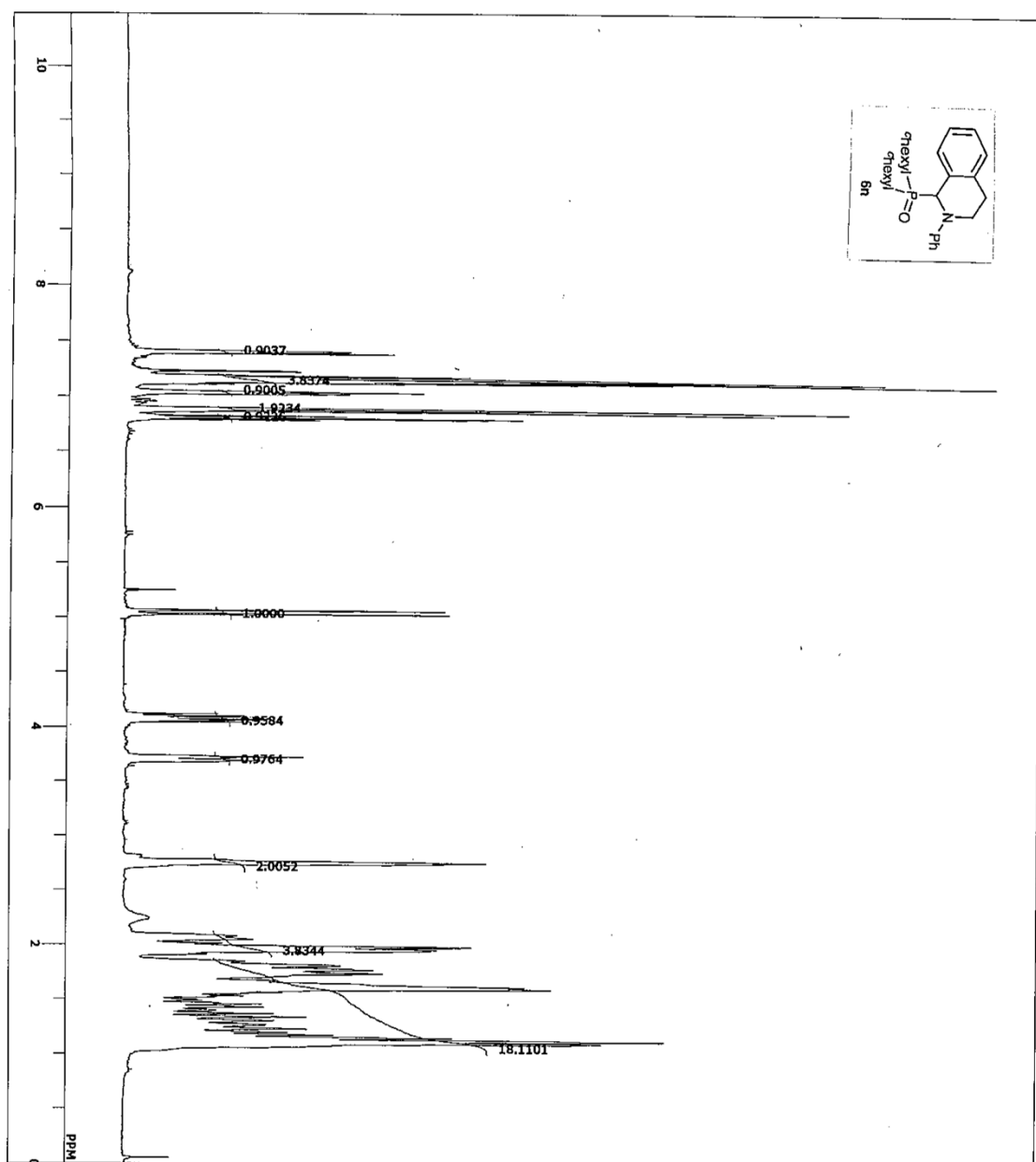
DFILE D:\Documents and Settings\Kobayashi\lab\Desktop\WY-16
 COMMENT 09-08-2013 09:55:19
 DATIM 13C
 OBNUC single_pulse_dec
 EXMOD 100.53 MHz
 OBRQ 5.35 KHz
 OBRF 5.86 Hz
 POINT 32768
 FREQ 31407.03 Hz
 SCANS 234
 ACQTM 1.0433 sec
 PM1 2.0000 sec
 PM2 3.50 usec
 IRNUC 1H 402.0 c
 CTMP CIRC3
 SLYNT 77.00 ppm
 EXREF 0.12 Hz
 BF 56
 RGAIN 56



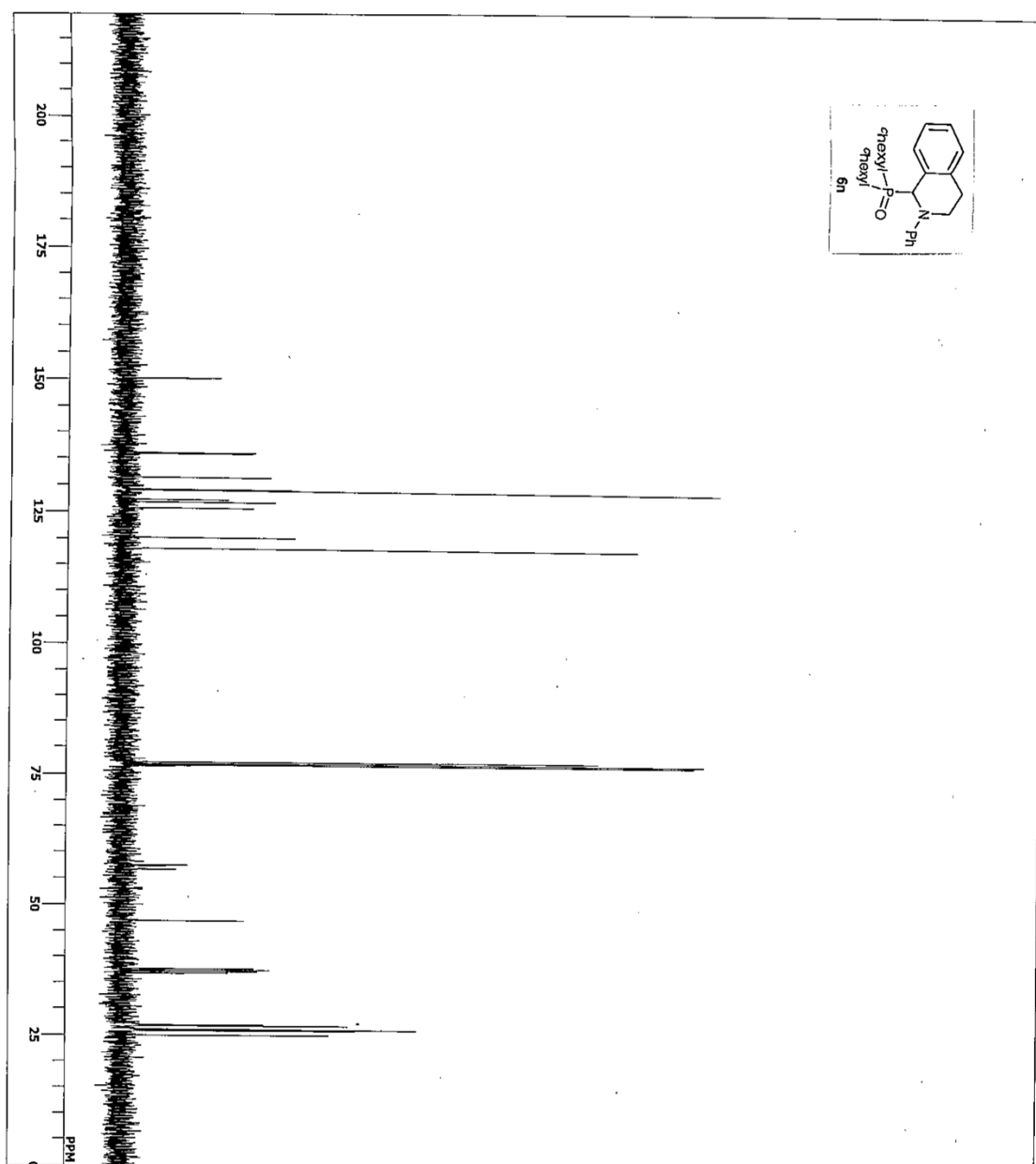
FILE D:\Documents and Settings\Kobayashi\lab\Desktop\KWT-16
 COUNT 31P
 DATE 09-08-2013 09:59:48
 NAME 31P
 EXMOD single, pulse, dec
 OBSFREQ 161.83 MHz
 OBSFREQ 4.69 KHz
 OBSFREQ 3.09 Hz
 POINT 32768
 FREQU 142045.45 Hz
 SCANS 61
 ACQTM 0.2307 sec
 PD 2.0000 sec
 PW1 5.80 usec
 IRNUC 1H
 CTMP 402.4 c
 SILVT CDCl3
 EXREF 0.00 ppm
 BF 0.42 Hz
 RGAIN 55



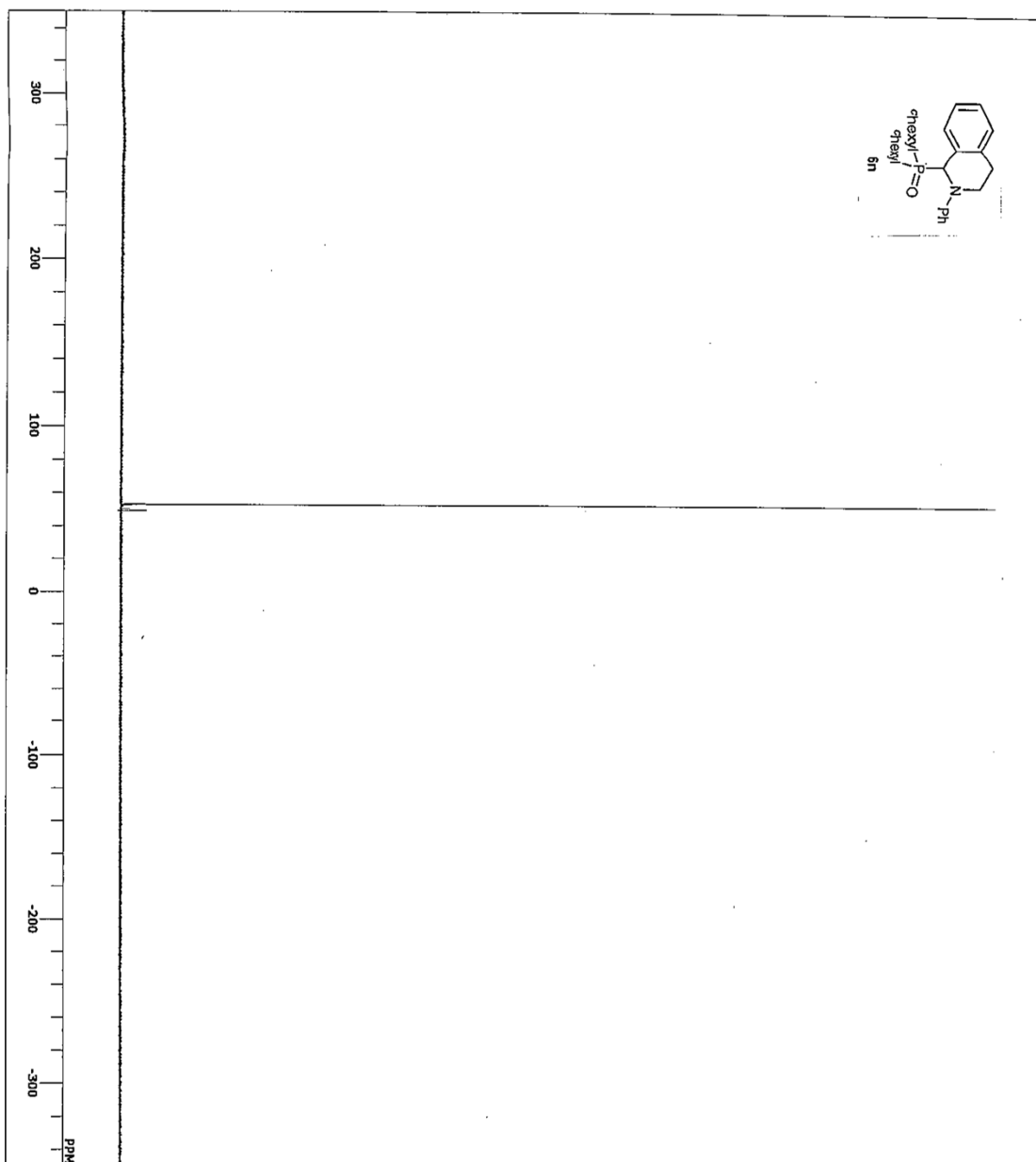
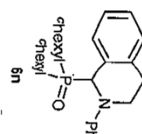
FILE D:\Documents and Settings\Kobayashi\lab\Desktop\WV-16
 COUNT 09-08-2013 10:03:20
 DATE 19F
 OBNUC single, pulse man
 EXMOD 376.13 MHz
 OBSER 3.43 KHz
 OBSIN 6.82 Hz
 POINT 32768
 FREOU 142045.45 Hz
 SCANS 8
 ACQTM 0.2307 sec
 PD 7.0000 sec
 PW1 6.70 usec
 IRNUC 19F
 CTMP 402.7 c
 SLVNT CDCl3
 EXREF -64.16 ppm
 BF 0.12 Hz
 RGAIN 50



D:\Documents and Settings\Kobayashi\Desktop\WY-16
 D:\Documents and Settings\Kobayashi\Desktop\WY-16
 09-08-2013 10:08:58
 1H
 single_pulse.ac2
 399.78 MHz
 4.19 KHz
 7.29 Hz
 16384
 7505.00 Hz
 2.1937 sec
 2.0000 sec
 6.50 urec
 403.2 c
 7.24 ppm
 0.12 Hz
 28
 RGAIN



DFILE D:\Documents and Settings\Kobayashi\lab\Desktop\WY-16
 COMMENT 09-08-2013 10:16:48
 DATIM 13C
 EXMOD single_pulse_dec
 OBRFQ 100.53 MHz
 OBSRT 5.35 KHz
 OBTIN 5.86 Hz
 POUTN 32768
 PREQU 31407.03 Hz
 SFASS 94
 ACQTM 1.6433 sec
 PD 2.000 sec
 PUL 3.50 usec
 IRNUC 1H 403.8 c
 CTMP CDCL3
 SLVNT 77.00 ppm
 EXREF 0.12 Hz
 BF 58
 RGAIN 58



FILE D:\Documents and Settings\Kobayashi\lab\Desktop\WV-16
 COMNT 09-08-2013 10:20:22
 DATIN 31P
 OBNUC single, pulse, dec
 EKMOP 161.83 MHz
 OFPRQ 4.69 kHz
 OBSET 3.09 Hz
 OBFIN 32768
 POINT 142045.45 Hz
 FREQU 28
 SCANS
 ACQTM 0.2307 sec
 PD 2.0000 sec
 PW1 5.80 usec
 IRNUC 1H 404.1 c
 CTEMP CDCL3 0.00 ppm
 SILNT 0.42 Hz
 EREF 54
 RGAIN

Part III: References

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