1	Sustainable route to produce scytonemin precursor using Escherichia coli
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### 17 **Protein Expression of ScyA, ScyB and ScyC.**

For the analysis of *scyA*, *scyB* and *scyC* expression, the recombinant plasmids pCDF-ScyA, pACYC-ScyB and pCDF-ScyAC were transformed into *E. coli* BL21 (DE3) and protein isolation from the recombinant strains were carried out as described in materials and methods. Then the protein expression was analyzed in SDS-PAGE.

	Gene name	Nucleotides (bp)	Amino acid (aa)	Protein size (kDa)
23	scyA	1875	625	68.7
	scyB	1062	354	38.9
24	scyC	969	323	35.5



- 25
- 26 Figure S1: Protein expression. Lane 1: pCDF-ScyA, pACYC-ScyB, pCDF-ScyAC, and M:
- 27 Marker.
- 28

# 29 Yellowish metabolites produced from *E. coli* SM2, SM3 and SM4 strains



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31 Figure S2: Cell pellets of *E. coli* recombinant strains and control strain cultured in 50 mL

32 of M9 minimal media (I) without and (II) with supplementation of 1 mM of tryptophan and

- 33 1 mM of tyrosine at 5 days. A) E. coli BL21 (DE3), B) E. coli SM1, C) E. coli SM2, D) E.
- 34 *coli* SM3, and E) *E. coli* SM4 strains.

# 36 UV absorbance spectra of compounds

- 37 The UV absorbance spectra of standard scytonemin, the monomer moiety of scytonemin,
- 38 and other compounds were given as follows:



40 Figure S3: UV absorbance spectra of scytonemin standard, compounds 4a, 4b, 3, 5, 6, 7, 8,
41 and 9.





46 Figure S4: A) HPLC analysis of biotransformation products from *E. coli* SM1
47 supplemented with 1 mM of tryptophan and tyrosine. Compound 2 has retention time of
48 14.2 min. B) UV absorbtion spectra of compound 2 and C) Mass analysis of compound 2.

### 50 Structural elucidation

### 51 A) NMR spectra of Compound 4

# 52 i) <sup>1</sup>H NMR spectra of 4



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# ii) <sup>13</sup>C NMR spectra of 4







#### iv) HMBC NMR spectra of 4

### HMBC NMR spectra of 4



### 64 v) HSQC spectra of 4

HSQC NMR spectra of 4



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# 67 vi) NOE spectra of 4

NOE spectra of 4





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# 72 B) NMR spectra of Compound 7

73 i) <sup>1</sup>H NMR spectra of 7





# 76 ii) <sup>13</sup>C NMR spectra of 7



13



### 82 iv) HMBC spectra of 7



### 85 v) HSQC spectra of 7



87 **Figure S6:** NMR spectra of compound **7**. i) <sup>1</sup>H-NMR, ii) <sup>13</sup>C-NMR, iii) COSY, iv) HMBC,

<sup>88</sup> and v) HSQC spectra of compound 7.

	Chemical shift (ppm)		
position	<sup>13</sup> C	$^{1}\mathrm{H}$	
1	216.36		
2	38.51	2.47, d (J = 20.88 Hz); 2.83, d (J = 20.88 Hz)	
3*	112.30		
4*	139.0		
5	118.60	7.24, d (J = 7.97 Hz)	
6	118.70	6.94, t (J = 7.42 Hz)	
7	121.00	7.07, t (J = 7.42 Hz)	
8	112.30	7.42, d (J = 8.24 Hz)	
9*	122.98		
10		10.92, s	
11*	139.50		
12	63.36		
13	36.34	3.30, d (J = 13.46 Hz); 3.48, d (J = 13.46 Hz)	
14	125.87		
15	130.55	6.45, d (J = 8.79 Hz)	
16	114.31	6.25, d (J = 8.79 Hz)	
17	155.34		
18		8.98, s	

**Table S1**. Assignment of <sup>1</sup>H and <sup>13</sup>C signals for compound **7** 

91 \*assignments of carbon signals were assisted by chemical shift estimation using
92 chemdraw software.

93

The <sup>13</sup>C spectrum confirmed the presence of a symmetric molecule, since only 16 94 95 carbon signals is present in a 36 carbon molecule (16 carbon signal plus 2 carbon signal 96 from the symmetric phenyl ring times 2 gives 36 carbon). The indole and phenyl structures 97 are readily apparent, since typical proton signals multiplicities and COSY correlations are 98 present. A ketone group resonating at 216.36 ppm is also present. The HSQC spectrum 99 detects one aliphatic quaternary carbon and two aliphatic CH<sub>2</sub> groups (one attached to the 100 phenyl group and another one in the ring). The only possible explanation is that molecule **3** 101 dimerized via the carbon attached to the phenyl group as depicted in Figure S6.1. All the acquired NMR data is consistent with the structure depicted in Figure S6.1 and all of the 102 103 carbon and proton signals were assigned in Table S1.





Figure S6.1. Structure of the compound 7 with atom numbering

# 107 C) NMR spectra of Compound 6

# 108 i) <sup>1</sup>H NMR spectra of compound 6.



# 111 ii) <sup>13</sup>C-NMR spectra of compound 6.



iii) COSY, spectra of compound 6.



### 117 iv) HMBC spectra of compound 6.

### HMBC NMR spectra of 6



### 120 v) HSQC spectra of compound 6.







	Chemical shift (ppm)			
position	<sup>13</sup> C	$^{1}\mathrm{H}$		
1	217.10			
2	20.22	3.38, d (J = 20.81 Hz);		
2	30.33	2.70, d (J = 20.81 Hz)		
3	na			
4	na			
5	118.70	7.36, d (J = 8.21 Hz)		
6	118.82	6.98, t (J =7.86 Hz)		
7	120.76	7.07, t (J = 7.69 Hz)		
8	118.70	7.36, d (J = 8.21 Hz)		
9	na			
10		11.34, s		
11	na			
12	56.62			
12	40.89	3.71, d (J = 13.32 Hz);		
15		3.39, d (J = 13.32 Hz)		
14	126.40			
15	130.39	6.60, d (J = 8.39 Hz)		
16	114.45	6.38, d (J = 8.46 Hz)		
17	155.40			
18		9.06, s		
19		11.14, s		
20	123.57	7.44, d (J = 2.51 Hz)		
21	na			
22	na			
23	118.70	6.71, nd		
24	118.70	6.71, nd		
25	120.88	6.97, nd		
26	111.46	7.33, d (J = 8.21 Hz)		
27	na			

Table S2. Partial assignment of <sup>1</sup>H and <sup>13</sup>C signals for compound 6

na – Not assigned

nd - can not determine (due to overlap)

127 128

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130 NMR data (1H,  ${}^{13}$ C APT, COSY, HSQC and HMBC) was acquired. The spectral data 131 showed signals that can be interpreted as two indole like structures, one phenyl group, one 132 ketone, two CH<sub>2</sub> groups and one quaternary aliphatic carbon. These structures with the 133 respective carbon and proton information are shown in Figure S7.1.

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Figure S7.1. Partial structures that appear to be present in compound 6 with carbon and proton chemical shift
information. Carbon chemical shifts are in black font, proton in red font. Cq – aliphatic quaternary carbon
atom.

140 It can be observed from the chemical shift values that there is considerable signal 141 overlap both in carbon and proton (noticeably three resonances at 118.70 ppm in carbon, 142 two resonances at 7.36 and at 6.71 in proton). The relatively high proportion of quaternary 143 carbons in these molecules does not allow for a straightforward sequential assignment, and 144 the only tool available to assign those resonances and to link the several pieces of the 145 structure comes from the HMBC data. Since the HMBC can detect connectivity 2, 3, 4 (and 146 sometimes even 5) bonds apart, in cyclic molecules like these the information becomes 147 ambiguous. Considering the available information the structure depicted in Figure S7.2 is 148 consistent with the data.





Figure S7.2. Possible structure of the compound 6 with atom numbering

151 The acquired NMR data is consistent with the structure depicted in Figure S7.2, 152 however, signal overlap precluded the complete analysis of the HMBC spectrum and 153 therefore the complete assignment of the carbon signals was not possible.

# 155 D) NMR spectra of Compound 8

156 i) <sup>1</sup>H NMR spectra of compound 8.



<sup>1</sup>H-NMR spectra of 8

ii) COSY spectra of compound 8



- iii) HSQC spectra of compound 8



### 167 iv) HMBC spectra of compound 8

### HMBC NMR spectra of 8



169 Figure S8: NMR spectra of compound 8. i) <sup>1</sup>H-NMR, ii) COSY, iii) HMBC, and iv) HSQC spectra of

<sup>170</sup> compound **8**.

	Chemical shift (ppm)				
position	<sup>13</sup> C	<sup>1</sup> H	position	<sup>13</sup> C	<sup>1</sup> H
1	215.98		1'	216.16	
2	38.37	2.82, d (J= 12.51 Hz); 2.47, (#)	2'	38.37	2.82 d (J= 12.51 Hz) ; 2.47 (#)
3*	112.25		3'*	112.25	
4*	139.05		4'*	139.05	
5	118.72	7.24, d (J= 7.52 Hz)	5'	118.72	7.24, d (J= 7.52 Hz)
6	118.88	6.94, t (J= 7.52 Hz)	6'	118.88	6.94, t (J= 7.52 Hz)
7	121.07	7.08, t (J= 7.52 Hz)	7'	121.07	7.08, t (J= 7.52 Hz)
8	112.39	7.42, d (J= 7.88 Hz)	8'	112.39	7.42, d (J= 7.88 Hz)
9*	122.85		9'*	122.85	
10		10.99 s	10'		10.96, s
11*	139.30		11'*	139.00	
12	63.39		12'	63.18	
13	36.13	3.51, d (J=13.57 Hz); 3.33, (#)	13'	36.22	3.62, d (J = 13.57 Hz); 3.44, d (13.57 Hz)
14	125.75		14'	133.43	
15	130.50	6.47, d (J= 8.51 Hz)	15'	130.50	6.96, d (J= 8.60 Hz)
16	114.32	6.26, d (J= 8.51 Hz)	16'	120.75	6.66, d (J= 8.60 Hz)
17	155.24		17'	148.61	
18		8.98, s	18'		
			19'	168.71	
			20'	20.66	2.11, s

172 **Table S3:** Assignment of <sup>1</sup>H and <sup>13</sup>C signals for compound number 8

\*assignments of carbon signals were assisted by chemical shift estimation using chemdraw software. # Peak structure cannot be determined because of overlap with solvent lines

A similar analysis was performed with this sample resorting to <sup>1</sup>H, COSY, HSQC 176 and HMBC. In this case the 1D <sup>13</sup>C APT spectrum was not acquired because of lack of 177 178 signal, the sample is not concentrated enough to allow direct carbon detection. However, 179 carbon data was obtained from the analysis of the indirect detection spectra (HSQC and 180 HMBC). This was possible partly because of the structural similarities between this 181 compound and the previously analysed sample (compound 7). From the gathered data an 182 indole and phenyl structures are apparent, with typical proton signals multiplicities and 183 COSY correlations. A ketone and a methyl signals are also present. As with compound 7, there is a CH<sub>2</sub> and a quaternary carbon in the five member ring and the link between the 184 185 two major portions of the molecule is done via that quaternary carbon. The NMR data is 186 consistent with the structure depicted in Figure S8.1.



Figure S8.1. Structure of the compound 8 with atom numbering

As expected from the structural similarity, the spectra of compound 8 present features which are similar to those of compound 7; however, in this case the symmetry of the molecule is broken. Some signals (like the ones coming from the indole group) overlap completely while others (like 15 and 15' or 16 and 16') are well differentiated and others still (like 1 and 1' or 12 and 12') are so close that it is almost impossible to distinguish them. Nevertheless, the analysis of the HSQC and HMBC together with the gathered knowledge from the assignment of compound 7 allowed to assign the proton and carbon resonances (Table S3).

- E) NMR spectra of Compound 9
- 201 i) <sup>1</sup>H NMR spectra of compound 9.



206 ii) COSY NMR spectra of compound 9.



COSY NMR spectra of 9

# 210 iii) HMBC NMR spectra of compound 9.

# HMBC NMR spectra of 9



- 213
- iv) HSQC NMR spectra of compound 9.



216 Figure S9: NMR spectra of compound 9. i) <sup>1</sup>H-NMR, ii) COSY, iii) HMBC, and iv) HSQC spectra of

- 217 compound **9**.
- 218

		Chemical shift (ppm)				
position	<sup>13</sup> C	<sup>1</sup> H	position	<sup>13</sup> C	$^{1}\mathrm{H}$	
1	216.24		1'	216.02		
2	38.48	2.82, d (J=12.51Hz) ; 2.45, (#)	2'	38.48	2.81, d (J=12.51Hz); 2.45, (#)	
3*	112.20		3'*	112.20		
4*	138.98		4'*	138.98		
5	118.72	7.23, d (J=7.52 Hz)	5'	118.72	7.23, d (J=7.52 Hz)	
6	118.82	6.94, t (J=7.52 Hz)	6'	118.82	6.94, t (J=7.52 Hz)	
7	121.08	7.07, t (J=7.52 Hz)	7'	121.08	7.07, t (J=7.52 Hz)	
8	112.27	7.42, d (J=7.88 Hz)	8'	112.27	7.42, d (J=7.88 Hz)	
9*	122.87		9'*	122.87		
10		11.00, s	10'		10.94, s	
11*	139.29		11'*	139.18		
12	63.34		12'	63.27		
13	36.17	3.51, d (J=13.57 Hz); 3.34 (#)	13'	36.92	3.61, d (J=13.57 Hz); 3.44, d (J=13.57 Hz)	
14	125.69		14'	136.04		
15	130.63	6.47, d (J=8.60 Hz)	15'	129.65	6.70, d (J=8.60 Hz)	
16	114.34	6.26, d (J=8.60 Hz)	16'	127.51	6.90, d (J=8.60 Hz)	
17	155.29		17'	126.08	6.91, d (J=8.60 Hz)	
18		8.98.8				

219 **Table S4** Assignment of <sup>1</sup>H and <sup>13</sup>C signals for compound 9

\*assignments of carbon signals were assisted by chemical shift estimation using chemdraw software. # peak structure cannot be determined because of overlap with solvent lines

223 A similar analysis was performed with compound 9 resorting to <sup>1</sup>H, COSY, HSQC and HMBC. Also, in this case the 1D <sup>13</sup>C APT spectrum was not acquired because of lack 224 225 of signal and carbon data was obtained from the analysis of the indirect detection spectra (HSQC and HMBC). The spectral features of this compound were extremely similar to 226 those of compound  $\mathbf{8}$  with the exception that the acetyl group is not present and that a 227 benzene ring like structure is visible. All the other characteristics, like the CH<sub>2</sub> group and 228 229 the quaternary carbon in the five member ring, are same. The NMR data is consistent with 230 the structure depicted in Figure S9.1.

Like compound **8**, the compound **9** is also not symmetrical. Some signals (like the ones coming from the indole group) overlap completely while others (like 15 and 15' or 16 and 16') are well differentiated and others still (like 1 and 1' or 12 and 12') are so close that it is almost impossible to distinguish them. Nevertheless, the analysis of the HSQC and HMBC together with the gathered knowledge from the assignment of compound 7 and 8

allowed to assign the proton and carbon resonances (Table S4).



# 3 F) NMR spectra of Compound 5







The NMR data showed signals that can be interpreted as two indole like structures, one phenyl group, one ketone, two  $CH_2$  groups and one quaternary aliphatic carbon. These structures with the respective carbon and proton information are shown in figure S 9.1.



257

Figure S10.1. Partial structures that appear to be present in compound 5 with carbon and proton chemical shift information. Carbon chemical shifts are in blue, proton in red. Cq – aliphatic quaternary carbon atom.

These partial structures are similar to those of compound **6**. Although both compounds **5** and **6** have same mass, they differ in retention time in HPLC and UV absorption spectra. Based upon these information, we proposed the structure of compound **5** could be the isomer of compound **6** shown in Figure S10.2.

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Figure S10.2. Proposed Structure of the compound 5.