## Supporting Information

# Acylation of cellulose in reversible ionic liquids

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#### 1. Experimental Details

#### **1.1 Materials**

Microcrystalline cellulose (MCC) with a degree of polymerization (DP) of 240 was obtained from Aladdin Commerce Reagent Co., Ltd. and dried at 60 °C for 24 h in vacuum oven before use. CO<sub>2</sub> with a purity of >99.999% was supplied from Beijing Bei Temperature Gas Factory. DMSO was purchased from Beijing Chemcial Reagent Co., Ltd. Methanol was purchased from Tianjing Damao Chemcial Reagent Factory. Acetic anhydride (Ac<sub>2</sub>O) was purchased from Sinopharm Chemcial Reagent Co., Ltd. 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU), propionic anhydride (Pr<sub>2</sub>O) and

butyric anhydride (Bu<sub>2</sub>O) were purchased from Aladdin Commerce Reagent Co., Ltd. and used as received. All chemical reagents were analytical reagents and used without further purification.

#### **1.2 Measurements**

The IR spectra of the samples were recorded with a Fourier transform IR spectrometer (FTIR PE-2000, the United States). The test specimens were prepared by the KBr-disk method.

<sup>1</sup>H NMR spectrum were recorded on a Bruker AV 500 spectrometer with 16 scans in DMSO-d<sub>6</sub> or CDCl<sub>3</sub>. The DS of cellulose ester was calculated by Eq. (1):

$$DS = \frac{7 * (I_{-CH3})}{3 * I_{H,AGU}}$$

Where  $I_{CH3}$  is the peak integral of methyl protons and  $I_{AGU}$  is the peak integral of anhydroglucose unit.

The <sup>13</sup>C NMR spectra of cellulose ester in DMSO-d<sub>6</sub> were recorded at 100 °C on a Bruker AV-500 instrument. Each spectrum was obtained with an accumulation of 1000 scans. Wide-angle X-ray powder diffraction (WAXD) was performed by X'pert Pro-1 X-ray diffractometer using Ni-filtered Cu KR radiation (40 kV, 30 mA) with 5°/min scanning rate at room temperature. Diffraction intensity was measured in a range of  $2\theta$ =5-60°.

### 1.3 Dissolution of cellulose in *in situ* formed [DBUH][O<sub>2</sub>COCH<sub>3</sub>]/DMSO (X<sub>RILs</sub>=0.2)

A mixture of MCC (1.3 g), DMSO (14.4 g, 184 mmol), methanol (1.5 g, 46 mmol) and 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) (7.0 g, 46 mmol) was added into a high pressure reactor, and then 0.5 MPa of CO<sub>2</sub> was introduced into the system to stabilize for 5 min till the pressure of CO<sub>2</sub> stabilize. Then the solution was heated to 60 °C for 3 h with vigorous magnetic stirring. When the temperature was decrease to

room temperature, the CO<sub>2</sub> was released slowly, and a 5 wt% of cellulose [DBUH]  $[O_2COCH_3]/DMSO (X_{RILs}=0.2)$  solution was obtained, and was ready for subsequent reactions. The dissolved cellulose can be regenerated by adding methanol in to the solution, and then washed by methanol (100 mL X 3) to remove the solvents and dried in a vacuum oven at 60 °C for about 24 h for WXRD analysis.

#### **1.4 Esterification Procedure**

A typical esterification procedure was used as follows: Acetic anhydride (2.86 g, 28 mmol) was dropped with a pipet into 10 g of 5 wt% of cellulose  $[DBUH][O_2COCH_3]$  (X<sub>RILs</sub>=0.2) solution in a flask equipped with a mechanical stirrer under N<sub>2</sub> atmosphere at room temperature, and then the reaction temperature was increased to 80 °C for 5 h with vigorous mechanical stirring. After the reaction, the cellulose acetate was precipitated by adding 5-fold amount of methanol, and separated by filtration. Cellulose acetate (0.63 g, 90% yield, DS=2.25) was obtained after subsequent wash with water (100 mL x 3) with distilled water, and dried in a vacuum oven at 60 °C for about 24 h.

#### **1.5 Supplemental Tables and Figures**

Code	Total DS	Distribution of substituents <sup>a</sup>		
		C <sub>6</sub>	$C_2$	C <sub>3</sub>
<b>CA-9</b>	0.98	0.71	0.16	0.11
<b>CA-10</b>	2.25	0.91	0.87	0.47
<b>CA-6</b>	2.51	0.99	0.89	0.63
<b>CA-7</b>	2.71	1.0	0.91	0.80

Table S1 Primary evaluation of the distribution trend of acetyl moiety among the three OH groups of AGU in CAs

<sup>a</sup>The results were primary calculated from the quantitative integration of acetyl methyl protons area of the <sup>1</sup>H NMR spectra.



Figure S1 Comparative <sup>1</sup>H NMR spectra of A: DBU+Methanol (1:1, molar ratio) and B: the formed [DBUH][O<sub>2</sub>COCH<sub>3</sub>] in DMSO-d6 after CO<sub>2</sub> capture



Figure S2 Comparative <sup>13</sup>C NMR spectra of A: DBU+Methanol (1:1, molar ratio) and B: the formed [DBUH] [O<sub>2</sub>COCH<sub>3</sub>] in DMSO-d6 after CO<sub>2</sub> capture



Figure S3 <sup>1</sup>H NMR spectrum of methyl acetate in CDCl<sub>3</sub>



Figure S4 <sup>13</sup>C NMR spectrum of methyl acetate in CDCl<sub>3</sub>



Figure S5 The flowchart of the whole procedure used in this study



Figure S6 Comparative <sup>1</sup>H NMR spectra of fresh DBU and recycled DBU.



Figure S7 Comparative <sup>13</sup>C NMR spectra of fresh DBU and recycled DBU.



Figure S8 <sup>1</sup>H NMR spectra of fresh DMSO and recycled DMSO.



Figure S9<sup>13</sup>C NMR spectra of fresh DMSO and recycled DMSO.