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Environmentally benign glycosylation of aryl pyranosides and aryl/alkyl furanosides demonstrating the versatility of thermostable CGTase from *Thermoanaerobacterium sp*.

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General Experimental Details

All reagents were purchased from commercial sources and were used without further purification unless noted. Thermonanerobacter sp. CGTase (Toruzyme® 3.0 L) was a gift of Novozymes A/S (Bagvaerd, Denmark). Unless otherwise stated, all reactions were monitored by TLC on Silica Gel 60 F₂₅₄. TLC spots were detected under 254 nm light or by staining with cerium ammonium molybdate solution. Column chromatography was performed on Silica Gel (40-63 µm). Optical rotations were measured at 20 °C. NMR spectra were recorded at 400 MHz for ¹H and 100 MHz for ¹³C. Chemical shifts are given in δ units (ppm) and referenced to MeOD (3.31 ppm). Coupling constants J were calculated in Hertz (Hz). Proton and carbon NMR peaks were unambiguously assigned by COSY (double quantum filtered with gradient pulse for selection), HSQC (gradient echo-anti echo selection and shape pulse) and HMBC (echo-anti echo gradient selection, magnitude mode) correlation experiments. For each oligosaccharide isolated, the reducing end (bearing PNP or alkyl chain) was quoted as "a", and the letter increased toward the non-reducing end (for example the sugar after was quoted as "b"). High Resolution Mass were measured by electrospray with a MS/MS ZabSpec TOF Micromass using *m*-nitrobenzylic alcohol as a matrix and accelerated caesium ions for ionization (Centre Regional des Mesures Physiques de l'Ouest, Université de Rennes 1).

S2

Kinetics of the enzymatic reaction with *p*NP glycosides

The synthesis ability of the CGTases from *Thermoanaerobacter sp.* was previously investigated and the reaction with the enzyme was optimized to efficiently produce the primary coupling product. Typically, CGTase (10 μ L, 2.34 mg/mL) was added to 1.5 mL microtubes containing β -cyclodextrin (100 mg, 0.1 mM) and various carbohydrates (0.2 mM) in 50 mM phosphate buffer, pH 6.0 (1 mL) at 50°C. The reaction mixtures were incubated in a thermomixer (Eppendorf) during 72 hours. 50 μ L samples were withdrawn at different intervals and diluted with 950 μ L of acetonitrile/water (50 : 50) followed by a filtration before chromatography analysis.

HPLC analysis

Chromatographic analysis were carried out on an Agilent 1200 HPLC system (Agilent Technologies, Mexico), equipped with a ChemStation software, a micro-vacuum degasser G1322A, a quartenary pump G1311A, an autosampler G1329A, a thermostatic column compartment G1316A), and a diode array detector G1315B. An analytical Alltech® PrevailTM carbohydrate ES column (Analytical, 4.6*250 mm, 5 µm, Grace) was used for chromatographic separation at 35 °C. Using acetonitrile (mobile phase A) and purified and distilled water (mobile phase B) at a flow rate of 1 mL / min, HPLC analysis started with 20 % B followed by gradient to 50% B over 40 min, then by a gradient step for 1 min back to 20 % B, before isocratic elution with starting conditions within 30 min. The injection volume of each sample was 10 μ L. Simultaneous monitoring was performed at 300, 368 and 400 nm.



Figure 1 : HPLC chromatograms of the reaction with pNP α -D-Glcp **1** after 72 hours incubation



Figure 2 : HPLC chromatograms of the reaction with pNP β -D-Glcp **2** after 72 hours incubation



Figure 3 : HPLC chromatograms of the reaction with pNP α -L-Araf 3 after 72 hours incubation



Figure 4 : HPLC chromatograms of the reaction with pN β -D-Galf **4** after 72 hours incubation

UPLC analysis

The samples collected during the development work were analyzed using an Acquity-LCT premier XE UPLC system from Waters equipped with a Acquity UPLC® BEH Amide column (BEH : Ethylene Bridged Hydride, 1.7 μ m 2.1*100 mm, Waters) at flow rate 0.2 mL /min. Column temperature was maintained at 35 °C. The mobile phases consisted of A (ACN /H2O, 9:1) and B (ACN/H2O, 3:7) both containing either 0.1 vol % sodium hydroxide. A linear gradient with increasing proportion of B (0 min = 10 % B, 5 min = 70 % B, 10 minutes = 10 % B) was used. The injected sample volume was 5 μ L and detected by a mass spectrometer with electrospray ionization. The instruments were operated in negative ion electrospray mode at a cone voltage of 40 V and capillary voltage of 28000.



Figure 5 : UPLC/MS analysis of the glucosylation reaction with methyl arabinofuranoside after 72 hours.



Figure 6: UPLC /MS analysis of the glucosylation reaction with hexyl arabinofuranoside after 72 hours.

Microwave assisted reaction between aryl glycoside and cyclodextrin in presence of CGTase

Microwave-assisted enzymatic reactions were performed in phosphate buffer only. CGTase (10 μ L, 2.34mg/mL) was added to the substrate solutions (1 mL) containing 50 mM Na₂HPO₄ buffer (pH 6.0), β -cyclodextrin (10%, 100 mg, 0.1 mM) and various carbohydrates (0.2 mol/L). The reaction mixture was introduced in the microwave equipment and irradiated with a variable power. The reactions were stopped by reducing the temperature to room temperature and filtered through 0.22 μ m Millipore express membrane.



Graph 1. Microwave power effect on trans-glycosylation and disproportionation yields on *p*NP α -Glc*p* **1**.



Graph 2. Microwave power effect on the trans-glycosylation and disproportionation yields on *p*NP β -Glc*p* **2**.

Transglycosylation reactions with glycoside acceptor in a preparative scale

Reaction conditions were implemented as previously described in a 1 mL final volume using a molar ratio glycoside/ β -cyclodextrin 3/1 and CGTase (10 μ L, 2.34 mg/mL). The resulting mixture was incubated at 50 °C during 72 h. Reaction mixture was lyophilized and the residue was purified by silica gel flash chromatrography using a eluant gradient AcOEt – (AcOH/H₂O, 1:1) from 90: 5 to 60 : 40. Elution of the products was monitored by UV detection and by TLC. Fractions corresponding to individual products were collected, evaporated and lyophilized.

Nitrophenyl β -D-maltoside (12).

This compound was obtained according to the described general procedure for transglycosylation with glycoside acceptor by incubation of of *p*-nitrophenyl β -D-glucopyranoside **1** (60 mg, 0.2 mmol) in the presence of the CGTase wt and was isolated in 7% yield (6.5 mg, 0.014 mmol) after purification Further elution yield the byproduct nitrophenyl β -D-isomaltoside (**13**) in 2% yield (2 mg, 4.3 µmol).

Spectroscopic data correspond to those described in the literature.^{1, 2}

12: $\delta_{H}(400 \text{ MHz}, \text{CD}_{3}\text{OD})$ 3. 97 – 3.30 (12H, m, H-2a, H-3a, H-4a, H-5a, H-6a, H-2b, H-3b, H-4b, H-5b, H-6b), 5.09 (1H, d, $J_{1b,2b}$ 3.2, H-1b), 5.21 (1H, t, $J_{1a,2a}$ 4.4, H-1a), 7.22 (2H, dd, Ph), 8.19 (2H, dd, Ph). δ_{C} (100 MHz, CD₃OD) 77.9 – 62.5, 80.7, 86.6, 102.7 (C-1b), 102.9 (C-1a), 117.8, 126.7. HRMS (ESI⁻) calcd. for C₁₈H₂₄NO₁₃ [M-H]⁻ 462,1326, found 462,1322.

13: $\delta_{H}(400 \text{ MHz}, \text{CD}_{3}\text{OD})$ 3. 97 – 3.30 (m, 12H, H-2a, H-3a, H-4a, H-5a, H-6a, H-2b, H-3b, H-4b, H-5b, H-6b), 4.80 (s, 1H, H-1b), 5.05 (d, 1H, $J_{1a,2a}$ 7.6 Hz, H-1a), 7.25 (d, 2H, *J* 9.2 Hz, Ph), 8.23 (d, 2H, *J* 9.2 Hz, Ph), δ_{C} (100 MHz, CD₃OD) 77.9 – 62.5, 94.6, 100.7 (C-1b), 101.7 (C-1a), 117.8, 126.9. HRMS (ESI⁻) calcd. for C₁₈H₂₄NO₁₃ [M-H]⁻ 462,1326, found 462,1324.

p-Nitrophenyl α-D-glucopyranosyl-(1,3)-α-L-arabinofuranoside (14)

This compound was obtained according to the described general procedure for transglycosylation with glycoside acceptor by incubation of of *p*-Nitrophenyl α -L-arabinofuranoside **3** (60 mg, 0.22 mmol) in the presence of the CGTase wt and was isolated in 17 % yield (14.5 mg) after purification. Further elution yield p-Nitrophenyl

 α -D-glucopyranosyl-(1,4)- α -D-glucopyranosyl-(1,3)- α -L-arabinofuranoside (8) in 10% yield (12.4 mg).

14: $R_f = 0.55$ (AcOEt/AcOH/H₂O, 7:2:2), $[\hat{1} \pm]_{D^{20}} + 10$ (c 1.0 MeOH). $\delta_H(400 \text{ MHz}, CD_3OD)$ 3.30 (1H, m, H-4b), 3.43 (1 H, dd, $J_{2b,1b}$ 4.0, $J_{2b,3b}$ 9.6, H-2b), 3.77-3.63 (4 H, m, H-3b, H-6b, H-5a), 3.87 (2 H, m, H-5b, H-6'b), 4.03 (1 H, dd, $J_{3a,2a}$ 3.2, $J_{3a,4a}$ 6.0, H-3a), 4.27 (1 H, m, H-4a), 4.59 (1 H, dd, $J_{2a,1a}$ 1.2, $J_{2a,3a}$ 3.2, H-2a), 4.96 (d, 1 H, $J_{1b,2b}$ 4.0, H-1b), 5.71 (1 H, d, $J_{1a,2a}$ 1.2 Hz, H-1a), 7.22 (2 H, d, J 9.20 Hz, H_{Ph}), 8.20 (2 H, d, J 9.20, H_{Ph}). δ_C (100 MHz, CD₃OD) 62.5 (C-5a), 62.6 (C-6b), 71.8 (C-4b), 73.4 (C-2b), 74.3 (C-5b), 74.8 (C-3b), 81.3(C-2a), 85.6 (C-4a), 86.8 (C-3a), 101.8 (C-1b), 107.7 (C-1a), 117.7, 126.6, 143.5, 163.3 (C-Ph). HRMS (ESI⁻) calcd. for $C_{17}H_{22}NO_{12}$ [M-H]⁻432.1142, found 432.1160.

15: $R_f = 0.32$ (AcOEt/AcOH/H₂O, 7:2:2), $[\hat{1} \pm]_{D^{20}} + 10.4$ (c 0.45 MeOH). δ_H (400 MHz, CD₃OD) 3.28 (1 H, m, H-4c), 3.48 (2 H, m, H-2b, H-2c), 3.55 (1 H, dd, J_{4b,3b} 9.2 Hz, H-4b), 3.72-3.64 (3 H, m, H-5c, H-6c, H-3c), 3.75 (2 H, m, H-5a), 3.83 (1 H, m, H-6'c), 3.85 (2 H, m, H-6b), 3.91 (1 H, d, J_{3b,4b} 9.2, H-3b), 4.01 (1 H, m, H-5b), 4.04 (1 H, dd, J_{3a,2a} 1.6, J_{3a,4a} 4.4, H-3a), 4.30 (1 H, dd, J_{4a,3a} 4.4, J_{4a,5a} 9.2, H-4a), 4.58 (1 H, d, J_{2a,3a} 1.6, H-2a), 4.97 (1 H, d, J_{1b,2b} 3.2, H-1b), 5.17 (1 H, d, J_{1c,2c} 3.2, H-1c), 5.73 (1 H, s, H-1a), 7.20 (2 H, d, J 9.2, H_{Ph}), 8.24 (2 H, d, J 9.2, H_{Ph}). δ_C (100 MHz, CD₃OD) 62.0 (C-6b), 62.6 (C-5a), 62.8 (C-6c), 71.5 (C-4c), 72.7 (C-5b), 73.0 (C-2b), 74.2 (C-2c), 74.7 (C-3b), 74.9 (C-5c), 75.1 (C-3c), 80.7 (C-2a), 82.0 (C-4b), 86.3 (C-4a), 87.0 (C-3a), 101.7 (C-1b), 103.1 (C-1c), 107.6 (C-1a), 117.5, 126.7, 143.5, 163.2 (C-Ph). HRMS (ESI⁻) calcd. for C₂₃H₃₂NO₁₇ [M-H]⁻ 594.1670, found 594.1690.

p-Nitrophenyl α -D-glucopyranosyl-(1,3)- β -D-galactofuranoside (16)

This compound was obtained according to the described general procedure for transglycosylation with glycoside acceptor by incubation of of *p*-Nitrophenyl β -D-galactofuranoside (60 mg, 0.02 mmol) in the presence of the CGTase wt and was isolated in 11 % yield (12 mg) after purification. Further elution yield *p*-Nitrophenyl α -D-glucopyranosyl-(1,4)- α -D-glucopyranosyl-(1,3)- β -D-galactofuranoside (**17**) in 7% yield (20 mg).

16: $R_f = 0.3$ (AcOEt/AcOH/H₂O, 7:2:2), $[I \pm]_{D^{20}} + 55$ (c 0.2 MeOH). δ_H (400 MHz, CD₃OD) 3.30 (1 H, m, H-4b), 3.43 (1 H, dd, J_{2b,1b} 4.0, J_{2b,3b} 10, H-2b), 3.65 (4 H, m, H-6b, H-6a, H-3b), 3.79 (1 H, m, H-5a), 3.86 (2 H, m, H-5b, H-6'b), 4.14 (1 H, dd, J_{2b,1b} 4.0, J_{2b,3b} 4.0, J_

 $J_{2a,3a}$ 3.6, $J_{3a,4a}$ 6.4, H-3a), 4.31 (1 H, m, H-4a), 4.58 (1 H, dd, $J_{2a,1a}$ 1.6, $J_{2a,3a}$ 3.6, H-2a), 4.98 (1 H, d, $J_{1b,2b}$ 4.0, H-1b), 5.70 (1 H, d, $J_{1a,2a}$ 1.6, H-1a), 7.22 (2 H, d, J 9.2, H_{Ph}), 8.21 (2 H, d, J 9.2, H_{Ph}). δ_{C} (100 MHz, CD₃OD) 62.7 (C-6b), 64.1 (C-6a), 71.8 (C-4b), 72.3 (C-5a), 73.4 (C-2b), 74.4 (C-5b), 74.9 (C-3b), 81.2 (C-2a), 84.5 (C-4a), 87.4 (C-3a), 102.0 (C-1b), 107.7 (C-1a), 117.7, 126.6, 143.6, 163.3 (C-Ph). HRMS (ESI⁻) calcd. for C₁₈H₂₄NO₁₃ [M-H]⁻462.1248, found 462.1260.

17: $R_f = 0.12$ (AcOEt/AcOH/H₂O, 7:2:2), $[I \pm]_{D^20} + 67$ (c 0.5 MeOH). δ_H (400 MHz, CD₃OD) 3.27 (1 H, m, H-4c), 3.47 (1 H, dd, J_{2c,1c} 3.6, J_{2c,3c} 9.6, H-2c), 3.49 (1 H, dd, J_{2b,1b} 3.6, J_{2b,3b} 10.0, H-2b), 3.54 (1 H, dd, J_{4b,3b} 8.8, J_{4b,5b} 10.0, H-4b), 3.70- 3.62 (5 H, m, H-3c, H-5c, H-6a, H-6c), 3.78 (1 H, dd, J_{6c,5c} 2.4 Hz, J_{6c,6'c}, H-6'c), 3.87-3.82 (3 H, m, H-5a, H-6b), 3.91 (1 H, dd, J_{3b,4b} 8.8, J_{3b,2b} 10.0, H-3b), 4.01 (1 H, m, H-5b), 4.15 (1 H, dd, J_{3a,2a} 3.2, J_{3a,4a} 5.6, H-3a), 4.34 (1 H, dd, J_{4a,5a} 3.6, J_{4a,3a} 5.6, H-4a), 4.57 (1 H, dd, J_{2a,1a} 1.6, J_{2a,3a} 3.2, H-2a), 4.99 (1 H, d, J_{1b,2b} 3.6, H-1b), 5.16 (1 H, d, J_{1c,2c} 3.6, H-1c), 5.72 (1 H, d, J_{1a,2a} 1.6, Ha-1), 7.16 (2 H, d, J 9.2, H_{Ph}), 8.24 (2 H, d, J 9.2, H_{Ph}), δ_C (100 MHz, CD₃OD) 62.0 (C-6b), 62.6 (C-5a), 62.8 (C-6c), 71.5 (C-4c, C-6a), 72.7 (C-5b, C-5a), 73.0 (C-2b), 74.2 (C-2c), 74.7 (C-3b), 74.9 (C-5c), 75.1 (C-3c), 80.7 (C-2a), 82.0 (C-4b), 86.3 (C-4a), 87.0 (C-3a), 101.7 (C-1b), 103.1 (C-1c), 107.6 (C-1a), 117.5, 126.7, 143.5, 163.2 (C-Ph). HRMS (ESI⁻) calcd. for C₂₄H₃₄NO₁₈ [M-H]⁻ 624.1776, found 624.1779.

Methyl α -D-glucopyranosyl-(1,3)- α -L-arabinofuranoside (18)

This compound was obtained according to the described general procedure for transglycosylation with glycoside acceptor by incubation of Methyl α -L-arabinofuranoside **11** (60 mg, 0.3 mmol) in the presence of the CGTase wt and was isolated in 14 % yield (16 mg) after purification. Further elution yield methyl α -D-glucopyranosyl-(1,3)- α -L-arabinofuranoside (**25**) in 11% yield (20 mg).

18: $R_f = 0.25$ (AcOEt/AcOH/H₂O, 7:2:2), $[\hat{1} \pm]_{D^{20}} + 15.6$ (c 1.0 MeOH). $\delta_H(400 \text{ MHz}, CD_3OD)$ 3.30 (1 H, m, H-4b), 3.36 (1 H, dd, $J_{2b,1b}$ 4.0, $J_{2b,3b}$ 9.6, H-2b), 3.37 (3 H, s, CH₃), 3.61-3.70 (2 H, m, H-3b, H-6b), 3.70 (1 H, dd, $J_{5a,4}$ 4.8, $J_{5a,5'a}$ 12.0, H-5a), 3.76 (1 H, dd, $J_{5'a,4} = 4.0$ Hz, $J_{5'a,5}$ 12.0, H-5'a), 3.79-3.85 (2 H, m, H-5b, H-6'b), 3.88 (1 H, dd, $J_{3a,2a}$ 3.2, $J_{3a,4a}$ 6.2, H-3a), 4.09 (1 H, m, H-4a), 4.19 (1 H, dd, $J_{2a,1a}$ 1.2, $J_{2a,3a}$ 3.2, H-2a), 4.79 (1 H, d, $J_{1a,2a}$ 1.2, H-1a), 4.88 (1 H, d, $J_{1b,2b}$ 4.0, H-1b). $\delta_C(100 \text{ MHz}, CD_3OD)$ 55.3 (CH₃), 62.7 (C-6b), 62.8 (C-5a), 72.9 (C-4b), 73.4 (C-2b), 74.1 (C-5b),

74.8 (C-3b), 81.6 (C-2a), 83.7 (C-4a), 86.7 (C-3a), 101.6 (C-1b), 110.6 (C-1a). HRMS (ESI-) calcd. for $C_{12}H_{21}O_{10}$ [M-H]⁻ 325.1135, found 325.1125.

25: $R_f = 0.07$ (AcOEt/AcOH/H₂O, 7:2:2), $[^{[1 \pm]}_{D^{20}} + 95$ (c 0.5 MeOH). δ_H (400 MHz, CD₃OD) 3.30 (1 H, m, H-4c), 3.37 (3 H, s, CH₃), 3.40-3.48 (2 H, m, H-2c, H-2b), 3.52 (1 H, m, H-4b), 3.58-3.70 (3 H, m, H-3c, H-5c, H-6c), 3.70 (1 H, dd, J_{5a,4a} 5.2, J_{5a,5'a} 12.0, H-5a), 3.76 (1 H, dd, J_{5'a,4a} 4.0, J_{5'a,5a} 12.0, H-5'a), 3.75-3.90 (5 H, m, H-3b, H-5b, H-6b, H-6'c), 3.90 (1 H, dd, J_{3a,2a} 3.2, J_{3a,4a} 6.4, H-3a), 4.10 (1 H, m, H-4a), 4.17 (1 H, dd, J_{2a,1a} 1.6, J_{2a,3a} 3.2, H-2a), 4.78 (1 H, d, J_{1a,2a} 1.6, H-1a), 4.90 (1 H, d, J_{1b,2b} 4.0, H-1b), 5.11 (1 H, d, J_{1c,2c} 3.6, H-1c). δ_C (100 MHz, CD₃OD) 55.3 (CH₃), 62.3 (C-6b), 62.7 (C-6c), 62.8 (C-5a), 71.6 (C-4c), 72.7 (C-5b), 73.5 (C-2b), 74.1 (C-2c), 74.2 (C-3b), 74.7 (C-5c), 75.1 (C-3c), 81.3 (C-2a), 81.9 (C-4b), 83.8 (C-4a), 86.7 (C-3a), 100.9 (C-1b), 102.9 (C-1c), 110.6 (C-1a). HRMS (ESI⁻) calcd. for C₁₈H₃₁O₁₅ [M-H]⁻ 487.1663, found 487.1655.

Ethyl α -D-glucopyranosyl-(1,3)- α -L-arabinofuranoside (19)

This compound was obtained according to the described general procedure for transglycosylation with glycoside acceptor by incubation of ethyl α -L-arabinofuranoside **12** (53 mg, 0.30 mmol) in the presence of the CGTase wt and was isolated in 26 % yield (25 mg) after purification. Further elution yield propyl α -D-glucopyranosyl-(1,4)- α -D-glucopyranosyl-(1,3)- α -L-arabinofuranoside (**26**) in 14% yield (20 mg).

19: $R_f = 0.30$ (AcOEt/AcOH/H₂O, 7:2:2), $[\hat{1} \pm]_D + 17.3$ (c 0.85 in MeOH). δ_H (400 MHz, CD₃OD) 1.20 (3 H, t, J 7.1, CH₃), 3.28 (1 H, dd, J_{4b,3b} 9.2, J_{4b,5b} 9.8, H-4b), 3.39 (1 H, dd, J_{2b,1b} 3.8, J_{2b,3b} 9.8, H-2b), 3.44-3-53 (1 H, m, OCH₂CH₃), 3.62 (1 H, dd, J_{3b,4b} 9.0, J_{3b,2b} 9.8, H-3b), 3.66 (1 H, dd, J_{6b,5b} 6.2, J_{6b,6'b} 11.4, H-6b), 3.72-3.81 (2 H, m, OCH₂CH₃, H-5b), 3.69 (1 H, dd, J_{5a,4a} 4.8, J_{5a,5'a} 12.0, H-5a), 3.76 (1 H, dd, J_{4a,5'a} 3.9, J_{5'a,5a} 12.0, H-5'a), 3.85 (1 H, dd, J_{6'b,5b} 2.3, J_{6'b,6b} 11.4, H-6'b), 3.88 (1 H, dd, J_{3a,2a} 3.5, J_{3a,4a} 6.4, H-3a), 4.11 (1 H, ddd, J_{4a,5'a} 3.9, J_{4a,5a} 4.5, J_{4a,3a} 6.4, H-4a), 4.20 (1 H, dd, J_{2a,1a} 1.7, J_{2a,3a} 3.5, H-2a), 4.89 (1 H, d, J_{1a,2a} 1.7, H-1a), 4.89 (1 H, d, J_{1b,2b} 3.8, H-1b). δ_C (100 MHz, CD₃OD) 15.4 (CH₃), 62.6 (C-6b), 62.8 (C-5a), 64.3 (OCH₂CH₃), 71.8 (C-4b), 73.5 (C-2b), 74.1 (C-5b), 74.9 (C-3b), 81.8 (C-2a), 83.4 (C-4a), 86.8 (C-3a), 101.2 (C-1b), 109.4 (C-1a). HRMS (ESI⁻) calcd. for C₁₃H₂₃O₁₀ [M-H]⁻ 339.1297, found 339.1298.

26: $R_f = 0.1$ (AcOEt/AcOH/H₂O, 7:2:2), $[\hat{1} \pm]_D + 63.3$ (c 0.9 in MeOH). δ_H (400 MHz, CD₃OD) 1.21 (3 H, t, J 6.9, CH₃), 3.23-3.30 (1 H, m, H-4c), 3.42-3.51 (3 H, m, H-2c, H-2b, O*CH*₂CH₃), 3.52 (1 H, dd, J 9.1, J 9.6, H-4b), 3.63 (1 H, dd, J 9.1, J 9.6, H-3c), 3.57-3.71 (2 H, m, H-5c, H-6c), 3.68 (1 H, dd, J_{5a,4a} 4.6, J_{5a,5'a} 11.9, H-5a), 3.72-3.82 (1 H, m, O*CH*₂CH₃), 3.75 (1 H, dd, J_{5'a,4a} 3.9, J_{5'a,5a} 11.9, H-5'a), 3.79-3.92 (5 H, m, H-5b, H-6b, H-3b, H-6'c), 3.88 (1 H, dd, J_{3a,2a} 3.2, J_{3a,4a} 6.0, H-3a), 4.12 (1 H, ddd, J_{4a,5'a} 3.9, J_{4a,5a} 4.6, J_{4a,3a} 6.0, H-4a), 4.20 (1 H, dd, J_{2a,1a} 1.7, J_{2a,3a} 3.2, H-2a), 4.89 (1 H, d, J_{1a,2a} 1.7, H-1a), 4.91 (1 H, d, J_{1b,2b} 4.4, H-1b), 5.15 (1 H, d, J_{1c,2c} 3.8, H-1c). δ_C (100 MHz, CD₃OD) 15.5 (CH₃), 61.9 (C-6b), 62.6 (C-6c), 62.7 (C-5a), 64.1 (*C*H₂CH₃), 71.3 (C-4c), 72.4 (C-5b), 72.9, 74.0 (C-2c, C-2b), 74.5 (C-3b, C-5c), 74.9 (C-3c), 81.2 (C-2a), 81.7 (C-4b), 83.5 (C-4a), 86.7 (C-3a), 100.8 (C-1b), 102.8 (C-1c), 109.1 (C-1a). HRMS (ESI⁻) calcd. for C₁₉H₃₃O₁₅ [M-H]⁻ 501.1825, found 501.1821.

Propyl α-D-glucopyranosyl-(1,3)-α-L-arabinofuranoside (20)

This compound was obtained according to the described general procedure for transglycosylation with glycoside acceptor by incubation of of propyl α -L-arabinofuranoside **13** (56 mg, 0.29 mmol) in the presence of the CGTase wt and was isolated in 21 % yield (22 mg) after purification. Further elution yield propyl α -D-glucopyranosyl-(1,4)- α -D-glucopyranosyl-(1,3)- α -L-arabinofuranoside (**27**) in 13% yield (20 mg).

20: $R_f = 0.38$ (AcOEt/AcOH/H₂O, 7:2:2), $[\hat{1} \pm]_{D^{20}} + 16.3$ (*c* 0.27 in MeOH). $\delta_H(400$ MHz, CD₃OD) 0.94 (3 H, t, J 7.8, CH₃), 1.60 (2 H, m, OCH₂CH₂), 3.28 (1 H, dd, J_{4b,3b} 8.9, J_{4b,5b} 9.8, H-4b), 3.25-3-43 (2 H, m, OCH₂CH₂, H-2b), 3.61 (1 H, dd, J_{3b,4b} 8.9, J_{3b,2b} 9.8, H-3b), 3.63-3.69 (2 H, m, H-6b, OCH₂CH₂), 3.69 (1 H, dd, J_{5a,4a} 4.7, J_{5a,5'a} 12.1, H-5a), 3.76 (1 H, dd, J_{5'a,4a} 3.9, J_{5'a,5a} 12.1, H-5'a), 3.77 (1 H, m, H-5b), 3.85 (1 H, dd, J_{6'b,5b} 2.2, J_{6'b,6b} 11.3, H-6'b), 3.87 (1 H, dd, J_{3a,2a} 3.2, J_{3a,4a} 6.7, H-3a), 4.11 (1 H, ddd, J_{4a,5'a} 3.9, J_{4a,5a} 4.7, J_{4a,3a} 6.7, H-4a), 4.21 (1 H, dd, J_{2a,1a} 1.7, J_{2a,3a} 3.2, H-2a), 4.88 (1 H, d, J_{1a,2a} 1.7, H-1a), 4.89 (1 H, d, J_{1b,2b} 3.9, H-1b). δ_C (100 MHz, CD₃OD) 11.0 (CH₃), 23.9 (OCH₂CH₂), 62.6 (C-6b), 62.8 (C-5a), 70.5 (OCH₂CH₂), 71.8 (C-4b), 73.5 (C-2b), 74.1 (C-5b), 74.9 (C-3b), 81.8 (C-2a), 83.3 (C-4a), 87.0 (C-3a), 101.3 (C-1b), 109.5 (C-1a). HRMS (ESI⁻) calcd. for C₁₄H₂₅O₁₀ [M-H]⁻ 353,1448, found 353,1437.

27: $R_f = 0.19$ (AcOEt/AcOH/H₂O, 7:2:2), $[\hat{1} \pm]_{b^{20}} + 60.5$ (*c* 0.4 in MeOH). δ_H (400 MHz, CD₃OD) 0.95 (3 H, t, J 7.4, CH₃), 1.60 (2 H, m, OCH₂CH₂), 3.24-3.30 (1 H, m, H-4c), 3.34-3.43 (1 H, m, OCH₂CH₂), 3.45 (2 H, dd, J_{2,1} 3.8, J_{2,3} 9.5, H-2b, H-2c), 3.53 (1 H, dd, J 9.2, 9.5, H-4b), 3.63 (1 H, t, J 9.5, H-3c), 3.62-3.72 (3 H, m, H-5c, H-6c, OCH₂CH₂), 3.69 (1 H, dd, J_{5a,4a} 4.6, J_{5a,5'a} 11.9, H-5a), 3.75 (1 H, dd, J_{5'a,4a} 3.6, J_{5'a,5a} 11.9, H-5'a), 3.78-3.91 (5 H, m, H-5b, H-6b, H-3b, H-6'c), 3.87 (1 H, dd, J_{3a,2a} 3.1, J_{3a,4a} 6.2, H-3a), 4.13 (1 H, ddd, J_{4a,5'a} 4.1, J_{4a,5a} 4.6, J_{4a,3a} 5.6, H-4a), 4.22 (1 H, dd, J_{2a,1a} 1.4, J_{2a,3a} 3.0, H-2a), 4.89 (1 H, d, J_{1a,2a} 1.4, H-1a), 4.91 (1 H, d, J_{1b,2b} 3.8, H-1b), 5.14 (1 H, d, J_{1c,2c} 3.8, H-1c). δ_C (100 MHz, CD₃OD) 11.1 (CH₃), 24.0 (CH₂CH₃), 61.9 (C-6b), 62.7 (C-6c), 62.9 (C-5a), 70.4 (OCH₂CH₂), 71.5 (C-4c), 72.5 (C-5b), 73.0, 73.1 (C-2c, C-2b), 74.7 (C-3b), 74.8 (C-5c), 75.1 (C-3c), 81.2 (C-2a), 81.9 (C-4b), 84.0 (C-4a), 87.2 (C-3a), 101.2 (C-1b), 103.1 (C-1c), 109.5 (C-1a). HRMS (ESI⁻) calcd. for C₂₀H₃₅O₁₅ [M-H] 515.1976, found 515.1970.

Butyl α-D-glucopyranosyl-(1,3)-α-L-arabinofuranoside (21)

This compound was obtained according to the described general procedure for transglycosylation with glycoside acceptor by incubation of butyl α -L-arabinofuranoside **14** (67 mg, 0.32 mmol) in the presence of the CGTase wt and was isolated in 19 % yield (23 mg) after purification. Further elution yield butyl α -D-glucopyranosyl-(1,4)- α -D-glucopyranosyl-(1,3)- α -L-arabinofuranoside (**28**) in 12% yield (20 mg).

21: $R_f = 0.5$ (AcOEt/AcOH/H₂O, 7:2:2), $[\hat{I} \pm]_{D^{20}} + 18.2$ (*c* 0.5 in MeOH). $\delta_H(400 \text{ MHz}, CD_3OD)$ 0.94 (3 H, t, J 7.4, CH₃), 1.40 (2 H, m, CH₂CH₃), 1.57 (2 H, m, OCH₂CH₂), 3.28 (1 H, dd, J_{4b,3b} 8.9, J_{4b,5b} 9.9, H-4b), 3.36-3-46 (1 H, m, OCH₂CH₂), 3.39 (1 H, dd, J_{2b,1b} 4, J_{2b,3b} 9.7, H-2b), 3.61 (1 H, dd, J_{3b,4b} 8.9, J_{3b,2b} 9.7, H-3b), 3.62-3.70 (2 H, m, H-6b, OCH₂CH₂), 3.69 (1 H, dd, J_{5a,4a} 4.6, J_{5a,5'a} 11.9, H-5a), 3.76 (1 H, dd, J_{5'a,4a} 3.8, J_{5'a,5a} 11.9, H-5'a), 3.74-3.80 (1 H, m, H-5b), 3.86 (1 H, dd, J_{6'b,5b} 2.5, J_{6'b,6b} 11.7, H-6'b), 3.87 (1 H, dd, J_{3a,2a} 3.4, J_{3a,4a} 6.5, H-3a), 4.10 (1 H, ddd, J_{4a,5'a} 3.8, J_{4a,5a} 4.6, J_{4a,3a} 6.5, H-4a), 4.20 (1 H, dd, J_{2a,1a} 1.8, J_{2a,3a} 3.4, H-2a), 4.87 (1 H, d, J_{1a,2a} 1.8, H-1a), 4.89 (1 H, d, J_{1b,2b} 4, H-1b). $\delta_C(100 \text{ MHz}, \text{ CD}_3\text{OD})$ 14.2 (CH₃), 20.4 (CH₂CH₃), 32.9 (OCH₂CH₂), 62.6 (C-6b), 62.8 (C-5a), 68.6 (OCH₂CH₂), 71.8 (C-4b), 73.5 (C-2b), 74.1 (C-5b), 74.9 (C-3b), 81.8 (C-2a), 83.4 (C-4a), 87.0 (C-3a), 101.3 (C-1b), 109.5 (C-1a). HRMS (ESI⁻) calcd. for C₁₅H₂₇O₁₀ [M-H]⁻ 367.1604, found 367.1609.

28: $R_f = 0.21$ (AcOEt/AcOH/H₂O, 7:2:2), $[I \pm]_{D^{20}} + 63.4$ (*c* 0.35 in MeOH). $\delta_H(400$ MHz, CD₃OD) 0.95 (3 H, t, J 7.3, CH₃), 1.41 (2 H, m, CH₂CH₃), 1.57 (2 H, m, OCH₂CH₂), 3.28 (1 H, t, J 9.4, H-4c), 3.40-3.47 (3 H, m, H-2b, H-2c, OCH₂CH₂), 3.52 (1 H, t, J 9.4, H-4b), 3.63 (1 H, t, J 9.4, H-3c), 3.64-3.72 (3 H, m, H-5c, H-6c, OCH₂CH₂), 3.69 (1 H, dd, J_{5a,4a} 4.5, J_{5a,5'a} 11.7, H-5a), 3.75 (1 H, dd, J_{5'a,4a} 4.0, J_{5'a,5a} 11.7, H-5'a), 3.78-3.91 (5 H, m, H-5b, H-6b, H-3b, H-6'c), 3.88 (1 H, dd, J_{3a,2a} 2.8, J_{3a,4a} 6.0, H-3a), 4.11 (1 H, ddd, J_{4a,5'a} 4.0, J_{4a,5a} 4.5, J_{4a,3a} 6.0, H-4a), 4.21 (1 H, dd, J_{2a,1a} 1.4, J_{2a,3a} 2.8, H-2a), 4.87 (1 H, d, J_{1a,2a} 1.4, H-1a), 4.90 (1 H, d, J_{1b,2b} 3.8, H-1b), 5.14 (1 H, d, J_{1c,2c} 3.7, H-1c). δ_C (100 MHz, CD₃OD) 14.2 (CH₃), 20.4 (CH₂CH₃), 32.8 (OCH₂CH₂), 62.0 (C-6b), 62.7 (C-6c), 62.9 (C-5a), 68.4 (OCH₂CH₂), 71.5 (C-4c), 72.6 (C-5b), 73.2, 73.3 (C-2c, C-2b), 74.7 (C-3b), 74.8 (C-5c), 75.1 (C-3c), 81.3 (C-2a), 81.9 (C-4b), 83.8 (C-4a), 87.2 (C-3a), 101.2 (C-1b), 103.1 (C-1c), 109.5 (C-1a). HRMS (ESI⁻) calcd. for C₂₁H₃₇O₁₅ [M-H]⁻529.2132, found 529.2134.

n-pentyl α -D-glucopyranosyl-(1,3)- α -L-arabinofuranoside (22)

This compound was obtained according to the described general procedure for transglycosylation with glycoside acceptor by incubation of butyl α -L-arabinofuranoside **15** (60 mg, 0.27 mmol) in the presence of the CGTase wt and was isolated in 19 % yield (20 mg) after purification. Further elution yield butyl α -D-glucopyranosyl-(1,3)- α -L-arabinofuranoside (**29**) in 11% yield (17 mg).

22: $R_f = 0.5$ (AcOEt/AcOH/H₂O, 7:2:2), $[I \pm]_{D^{20}} + 20.6$ (*c* 0.7 in MeOH). δ_H (400 MHz, CD₃OD) 0.92 (3 H, t, J 6.9, CH₃), 1.30-1.40 (4 H, m, C₂H₄CH₃), 1.53-1.65 (2 H, m, OCH₂CH₂), 3.29 (1 H, dd, J_{4b,3b} 9.0, J_{4b,5b} 9.9, H-4b), 3.36-3-45 (1 H, m, OCH₂CH₂), 3.39 (1 H, dd, J_{2b,1b} 3.9, J_{2b,3b} 9.7, H-2b), 3.62 (1 H, dd, J_{3b,4b} 9.0, J_{3b,2b} 9.7, H-3b), 3.62-3.75 (2 H, m, H-6b, OCH₂CH₂), 3.72 (1 H, dd, J_{5a,4a} 4.3, J_{5a,5'a} 12.0, H-5a), 3.76 (1 H, dd, J_{5'a,4a} 3.6, J_{5'a,5a} 12.0, H-5'a), 3.73-3.81 (1 H, m, H-5b), 3.85 (1 H, dd, J_{6'b,5b} 2.5, J_{6'b,6b} 11.9, H-6'b), 3.85 (1 H, dd, J_{3a,2a} 3.5, J_{3a,4a} 6.3, H-3a), 4.10 (1 H, ddd, J_{4a,5'a} 3.6, J_{4a,5a} 4.3, J_{4a,3a} 6.3, H-4a), 4.21 (1 H, dd, J_{2a,1a} 1.6, J_{2a,3a} 3.5, H-2a), 4.87 (1 H, d, J_{1a,2a} 1.6, H-1a), 4.89 (1 H, d, J_{1b,2b} 3.9, H-1b). δ_C (100 MHz, CD₃OD) 14.4 (CH₃), 23.5, 29.5 (C₂H₄CH₃), 30.4 (OCH₂CH₂), 62.6 (C-6b), 62.8 (C-5a), 68.9 (OCH₂CH₂), 71.8 (C-4b), 73.5 (C-2b), 74.1 (C-5b), 74.9 (C-3b), 81.8 (C-2a), 83.4 (C-4a), 87.0 (C-3a), 101.3 (C-1b), 109.5 (C-1a). HRMS (ESI⁻) calcd. for C₁₆H₂₉O₁₀ [M-H]⁻ 381.1766, found 381.1765.

29: $R_f = 0.21$ (AcOEt/AcOH/H₂O, 7:2:2), $[^{\hat{1} \pm]_{D}20} + 57.1$ (*c* 0.7 in MeOH). δ_H (400 MHz, CD₃OD) 0.93 (3 H, t, J 7.1, CH₃), 1.30-1.42 (4 H, m, C₂H₄CH₃), 1.53-1.65 (2 H, m, OCH₂CH₂), 3.28 (1 H, t, J 9.1, H-4c), 3.37-3.47 (3 H, m, H-2b, H-2c, OCH₂CH₂), 3.52 (1 H, t, J 9.4, H-4b), 3.63 (1 H, t, J 9.0, H-3c), 3.59-3.72 (3 H, m, H-5c, H-6c, OCH₂CH₂), 3.69 (1 H, dd, J_{5a,4a} 5.0, J_{5a,5'a} 11.7, H-5a), 3.76 (1 H, dd, J_{5'a,4a} 3.9, J_{5'a,5a} 11.7, H-5'a), 3.79-3.92 (5 H, m, H-5b, H-6b, H-3b, H-6'c), 3.88 (1 H, dd, J_{3a,2a} 3.0, J_{3a,4a} 6.2, H-3a), 4.11 (1 H, ddd, J_{4a,5'a} 4.0, J_{4a,5a} 4.6, J_{4a,3a} 6.2, H-4a), 4.20 (1 H, dd, J_{2a,1a} 1.5, J_{2a,3a} 3.0, H-2a), 4.87 (1 H, d, J_{1a,2a} 1.5, H-1a), 4.90 (1 H, d, J_{1b,2b} 3.8, H-1b), 5.14 (1 H, d, J_{1c,2c} 3.8, H-1c). δ_C (100 MHz, CD₃OD) 14.5 (CH₃), 23.5, 29.5 (C₂H₄CH₃), 30.4 (OCH₂CH₂), 62.0 (C-6b), 62.7 (C-6c), 62.8 (C-5a), 68.7 (OCH₂CH₂), 71.5 (C-4c), 72.6 (C-5b), 73.1, 74.3 (C-2c, C-2b), 74.7 (C-3b), 74.8 (C-5c), 75.1 (C-3c), 81.4 (C-2a), 81.9 (C-4b), 83.7 (C-4a), 87.2 (C-3a), 101.2 (C-1b), 103.1 (C-1c), 109.5 (C-1a). HRMS (ESI⁻) calcd. for C₂₂H₃₉O₁₅ [M-H]⁻ 543.2294, found 543.2300.

n-Hexyl α -D-glucopyranosyl-(1,3)- α -L-arabinofuranoside (23)

This compound was obtained according to the described general procedure for transglycosylation with glycoside acceptor by incubation of 60 mg (0.26 mmol) of *n*-hexyl α -L-arabinofuranoside and in the presence of the CGTase wt and was isolated in 25 % yield (25 mg) after purification. Further elution yield *n*-hexyl α -D-glucopyranosyl-(1,3)- α -L-arabinofuranoside (**30**) in 10% yield (13.6 mg).

23: $R_f = 0.5$ (AcOEt/AcOH/H₂O, 7:2:2), $[I \pm]_{D^{20}} + 22.5$ (c 1.0 MeOH). $\delta_H(400 \text{ MHz}, CD_3OD)$ 0.91 (3 H, t, *J* 6.8, CH₃), 1.32 (6 H, m, C₃H₆), 1.58 (2 H, m, OCH₂C*H*₂), 3.28 (1 H, dd, J_{4b,3b} 8.4, J_{4b,5b} 10.0, H-4b), 3.40 (1 H, dd, J_{2b,1b} 4.0, J_{2b,3b} 9.6, H-2b), 3.42 (1 H, m, OCH₂CH₂), 3.61 (1 H, dd, J_{3b,4b} 8.4, J_{3b,2b} 9.6, H-3b), 3.64-3.68 (2 H, m, H-6b, OCH₂CH₂), 3.69 (1 H, dd, J_{5a,4a} 4.2, J_{5a,5'a} 12.4, H-5a), 3.72 (1 H, dd, J_{5'a,4a} 4.0, J_{5'a,5a} 12.4, H-5'a), 3.82 (1 H, dd, J_{5b,6b} 2.4, J_{5b,4b} 10.0, H-5b), 3.84 (1 H, m, H-6'b), 3.86 (1 H, dd, J_{3a,2a} 3.6, J_{3a,4a} 6.4, H-3a), 4.09 (1 H, m, H-4a), 4.20 (1 H, dd, J_{2a,1a} 1.6, J_{2a,3a} 3.6, H-2a), 4.88 (1 H, d, J_{1a,2a} 1.6, H-1a), 4.90 (d, 1H, J_{1b,2b} 4.0, H-1b). δ_C (100 MHz, CD₃OD) 14.4 (CH₃), 32.8, 30.6, 26.9, 23.7 (C₄H₈), 62.6 (C-6b), 62.7 (C-5a), 68.9 (OCH₂CH₂), 71.8 (C-4b), 73.5 (C-2b), 74.1 (C-5b), 74.9 (C-3b), 81.8 (C-2a), 83.3 (C-4a), 87.0 (C-3a), 101.3 (C-1b), 109.5 (C-1a). HRMS (ESI⁻) calcd. for C₁₇H₂₉O₁₀ [M-H]⁻ 395.1917, found 395.1880.

30: $R_f = 0.3$ (AcOEt/AcOH/H₂O, 7:2:2), $[\hat{1} \pm]_{D^{20}} + 56.5$ (c 0.2 MeOH). $\delta_H(400 \text{ MHz}, CD_3OD)$ 0.91 (3 H, t, CH₃), 1.33 (6 H, m, C₃H₆), 1.58 (2 H, m, OCH₂CH₂), 3.28 (1 H, m, H-4c), 3.35-3.46 (3 H, m, H-2b, H-2c, OCH₂CH₂), 3.52 (1 H, m, H-4b), 3.62-3.74 (4 H, m, H-3c, H-5c, H-6c, OCH₂CH₂), 3.70 (1 H, dd, J_{5a,4a} 5.2, J_{5a,5'a} 12.0, H-5a), 3.76 (1 H, dd, J_{5'a,4a} 4.0, J_{5'a,5a} 12.0, H-5'a), 3.86-3.81 (5 H, m, H-3b, H-5b, H-6b, H-6'c), 3.90 (1 H, dd, J_{3a,2a} 2.8, J_{3a,4a} 6.4, H-3a), 4.10 (1 H, m, H-4a), 4.17 (1 H, dd, J_{2a,1a} 1.2, J_{2a,3a} 2.8, H-2a), 4.79 (1 H, d, J_{1a,2a} 1.2, H-1a), 4.90 (1 H, d, J_{1b,2b} 4.0, H-1b), 5.13 (1 H, d, J_{1c,2c} 3.6, H-1c). $\delta_C(100 \text{ MHz}, CD_3OD)$ 14.4 (CH₃), 32.7, 30.6, 26.9, 23.7 (C₄H₈), 61.9 (C-6b), 62.7 (C-6c), 62.8 (C-5a), 68.7 (OCH₂CH₂), 71.4 (C-4c), 72.6 (C-5b), 73.1 (C-2c), 74.2 (C-2b), 74.6 (C-3b), 74.7 (C-5c), 75.0 (C-3c), 81.4 (C-2a), 81.9 (C-4b), 83.6 (C-4a), 87.0 (C-3a), 101.1 (C-1b), 101.3 (C-1c), 109.5 (C-1a). HRMS (ESI⁻) calcd. for C₂₃H₄₁O₁₅ [M-H]⁻ 557.2445, found 557.2365.

n-Octyl α -D-glucopyranosyl-(1,3)- β -D-galactofuranoside (24)

This compound was obtained according to the described general procedure for transglycosylation with glycoside acceptor by incubation of 100 mg (0.34 mmol) of *n*-octyl β -D-galactofuranoside and in the presence of the CGTase wt and was isolated in 5% yield (7.5 mg) after purification. Further elution yield *n*-octyl α -D-glucopyranosyl-(1,4)- α -D-glucopyranosyl-(1,3)- β -D-galactofuranoside (**31**) in 3% yield (6.3 mg).

24: $R_f = 0.56$ (AcOEt/AcOH/H₂O, 7:2:2), $[^{\hat{I} \pm}]_{D^{20}} + 32.7$ (c 1.0 MeOH). δ_H (400 MHz, CD₃OD) 0.89 (3H, t, J 6.8, CH₃), 1.28-1.32 (10H, m, CH₂), 1.60-1.63 (2H, m, OCH₂CH₂), 3.28 (1 H, m, H-4b), 3.42 (1 H, dd, J_{2b,1b} 3.8, J_{2b,3b} 11, H-2b), 3.46 (1H, t, J 6.7, OCH₂), 3.53-3.65 (5H, m, H-5a, H6a, H-3b, H-6b), 3.72 (1H, dd, J 5.0, H-4a), 3.80 (1H, dt, J 6.7, J 9.5, OCH₂), 3.89 (2 H, m, H-5b, H-6'b), 3.94 (1H, m, H-2a), 4.12 (1 H, dd, J_{2a,3a} 3.2, J_{3a,4a} 6.5, H-3a), 4.86 (1H, d, J_{1a,2a} 1.5, H-1a), 4.98 (1 H, d, J_{1b,2b} 4.0, H-1b). δ_C (100 MHz, CD₃OD) 15.2 (CH₃), 24.4, 31.4, 31.2, 31.1, 27.9 [(CH₂)₅CH₃], 33.7 (OCH₂CH₂), 62.7 (C-6b), 65.4 (C-6a), 69.6 (OCH₂), 71.8 (C-4b), 73.2 (C-5a), 73.4 (C-2b), 74.4 (C-5b), 74.9 (C-3b), 84.0 (C-2a), 84.9 (C-4a), 87.4 (C-3a), 102.0 (C-1b), 110.0 (C-1a). HRMS (ESI⁻) calcd. for C₂₀H₃₇O₁₁ [M-H]⁻ 453,2336, found 453,2332.

31: $R_f = 0.4$ (AcOEt/AcOH/H₂O, 7:2:2), $[\hat{l} \pm]_{D^{20}} + 56.5$ (c 0.2 MeOH). δ_H (400 MHz, CD₃OD) 0.90 (3H, t, J 7, CH₃), 1.30-1.35 (10H, m, CH₂), 3.50 (1 H, dd, J_{2b,1b} 3.6,

J_{2b,3b} 10.0, H-2b), 1.61-1.63 (2H, m, OCH₂CH₂), 3.27 (1 H, m, H-4c), 3.45-3-47 (2H, m,H-2c, OCH₂), 3.52-3.70 (7H, m, H-5a, H6a, H-4b, H-3c, H-5c, H-6c), 3.72 (1H, dd, J 5.0, H-4a), 3.78 (1 H, dd, J_{6c,5c} 2.4 Hz, J_{6c,6'c} 12, H-6'c), 3.80 (1H, dt, J 6.7, J 9.5, OCH₂), 3.85-3.92 (3 H, m, H-3b, H-6b), 3.98-4.05 (2H, m, H-2a, H-5b), 4.15 (1 H, dd, J_{3a,2a} 3.2, J_{3a,4a} 5.6, H-3a), 4.88 (1H, d, J_{1a,2a} 1.5, H-1a), 4.99 (1 H, d, J_{1b,2b} 3.6, H-1b), 5.16 (1 H, d, J_{1c,2c} 3.6, H-1c) δ_{c} (100 MHz, CD₃OD) 15.1 (CH₃), 24.5, 27.9, 31.2, 31.6, 31.7 [(CH₂)₅CH₃], 34.0 (OCH₂CH₂), 62.2 (C-6b), 62.5 (C-6c), 65.5 (C-6a), 69.3 (OCH₂), 71.5 (C-4c), 72.7 (C-5b), 73.0 (C-2b), 73.2 (C-5a), 74.2 (C-2c), 74.7 (C-3b), 74.9 (C-5c), 75.1 (C-3c), 82.0 (C-4b), 84.0 (C-2a), 84.9 (C-4a), 87.4 (C-3a), 101.7 (C-1b), 103.1 (C-1c), 110.0 (C-1a). HRMS (ESI⁻) calcd. for C₂₆H₄₇O₁₆ [M-H]⁻ 615,2864, found 615,2860.

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NMR characterisation of isolated compounds 14 and 15 from action of CGTase in presence of cyclodextrin and *p*NP Araf 3



¹³C NMR spectrum of isolated disaccharide **14**.



2D NMR COSY spectrum of isolated disaccharide 14.



2D NMR HSQC spectrum of isolated disaccharide 14.



2D NMR HMBC spectrum of isolated disaccharide 14.



¹H NMR spectrum of isolated trisaccharide **15**.





2D NMR HMBC spectrum of isolated trisaccharide 15.

NMR spectrum of isolated compounds 16-31



































S42



























