

Probing Dynamic Cell-Substrate Interactions using Photochemically Generated Surface-Immobilized Gradients: Application to Selectin-Mediated Leukocyte Rolling

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Electronic Supplementary Information (ESI)

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Table S1 Summary of contact angles following silanization with TESA and functionalization with benzophenone (BP). The contact angles reported for each batch are the average (\pm standard deviation) of at least four substrates.

| | H ₂ O Contact Angle, Batch 1 | H ₂ O Contact Angle, Batch 2 | H ₂ O Contact Angle, Batch 3 | Average of 3 batches |
|--------------------------|---|---|---|----------------------|
| TESA silanization | 48.0 (\pm 1.4) | 42.7 (\pm 1.1) | 45.2 (\pm 2.6) | 45.2 (\pm 2.6) |
| +BP | 57.5 (\pm 7.7) | 59.6 (\pm 0.9) | 57.3 (\pm 3.9) | 58.1 (\pm 3.9) |

Table S2 Summary of gradient generation conditions used to generate one-component gradients of P-selectin and PSGL-1.

| Conditions for generation of one-component gradients used in Jurkat cell flow assays | | | | | | |
|---|-----------------------------|-----------------------------|---------------------------|--------------------|-------------------------------------|-------------------------------|
| Protein | concentration (μ g/mL) | power (mW/cm ²) | total exposure time (sec) | gradient size (mm) | size of gradient used in assay (mm) | # positions analyzed in assay |
| P-selectin | 5 | 14 | 24 | 4.8 | 2.4 | 8 |
| PSGL-1 | 10 | 14 | 24 | 4.8 | 3 | 7 |

| Conditions for generation of one-component gradients used in HL-60 cell flow assays | | | | | | |
|--|-----------------------------|-----------------------------|---------------------------|--------------------|-------------------------------------|-------------------------------|
| Protein | concentration (μ g/mL) | power (mW/cm ²) | total exposure time (sec) | gradient size (mm) | size of gradient used in assay (mm) | # positions analyzed in assay |
| P-selectin | 10 | 14 | 30 | 4.8 | 4.2 | 10 |
| PSGL-1 | 10 | 14 | 24 | 4.8 | 4.2 | 7 |

Table S3 Summary of data and associated approximations from one-component gradients of P-selectin and PSGL-1. Protein site density was determined by converting data from fluorescence assays to site density using data from fluorescence-radioactivity correlation studies (see **Fig. 2** in text).

| Substrates for Jurkat cell flow assays | | | Substrates for HL-60 cell flow assays | | |
|--|--------------------|---------------|---------------------------------------|--------------------|---------------|
| P-selectin site density | standard deviation | approximation | P-selectin site density | standard deviation | approximation |
| (molecules/ μm^2) | | | (molecules/ μm^2) | | |
| 2392.03 | 364.52 | 2400 | 958.43 | 87.69 | 1000 |
| 2180.13 | 502.35 | 2200 | 770.42 | 152.76 | 800 |
| 1782.28 | 525.53 | 1800 | 657.79 | 130.43 | 650 |
| 1384.13 | 396.81 | 1400 | 545.16 | 102.76 | 550 |
| 1026.76 | 311.67 | 1000 | 448.74 | 84.59 | 450 |
| 669.1 | 92.73 | 650 | 352.32 | 61.69 | 350 |
| 425.96 | 28.84 | 450 | 286.76 | 50.21 | 280 |
| 182.81 | 12.38 | 180 | 221.19 | 0.73 | 220 |
| | | | 182.78 | 22.78 | 180 |
| | | | 144.37 | 17.99 | 140 |

| PSGL-1 site density | standard deviation | approximation | PSGL-1 site density | standard deviation | approximation |
|-------------------------------|--------------------|---------------|-------------------------------|--------------------|---------------|
| (molecules/ μm^2) | | | (molecules/ μm^2) | | |
| 5899.34 | 1313.66 | 6000 | 4339.32 | 1032.91 | 4500 |
| 4646.76 | 777.97 | 5000 | 3860.19 | 971.11 | 4000 |
| 3927.47 | 712.62 | 4000 | 3470.98 | 767.19 | 3500 |
| 3424.61 | 721.77 | 3500 | 2697.33 | 738.54 | 2500 |
| 2581.96 | 544.16 | 2500 | 1718.97 | 516.5 | 1500 |
| 1739.31 | 365.04 | 1800 | 1125.06 | 397.79 | 1000 |
| 836.41 | 175.5436 | 800 | 123.08 | 84.66 | 100 |

Fig. S1 Linescans for one-component gradient substrates prepared for flow assays with HL-60 cells. BP-modified substrates present immobilized (a) P-selectin and (b) PSGL-1 on BP-modified substrates. Gradients were characterized in a fluorescence assay and fluorescence data was converted to site density using the results from the fluorescence-radioactivity correlation studies. Three representative gradient lines scans are shown for each immobilized protein gradient.

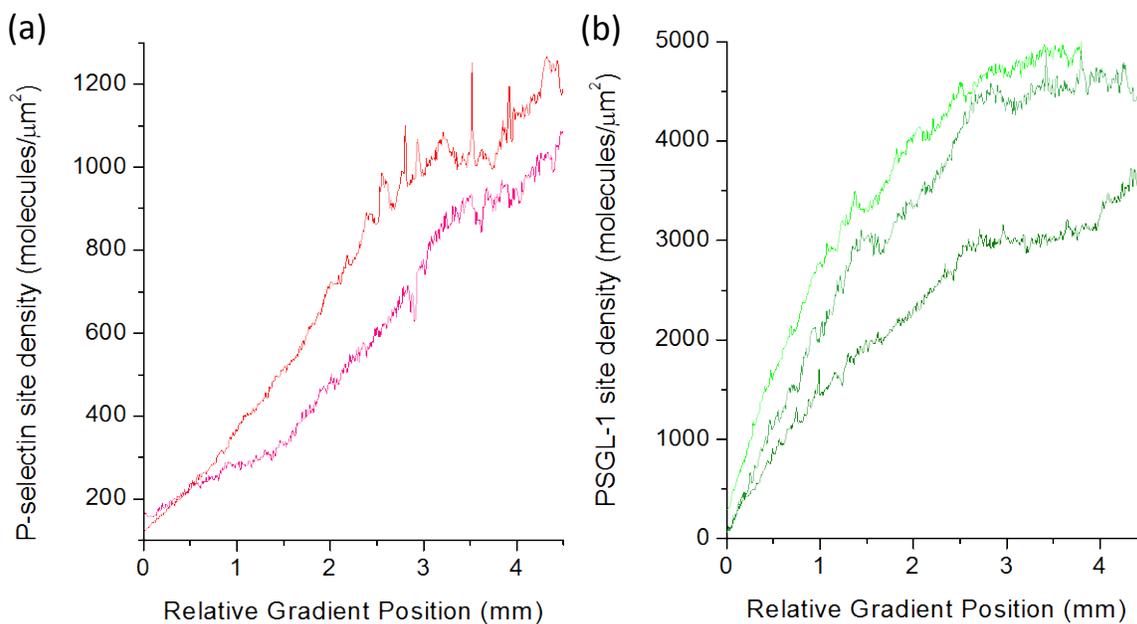


Table S4 The critical velocity (v_{crit}) was determined by analyzing non-interacting cells along the wall of the flow chamber and determining their average velocity at each shear stress. The values and associated error are reported above for $n=8+$ cells. At flow rates greater 250 $\mu\text{L}/\text{min}$, non-interacting cells were traveling to be analyzed as they appeared as streaks in the images.

| Flow rate ($\mu\text{L}/\text{min}$) | Shear stress (dyn/cm^2) | n | Critical velocity ($\mu\text{m}/\text{sec}$) | Standard deviation | 95% C.I. |
|--|---|-----|--|--------------------|----------|
| 10 | 0.062 | 9 | 21.7 | 3.5 | 2.3 |
| 25 | 0.155 | 7 | 43.4 | 5.7 | 4.2 |
| 50 | 0.31 | 8 | 97.6 | 21.9 | 15.2 |
| 75 | 0.465 | 23 | 139 | 32.7 | 13.4 |
| 100 | 0.62 | 10 | 340.9 | 99 | 61.3 |
| 150 | 0.93 | 20 | 417.3 | 102.3 | 44.9 |
| 250 | 1.55 | n/a | Not determined | n/a | n/a |