

Supplementary Information

Breast tumour initiating cell fate are regulated by microenvironmental cues from extracellular matrix

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Table S1. Water contact angle measurements.

ECM substrates	Water contact angle (Deg)
Coll-I	35.7 ± 2
Fn	48.2 ± 2
PDL/Lam-I	54.4 ± 3
PDL	35.1 ± 2

Table S2. Primer sequences for real time PCR.

Gene Name	Forward Sequence 5'-3'	Reverse Sequence 5'-3'
<i>Gapdh</i>	TGCCGCCTGGAGAAACCTGC	CAGCCCCGGCATCGAAGGTG
<i>Ck18</i>	TCCGCGCCCAGTATGAAGCG	GAGGGTGCGTCTCAGCTCCG
<i>Ck14</i>	CGCCGTCTGCTGGAGGGAGA	GGAGACCACCTGCCATCGTGC
<i>Sma</i>	GGCTCCATCCTGGCTTCGCT	GGAGGCGCTGATCCACAAAACG
<i>Abcg2</i>	TTCACCTGTATATTATACTTCATGTTAGGACTG	CGATTGTCATGAGAAGTGTTGCT
<i>Tp63</i>	ACAGACTGCAGCATTGTCAGTTTC	CTTCAGACTTGCCAAATCATCCA
<i>Aldh1</i>	GCTGGGGTGGTGTGGGTT	GACCATGTTACCCAGTTCTCTTC
<i>Gli1</i>	TCGGACCCACTCCAATGAGAA	CTCGATGCCGCTTGGTCAC
<i>CD133</i>	TTATATGGTGTTCACAATCCTGTTATGAC	GCTTCCATGTTGACTATCTTGTTGTTTC

<i>Foxc2</i>	GTGTCCACTGGATAAGGTTTCGTCT	CATCTACTGTACAAAGCCATGCACTTC
<i>Il6</i>	AAATCGTGGAAATGAGAAAAGAGTTGT	TTTCCTGATTATATCCAGTTTGGTAGCA
<i>Snail</i>	GCCTTGTGTCTGCACGACCTG	TCTTCACATCCGAGTGGGTTTG
<i>Twist</i>	CCACGAGCGGCTCAGCTAC	CCTTCTCTGGAAACAATGACATCTAGG
<i>Pai</i>	AGGTAAACGAGAGCGGCACAGT	GCCCATGAAGAGGATTGTCTCTGTC
<i>Cdh2</i>	GCTGATCCTTGTTCATGTTTGTG	GCTGGCTCAAGTCATAGTCCTGGT
<i>Zeb1</i>	AGACACAAATATGAGCACACAGGTAAGA	GCTTGCCACACTTGTCACATTG
<i>CD49f</i> <i>(Ia6)</i>	GGCTCTATTAGTGTTTTTACTGTGGAAGTG	AATACTATGCATCGGAAGTAAGCCTCTC
<i>CD61</i> <i>(Iβ3)</i>	GAGCCAAGTGGGACACAGCA	CGTCATCTGAAGATGGTCTCATTAAGT
<i>Ia1</i>	AACTGGATGGTCATCTTCTGATAATGT	CTGGATTGTGCCTCTTTTGAGAAC
<i>Ia2</i>	TAACATCACCTGCTGGCTGAAAG	CTGCACAGTCTGGAAAGTCGAAG
<i>Ia5</i>	CCCCAAAAGAAACTTCAGGTGG	TTTGAAGAAGCCGAGCTTGTAGAG
<i>Iav</i>	GGATCTCTTCAACTCTACACTAGTCACC	CGTTTGAAAAAGCCCATCCTG
<i>Iβ1</i>	CAAGTGGGACACGGGTGAA	CTACTGTGACTAAGATGCTGCTGCTG
<i>Iβ4</i>	GAGCCCTTCCTCATGGATGGT	GAACTGTTGGTCCATATGAGTGCTG

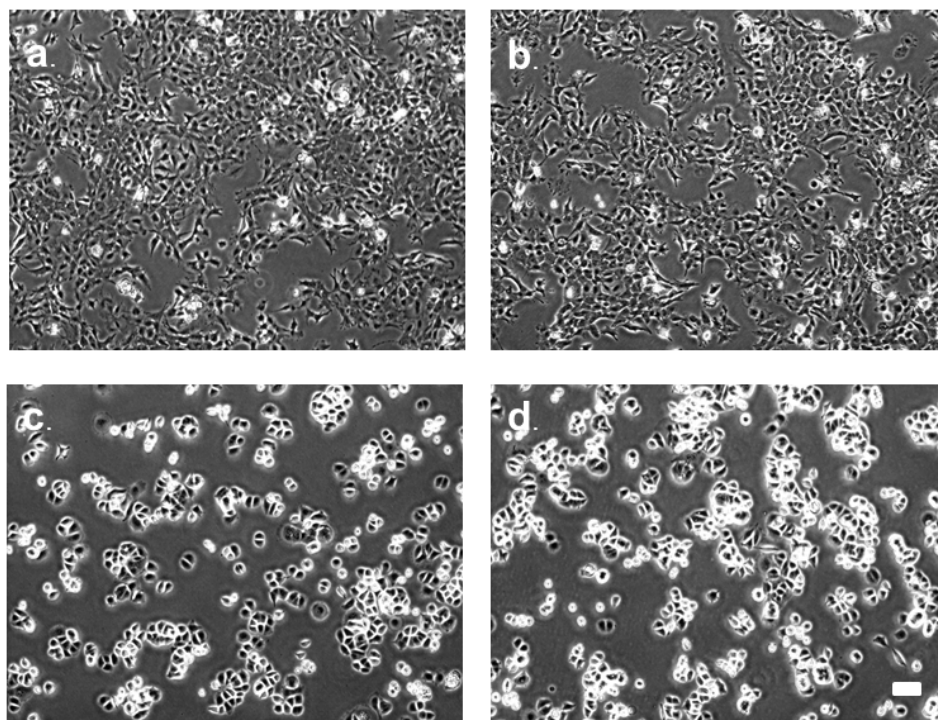


Fig. S1. Phase contrast micrographs of H6O5 cells on (a) Poly-lysine coated TCP, (b) on TCP and MCF-7 cells on (c) Poly-lysine coated TCP, (d) on TCP. No difference in morphology was observed between cells on Poly-lysine coating and TCP. Scale bar = 50 μm .

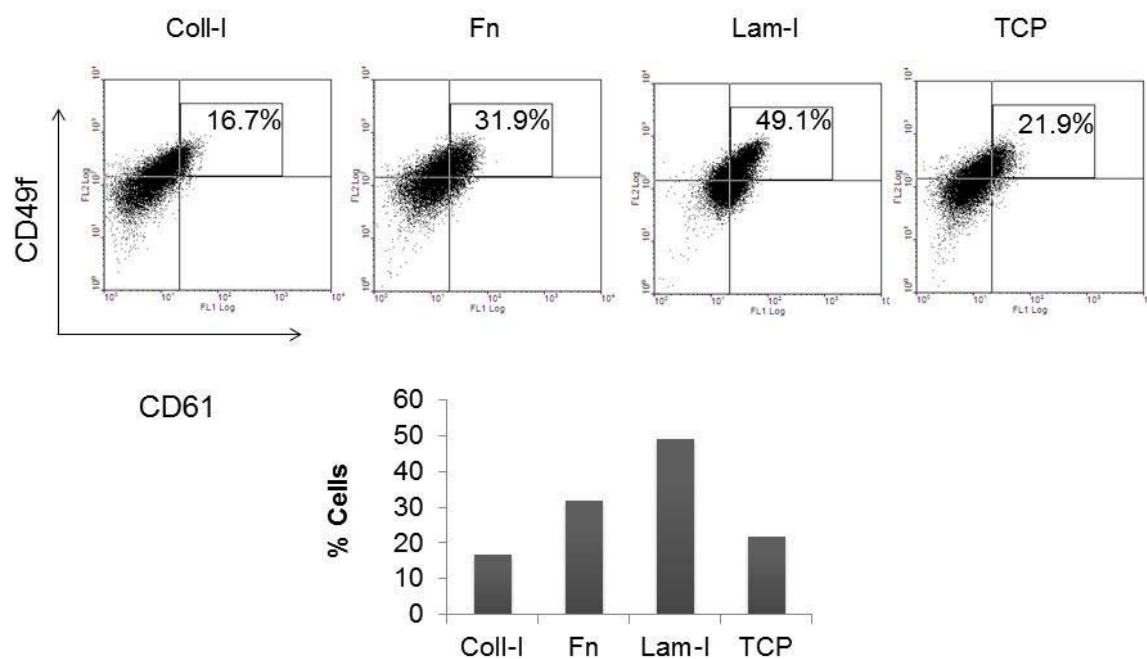


Fig. S2. FACS analysis showed that Lam-I and Fn maintained the majority of the TIC population whereas Coll-I and TCP cell cultures exhibited significant loss of TICs after two weeks of culture on coated substrates. The FACS analysis data were also plotted as a bar graph (the bottom panel).

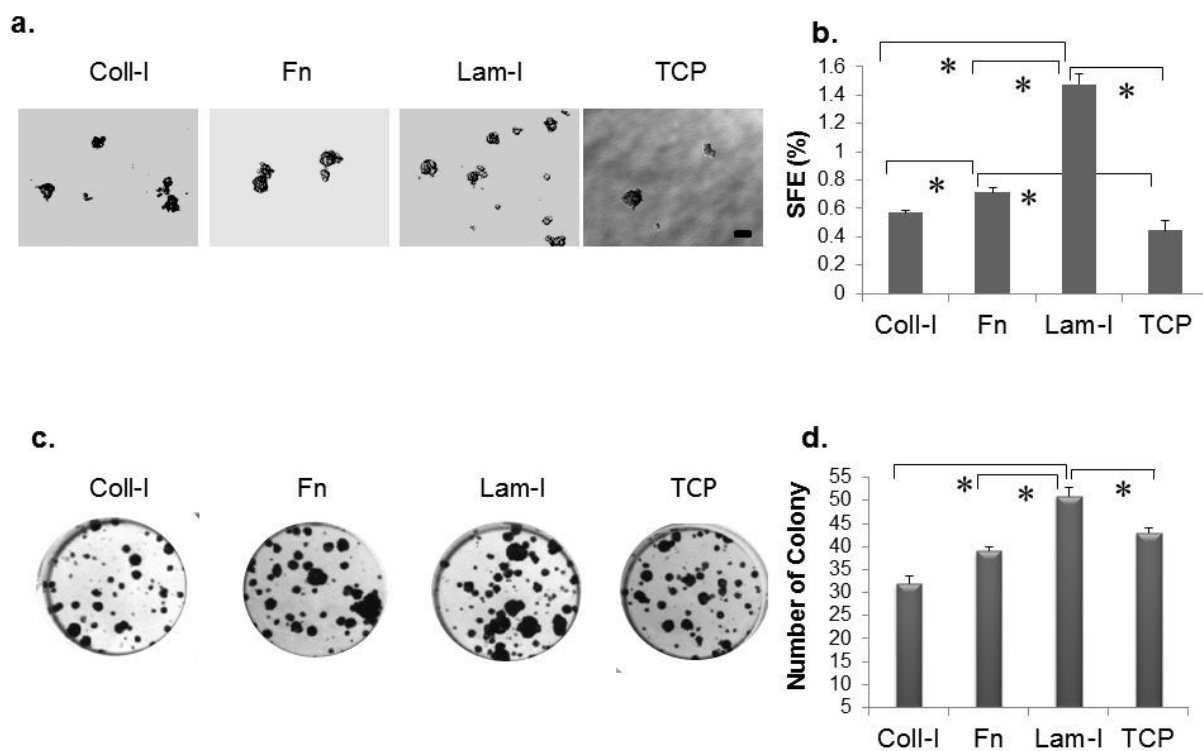


Fig. S3. TICs exhibit greatest self-renewal characteristics in Lam-I after two weeks of culture. (a) *In vitro* tumorsphere formation analysis of TICs retrieved from Coll-I, Fn, Lam-I and TCP. Scale bar = 50 μ m. (b) Plot showing sphere formation efficiency (SFE) for each substrate. (c) *In vitro* clonogenic assay of TICs pre-cultured on Coll-I, Fn, Lam-I and TCP substrates. (d) Plot showing colony numbers for each substrate. All measurements were done in triplicates and data are indicated as mean \pm standard deviation. (*) $p < 0.05$ indicates a statistically significant difference.

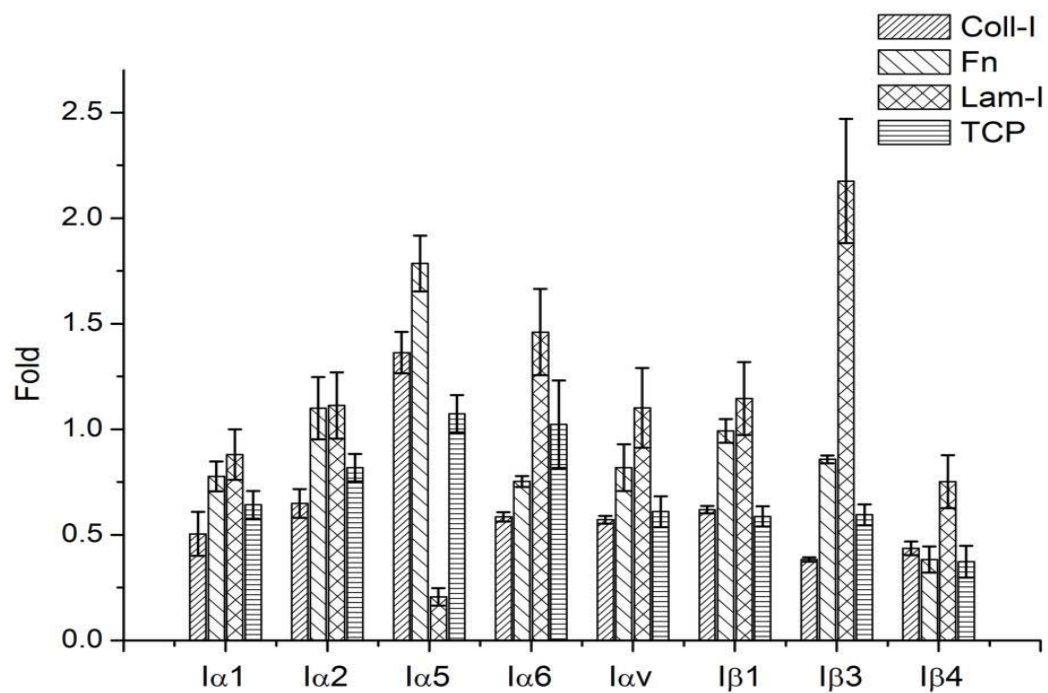


Fig. S4. Expression analysis of integrins in sorted cells cultured on Coll-I Fn, Lam-I and TCP substrates for one week using quantitative real-time reverse transcription PCR analysis. Each expression was normalized by using *Gapdh* as the internal control.

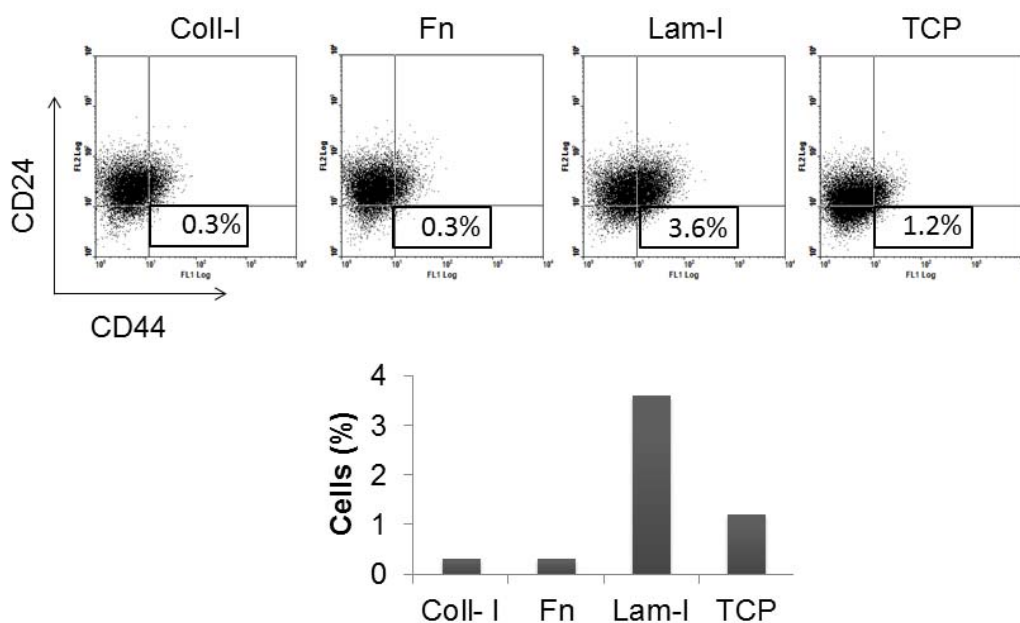


Fig. S5. FACS analysis for CD44^{high}CD24^{low} population in MCF-7 cell line after one week of culture on coated substrates, showed increase in TICs population in Lam-I whereas Coll-I and Fn cultures exhibited significant loss of TICs. The FACS analysis data were also plotted as a bar graph (the bottom panel). MCF-7 is a human breast cancer cell line reported to have very low population of CD44^{high}CD24^{low} that identified as a cancer stem cell like population.