Supporting information

Imidazolide monolayers for reactive microcontact printing⁺

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Characterization of SAMs

Table 1 Advancing $\theta_a(^{o})$ and receding $\theta_r(^{o})$ water contact angles, atomic ratios of C and N from XPS of SAMs shown in Figure 1.

SAMs	$\theta_a(^{o})$	$\theta_r(^{o})$	C:N (XPS)	C:N (calcd)
NH ₂ SAM	68 ± 2	55 ± 2	2.7 ± 0.2	2.5
IM SAM	52 ± 2	27 ± 1	2.5 ± 0.2	2.3

The XPS measurements show a small carbon contamination of the amino (NH₂ SAM) and imidazolide SAMs (IM SAM). The change in C/N ratio between NH₂ SAM and IM SAMs is too small to provide a definite conclusion about the success of formation of the imidazolide SAM. Stronger evidence for the formation of imidazolide groups on the IM SAM stems from the XPS narrow scans of the carbon 1s region. The new peaks at 287.8 and 288.9 eV in the C_{1s} spectrum of the IM SAM are consistent with and characteristic for the N-C(O)-N carbon and N-C-N of the imidazolide intermediate.



Reaction rate of ADF and Atto520

Fig. 1 Reaction rate of ADF and Atto520 printed onto IM SAM and NH₂ SAM

Printing time (min)

Kinetic printing of Atto520

Different printing times were applied ranging from 5 min to 2 h and the substrates were vigorously rinsed and sonicated with acetate buffer for 10 min directly after printing. The intensity of the patterns printing with **IM SAMs** increased in time and leveled off after 1h. Instead, the intensity profiles of the $NH_2 SAMs$ showed consistent relatively weak pattern, which might come from the physisorption of Atto520.



Fig. 2 Fluorescence microscopy images (884 μm x 666 μm) of 100 μm Atto520 printed dots onto **IM** (A,B,C,D,E,F) and **NH₂ SAMs** (G,H,I,J,K,L) for 5 min, 15 min, 30 min, 50 min, 1h and 2h.

Alkyl-chain vs. Fluoroalkyl-chain molecules for reverse microcontact printing

Octadecylamine was used as non-fluorescent ink to compare with the printing result of heptadecafluoroundecylamine. The ink was prepared in acetonitrile (1M) and follows the same procedure for heptadecafluoroundecylamine. A stamp was inked with the solution for 30 s, dried under nitrogen flow, and brought into conformal contact with imidazolide-terminated glass slides for 1h. After 1 h, the stamp was removed and the substrate rinsed with a copious volume of dichloromethane and blown dry with nitrogen. The remaining area was incubated with 0.1 mM Atto520 prepared in MilliQ water overnight. The reverse pattern was not clearly observed compared with heptadecafluoroundecylamine.



Fig. 3 Fluorescence microscopy image (884 μ m x 666 μ m) of Octadecylamine (1 M in acetonitrile) printed 100 μ m diameter dots onto **IM SAM** for 1h. The remaining area was filled by exposing samples to Atto520 in solution overnight (0.1mM in MilliQ water).

boc-protected ADF printing on IM-SAM

The reference molecule boc-ADF was synthesized (see next page) to demonstrate covalent binding between the introducing molecules and **IM SAM** is required for immobilization of the microcontact printed molecules. A 1mM of the boc-protected molecules (boc-ADF) was prepared in acetonitrile and printed onto **IM SAM** for 20 min. As shown in the **Figure 4**, the pattern is completely removed after rinsing and sonicating with THF. These data corroborate covalent binding is occurring in the printing of the non-protected ADF on **IM SAM**.



Fig. 4 Fluorescence microscopy image (884 μ m x 666 μ m) of boc-ADF (1 mM in acetonitrile) printed 100 μ m diameter dots onto **IM SAM** for 20 min before (**A**) and after (**B**) THF rinsing.

Synthesis of boc-ADF

4-(2-aminoethylamino)-7-(N,N-dimethylsulfamoyl)benzofurazan (25mg, 0.0876mmol) was dissolved in 25 ml of water/chloroform (50:50 v/v) in the presence of sodium hydroxide (5.25mg, 0.0131mmol). After stirring for 15 min, di-*tert*-butyl dicarbonate (Boc₂O) (19.12mg, 0.0876mmol) was added at 0 °C, and the reaction mixture was allowed to stand overnight at room temperature. The reaction was extracted with 4 x 50ml of chloroform. The organic layer was dried over MgSO₄ and the solvent was evaporated under reduced pressure. The residue was purified by column chromatography (SiO₂, diclomethane/methanol = 98:2) to give the desired product in 63 % yield as a yellow powder (15.75mg).1H NMR (300 MHz, CDCl₃, 20°C,TMS) δ : 7.83 (d, J=7.8 Hz, 1H), 6.06(d, J=7.8 Hz, 1H), 3.43 (m, 2H, CH₂), 3.12 (m, 2H, CH₂), 2.79 (s, 6H, CH₃), 1.39 (s, 9H, CH₃) ppm.



Fig. 5 Molecular structure of ADF and its reaction with di*-tert*-butyl dicarbonate (Boc₂O) for the preparation of boc-ADF.