

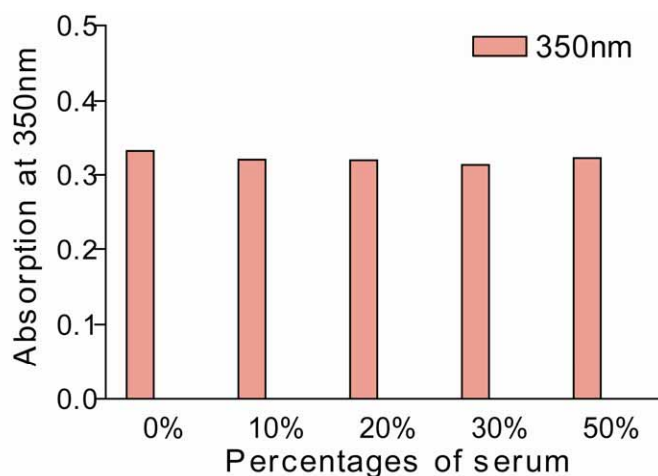
## Supporting Information

### Stability, Toxicity and Differential Cellular Uptake of Protein Passivated-Fe<sub>3</sub>O<sub>4</sub> Nanoparticles

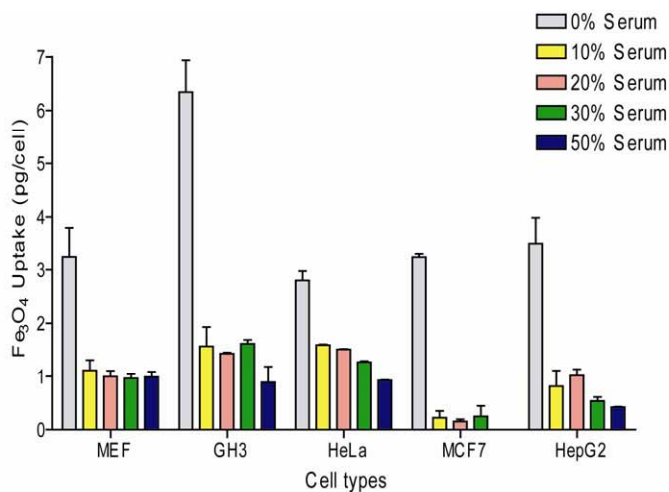
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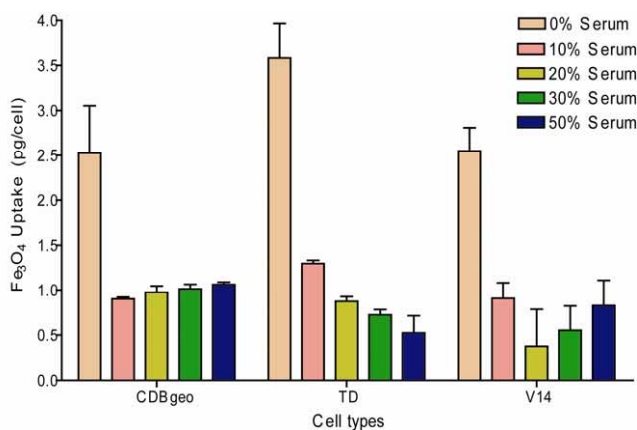
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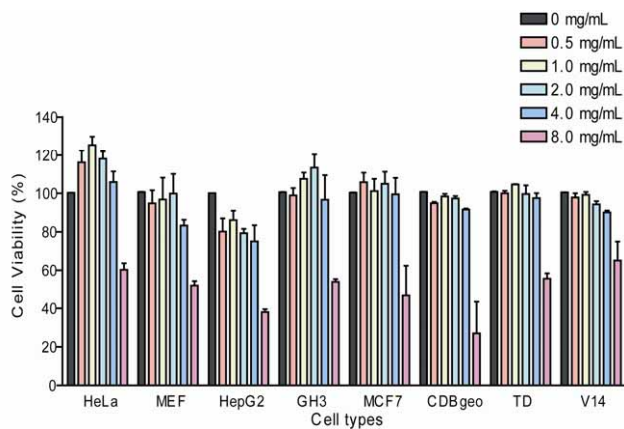
**Figure S-1.** Stability of the BSA coated Fe<sub>3</sub>O<sub>4</sub> nanoparticles at 6 h in the presence of different percentages of serum as evidenced from absorption graph at 350 nm. The BSA coated Fe<sub>3</sub>O<sub>4</sub> nanoparticles (4mg/mL) were dispersed in DMEM media supplemented with 0-50% of fetal bovine serum (FBS).



**Figure S2.** Uptake of the BSA-coated Fe<sub>3</sub>O<sub>4</sub> nanoparticles in different cell types in absence and presence of different percentages of serum. Cells ( $1 \times 10^6$ ) were incubated with BSA-coated Fe<sub>3</sub>O<sub>4</sub> nanoparticles (2mg/mL per well) for 6h. The particle uptake was quantified by Prussian blue assay.



**Figure S3.** Uptake of the BSA-coated Fe<sub>3</sub>O<sub>4</sub> nanoparticles in different cell types in absence and presence of different percentages of serum. Cells ( $1 \times 10^6$ ) were incubated with BSA-coated Fe<sub>3</sub>O<sub>4</sub> nanoparticles (2mg/mL per well) for 6h. The particle uptake was quantified by Prussian blue assay.



**Figure S-4.** Cell viabilities of BSA-coated Fe<sub>3</sub>O<sub>4</sub> nanoparticles in different cell types at various concentrations of nanoparticles. Cells (10, 000) were incubated with BSA-coated Fe<sub>3</sub>O<sub>4</sub> nanoparticles for 6h and cell viability was calculated by Alamar blue assay.