Organic functionalizsation of ultradispersed nanodiamond: Synthesis and applications

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Supplementary material

Thermogravimetric analysis (TGA)

TGA measurements were made in Al_2O_3 crucibles in an atmosphere of nitrogen at a heating rate of 10 K min⁻¹ using a TGA/SDTA851e thermobalance.



Thermogravimetric analysis of ND



Thermogravimetric analysis of ND-1



Thermogravimetric analysis of ND-2





Thermogravimetric analysis of ND-4



Thermogravimetric analysis of ND-5





Thermogravimetric analysis of ND-7







Thermogravimetric analysis of ND-9



Thermogravimetric analysis of ND-10



Thermogravimetric analysis of ND-11



Thermogravimetric analysis of ND-12

General Experimental

All commercial reagents were used without further purification. Solvents were dried according to standard procedures. Raw ND with >95% purity was purchased from Nanostructured and Amorphous Materials, Inc., USA. Transmission FTIR spectra were obtained from KBr pellets using a Perkin Elmer Paragon 1000 FTIR spectrometer, a JASCO FT/IR 410 spectrometer and an ABB FTLA2000 FTIR spectrometer with a MCT detector (samples in vacuum, $\sim 10^{-6}$ torr). TGA measurements were made in Al₂O₃ crucibles in an atmosphere of nitrogen at a heating rate of 10 K min⁻¹ using a TGA/SDTA851e thermobalance. RGA measurements were made using a DYCOR LC series Residual Gas Analyzer (LC 200 series with UHV system, experiment background pressure: 2×10^{-7} torr). Energy-dispersive X-ray spectroscopy (EDS) was performed at SEM/EDS, JEOL JSM-7000F. X-ray diffraction analysis of the samples was carried out using an X-ray diffractometer (XRD D8 Advanced, Bruker) with Cu-Ka radiation. X-Ray Photoelectron Spectroscopy (XPS, Thermo K-Alpha) was carried out using an Al-Ka radiation source and electron energy analyzer (Energy resolution, Ag 3d5/2 peak 0.5 eV, PET 0=C-0 peak 0.85eV). The powder was pressed into an indium foil as a carrier for the measurements. ¹H NMR spectra were obtained on 400 MHz, and ¹³C NMR spectra were obtained at 100.6 MHz using a Bruker NMR spectrometer. Chemical shifts (δ) are reported in ppm relative to CDCl₃ (7.26 and 77.0 ppm respectively). Electrochemical measurements were made using a model CHI420 electrochemical workstation that was controlled by a personal computer. A three-electrode system was used to make the measurements, with a bare glassy carbon electrode as the working electrode, a saturated calomel electrode (SCE) as the reference electrode and a platinum wire as the auxiliary electrode. The measurements were made using 5.00 mL of buffer solutions. Enantiomeric excesses were determined using Lab Alliance Series III high performance liquid chromatograph (HPLC) with a chiracel OD-H chiral column (Daicel Chemical Industries, LTD). Optical rotations were measured using a JASCO P-1010 polarimeter at the indicated temperature using a sodium lamp (D line, 589 nm). Flash column chromatography was performed using MN silica gel 60 (70–230 mesh) that was purchased from Macherey-Nagel.



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Fig. 6 FT-IR spectrum of ND-2







Fig. 9¹³ C-NMR spectrum of 6-(chloro-hexyloxy)-tetrahydropyran







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Fig. 13 ¹H-NMR spectrum of ND-4

















Fig. 21 FT-IR spectrum of ND-16




































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Fig. 37¹³C-NMR spectrum of 3

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Fig. 41 ¹H-NMR spectrum of 4



Fig. 42 13 C-NMR spectrum of 4





Fig. 44 ¹H-NMR spectrum of 5

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Fig. 45 13 C-NMR spectrum of 5





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Fig. 50 ¹H-NMR spectrum of 7



Fig. 51 ¹³C-NMR spectrum of 7



Fig. 52 FT-IR spectrum of 7





Fig. 54 ¹³C-NMR spectrum of 8







Current Data Parameters NAME EXPNO 2 PROCNO 1	F2 - Acquisition Farameters Date14.46 Time14.46 INSTRUM 5 mm QNP 1H/13 FULFROG 590930 FULFROG 59930 SOLVENT CDC13 NS 25530 203 Hz 55536 sec CDC13 NS 25510.203 Hz 1.2845556 sec RG 1.2845556 sec RG 1.28455556 sec RG 1.284555555556 sec RG 1.284555555555555555555555555555555555555	NUC1 CHANNEL fl 13C 13C 9.80 usec PL1 9.80 usec PL1 100.6248425 MHz	CPDPRG2 CHANNEL f2	F2 - Processing parameters si 327690 MHz SF 100.6127690 MHz SSB 200 Hz GB 2.00 Hz GB	Ppm 1.00
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Fig. 57 ¹³C-NMR spectrum of 9







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Fig. 60¹³C-NMR spectrum of 10



Fig. 63 HPLC analysis of 2-[Hydroxy-(4-nitro-phenyl)-methyl]-cyclohexanone (4). (Chiralcel OD-H column, hexane:2-propanol = 9:1, 0.5 mL/min, 254 nm UV detector)

a) 82.3% ee (Table 1; entry 14)



b) racemic (4)



Fig. 64 HPLC analysis of 2-[Hydroxy-(2-nitro-phenyl)-methyl]-cyclohexanone (5).
(Chiralcel OD-H column, hexane:2-propanol = 9:1, 0.5 mL/min, 254 nm UV detector)
a) 87.3% ee (Table 2; entry 1)















Fig. 66 HPLC analysis of 3-[Hydroxy-(2-oxo-cyclohexyl)-methyl]-benzonitrile (7). (Chiralcel OD-H column, hexane:2-propanol = 9:1, 0.5 mL/min, 254 nm UV detector)

a) 71.1% ee (Table 2; entry 3)






Fig. 67 HPLC analysis of 4-[Hydroxy-(2-oxo-cyclohexyl)-methyl]-benzonitrile (8).

(Chiralcel OD-H column, hexane:2-propanol = 9:1, 0.5 mL/min, 254 nm UV detector) a) 72.9% ee (Table 2; entry 4)







Fig. 68 HPLC analysis of 2-[(**4-Chloro-phenyl**)-**hydroxy-methyl**]-**cyclohexanone (9).** (Chiralcel OD-H column, hexane:2-propanol = 9:1, 0.5 mL/min, 254 nm UV detector) **a) 66.3% ee** (Table 2; entry 5)



b) racemic (9)



Fig. 69 HPLC analysis of 2-[Hydroxy-(2-methoxy-phenyl)-methyl]-cyclohexanone (10). (Chiralcel OD-H column, hexane:2-propanol = 9:1, 0.5 mL/min, 254 nm UV detector)

a) 68.9% ee (Table 2; entry 6)



b) racemic (10)

