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Electronic Supplementary Information (ESI)

Novel and Efficient MR active aqueous colloidal Fe₃O₄ nanoassemblies

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Fig. S1. XRD patterns of (a) Fe_3O_4 MNNA and (b) Fe_3O_4 MNP samples (position and relative intensities of all diffraction peaks well matched with those from the JCPDS card 75-1609 of magnetite).

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Fig. S2. (a) Low magnification TEM micrograph and (b) large area SEM micrograph of Fe_3O_4 MNNA samples. Insets of (a) shows the size distribution of nanoparticles assembled in a single Fe_3O_4 MNNA and inset of (b) shows the size distribution of Fe_3O_4 MNNAs.



Fig. S3. (a) Electron diffraction pattern of Fe_3O_4 MNNA and (b) high resolution TEM image of Fe_3O_4 MNNA sphere (yellow coloured arrow marks indicate the direction of orientation of nanoparticles assembled in MNNA and inset shows the magnified image of the square marked region). The electron diffraction pattern of Fe_3O_4 MNP was also similar to that of Fe_3O_4 MMNA.



Fig. S4. DLS measurements of Fe₃O₄ MNNA, Fe₃O₄ MNP and ferumoxytol in aqueous medium showing mean hydrodynamic diameter of 90, 17 and 25 nm ($\sigma < 5$ %), respectively (x-axis in log scale)



Fig. S5. FTIR spectra of amine functionalization of Fe_3O_4 MNNA, bare Fe_3O_4 MNNA (control experiment: Fe_3O_4 MNNA synthesized in absence of EDA following a similar process) and ethylenediamine.

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Fig. S6. A schematic representation of the effect of magnetic field on MR contrast properties of Fe₃O₄ MNP and Fe₃O₄ MMNA



Fig. S7. Percentage viability of HeLa cells after 24 h incubation of stable aqueous suspension of amine functionalized Fe₃O₄ MNNA (control: viability of HeLa cells without Fe₃O₄ MNNA). The results are shown as mean \pm standard deviation (n = 4).