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Supporting Information for

Smartly designed photoreactive silica nanoparticles and their reactivity.

by

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Supporting information

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List of abbreviations:

AFM	atomic force microscopy
APTES	(3-aminopropyl)triethoxysilane
CDI	1,1'-carbonyldiimidazole
CP-MAS	cross polarization-magic angle spinning
DLS	dynamic light scattering
DMAP	para-dimethylaminopyridine
DMSO	dimethylsulfoxide
DSS	4,4-dimethyl-4-silapentane-1-sulfonic acid
EtOH	ethanol
FE	field emission
FIB	focused ion beam
FTIR	fourier transform infrared spectroscopy
HRMS	high resolution mass spectra
NMR	nuclear magnetic resonance
RT	room temperature
SEM	scanning electron microscope
ssNMR	solid state nuclear magnetic resonance
TEOS	tetraethyl orthosilicate
THF	tetrahydrofurane
TMS	tetramethylsilane
TEM	transmission electron spectroscopy
UV	ultra violet
XPS	X-ray photoelectron spectroscopy

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Literature compounds:

4-azidobenzoic acid

The 4-azidobenzoic acid was prepared according to a literature procedure.^[1]

4-azido-2,3,5,6-tetrafluorobenzoic acid



The 4-azido-2,3,5,6-tetrafluorobenzoic acid was prepared according to a literature procedure.^[2]

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Experimental Procedures:

Synthesis of 4-azido-N-(3-(triethoxysilyl)propyl)benzamide 1 (PATES)



4-azidobenzoic acid

Yellow oil; TLC, $R_f = 0.24$ (eluent: acetone /*n*-hexane: 85/15); v_{max} (KBr easy diff)/cm⁻¹ 3322 (NH), 2973 and 2885 (CH stretching), 2927 (CH₂ stretching), 2124 (azide), 1639 (C=O amide), 1549 and 1287 (C=C), 1287 (C-O), 1445 and 765 (CH₂ bending), 1390 (Si-OCH₂CH₃), 1100 (Si-O),956 (C-O); $\delta_{\rm H}$ ¹H NMR (300 MHz; DMSO- d_6) 8.48 (1H, br t, NH), 7.91 (2H, d, J = 8.7 Hz, Ar-H), 7.17 (2H, d, J = 8.7 Hz, Ar-H), 3.74 (6H, q, J = 6.9 Hz, O-CH₂-CH₃), 3.27-3.21 (2H, m, NH-CH₂-CH₂), 1.64-1.54 (2H, m, CH₂-CH₂-CH₂), 1.13 (9H, t, J = 6.9 Hz, O-CH₂-CH₃), 0.61-0.56 (2H, m, CH₂-Si); $\delta_{\rm C}$ ¹³C NMR (75 MHz, [D6] DMSO, TMS) 165.1 (C), 142.0 (C), 131.3 (C), 129.0 (CH), 118.7 (CH), 57.7 (CH₂), 42.0 (CH₂), 22.7 (CH₂), 18.1 (CH₃), 7.4 (CH₂); UV/Vis λ_{max} (EtOH)/nm (270); HRMS (DCI+CH4) m/z calcd for $C_{16}H_{26}N_4O_4Si$ $(MH)^+$ 366.4875 found 366.1723.

Synthesis of 4-azido-2,3,5,6-tetrafluorobenzoyl chloride



The 4-azido-2,3,5,6-tetrafluoroenzoic acid (2.20 g, 9.36 mmol) was dissolved in dry CH₂Cl₂ (10 mL). In order to dissolve 4-azido-2,3,5,6-tetrafluoroenzoic acid completely dry DMF (0.3 mL) was added to a reaction mixture. Thus oxalyl chloride (3.6 mL, 42.12 mmol) was added and reaction was stirred at RT for 2h during with time gas evolution was observed. After reaction completion the solvents were evaporated to give 4-azido-2,3,5,6-tetrafluorobenzoyl chloride as yellow oil which was directly used in the next step without further purification.

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Synthesis of 4-azido-2,3,5,6-tetrafluoro-N-(3-(triethoxysilyl)propyl)benzamide

2 (PFPATES)



Yellow oil; TLC, Rf = 0.37 (eluent: ether/n-hexane: 60/40); v_{max} (KBr easy diff)/cm⁻¹ 3280 (NH), 3080 and 2884 (CH stretching), 2930 (CH₂ stretching), 2133 (azide), 1650 (C=O amide), 1392 (Si-OCH₂CH₃), 1100 (Si-O), 958 (C-O), 777 (CH₂ bending); $\delta_{\rm H}$ ¹H NMR (300 MHz, CDCl₃) 6.42 (1H, br t, NH), 3.81 (6H, q, J = 6.9 Hz, O-<u>CH₂-CH₂), 3.51-3.44 (2H, m, NH-<u>CH₂-CH₂), 1.81-1.71 (2H, m, CH₂-<u>CH₂-CH₂), 1.21 (9H, t, J = 6.9 Hz, O-CH₂-<u>CH₃), 0.73-0.66 (2H, m, CH₂-Si); $\delta_{\rm C}$ ¹³C NMR (75 MHz; CDCl₃) 157.4 (C), 145.7 (C), 142.1 (C), 138.7 (C), 112.1 (C), 58.5 (CH₂), 42.4 (CH₂), 22.5 (CH₂), 18.2 (CH₃), 7.6 (CH₂); $\delta_{\rm F}$ ¹⁹F NMR (188 MHz; CDCl₃) 141.3-141.5 (m, 2F), 150.9-151.1 (m, 2F); UV/Vis λ_{max} (EtOH)/nm (260); HRMS (DCI+ CH₄) *m/z* calcd for C₁₆H₂₂F₄N₄O₄Si (MH-EtOH)⁺ 393.3810 found 393.1006.</u></u></u></u>





¹HNMR was identical to previously reported. ^[3]

White solid; TLC, $R_f = 0.68$ (eluent: acetone /*n*-hexane: 85/15); m.p. 90-91°C; v_{max} (KBr easy diff)/cm⁻¹ 3326 (NH), 2975 and 2885 (CH stretching), 2928 (CH₂ stretching), 1663 (C=O ketone), 1631 (C=O amide), 1553 (C=C), 1445 and 790 (CH₂ bending), 1390 (Si-OCH₂CH₃), 1300 (C-H bending), 1278 and 1108 (C-O), 1167 (C-N), 1080 (Si-O) cm⁻¹; δ_H ¹H NMR (300 MHz, [D₆] DMSO, TMS) 8.68 (1H, br t, NH), 8.02-7.98 (2H, m, Ar-H), 7.78-7.68 (5H, m, Ar-H),

7.61-7.56 (2H, m, Ar-H), 3.77 (6H, q, J = 9.6 Hz, O-<u>CH₂</u>-CH₃), 3.35-3.25 (2H, m, NH-<u>CH₂</u>-CH₂), 1.67-1.57 (2H, m, CH₂-<u>CH₂</u>-CH₂), 1.15 (9H, t, J = 9.6 Hz, O-CH₂-<u>CH₃</u>), 0.65-0.59 (2H, m, <u>CH₂</u>-Si); $\delta_{\rm C}$ ¹³C NMR (75 MHz, [D₆] DMSO, TMS) 195.4 (C), 165.3 (C), 139.0 (C), 138.0 (C), 136.7 (C), 133.0 (CH), 129.7 (CH), 129.5 (CH), 128.7 (CH), 127.3 (CH), 57.7 (CH₂), 42.1 (CH₂), 22.7 (CH₂), 18.2 (CH₃), 7.5 (CH₂); UV/Vis $\lambda_{\rm max}$ (EtOH)/nm (260); HRMS (DCI+ CH₄) m/z calcd for C₂₃H₃₁NO₅Si (MH)⁺ 429.5814 found 430.248.



¹H- and ¹³C- NMR Spectra:

Figure 1. ¹H NMR spectrum of **1** (PATES)



Figure 2. ¹³C NMR spectrum of **1** (PATES)





Figure 3. ¹H NMR spectrum of **2** (PFPATES)





Figure 4. ¹⁹ F NMR spectrum of 2 (PFPATES)



Figure 5. TGA of a) SiO₂, b) SiO₂@BPh, c) SiO₂@PFPA, d) SiO₂@PA NPs. TGA was performed in air at a heating rate of 10 °C/min





Figure 6. a) ¹³C CP/MAS NMR spectrum of the SiO₂@PFPA NPs; b) ¹³C NMR (75 MHz; CDCl₃) spectrum of **2** (PFPATES)



Figure 7. a) ¹³C CP/MAS NMR spectrum of the SiO₂@BPh NPs; b) ¹³C NMR (75 MHz; $[D_6]$ DMSO) spectrum of **3** (BPhTES)

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UV absorbance spectra:



Figure 8. UV/Vis spectra of the 3 (BPhTES) and SiO₂@BPh NPs



Figure 9. UV/Vis spectra of the 2 (PFPATES) and SiO₂@PFPA NPs



Figure 10. UV/Vis spectra of the 1 (PATES) and SiO₂@PA NPs



FT-IR spectra:

Figure 11. FTIR spectra of 1) SiO₂@BPh, 2) SiO₂@PA, and 3)SiO₂@PFPA NPs

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Size distribution graphs (according to TEM):

Figure 12. Size distribution of the SiO₂@BPh NPs



Figure 13. Size distribution of the SiO₂@PFPA NPs



Figure 14. Size distribution of the SiO₂@PFPA NPs



Figure 15. Size distribution of the SiO₂ NPs

SEM tilt images:



Figure 16. SEM tilt image of PC-SiO₂@PA film



Figure 17. SEM tilt image of PC-SiO₂@PFPA film



Figure 18. SEM tilt image of PC-SiO₂@BPh film



Figure 19. FE-SEM image of PC-SiO₂@PFPA modified film (the analysis was performed without gold conducting coating)

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AFM images:



Figure 20. AFM image of PC-SiO₂@PA film



Figure 21. AFM image of PC-SiO₂@PFPA film



Figure 22. AFM image of PC-SiO₂@BPh film

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AFM NP manipulation images:

Figure 23. AFM image of PC-SiO₂@PA film : a) before and b) after NP manipulations. Particle 1 was moved with vertical tip depth of Z = -40nm and particle 2 with Z = -30nm both particles did not moved and scratches can be seen for both cases.



Figure 24. AFM image of PC-SiO₂@PFPA film : a) before and b) after NP manipulations. Particle 1 was moved with vertical tip depth of Z = -20nm and Z = -30, particle 2 with Z = -30nm and particle 3 with Z = -40nm all three particles did not moved and a dip scratch can be seen for 3 and a hole in the place where the tip stopped for 2.

Figure 25. AFM image of PC-SiO₂@BPh film: a) before and b) after NP manipulations. Particle 1 was moved with vertical tip depth of Z = -30 and Z = -40nm and particle 2 with Z = -40nm, Z = -50nm and Z = -60nm both particles did not moved and scratch can be seen for 1.

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Neat PC film



Figure 26. SEM image of neat PC film



Figure 27. AFM image of neat PC film



Figure 28. XPS spectra of neat PC film



Figure 29. XPS spectra of PC-SiO₂ film



Figure 30. XPS spectra of PC-SiO₂@PFPA film



Figure 31. XPS spectra of PC-SiO₂@BPh film



Figure 32. XPS spectra of PC-SiO₂@PA film



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Figure 33. DIC filter images of A) PC-SiO₂@PA ; B) PC-SiO₂@PFPA; C) PC-SiO₂@BPh, and D) PC-SiO₂ films



Figure 34. merged GFP, CFP, and DIC filters fluoresence images of A) PC-SiO₂@PA ; B) PC-SiO₂@PFPA; C) PC-SiO₂@BPh, and D) PC-SiO₂ films

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- [3] H. Li, G. McGall, *Frontiers in Biochip Technology*, Springer, 2006.