

Electronic Supplementary Information

Sensing of Linear Alkylammonium Ions by a 5-Pyrenoylamido-Calix[5]arene Solution and Monolayer Using Luminescence Measurements

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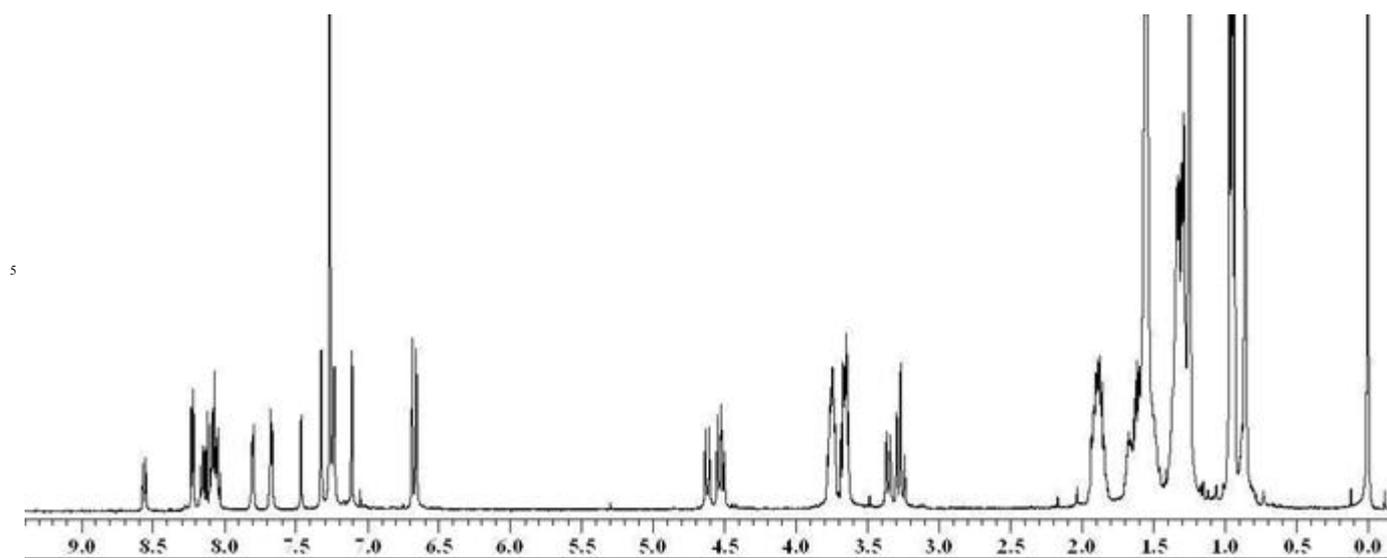


Figure S1. ^1H NMR spectrum of the compound **2** in CDCl_3 .

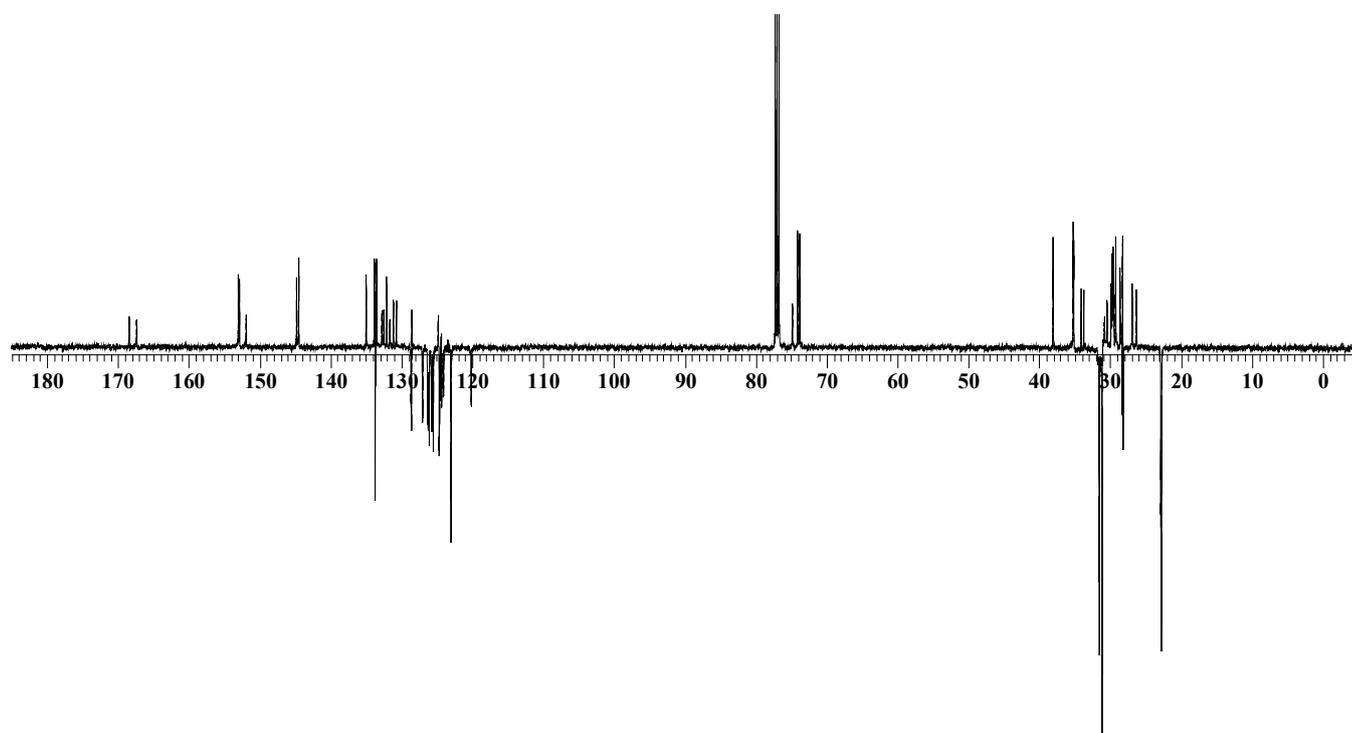


Figure S2. APT spectrum of the compound **2** in CDCl_3 .

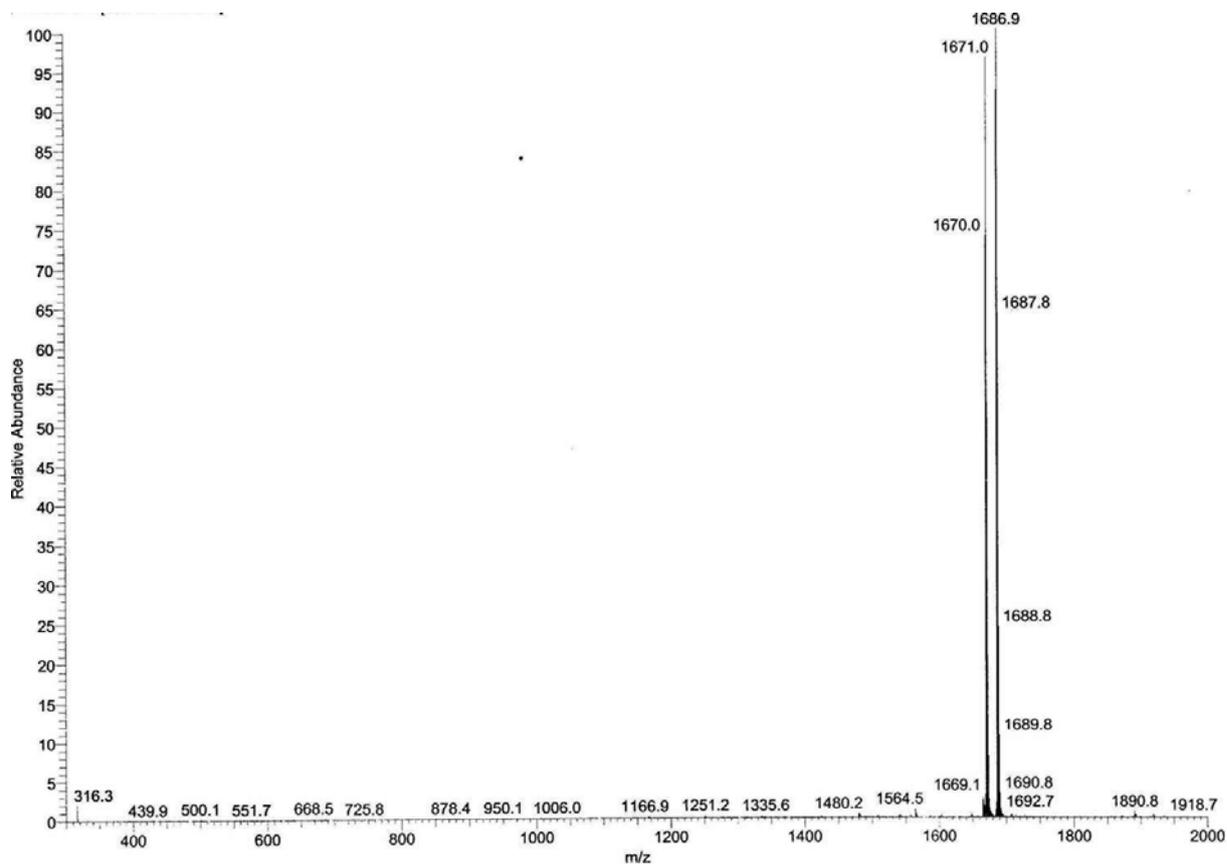


Figure S3. ESI-MS of the compound 2.

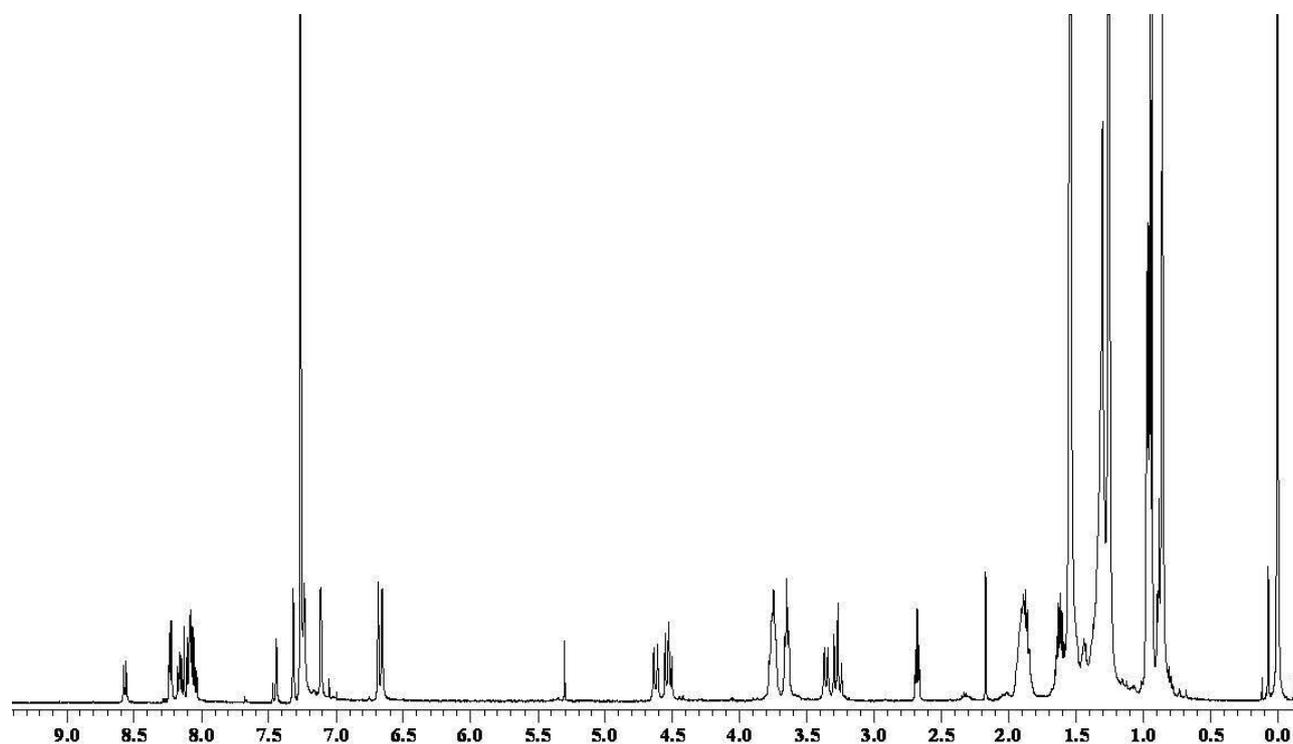


Figure S4. ^1H NMR spectrum of PyC5-NH₂ in CDCl₃.

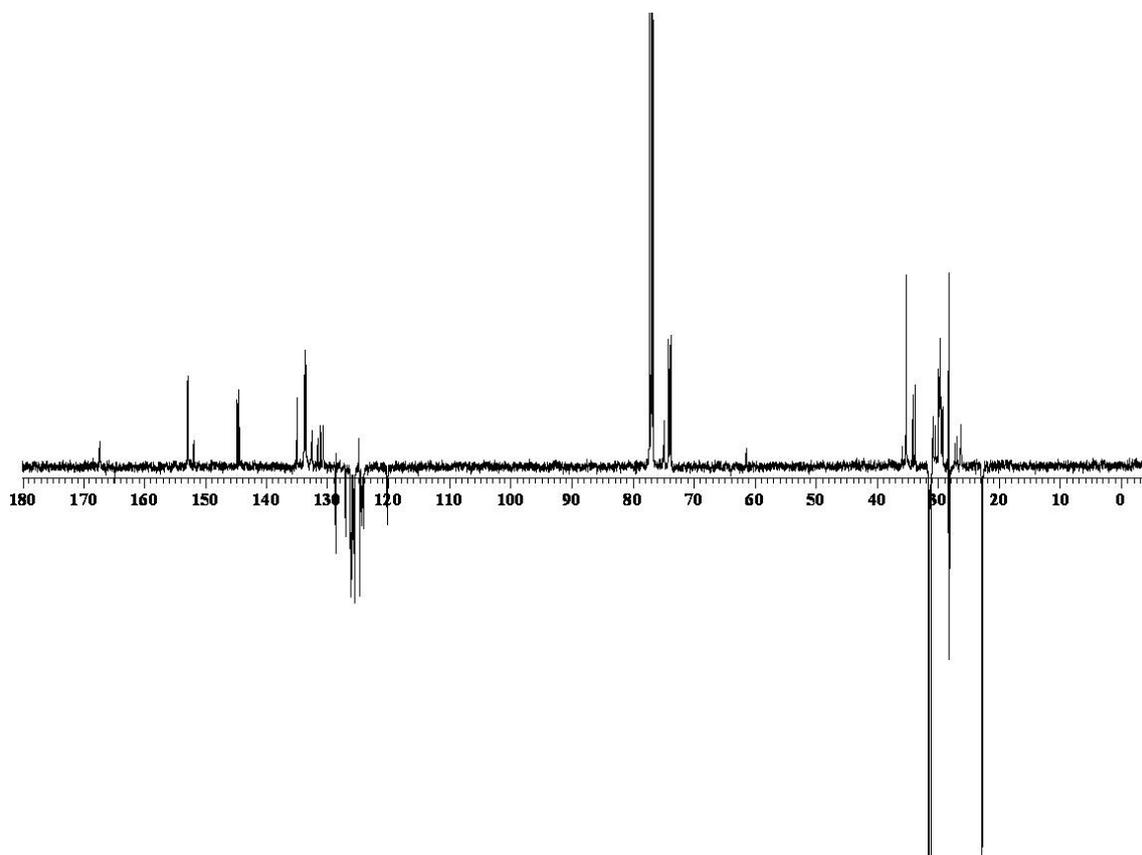


Figure S5. APT spectrum of **PyC5-NH₂** in CDCl₃.

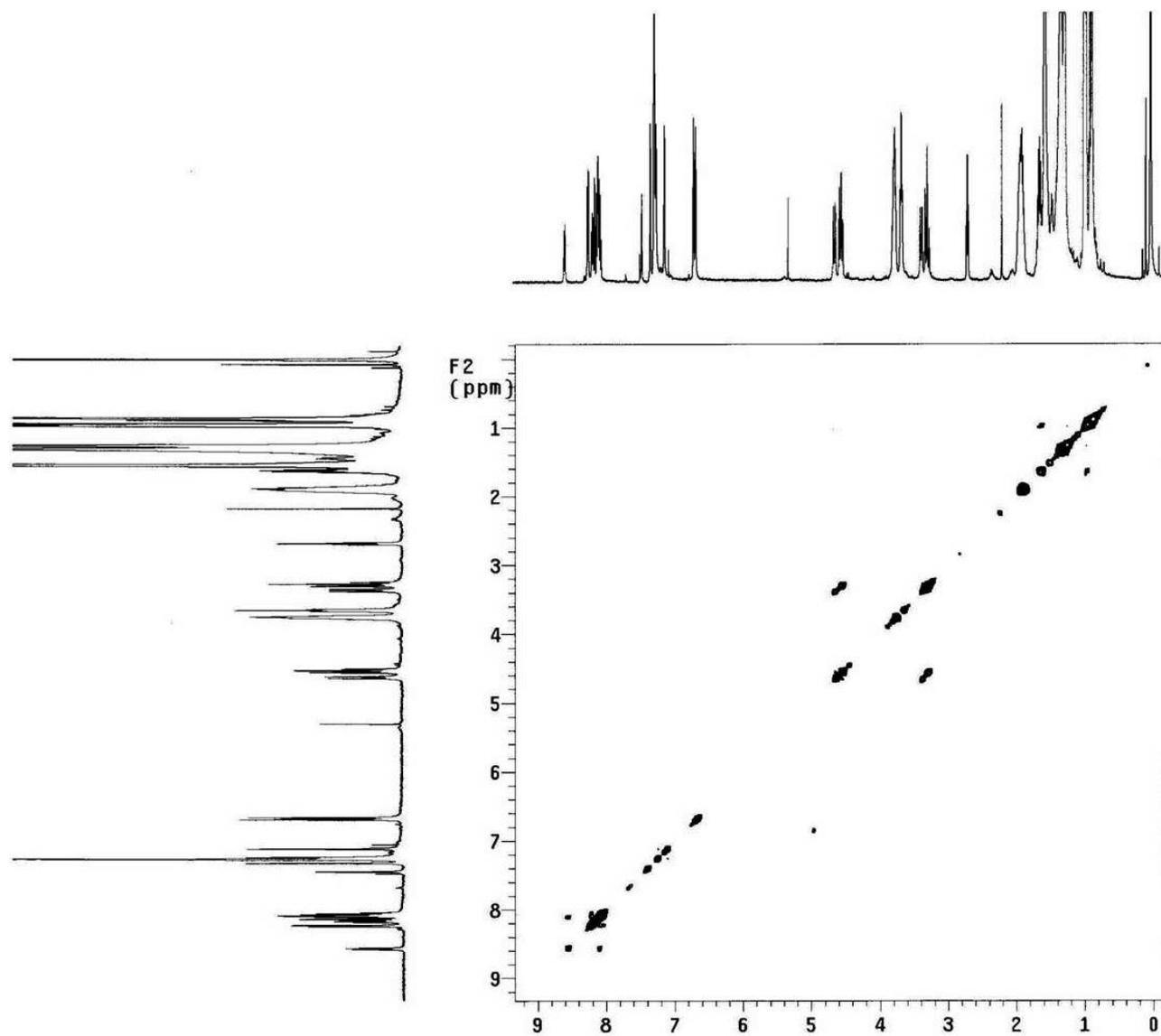


Figure S6. 2D-gCOSY spectrum of **PyC5-NH₂** in CDCl₃.

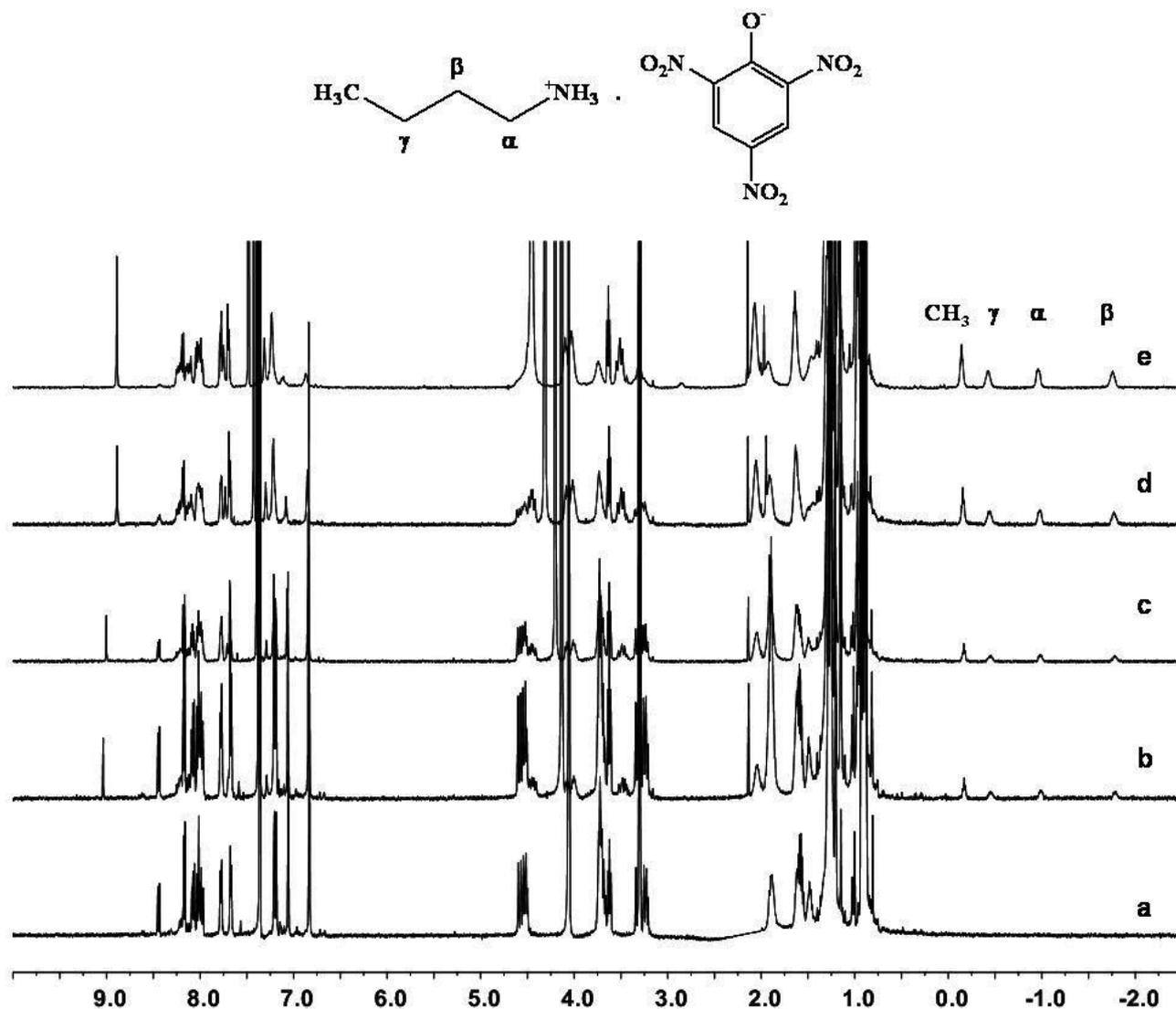


Figure S7. ^1H NMR titration of $n\text{-Bu}\cdot\text{HPic}$ with compound **3**. Constant concentration of host **3** (2.00×10^{-3} M) with addition of various concentrations ($0\text{--}4.00\times 10^{-3}$ M) of guest $n\text{-Bu}\cdot\text{HPic}$.
a) compound **3**; b) $[\text{H}]/[\text{G}]$ 4:1; c) $[\text{H}]/[\text{G}]$ 2:1; d) $[\text{H}]/[\text{G}]$ 1:1; e) $[\text{H}]/[\text{G}]$ 1:2. Signals for the free $n\text{-Bu}\cdot\text{HPic}$ guest are absent in the expected 0.8–3 ppm range. In contrast, they appear in the 0– -2 ppm range, thus confirming the endo-complexation of the guest. In addition, an evident intensity increase of these signals parallels the increasing in the guest concentration.

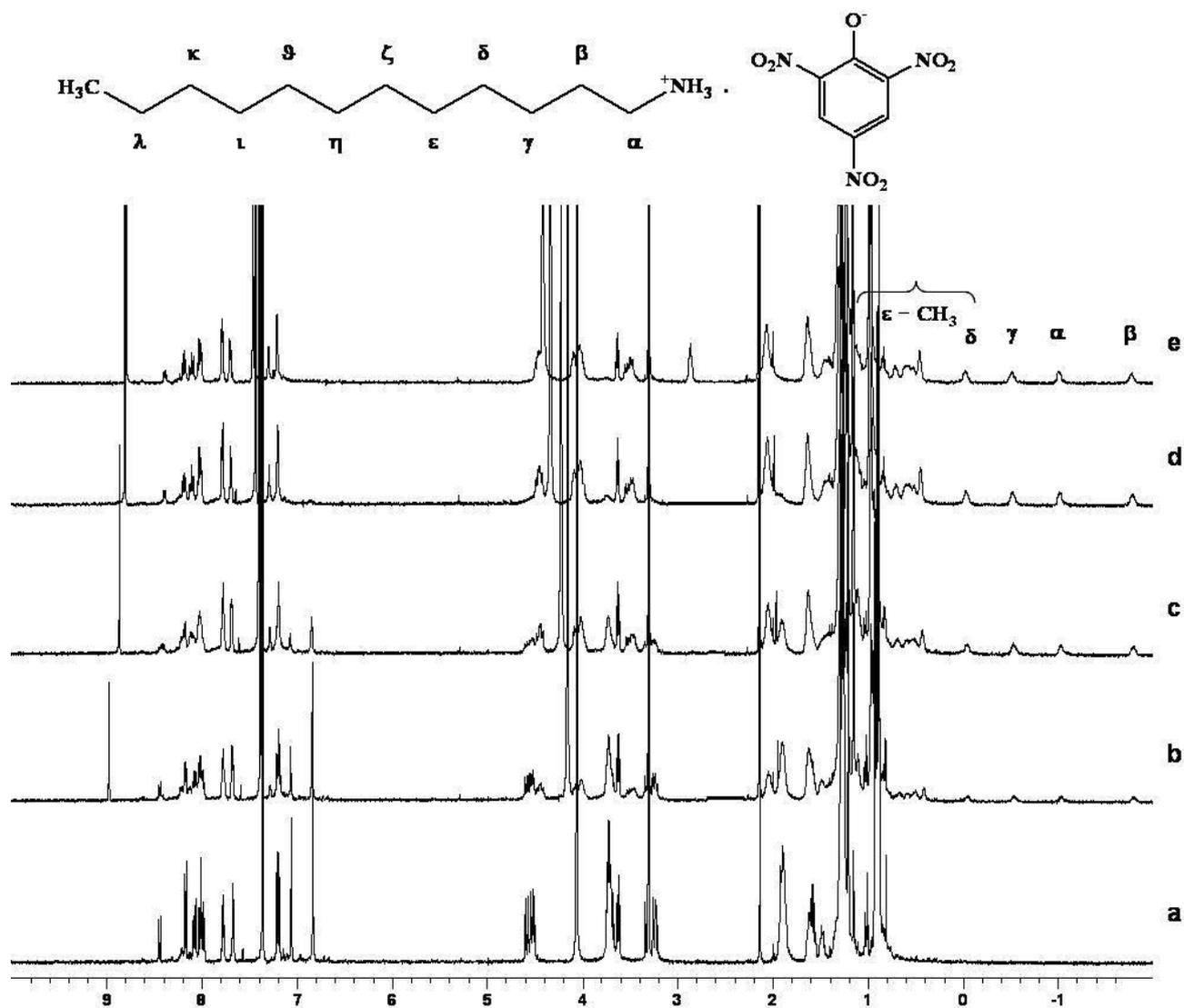


Figure S8. ^1H NMR titration of $n\text{-C}_{12}\text{NH}_2\cdot\text{HPic}$ with compound **3**. Constant concentration of host **3** (2.00×10^{-3} M) with addition of various concentrations ($0\text{--}4.00\times 10^{-3}$ M) of guest $n\text{-C}_{12}\text{NH}_2\cdot\text{HPic}$.

a) compound **3**; b) $[\text{H}]/[\text{G}]$ 4:1; c) $[\text{H}]/[\text{G}]$ 2:1; d) $[\text{H}]/[\text{G}]$ 1:1; e) $[\text{H}]/[\text{G}]$ 1:2. Signals for the free $n\text{-C}_{12}\text{NH}_2\cdot\text{HPic}$ guest are absent in the expected 0.8-3 ppm range. In contrast, they appear in the 0-2 ppm range, thus confirming the endo-complexation of the guest. In addition, an evident intensity increase of these signals parallels the increasing in the guest concentration.

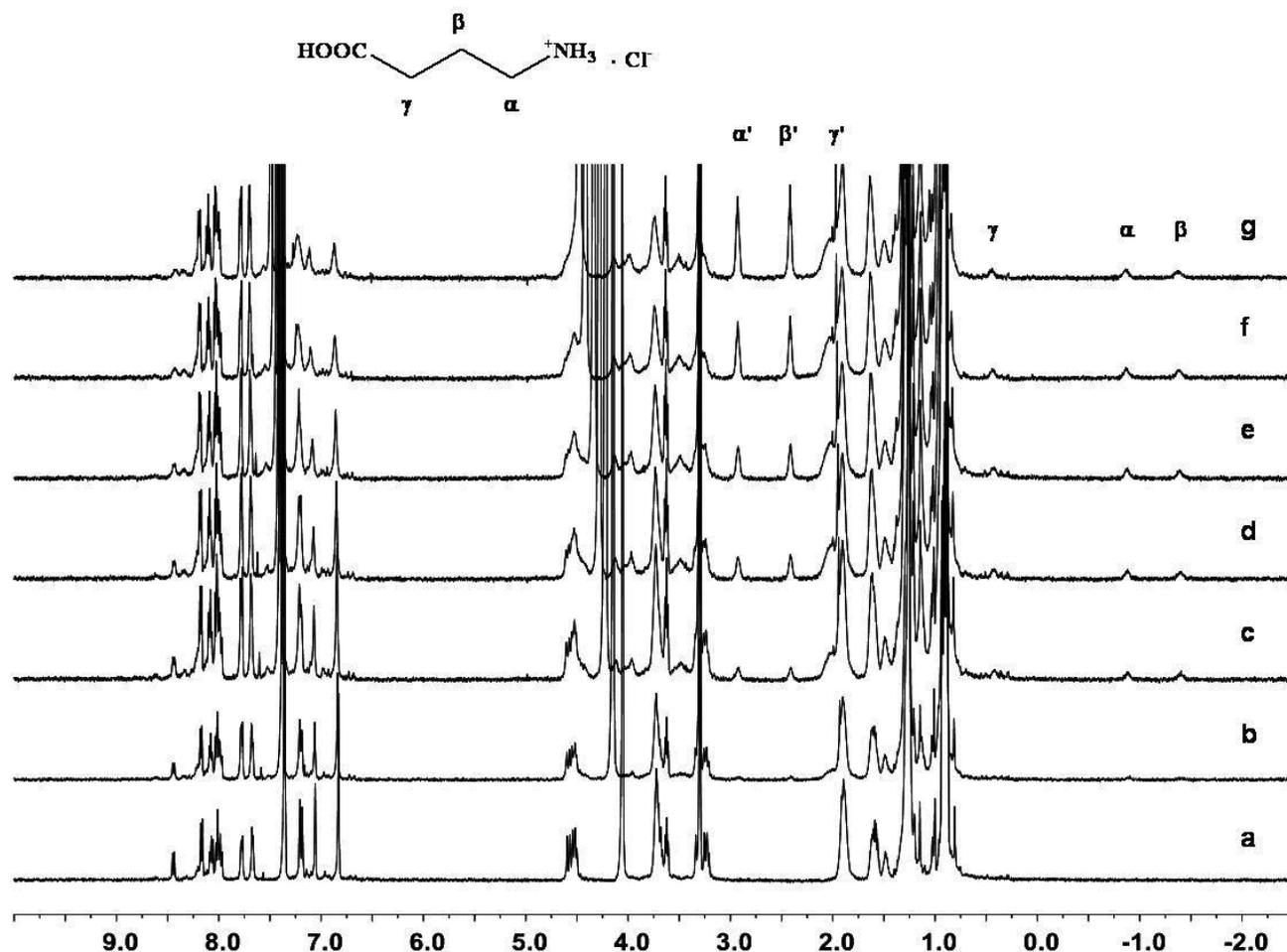


Figure S9. ^1H NMR titration of **GABA·HCl** with compound **3**. Constant concentration of host **3** (2.00×10^{-3} M) with addition of various concentrations (0 – 4.00×10^{-3} M) of guest **GABA·HCl**.
s a) compound **3**; b) $[\text{H}]/[\text{G}]$ 4:1; c) $[\text{H}]/[\text{G}]$ 2:1; d) $[\text{H}]/[\text{G}]$ 1:0.75; e) $[\text{H}]/[\text{G}]$ 1:1; f) $[\text{H}]/[\text{G}]$ 1:1.5; g) $[\text{H}]/[\text{G}]$ 1:2. Signals for the free **GABA·HCl** guest are present in the expected 2.0-3.0 ppm range (α' β' γ'). In addition, signals of the complexed guest appear in the 0.5– -1.5 (α , β , γ) ppm range, thus confirming partial endo-complexation of this guest.

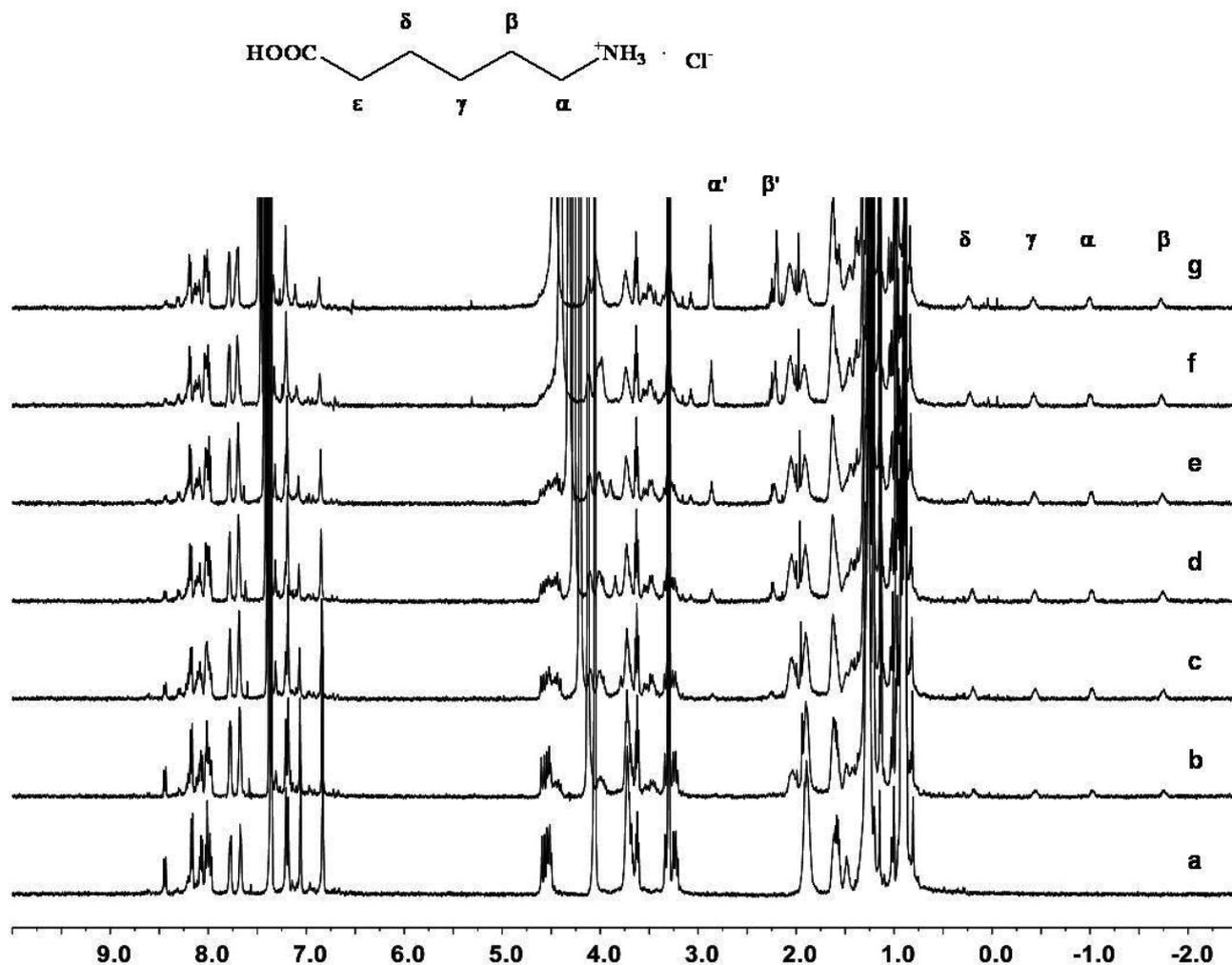


Figure S10. ^1H NMR titration of ϵ -Ahx·HCl with compound 3. Constant concentration of host 3 (2.00×10^{-3} M) with addition of various concentrations (0 – 4.00×10^{-3} M) of guest ϵ -Ahx·HCl. a) compound 3; b) [H]/[G] 4:1; c) [H]/[G] 2:1; d) [H]/[G] 1:0.75; e) [H]/[G] 1:1; f) [H]/[G] 1:1.5; g) [H]/[G] 1:2. Signals for the free ϵ -Ahx·HCl guest are present in the expected 1.0–3.0 ppm range (α' , β' , γ' , δ' , ϵ'). In addition, signals of the complexed guest appear in the 1.0 – -2.0 (α , β , γ , δ , ϵ) ppm range, thus confirming partial endo-complexation of this guest.

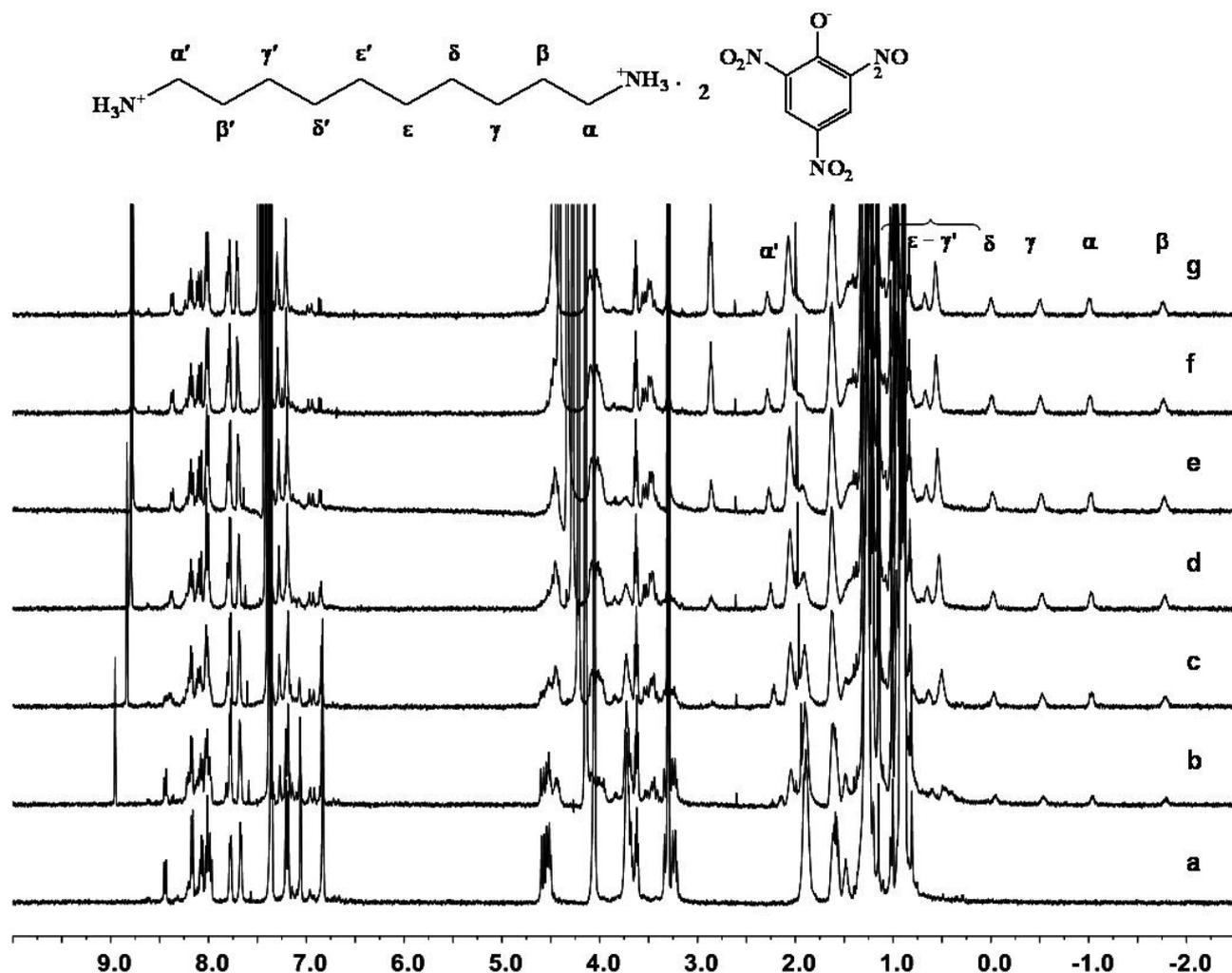


Figure S11. ^1H NMR titration of $\text{C}_{10}\cdot 2\text{NH}_2\cdot \text{HPic}$ with compound **3**. Constant concentration of host **3** (2.00×10^{-3} M) with addition of various concentrations ($0\text{--}4.00\times 10^{-3}$ M) of guest $\text{C}_{10}\cdot 2\text{NH}_2\cdot \text{HPic}$.

a) compound **3**; b) $[\text{H}]/[\text{G}]$ 4:1; c) $[\text{H}]/[\text{G}]$ 2:1; d) $[\text{H}]/[\text{G}]$ 1:0.75; e) $[\text{H}]/[\text{G}]$ 1:1; f) $[\text{H}]/[\text{G}]$ 1:1.5;

g) $[\text{H}]/[\text{G}]$ 1:2. Titration experiments of **2** with $\text{C}_{10}\cdot 2\text{NH}_2$, give rise to changes in the ^1H NMR spectra that are consistent with the formation of a single complexed species, corresponding to the 1:1 host–guest complex, at any $[\text{H}]/[\text{G}]$ ratios. This conclusion is validated by the appearing in the spectra of a triplet centered at $\delta=2.25$ ppm for $\text{C}_{10}\cdot 2\text{NH}_2$, since the addition of the first aliquot of the guest salt.

This triplet is assigned to the α' - CH_2 protons of the guest and judged of diagnostic value for the formation of the 1:1 host–guest complex.

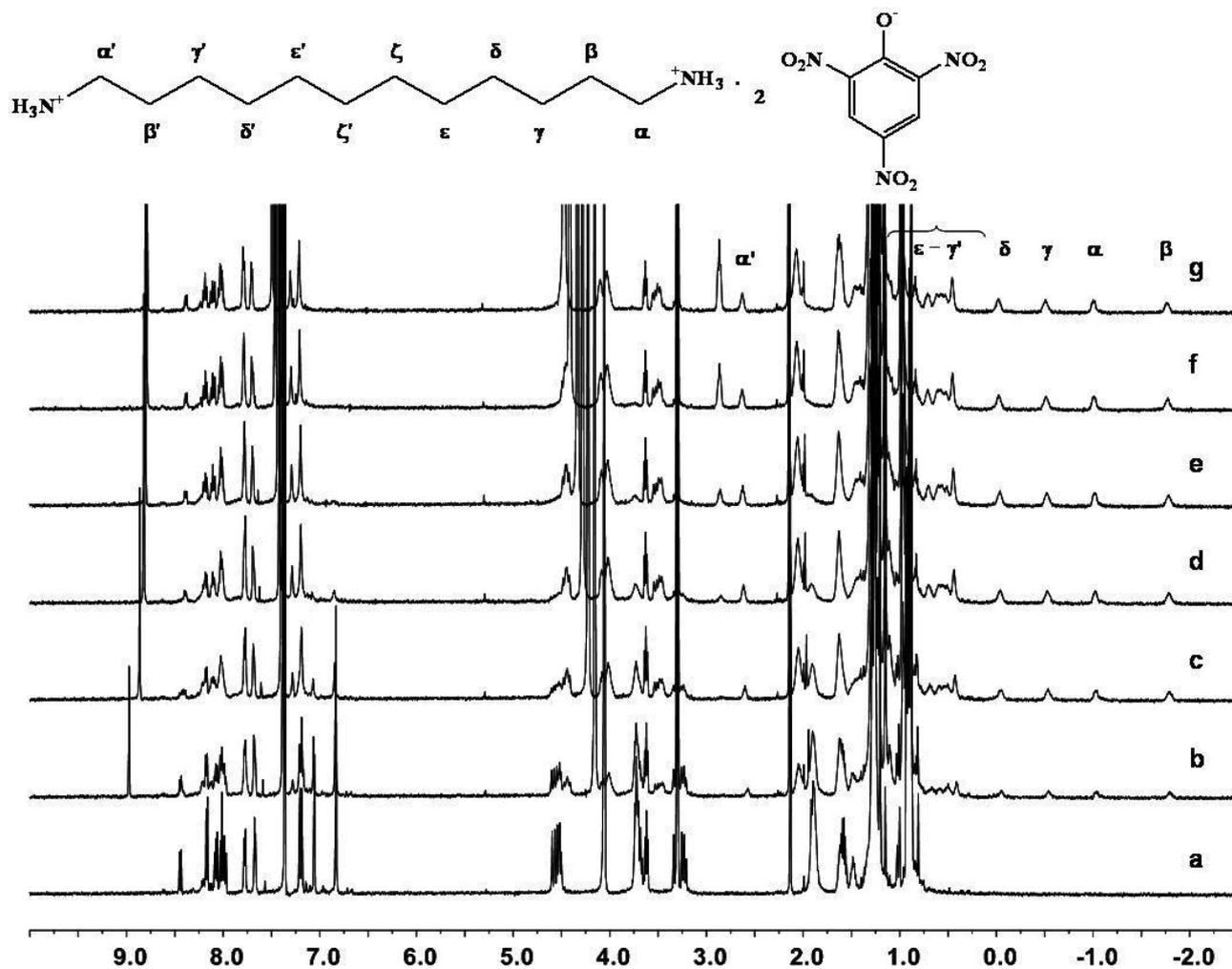


Figure S12. ^1H NMR titration of $\text{C}_{12}\cdot 2\text{NH}_2\cdot \text{HPic}$ with compound **3**. Constant concentration of host **3** (2.00×10^{-3} M) with addition of various concentrations ($0\text{--}4.00\times 10^{-3}$ M) of guest $\text{C}_{12}\cdot 2\text{NH}_2\cdot \text{HPic}$.

a) compound **3**; b) $[\text{H}]/[\text{G}]$ 4:1; c) $[\text{H}]/[\text{G}]$ 2:1; d) $[\text{H}]/[\text{G}]$ 1:0.75; e) $[\text{H}]/[\text{G}]$ 1:1; f) $[\text{H}]/[\text{G}]$ 1:1.5; g) $[\text{H}]/[\text{G}]$ 1:2. Titration experiments of **2** with $\text{C}_{12}\cdot 2\text{NH}_2$, give rise to changes in the ^1H NMR spectra that are consistent with the formation of a single complexed species, corresponding to the 1:1 host–guest complex, at any $[\text{H}]/[\text{G}]$ ratios. This conclusion is validated by the appearing in the spectra of a triplet centered at $\delta=2.63$ ppm for $\text{C}_{12}\cdot 2\text{NH}_2$, since the addition of the first aliquot of the guest salt. This triplet is assigned to the α' - CH_2 protons of the guest and judged of diagnostic value for the formation of the 1:1 host–guest complex.

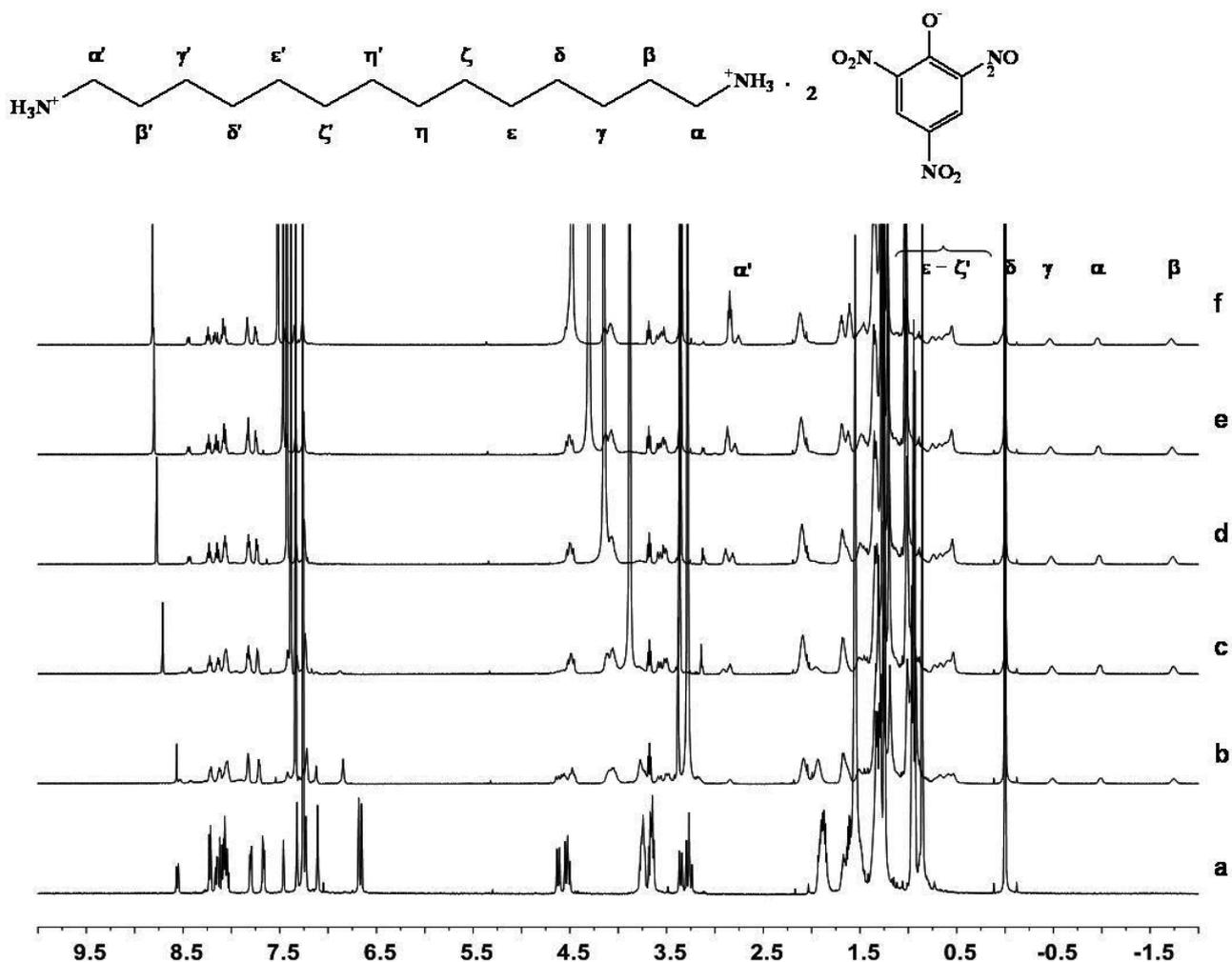


Figure S13. ^1H NMR titration of $\text{C}_{14}\cdot 2\text{NH}_2\cdot \text{HPic}$ with compound **3**. Constant concentration of host **3** (2.00×10^{-3} M) with addition of various concentrations ($0\text{--}4.00\times 10^{-3}$ M) of guest $\text{C}_{14}\cdot 2\text{NH}_2\cdot \text{HPic}$.
a) compound **3**; b) $[\text{H}]/[\text{G}]$ 4:1; c) $[\text{H}]/[\text{G}]$ 2:1; d) $[\text{H}]/[\text{G}]$ 1:0.75; e) $[\text{H}]/[\text{G}]$ 1:1; f) $[\text{H}]/[\text{G}]$ 1:2. Titration experiments of **2** with $\text{C}_{14}\cdot 2\text{NH}_2$, give rise to changes in the ^1H NMR spectra that are consistent with the formation of a single complexed species, corresponding to the 1:1 host–guest complex, at any $[\text{H}]/[\text{G}]$ ratios. This conclusion is validated by the appearing in the spectra of a triplet centered at $\delta=2.75$ ppm for $\text{C}_{14}\cdot 2\text{NH}_2$, since the addition of the first aliquot of the guest salt. This triplet is assigned to the α' - CH_2 protons of the guest and judged of diagnostic value for the formation of the 1:1 host–guest complex.

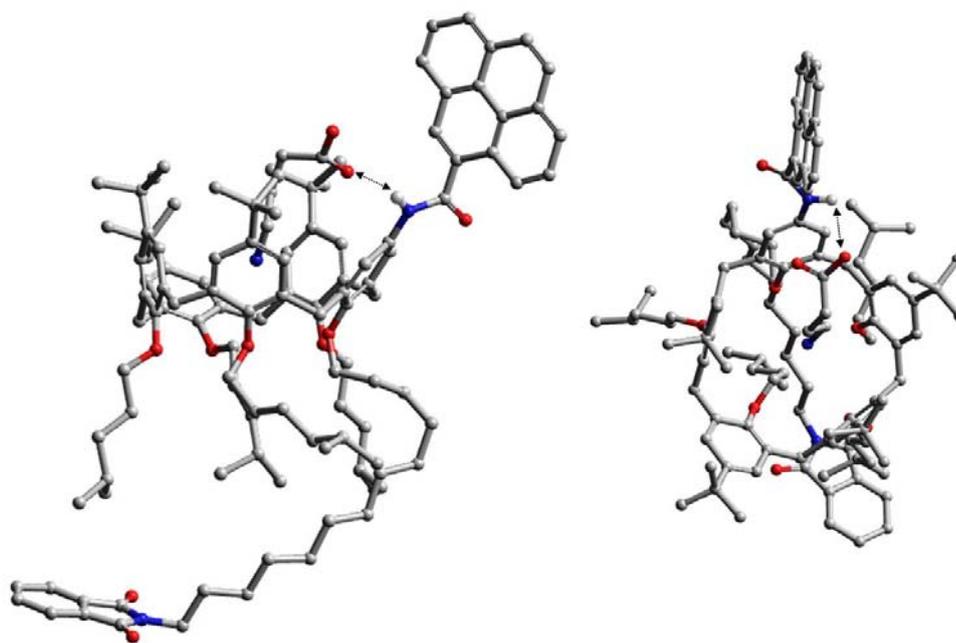


Figure S14. Optimized structures of the 1:1 complexes of **3** with **GABA**. Side view (left) and top view (right). (Hydrogens omitted for clarity).

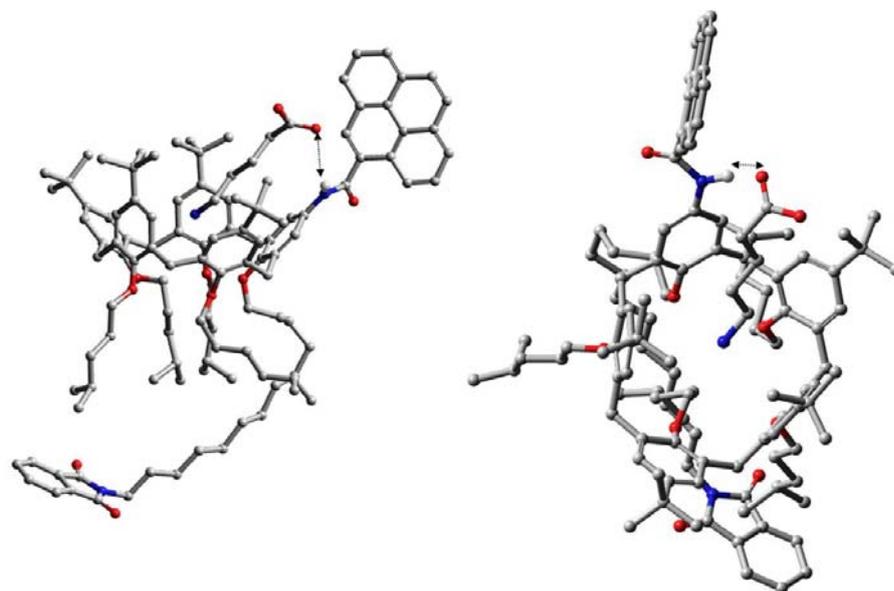


Figure S15. Optimized structures of the 1:1 complexes of **3** with **ε-Ahx**. Side view (left) and top view (right). (Hydrogens omitted for clarity).

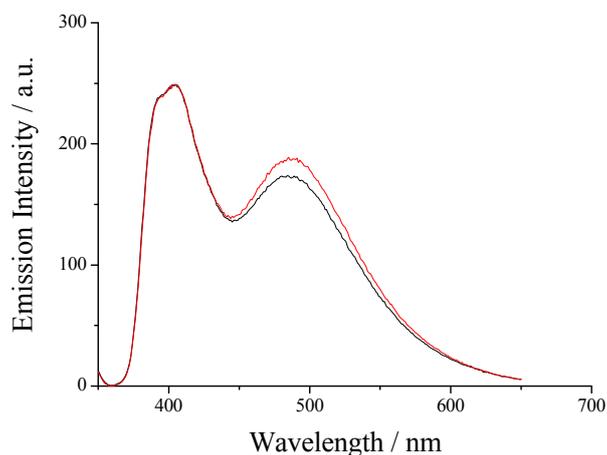


Figure S16. Fluorescence spectra of the **PyC5-NH₂** 1.0×10^{-5} M $\text{CHCl}_3/\text{CH}_3\text{OH}$ (50:50 v:v) solution (lower intensity) and upon addition of a stoichiometric amount of HCl (higher intensity).

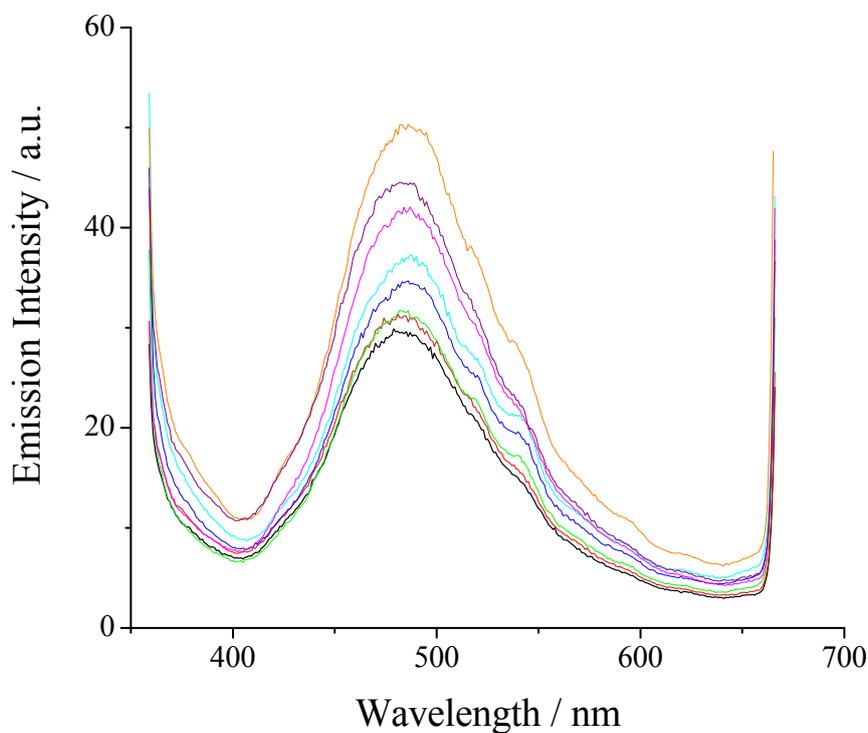


Figure S17. Fluorescence spectra of a representative **PyC5-NH₂-SAM** and upon 10 min immersion in $\text{CHCl}_3/\text{CH}_3\text{OH}$ (50:50 v:v) **C₁₂NH₂·HCl** solutions. Increasing emission intensities refer to the following increasing **C₁₂NH₂·HCl** concentrations: 0, 10, 20, 50, 70, 100, 150 and 200 ppm; $\lambda_{\text{exc}} = 343$ nm.