

Supporting information for:

Hierarchical Magnetic Core-Shell Nanoarchitectures: Non-linker Reagent Synthetic Route and Applications in a Biomolecule Separation System

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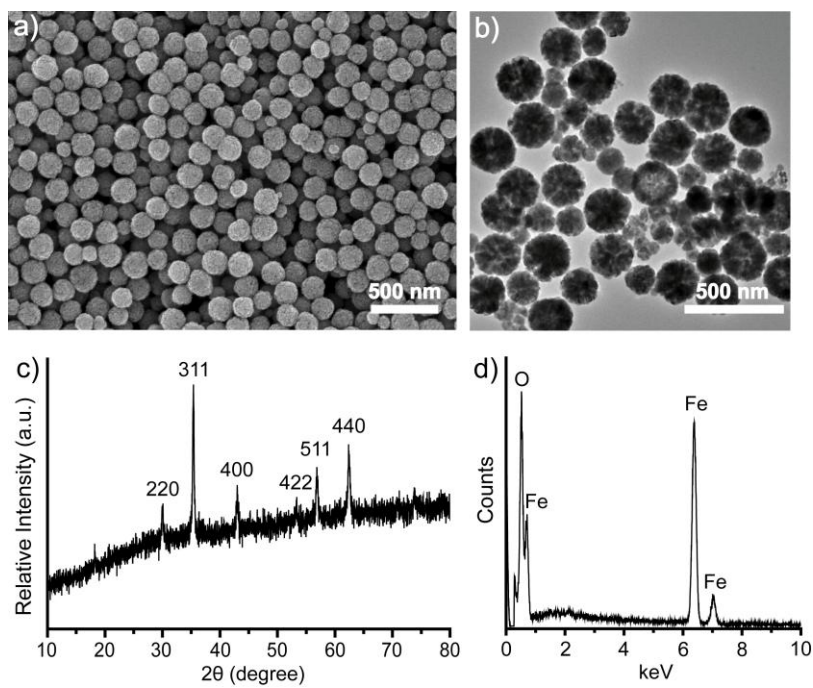


Figure S1. SEM image (a), TEM image (b), wide-angle XRD pattern (c) and EDS spectrum (d) of magnetic Fe_3O_4 nanoparticles.

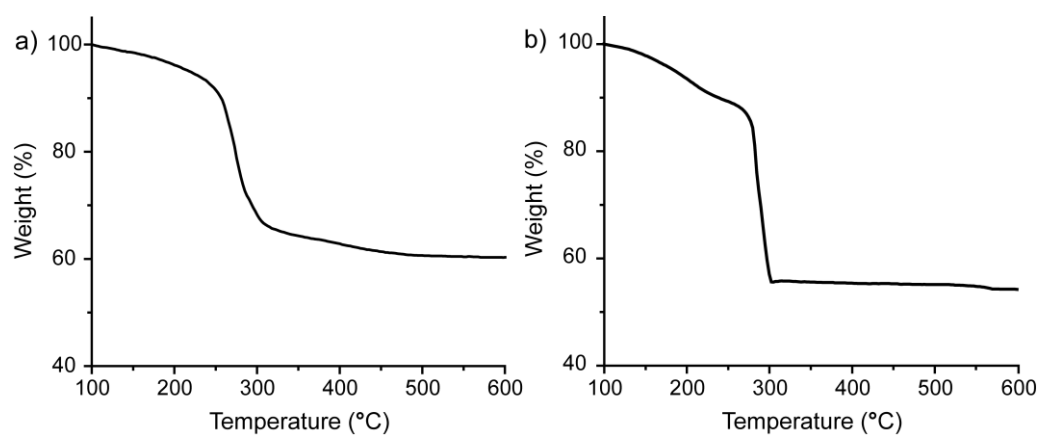


Figure S2. TGA curves of α -Ni(OH)₂ (a) and amorphous Co(OH)₂ (b) synthesized through the same solvothermal method. The results present above were applied to confirm the calcining temperature and duration.

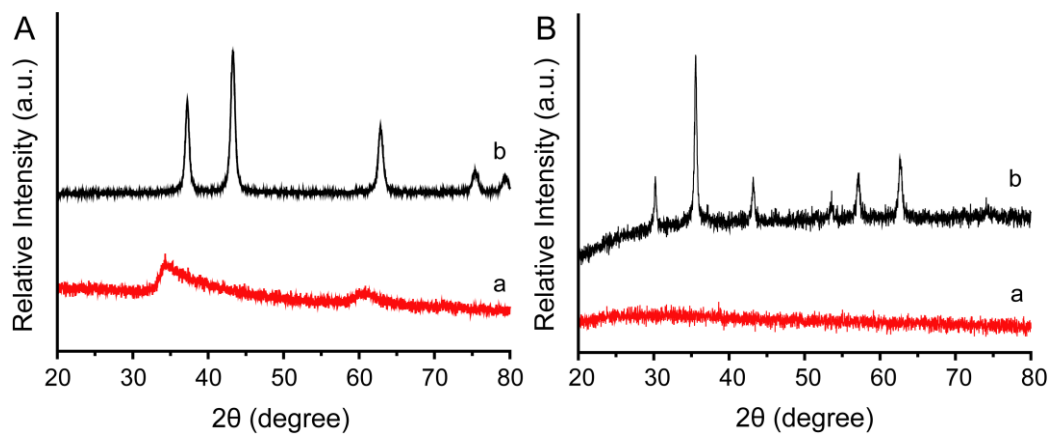


Figure S3. A: Wide-angle XRD patterns of α -Ni(OH)₂ composite (a) and NiO coating materials(b). B: Wide-angle XRD patterns of amorphous Co(OH)₂ composite (a) and Co₃O₄ coating materials (b). Control experiments were carried out to evidence the detailed components of the coating shells.

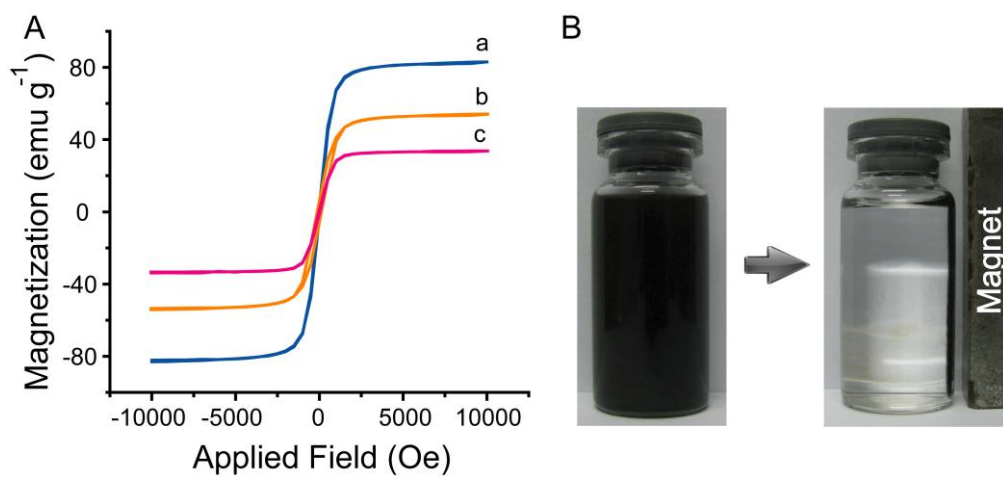


Figure S4. A: Room-temperature field-dependent magnetization curves of a) Fe₃O₄, b) Fe₃O₄@NiO, and c) Fe₃O₄@Co₃O₄. B: photograph shows the strong attraction of the as-prepared Fe₃O₄@NiO heteronanostructures suspended in water toward an assistant magnet.

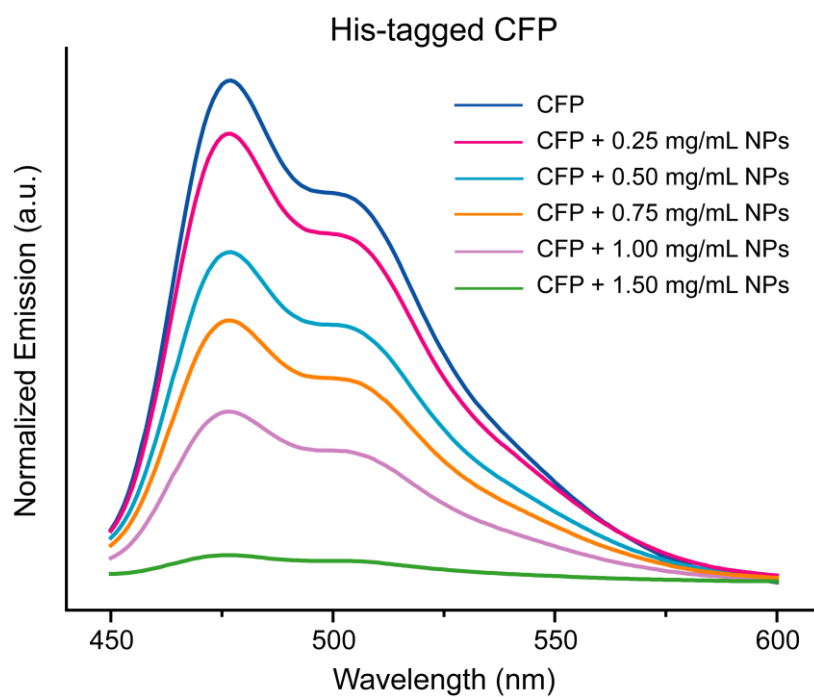


Figure S5. The fluorescence spectra corresponding to the change of emission intensity during the His-tagged CFP capturing experiments incubated with different concentrations of $\text{Fe}_3\text{O}_4@\text{NiO}$ microspheres.

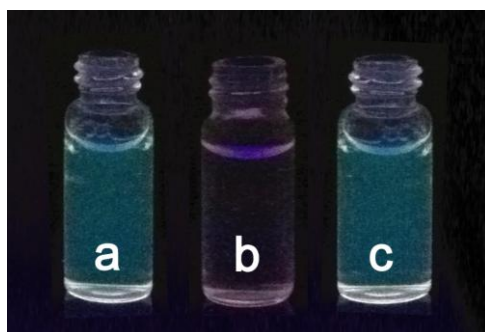


Figure S6. Fluorescent images from the solutions of His-tagged CFP (a) before and (b) after treatment with $\text{Fe}_3\text{O}_4@\text{NiO}$ microspheres and (c) after treating protein-bound $\text{Fe}_3\text{O}_4@\text{NiO}$ with imidazole solution (irradiation upon UV light).

| Cycle | Practical Fluorescent Percentage (%) | Normalized Fluorescent Percentage (%) |
|-------|--------------------------------------|---------------------------------------|
| 1 | 91.39 | 100 |
| 2 | 90.93 | 99.50 |
| 3 | 90.28 | 98.79 |
| 4 | 89.11 | 97.50 |
| 5 | 89.59 | 98.03 |

Table S1. Detailed magnetic purification and recycling efficiency of Fe₃O₄@NiO microspheres for selective separation of His-tagged CFP from cell lysate. Normalized fluorescent percentages are calculated from practical fluorescent percentage by recognizing the value of the first cycle as 100%.