

1                           **Supporting Information**

2

3                           **Asymmetric and Symmetric PCR of Gold Nanoparticles: A Pathway to**  
4                           **Scaled-Up Self-Assembly with Tunable Chirality**

5                           Yuan Zhao<sup>1</sup>, Liguang Xu<sup>1</sup>, Hua Kuang<sup>1</sup>, Libing Wang<sup>1,2\*</sup>, Chuanlai Xu<sup>1\*</sup>

6                           <sup>1</sup>*School of Food Science & Technology, State Key Lab of Food Science & Technology, Jiangnan University,*  
7                           *Wuxi 214122, China*

8                           <sup>2</sup>*Technological Centre of Hunan Entry-Exit Inspection and Quarantine Bureau, Changsha, 410001, China*

9

10

11                           **Captions:**

12                           **Table S1.** The nucleotide sequences of template and the primers.

13                           **Fig. S1** TEM images of gold nanoparticles with the diameter at  $25\pm3$  nm (A) and  $10\pm2$  nm  
14                           (B).

15                           **Fig. S2** The electrophoresis results of the GNPs-primer conjugation.

16                           **Fig. S3** The electrophoresis results of assembled structures by PCR at 40 cycles.

17                           **Fig. S4** CD spectra of the control groups including small and big GNPs, dsDNA and the  
18                           mixtures.

19                           **Fig. S5** UV-vis spectra of superstructures in asymmetric PCR at different cycles (150 bp).

20

21

22

23

---

\* Corresponding author Tel: +86-510-85329076; fax: +86-510-85329076; E-mail:[wanglb@hnciq.gov.cn](mailto:wanglb@hnciq.gov.cn), [xcl@jiangnan.edu.cn](mailto:xcl@jiangnan.edu.cn)

1

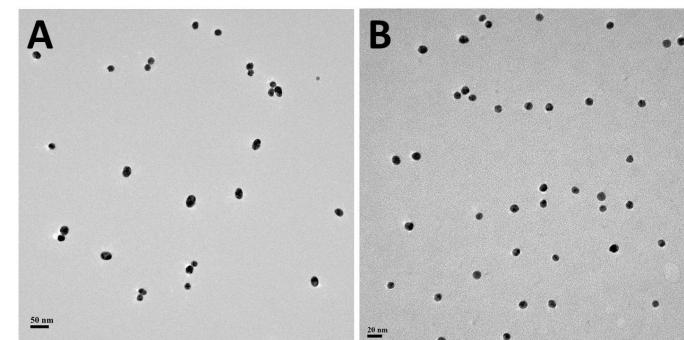
2 **Table S1.** The nucleotide sequences of template and the primers.  
3

Name Template	Nucleotide sequences (5' to 3') Plasmid λDNA
F-primer (100bp)	-SH-ATGAAACGGCAGGCAGAACAGG
R-primer (100bp)	-SH-ACAGGGACATGCCACCAAGAAA
F-primer (152bp)	-SH-GGAGGGCGTAACCGACAA
R-primer (152bp)	-SH-CCGCAGCAAACTCACCATT

4

5

6

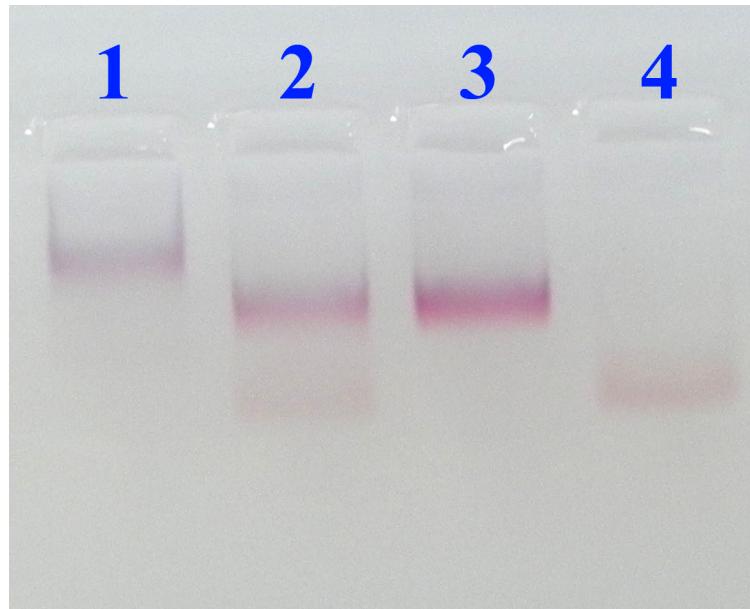


7

8 **Fig. S1** TEM images of gold nanoparticles with the diameter at  $25 \pm 3$  nm (A) and  $10 \pm 2$  nm

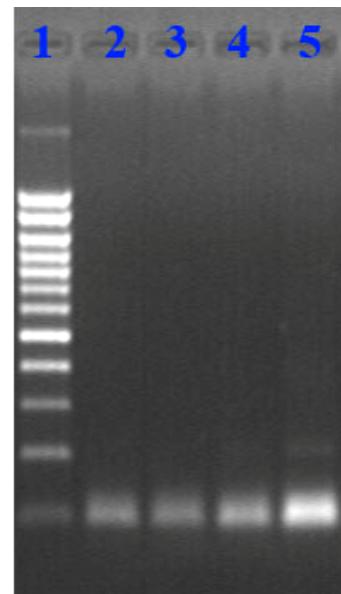
(B). The bar in A) was 50nm and in B) was 20nm.

9



**Fig. S2** The electrophoresis results of the GNPs-primer conjugation.

Lane 1: PCR products; lane 2: mixture of big GNPs-primers and small GNPs-primers; lane 3: big GNPs-primer conjugates; lane 4: small GNPs-primer conjugates.  
From Figure S2, the bright-field image of the gel shows the mobility of GNPs in different statuses. Lane 1 is the result of PCR product, which is composed of many big and small GNPs in the structures. Therefore, the mobility is the slowest. As to the short sequence of the forward and backward primers conjugated to the small and big GNPs, therefore, the mobility of the GNPs-primer conjugates is a little faster.



2           **Fig. S3** The electrophoresis results of assembled structures by PCR at 40 cycles.

3       Lane 1: standard marker; lane 2 and 3, the 1 fold PCR products; lane 4: 5-fold concentrated  
4       PCR products; lane 5: 10-fold concentrated PCR products. Firstly, from the dark field image  
5       of the gel, it is definitely confirmed that the PCR is carried out successfully.

6

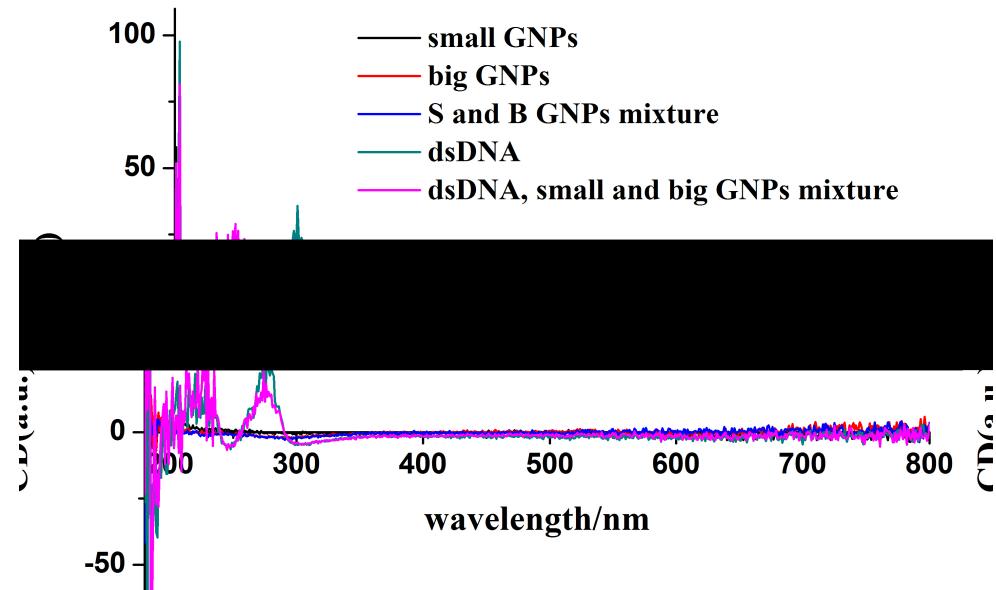
7

8

9

10

11



1

2

3 **Fig. S4** CD spectra of the control groups including small and big GNPs, dsDNA and the  
4 mixtures.

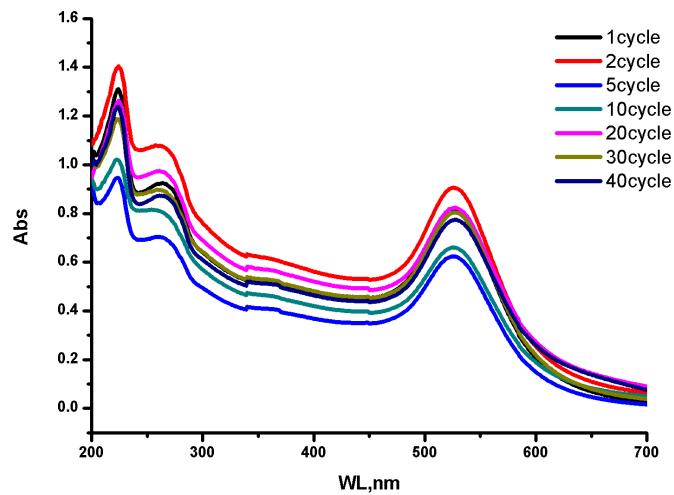
5

6

7

8

9



1

2 **Fig. S5** UV-vis spectra of superstructures in asymmetric PCR at different cycles (150 bp).

3