Electronic Supplementary Information for

Fe₃O₄@MOF core-shell magnetic microspheres with a designable metal-organic framework shell

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Experimental

Instruments

The powder X-ray diffraction (PXRD) patterns of the samples were collected using an X-ray diffractometer with Cu target (36 kV, 25 mA) from 5 to 65°. Analyses of the morphology and chemical composition of the samples were conducted by a Hitachi S-4800 field emission scanning electron microscope (FE-SEM) and a JEOL JEM 2100 transmission electron microscope at 200 kV, respectively. Nitrogen sorption-desorption isotherms were obtained at 77 K on a Micromeritics ASAP 2020M+C analyzer. Elemental analysis of C, H and N was carried out on a Vario ELIII, and the Cu and Fe contents were determined by atomic absorption spectrometry using a Thermo Elemental, USA M6 spectrometer. The magnetization curves were measured at room temperature under a varying magnetic field from -10000 to 10000 Oe on a BHV-55 vibrationg sample magnetometer (VSM).

Synthesis of core–shell magnetic Fe₃O₄@MOF microspheres

 Fe_3O_4 nanoparticles were synthesized by a solvothermal method as reported previously.^{2a} Mercaptoacetic acid (MAA)-functionalized Fe_3O_4 nanoparticles were prepared according to the following process. In a typical procedure, 0.05 g Fe_3O_4 was

added to 10 mL of ethanol solution of mercaptoacetic acid (0.29 mM) under shaking for 24 h. The product was recovered by an external magnetic field and washed several times with distilled water and ethanol, then re-dispersed in ethanol. 0.05 g MAA-functionalized Fe₃O₄ core synthesized as described above was dispersed in 4 mL of Cu(CH₃COO)₂·H₂O (for Fe₃O₄@[Cu₃(btc)₂] microspheres, or FeCl₃·6H₂O for Fe₃O₄@MIL-100(Fe) microspheres) ethanol solution (10 mM) for 15 minutes and then in 4 mL of benzenetricarboxylic acid (H₃btc) ethanol solution (10 mM) for 30 minutes at 25 °C or 70 °C. Between each step the microspheres were separated by magnetic decantation and washed with ethanol. After a given number of cycles the samples were washed with ethanol, and dried under vacuum at 150 °C.

Table S1.

Porosity properties, Fe₃O₄ contents, saturation magnetization values of Fe₃O₄@[Cu₃(btc)₂] **A1** (25 °C, 25 cycles), **A2** (25 °C, 50 cycles), **A3** (70 °C, 50 cycles) and Fe₃O₄@MIL-100(Fe) **B1** (25 °C, 50 cycles) core–shell magnetic microspheres.

	A1	A2	A3	B 1
Fe ₃ O ₄ Content [wt%]	46.1	30.0	16.1	33.17
$S_{\text{BET}}[\text{m}^2\text{g}^{-1}]$	230.79	550.93	668.15	523.32
$V [m^3 g^{-1}]$	0.14	0.32	0.49	0.31
$M_{\rm s}$ [emu g ⁻¹]	36.51	24.42	14.23	38.63



Fig. S1 PXRD patterns simulated from the crystallographic data of (a) HKUST-1 and Fe₃O₄@[Cu₃(btc)₂] core–shell microspheres (b) **A1** (25 °C, 25 cycles), (c) **A2** (25 °C, 50 cycles) and (d) **A3** (70 °C, 50 cycles).



Fig. S2 SEM images of the initial Fe₃O₄ nanospheres (a) at low magnification, and (b) at high magnification.



Fig. S3 SEM images of Fe₃O₄@[Cu₃(btc)₂] core–shell magnetic microspheres after (a) 10, (b) 20, (c) 25, (d) 30, (e) 40, and (f) 50 assembly cycles at 25 °C.



Fig. S4 SEM images of $Fe_3O_4@[Cu_3(btc)_2]$ core-shell magnetic microspheres prepared at 70 °C after (a) 20, (b) 30, (c) 40 and (d) 50 assembly cycles.



Fig. S5 TEM images of $Fe_3O_4@[Cu_3(btc)_2]$ core–shell microsphere after (a) 20, (b) 30, (c) 40, and (d) 50 assembly cycles at 70 °C, (e) HRTEM image of the MOF shell, and (f) the selected area electron diffraction pattern.



Fig. S6 PXRD patterns of simulated from the crystallographic data of MIL-100(Fe) and Fe₃O₄@MIL-100(Fe) core–shell magnetic microspheres after 50 assembly cycles at 25 °C.



Fig. S7 Nitrogen sorption-desorption isotherms of $Fe_3O_4@[Cu_3(btc)_2]$ microspheres (a) A1 (25 °C, 25 cycles), (b) A2 (25 °C, 50 cycles) and (c) A3 (70 °C, 50 cycles) measured at 77 K.



Fig. S8 TGA under air of (a) Fe₃O₄@[Cu₃(btc)₂] microspheres A3 (70 °C, 50 cycles) and (b) Fe₃O₄@MIL-100(Fe) B1 (25 °C, 50 cycles).