





Toh *et al.*, 2006, Supplementary Figure 2

Supplementary Figure Legends

Supplementary Figure 1. Geometrical designs of micropillars affect the clogging tendency during cell immobilization process. Hepatocytes at cell density of 1.5×10^6 cells ml^{-1} were dynamically seeded into a microfluidic channel at 0.5 ml hr^{-1} . (a) $50 \mu\text{m} \times 30 \mu\text{m}$ semi-circular design (b) $30 \mu\text{m} \times 50 \mu\text{m}$ elliptical design.

Supplementary Figure 2. The density of complex coacervated matrix can be modulated by varying laminar flow complex coacervation conditions. (a) – (d) By changing the relative flow rates of polyelectrolytes. (a) - (b) are flow profiles of a pair of polyelectrolytes (designated P1 and P2) in a microfluidic channel with immobilized cells when their relative flow rates ($P2/P1$) was < 5 and > 50 respectively. P1 and P2 were simulated with 0.2 % FITC solution and 1X PBS respectively. (c) - (d) show the complex coacervated matrix formed when P1 was represented by 1.5 mg ml^{-1} of Alexa Fluor-532 (AF-532) labeled methylated collagen and P2 by 3 % HEMA-MMA-MAA terpolymer solution at $P2/P1 < 5$ and $P2/P1 > 50$ respectively. At high $P2/P1$, the extent of complex coacervation was decreased due to limiting amount of P1, resulting in a loose matrix. (e) By changing the concentration of polyelectrolytes. The density of complex coacervated matrix was augmented by changing the concentration of AF-532-labeled methylated collagen (P1) from 1.5 mg ml^{-1} to 3.0 mg ml^{-1} while maintaining at the same $P2/P1$ as (d). Images in (c) – (e) were maximum projections of $30 \mu\text{m}$ optical sections. Hepatocytes nuclei were counterstained with SYTOX Green (Molecular Probes, USA).

Supplementary video. Cells are damaged during dynamic cell immobilization process. Hepatocytes in culture medium dosed with $50 \mu\text{g ml}^{-1}$ Propidium iodide (PI) were dynamically seeded into the 3D-

μ FCCS at a flow rate of 1 ml hr^{-1} . Cells that turned fluorescent were indicative of damaged cell membrane sustained during the seeding process allowing PI to enter the cells.