A micro surface tension alveolus (MISTA) in a glass microchip (Electronic Supplementary Information)

Xing Yue (Larry) Peng

E-mail: xypeng@xmu.edu.cn

5 Department of Biology, Xiamen University, Xiamen, Fujian, China 361005. Fax: 86-592-2181386; Tel: 86-592-2181386

Experimental

Microchip fabrication

- Photolithographic and wet chemical etching techniques were ¹⁰ used to fabricate the microstructure (Fig.S1) on a 1.6mm thick, 60 mm square glass substrate (type SG2506 glass substrate with chromium and S-1805 photoresist obtained from Shaoguang Microelectronics Corp., Changsha, China). The etching speed of dilute HF/NH4F was 2µm/min and the
- ¹⁵ precise etching depth was controlled by inspection under a light microscope (Fig.S2). The glass bonding temperature was 560°C. We developed a reusable bonding method: detergent and water washed the cover glass and the etched substrate glass, and a steel clamp was used to keep them tightly ²⁰ together during the experiment (Fig. S3). After the
- experiment, the cover glass was removed and washed ready for the next experiment.



Fig.S1 The photomask of the microchip for gas exchange experiments.







Fig.S3 Pressure encapsulation for a two-layer glass chip.

30 Gas pressure control

Cylinders of N₂, O₂, and CO₂ & Air were used in the experiments. The pressure was adjusted precisely at $0 \sim 10 \pm 0.1$ kPa (Anthone Elec. Ltd., Xiamen, China). The gases were connected to a gas switch (BD Connecta, ref 394601, Becton ³⁵ Dickinson, Sweden) and a 1mm diameter plastic tube connected the switch to the gas inlet of the microchip.

Extraction and de-oxygen of chicken Hb

Newly collected chicken blood was centrifuged (900g) for the collection of red cells. An equal volume of water was added ⁴⁰ for effective ultrasonic cell crushing. To preserve the chicken Hb, the temperature was kept below 40 °C by stopping the ultrasonic treatment and keeping the preparation on ice. The mixture of chicken Hb and cell debris was centrifuged (6000g) to clarify the solution for experiments. Water was de-⁴⁵ oxygenated by a N₂ stream for 2 days at room temperature N

 $(N_2$ -water). We added 200µl Hb (blood volume) to 40ml N_2 water and continued the N_2 stream for another day. The purpose of the N_2 stream is to remove O_2 or other gases from the water²⁷ and Hb. The de-oxygenated Hb solution was now

so ready for spectrum scan. We used the high concentration deoxygenated Hb-solution (50% blood red cell) as an on-chip O_2 and CO_2 probe.

Image processing

The curvature radius of the MISTA front was measured as 55 follows (Fig.S4): we drew the outline of the MISTA front; measured the chord length (L) of the arc; measured the height of the arc from the chord (H). The radius (R) is given as: $R = (L^2 + 4H^2) / 8H$ (2)

Microscopic images were recorded automatically at intervals $_{60}$ of 0.2 \sim 60 s by a colour CCD (DH-HV3103UC, China Daheng

25

Group, Inc. China) (Fig.S5)). To enhance the image, background was subtracted (Fig.S8d) from the original image (the background was the image before the experiment), phase reversed and adjusted to high contrast. Software was ⁵ developed to extract the diffusion profile from time-lapse images. This software picked RGB colour on each image at different distances from the MISTA following the diffusion direction and combined all the RGB data into one image, with the x-axis as time and the y-axis as distance (Fig.3b & ¹⁰ Fig.S10).



Fig.S4 The measurement and calculation of curvature radius.



Fig.S5 The RGB quantum efficiencies of the CCD used in the experiments.

Results and discussion

The influence of pressure on the curvature radius of MISTA

Fig.S6 is a graph data of overlapping outlines extracted from experiments. It shows how the pressure difference bends the ²⁰ surface of MISTA. When a series of MISTA are embedded along a wall, the pressure difference bends all the MISTA, simultaneously (Fig.S7a-c). The to and fro effect because of the friction is depicted in Fig.S7d.



15



Fig.S6 Overlapping outlines the curvature of a MISTA changed to gas pressure in the process of increasing pressure (blue) and decreasing pressure (red). The curvature calculations were based on these outlines.



Fig.S7 The changing shape of a MISTA and its curvature measurements to differential pressures. (**a**–**c**) Increasing pressure from gas pushed two MISTAs into the liquid. (**d**) The background image showed a MISTA. The two blue arrows indicated the increasing pressure and the increasing curvature. The two red arrows indicated the decreasing pressure and the

40 curvature. The two red arrows indicated the decreasing pressure and the decreasing curvature.



Fig.S8 The colour of hemoglobin (Hb) as an indicator to dissolved O_2 and CO_2 . (a) The Hb (5% blood) showed different colours in Air-water, N₂-water, O₂-water and CO₂-water. (b) The diffusions of O_2 and CO_2 via gas-

liquid interface (a) or PDMS membrane (b) were measured by a spectrophotometer (c). The spectra of N₂-Hb (5% blood, O₂ or CO₂ was removed from Hb by N₂), O₂-Hb, CO₂-Hb were different. The spectrum of O₂-Hb under PDMS membrane during 2h O₂ diffusion (O₂-HbP in b) was similar to that of O₂-Hb under no membrane gas-liquid interface

during five-minute O₂ diffusion (the grey lines between red and black lines representing measurements every three minutes). (d) These spectrum differences in microchannel on chip were inspected by colour

CCD camera under microscope and the CCD images were enhanced by a RGB (red, green and blue value) colour process. BK: background colour.



Fig.S9 Absorbency ratio (A542nm+A577nm)/A562nm) of Hb was increasing because of oxygen diffusion from the air. Circles represented oxygen entering the Hb solution (0.5% blood red cell) directly and crosses represented oxygen in the air entered the Hb solution via a PDMS
 membrane (600 µm thick).



Fig.S10 A series of rapid switching between N₂ and O₂ showed by a RGB colour profile near a MISTA. The red double head arrow on the right depicted the position of the profile.

25 The Hb as a probe of both dissolved O2 and CO2

We have developed the Hb as a probe of both O_2 and CO_2 (Fig.S8). The experimental detail about the colours and the spectrums of O_2 -Hb and CO_2 -Hb can be found in Fig.S8a-c. Fig.S8d shows the processes of the colour images of the ³⁰ experiments.

The diffusion of O2 and CO2 through a PDMS membrane

Results about the diffusion of O2 and CO2 through a PDMS membrane can be found in Fig.S8c and Fig.S9.

Rapid switching of gases

³⁵ Fig.S10 shows the image records inside a MISTA with rapid N₂-O₂ gas shifting.

The simulation of convective/diffusive mass transfer from a MISTA

To evaluate the convective or diffusive mass transfer, we $_{40}$ simulated the O₂ and CO₂ diffusion in the Hb solution from a MISTA into the microchannel. The design of the MISTA diffusion channel in the experiments was narrow and the diffusion in this channel acts like one dimensional diffusion. The liquid in the microchannel is stationary and the model

⁴⁵ includes only diffusive mass transfer. The gas phase is mobile and the convective mass transfer of gas enables a constant saturated boundary at the MISTA gas-liquid surface. So the simulation set a saturated O₂ or CO₂ in the process of calculation. The calculations are based on Fick's first law.^{28, 29}

$$^{50} J = -D \frac{\partial \phi}{\partial x} \tag{2}$$

where J is the diffusion flux, D is the diffusion coefficient, Φ is the concentration and x is the position.

Fick's first law predicts how diffusion causes the concentration field to change with time:

$$^{55} \frac{\partial \phi}{\partial t} = D \frac{\partial^2 \phi}{\partial x^2}$$
(3)

where t is time.

Eqn (4) is a solution of eqn (3) in one dimension:

$$n(x,t) = n(0) \operatorname{erfc}\left(\frac{x}{2\sqrt{Dt}}\right) \tag{4}$$

where *erfc* is the complementary error function, which is ⁶⁰ defined in term of the error function:

$$erfc(x) = 1 - erf(x) = \frac{2}{\sqrt{\pi}} \int_{x}^{\infty} e^{-t^{2}} dt$$
 (5)



Fig.S11 The comparisons of the Fick's first law simulation model in this paper by a solution of Fick's second law.

One approximation of the erf(x) is given by

$${}^{5} erf^{2}(x) \approx 1 - \exp(-x^{2} \frac{4/\pi + ax^{2}}{1 + ax^{2}})$$
(6)

where

$$a = -\frac{8(\pi - 3)}{3\pi(\pi - 4)} \tag{7}$$

- Eqn (4), (6) and (7) are the approximate solution to predicts ¹⁰ how diffusion causes the concentration field to change with time. They are used to validate the simulation. Only when the model calculations are confrimed, can Hb be incorporated into the model.
- Fig.S11 compares the simulation results by Fick's first law 15 and the solutions by Fick's second law. These coincident lines proved the correct of the simulation program and the Hb was hereafter incorporated into the model. Parameters are shown in Table 1.

Table 1 parameters in the simulation

Length from MISTA	0~1000	(µm)
Precision	1	(µm)
Time step	10-4	(s)
Diffusion coefficient of O2 in water	1.97×10^{3}	$(\mu m^2/s)$
Diffusion coefficient of CO ₂ in water	1.77×10^{3}	$(\mu m^2/s)$
Saturated dissolved O2 in water	1.34 x10 ⁻¹⁸	(mol/µm ³)
Saturated dissolved CO2 in water	3.92×10^{-17}	$(mol/\mu m^3)$
Saturated dissolved O2-Hb	6.06x10 ⁻¹⁸	(mol/µm ³)
Saturated dissolved CO2-Hb	2.24 x10 ⁻¹⁸	$(mol/\mu m^3)$

- ²⁰ In the process of simulation, the flux of O_2 and CO_2 are firstly figured out by Fick's first law for the total mass of O_2 and CO_2 at each position. With the equilibrium curve of CO_2 -Hb (Fig.S12), we have the concentrations of dissolved CO_2 and CO_2 -Hb. With the equilibrium curve of O_2 -Hb (Fig.S13) and
- $_{25}$ the concentration of CO₂, we have the concentrations of dissolved O₂ and O₂-Hb. We now go to the next step for the new flux of O₂ and CO₂ with the new concentrations O₂ and CO₂.

The equilibrium curve of CO_2 -Hb is defined by eqn (8):

$$y(x) = -1.3151 x^4 + 4.1554 x^3 - 5.0529 x^2 + 3.1997 x$$
(8)



Fig.S12 The equilibrium curve of CO₂-Hb in the computer simulation of diffusion from MISTA. Data was extracted from a image on Dr. Charles L. Webber's website (http://www.meddean.luc.edu/lumen/medEd/MEDICINE/PULMONAR/physio/pf4.htm).





The equilibrium curve of O_2 -Hb is defined²⁵ by eqn (9):

$${}^{40} y(x) = \frac{x(1+x)^{n-1} + x^n((1+2ax)(1+ax)^{n-1}-1)}{2((1+x)^n + x^n(1+ax)^n - 1))}$$
(9)

where n=2 and *a*, the parameter of Bohr Effect²⁴, fitting eqn (9), is defined by eqn (10):

$$a = \frac{76 \times 3.5}{P_{co} + 5} \tag{10}$$

where P_{CO_2} is partial pressure of CO₂ defined by eqn (11)

⁴⁵
$$P_{CO_2} = \frac{76C_{CO_2}}{\hat{C}_{CO_2}}$$
 (11)

where C_{CO_1} is the concentration of dissolved CO₂ and \hat{C}_{CO_2} is

the saturated dissolved CO₂.

Fig.S14 and Fig.S15 (next pages) show the main result of the simulation (see also ESI movies about the simulation).



Fig.S14 A Computer simulation of convective/diffusive mass transfer from a MISTA. A-F Concentrations of dissolved oxygen and O_2 -Hb at 100s. D,D/10 and D/50 represent diffusion coefficient of O_2 in water (D), one tenth of D and fiftieth of D, respectively. 1x blood, 0.5x blood and water represent151516171819191010111213141515151617181919191010111213141515161718191919101010111213141515161717181919191010101112131415151616171718181919191919191919191919101010101112<t



Fig.S15 A Computer simulation of convective/diffusive mass transfer from a MISTA. **A-F** Concentrations of dissolved oxygen, Hb-O₂, dissolved CO₂ and CO₂-Hb at 300s, just 30s after CO₂ fillings the MISTA (at 270s). D, D/10 and D/50 represent diffusion coefficient of O₂ in water (D), one tenth of D and fiftieth of D, respectively. 1x blood, 0.5x blood and water represent the concentration of Hb of one time, 0.5 time of blood concentration and no Hb, respectively.

References

- 27 X. Y. Peng and Z. Z. Hong, Marine Sciences, 2004, 28(8), 23-27. (in chinese)
- 28 A. Fick, Poggendorff's Annel. Physik. 1855, 94, 59.
 5 29 A. Fick, Phil. Mag. 1855, 10, 30.

10