
Electronic Supplementary Information for:

Co-Axial Capillaries Microfluidic Device for Synthesizing Size- and Morphology-Controlled Polymer Core-Polymer Shell Particles

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Operating conditions:

Fig. 2: Top, $Q_I = 0.0012 \text{ mL}\cdot\text{min}^{-1}$; $Q_M = 0.005 \text{ mL}\cdot\text{min}^{-1}$; $Q_O = 0.06 \text{ mL}\cdot\text{min}^{-1}$. Bottom, $Q_I = 0.0035 \text{ mL}\cdot\text{min}^{-1}$; $Q_M = 0.004$
10 $\text{mL}\cdot\text{min}^{-1}$; $Q_O = 0.05 \text{ mL}\cdot\text{min}^{-1}$. Prepared in system I.

Fig. 3: (a) $Q_O = 0.07 \text{ mL}\cdot\text{min}^{-1}$ and $Q_M = 0.005 \text{ mL}\cdot\text{min}^{-1}$ in microfluidic system I; $Q_O = 0.06 \text{ mL}\cdot\text{min}^{-1}$ and $Q_M = 0.005 \text{ mL}\cdot\text{min}^{-1}$
in microfluidic system II; $\mu_O = 1500 \text{ cP}$; (b) $Q_O = 0.07 \text{ mL}\cdot\text{min}^{-1}$ and $Q_I = 0.0012 \text{ mL}\cdot\text{min}^{-1}$ in microfluidic system I; $Q_O = 0.06$
 $\text{mL}\cdot\text{min}^{-1}$ and $Q_I = 0.0032 \text{ mL}\cdot\text{min}^{-1}$ in microfluidic system II; $\mu_O = 1500 \text{ cP}$; (c) $Q_M = 0.005 \text{ mL}\cdot\text{min}^{-1}$ and $Q_I = 0.0012 \text{ mL}\cdot\text{min}^{-1}$
15 in microfluidic system I; $Q_M = 0.005 \text{ mL}\cdot\text{min}^{-1}$ and $Q_I = 0.0032 \text{ mL}\cdot\text{min}^{-1}$ in microfluidic system II; $\mu_O = 1500 \text{ cP}$; (d) $Q_O = 0.07$
 $\text{mL}\cdot\text{min}^{-1}$, $Q_M = 0.005 \text{ mL}\cdot\text{min}^{-1}$ and $Q_I = 0.0012 \text{ mL}\cdot\text{min}^{-1}$ in microfluidic system I; $Q_O = 0.06 \text{ mL}\cdot\text{min}^{-1}$, $Q_M = 0.005 \text{ mL}\cdot\text{min}^{-1}$
and $Q_I = 0.0032 \text{ mL}\cdot\text{min}^{-1}$ in microfluidic system II; $\mu_O = 1500 \text{ cP}$.

Fig. 5: $\mu_O = 1500 \text{ cP}$, $Q_O = 0.07 \text{ mL}\cdot\text{min}^{-1}$; $Q_M = 0.006 \text{ mL}\cdot\text{min}^{-1}$; $Q_I = 0.004 \text{ mL}\cdot\text{min}^{-1}$ (a,a'); $Q_I = 0.0006 \text{ mL}\cdot\text{min}^{-1}$ (b,b'); $Q_I =$
20 $0.0008 \text{ mL}\cdot\text{min}^{-1}$ (c,c') in microfluidic system I, $\Delta = -650 \text{ }\mu\text{m}$.

Table ES1.1 Sizes of the capillaries and experimental materials

		System I			System II			System III			System IV			System V			System VI		
Capillary size		Inner diameter	Outer diameter	Material	Inner diameter	Outer diameter	Material	Inner diameter	Outer diameter	Material	Inner diameter	Outer diameter	Material	Inner diameter	Outer diameter	Material	Inner diameter	Outer diameter	Material
	Inner capillary	20 µm	90 µm	Glass	75 µm	360 µm	PEEK	100 µm	165 µm	Glass	150 µm	360 µm	PEEK	100 µm	360 µm	PEEK	100 µm	165 µm	Glass
	Middle capillary	150 µm	360 µm	PEEK	700 µm	850 µm	Glass	300 µm	760 µm	PTFE	700 µm	850 µm	Glass	530 µm	670 µm	Glass	250 µm	360 µm	Glass
	Outlet tubing	1600 µm	3200 µm	PTFE	1600 µm	3200 µm	PTFE	1600 µm	3200 µm	PTFE	1600 µm	3200 µm	PTFE	1600 µm	3200 µm	PTFE	1600 µm	3200 µm	PTFE
Inner fluid phase		acrylamide, 10 wt %; NN ² -methylene-bisacrylamide, 1.5 wt %; Ammonium persulfate, 0.15 wt %; viscosity: 1.45 cP			acrylamide, 10 wt %; NN ² -methylene-bisacrylamide, 1.5 wt %; Ammonium persulfate, 0.15 wt %; viscosity: 1.45 cP			acrylamide, 10 wt %; NN ² -methylene-bisacrylamide, 1.5 wt %; Ammonium persulfate, 0.15 wt %; viscosity: 1.45 cP			acrylamide, 10 wt %; NN ² -methylene-bisacrylamide, 1.5 wt %; Ammonium persulfate, 0.15 wt %; viscosity: 1.45 cP			acrylamide, 10 wt %; NN ² -methylene-bisacrylamide, 1.5 wt %; Ammonium persulfate, 0.15 wt %; viscosity: 1.45 cP			distilled water		
Middle fluid phase		tripropylenglykol-diacrylate, 93 wt %; Span80, 3.5% 1-hydroxycyclohexyl phenyl ketone; viscosity: 14.2 cP			tripropylenglykol-diacrylate, 93 wt %; Span80, 3.5% 1-hydroxycyclohexyl phenyl ketone; viscosity: 14.2 cP			tripropylenglykol-diacrylate, 93 wt %; Span80, 3.5% 1-hydroxycyclohexyl phenyl ketone; viscosity: 14.2 cP			tripropylenglykol-diacrylate, 93 wt %; Span80, 3.5% 1-hydroxycyclohexyl phenyl ketone; viscosity: 14.2 cP			tripropylenglykol-diacrylate, 93 wt %; Span80, 3.5% 1-hydroxycyclohexyl phenyl ketone; viscosity: 14.2 cP			tripropylenglykol-diacrylate, 93 wt %; Span80, 3.5% 1-hydroxycyclohexyl phenyl ketone; viscosity: 14.2 cP		
Outer fluid phase		aqueous solution of methyl cellulose			aqueous solution of methyl cellulose			aqueous solution of methyl cellulose			aqueous solution of methyl cellulose			aqueous solution of methyl cellulose			aqueous solution of methyl cellulose		
		No.	viscosity (cP)		No.	viscosity (cP)		No.	viscosity (cP)		No.	viscosity (cP)		No.	viscosity (cP)		No.	viscosity (cP)	
		1	500		1	500		1	1300		1	1300		1	1300		1	1300	
		2	800		2	800		-	-		-	-		-	-		-	-	
		3	1100		3	1100		-	-		-	-		-	-		-	-	
		4	1400		4	1400		-	-		-	-		-	-		-	-	
		5	1600		5	1600		-	-		-	-		-	-		-	-	
		6	2000		6	2000		-	-		-	-		-	-		-	-	

Table ESI 2. Maximum standard errors (SE_{\max} , given in μm) of experimental data plotted in Figure 3

Graph	Data symbol					
	●	○	★	▲	△	■
3a	12.65	9.09	12.87	8.33	7.14	8.69
3b	11.77	10.52	11.08	5.45	9.36	9.33
3c	13.21	9.74	10.39	10.02	7.77	9.91
3d	10.63	13.92	11.6	7.38	6.64	6.37

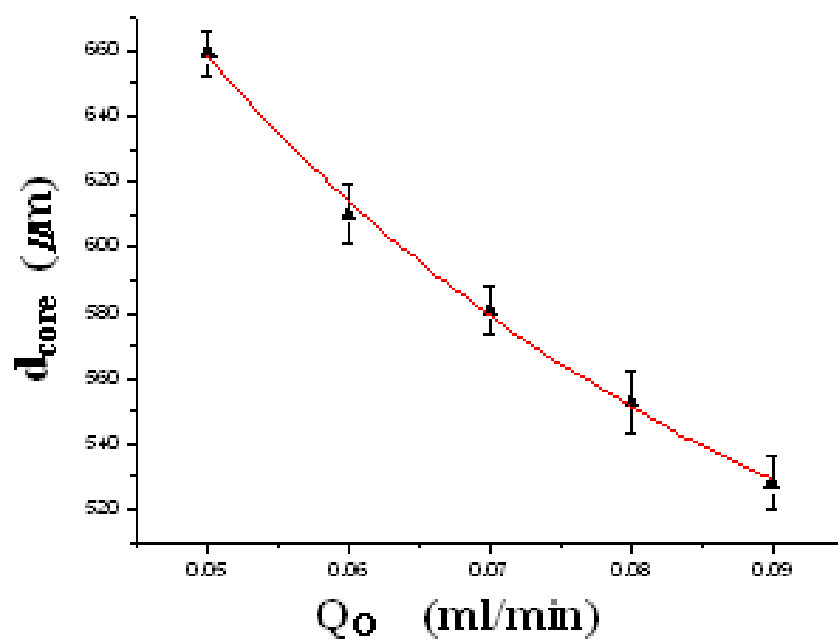


Fig. ESI.1 Variation of the core droplet diameter with respect to the outer fluid flow rate at constant Q_o/Q_M .

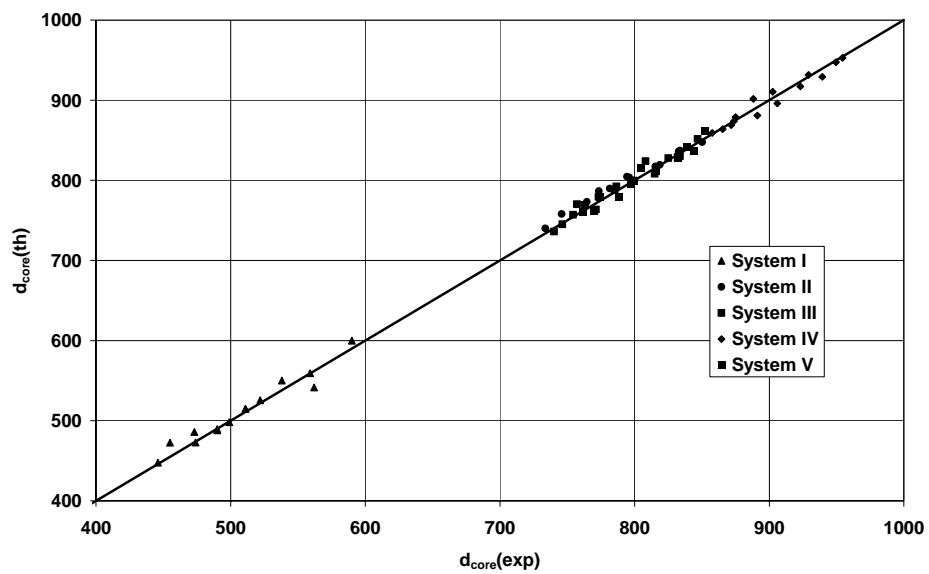


Fig. ESI.2 Comparison between theoretical (th) and experimental (exp) core droplet diameter for the 5 systems tested. Solid line represents the first bisector.

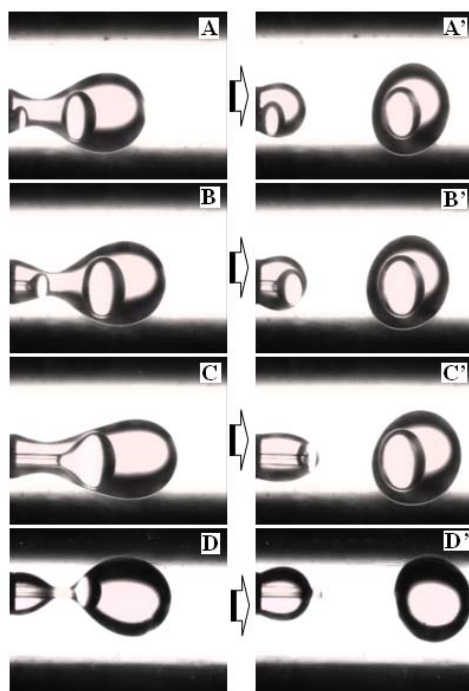


Fig. ESI.3 Snapshots of the formation of core-shell droplets for different positive capillaries' tips relative position, $\Delta=0$ (A,A'); $\Delta=146 \mu\text{m}$ (B,B'); $\Delta=360 \mu\text{m}$ (C,C'); $\Delta=480 \mu\text{m}$ (D,D'). Experimental conditions: $Q_O = 0.07 \text{ mL}\cdot\text{min}^{-1}$; $Q_M = 0.006 \text{ mL}\cdot\text{min}^{-1}$; $Q_I = 0.0012 \text{ mL}\cdot\text{min}^{-1}$ in microfluidic system I, $\mu_O = 1500 \text{ cP}$.

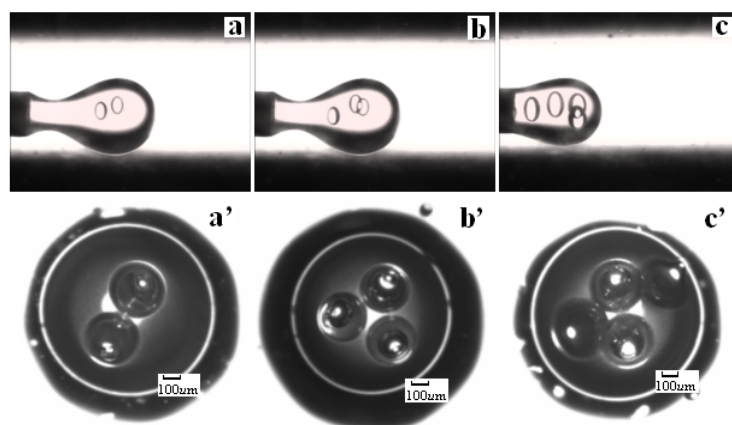


Fig. ESI.4 Optical microscopy images of the multiple core droplet formation in a co-axial capillaries microfluidic device with a negative capillaries' tips relative position (top) and the multiple poly(acrylamide) core-poly(tripropylenglycol-diacrylate) shell particles obtained (bottom), core being labeled with an aqueous dye (Nigrosin, Aldrich). Experimental conditions: $\mu_0 = 1500$ cP, $Q_0 = 0.07$ mL.min⁻¹; $Q_M = 0.006$ mL.min⁻¹; $Q_I = 0.004$ mL.min⁻¹(a,a'); $Q_I = 0.0006$ mL.min⁻¹(b,b'); $Q_I = 0.0008$ mL.min⁻¹(c,c') in microfluidic system I, $\Delta = -650$ μ m.

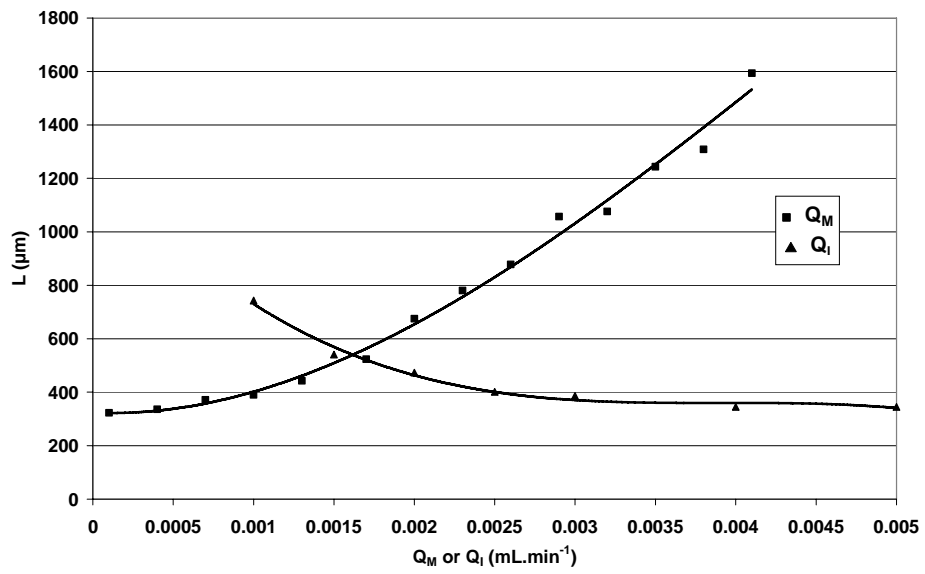


Fig. ES1.5 Effects of the inner and middle fluid flow rates on the length of rod-like polymer particles obtained with microfluidic system VI ($Q_i = 0.0023$ $\text{mL}\cdot\text{min}^{-1}$ when Q_M is varying from 0.0001 $\text{mL}\cdot\text{min}^{-1}$ to 0.006 $\text{mL}\cdot\text{min}^{-1}$; $Q_M = 0.0045$ $\text{mL}\cdot\text{min}^{-1}$ when Q_i is varying from 0.0025 $\text{mL}\cdot\text{min}^{-1}$ to 0.014 $\text{mL}\cdot\text{min}^{-1}$; $Q_o = 0.05$ $\text{mL}\cdot\text{min}^{-1}$).

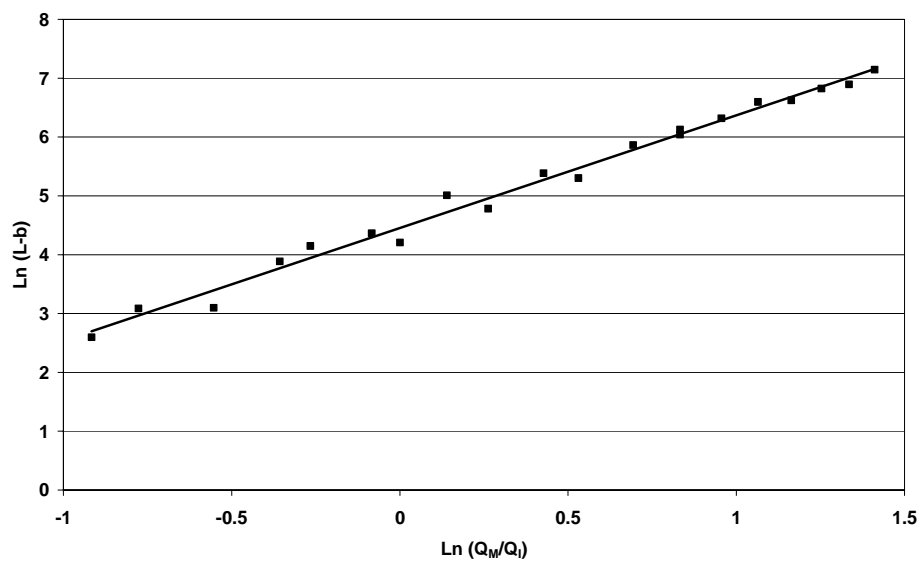


Fig. ESI.6 Variations of the length of rod-like polymer particles with respect to the ratio between the middle and inner fluid flow rates. Solid line is derived from equation 4.