

SUPPLEMENTARY INFORMATION

Quantitative Mapping of Aqueous Microfluidic Temperature with Sub-degree Resolution Using Fluorescence Lifetime Imaging Microscopy

Emmelyn M. Graham^{1,2,#}, Kaoru Iwai,³ Seiichi Uchiyama,⁴ A. Prasanna de Silva⁵, Steven W. Magennis^{*,2,##} and Anita C. Jones,^{*,1,2}

¹School of Chemistry, The University of Edinburgh, Edinburgh EH9 3JJ, U.K.;

²Collaborative Optical Spectroscopy, Micromanipulation and Imaging Centre (COSMIC), The University of Edinburgh, Edinburgh, EH9 3JZ, U.K.; ³Department of Chemistry, Faculty of Science, Nara Women's University, Kitauoya-Nishimachi, Nara 630-8506, Japan; ⁴Graduate School of Pharmaceutical Sciences, The University of Tokyo, 3-1, 7-Chome, Hongo, Bunkyo-ku, Tokyo 113-0033, Japan; ⁵School of Chemistry and Chemical Engineering, Queen's University, Belfast, BT9 5AG, U.K.

[#]Present address: TUV NEL Ltd, East Kilbride, Glasgow, G75 0QF, U.K.

^{##}Present address: School of Chemistry and Photon Science Institute, The University of Manchester, Alan Turing Building, Oxford Road, Manchester, M13 9PL, U.K.

Tables S1 and S2

Fig. S1

Temperature /°C	Lifetimes /ns			A-Factor			τ_{av}	χ^2
	τ_1	τ_2	τ_3	A ₁	A ₂	A ₃		
22.9	1.80	3.77	11.06	0.55	0.40	0.05	3.03	1.04
29.1	1.85	4.03	11.60	0.51	0.42	0.07	3.46	1.05
30.5	1.89	4.09	11.64	0.50	0.43	0.07	3.53	1.06
31.1	1.83	4.10	12.74	0.44	0.46	0.10	3.97	1.07
31.6	1.94	4.49	14.47	0.43	0.38	0.19	5.33	1.19
31.9	2.10	5.92	15.39	0.39	0.30	0.31	7.36	1.08
32.2	2.33	7.40	15.67	0.25	0.27	0.48	10.13	1.07
32.7	2.10	7.16	15.53	0.16	0.28	0.56	11.00	1.06
33.1	2.47	7.95	15.68	0.12	0.29	0.59	11.84	1.04
33.7	2.24	8.23	16.20	0.09	0.31	0.61	12.55	1.01
34.3	2.36	8.21	15.66	0.06	0.29	0.64	12.64	1.00
34.7	2.50	8.53	15.71	0.06	0.30	0.64	12.77	1.02
35.2	2.33	7.55	15.37	0.03	0.26	0.71	12.90	1.05
36.3	2.91	9.09	16.29	0.05	0.32	0.64	13.37	1.00
37.3	2.32	8.35	15.99	0.03	0.28	0.69	13.41	1.05
38.3	3.26	8.04	15.95	0.01	0.29	0.70	13.51	1.03

Table S1. Decay parameters for TCSPC data obtained by fitting a 3-exponential function to individual decay curves (i.e. not global analysis). The fluorescence decay curves were analysed by tail-fitting (i.e. without convolution with the instrument response function), from 1 ns after the peak of the decay to correspond to the conditions used to fit the FLIM decays. The average lifetimes were used to generate the calibration curve shown in Figure 4 .

Global lifetimes: $\tau_1 = 2.16$ ns $\tau_2 = 6.02$ ns $\tau_3 = 15.1$ ns

Global $\chi^2 = 1.026$

Temperature /°C	A ₁	A ₂	A ₃	Local χ^2
22.9	0.88	0.11	0.01	1.00
29.1	0.82	0.17	0.01	1.04
30.5	0.81	0.18	0.01	1.04
31.1	0.77	0.20	0.03	1.07
31.6	0.70	0.20	0.10	1.12
31.9	0.57	0.22	0.21	1.05
32.2	0.34	0.23	0.43	1.04
32.7	0.24	0.25	0.51	1.04
33.1	0.14	0.27	0.59	1.04
33.7	0.14	0.19	0.67	1.06
34.3	0.03	0.28	0.69	1.02
34.7	0.01	0.29	0.70	1.04
35.2	0.00	0.30	0.70	1.06
36.3	0.05	0.19	0.76	1.05
37.3	0.03	0.19	0.78	1.05
38.3	0.02	0.20	0.78	1.04

Table S2. Decay parameters for TCSPC data obtained from global analysis of the decays. Decays were analysed by iterative reconvolution with the instrument response function to obtain precise decay parameters for the photophysical interpretation of the temperature dependence. These are the decay parameters presented in Figure 3.

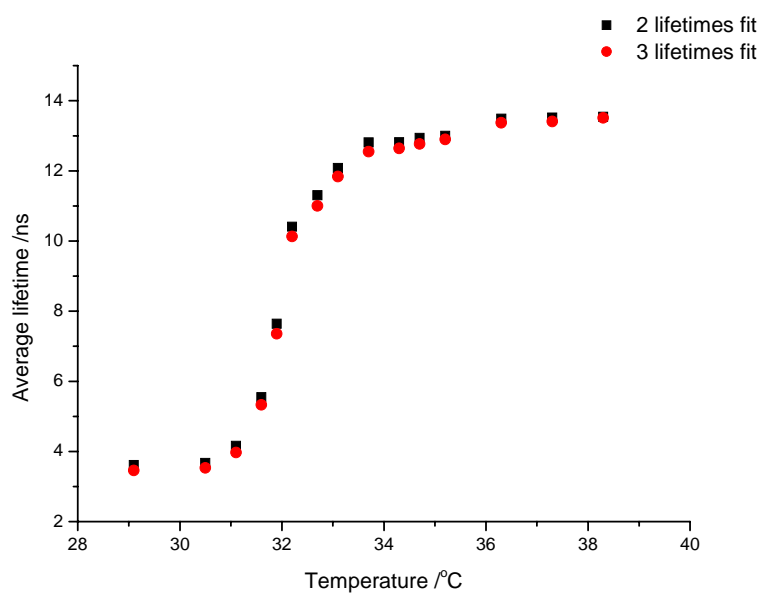


Figure S1: The average lifetimes determined by fitting two or three lifetimes to individual TCSPC decay curves, plotted as a function of temperature.