Application of "click" chemistry to the production of DNA microarrays

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Contents:

(1)	General remarks	1
(2)	^{1}H ^{1}S ^{2}S ^{1}NMR and FT-IR spectra for 1	2
(3)	¹ H ¹³ C ³¹ P NMR spectra for 2	7
(4)	Image microarrays after printing	
(5)	Image CPG from fluorescent microscope	
(6)	Analysis of oligonucleotides by capillary el	ectrophoresis and MALDI-TOF analysis13
(7)	Calculated the level of loading of CPG with	1 functional groups20
(8)	Microscope image of the CPG particles with	h length bar21

(1) General remarks:

All reagents (analytical grade) were obtained from commercial suppliers and used without further purification. Anhydrous benzene, toluene and CH₂Cl₂ were freshly distilled from P₂O₅ and CaH₂, respectively. All other anhydrous solvents and liquid reagents were dried through storage over activated 4Å (EtOH, Et₃N, pyridine,) or 3Å (MeOH, MeCN) molecular sieves.

Progress of reaction was monitored by Thin Layer Chromatography and ³¹P NMR spectra while purification was effected by column chromatography, using silica gel (60-120 mesh).

¹H, ¹³C, ²⁹Si NMR. The NMR spectra were recorded at 298 K on a Bruker Advance DRX 400 spectrometer operating at frequencies 400.13201 MHz (¹H), 100.62281 MHz (¹³C) and 79.45750 MHz (²⁹Si). ³¹P NMR spectra were recorded at on Varian Unity 300 spectrometer operating at frequencies 121 MHz; 5% H₃PO₄ in D₂O as an external

In all spectra for $1\ \text{CDCl}_3$ was used and for $2\ \text{DMSO}$ was used as the solvent.

FT-IR spectra were recorded on a Bruker Tensor 27 Fourier transform spectrometer equipped with a SPECAC Golden Gate diamond ATR unit. In all cases, 16 scans at a resolution of 2 cm^{-1} at RT were collected for the spectrum.

Mass spectra of analyzed compound 1 was collected on Varian 4000 GC/MS chromatographic system equipped with Varian VF Factor-Four – 5ms, 30 m long capillary column, split/splitlles type injection chamber, MS (ion trap) detector and autosampler. Mass spectra was recorded in a range 50 to 1000 m/z and electron impact (EI) ionization technique was used.

¹H NMR of compound **1**



2

Fig ¹³C NMR of compound 1



3

Fig ²⁹Si NMR of compound **1**



Fig FT-IR of compound 1



Fig GC-MS of compound 1



Electronic Supplementary Material (ESI) for Lab on a Chip This journal is C The Royal Society of Chemistry 2012

Fig ¹H NMR of compound 2



Fig $^{13}\mathrm{C}$ NMR of compound 2



Fig uncoupled ³¹P NMR a of compound 2



Fig 31 P NMR couple with hydrogen of compound 2



Image microarrays after printing





Fig 0.2% concentration of azidofunctional silane



Image CPG from fluorescent microscope

Fig . Effect of treatment azide-functionalized CPG with ODN-3 and labeled oligomer MIX-F The green fluorescence was visualized by fluorescent microscope







Analysis of oligonucleotides by capillary electrophoresis and MALDI-TOF analysis

ODN-1



MS MALDI calculated mass 6661,467; found m/z M- 1457,775 m/z; 1479,794 [M+Na] m/z; 1495,774 m/z [M+K]

ODN-2



MS MALDI calculated mass 1605; found m/z 1605,476 m/z; 1627,501 m/z [M+Na]+; 1643,543 m/z [M+K]+

ODN-3



MS MALDI calculated mass 3182,2; found m/z 3182,081 m/z;





MS MALDI calculated mass 545.4; found m/z 545,189 m/z;

ODN-5

capillary electrophoresis



MALDI-TOF MS



MS MALDI calculated mass 6661,467; found m/z 6661,6172 m/z

ODN-6



MS MALDI calculated mass 6972.959; found m/z 6973.708 m/z;

ODN-7



MALDI-TOF MS



MS MALDI calculated mass 8820,717; found m/z 8820,041 m/z

OND - 8



MS MALDI calculated mass 7511,3; found m/z 7511,056 m/z;

Calculated the level of loading of CPG with functional groups

 Table 1. The rate of loading calculated on the basis of UV measurements at 504 nm

 wavelength.

Support	Mass [mg]	1 attach	2 attach	Absolute loading [µmol/g]	Background-corrected loading [µmol/g]
1	6.00	1.36	1.24	32.48	27.21
2	7.84	1.73	1.57	31.45	27.42
3	7.22	1.63	1.43	32.33	27.96
blank	7.58	0.22	0.25	4.16	-
Average loading of support			ort	31.96	27.31

Table 2. The rate of loading calculated on the basis of UV measurements.

Probe	Mass [mg]	Reaction time [h]	Absorbance	Loading of support [µmol/g]
Ι	2.45	1	0.546	31.837
II	3.49	2	0.753	30.823
III	5.93	12	1.247	30.041

Microscope image of the CPG particles with length bar



