

Electronic Supplementary Information (ESI) for Lab on a Chip

Microfluidic Sensing Devices Employing *In-Situ*-Formed Liquid Crystal Thin Film for Detection of Biochemical Interactions

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Supplementary Methods

Materials

Liquid crystal, 5CB and 1-decanethiol [$\text{CH}_3(\text{C}_9\text{H}_{18})\text{SH}$], 96%, L- α -dilauroyl phosphatidylcholine (L-DLPC), N,N-dimethyl-N-octadecyl-3-amino-propyltrimethoxysilyl chloride (DMOAP) and phospholipase A₂ (PLA₂) from *Naja mossaambica mossaambica* were purchased from Sigma-Aldrich (St. Louis, MO, USA). N-hexadecyl mercaptan [$\text{CH}_3(\text{C}_{15}\text{H}_{30})\text{SH}$], 92% was purchased from Acros Organics (Geel, Belgium). Glass slides were purchased from Fisher Scientific (Pittsburgh, PA, USA). Microfab NI 100 make-up solution and Microfab NI 100 wetting agent were purchased from Enthone-OMI (West Haven, CT, USA). High purity Ni gauze, #39704, was purchased from Alfa Aesar (Ward Hill, MA, USA). Positive photoresist (PR), AZ P4620, was purchased from Clariant Corporation (Somerville, NJ, USA). Double adhesive spacer (250- μm thick) was acquired from 3M Co. (St. Paul, MN, USA). Ethyl vinyl acetate microbore tubings were purchased from Cole Parmer Co. (Vernon Hills, IL, USA).

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Acquisition and analysis of microscopic images

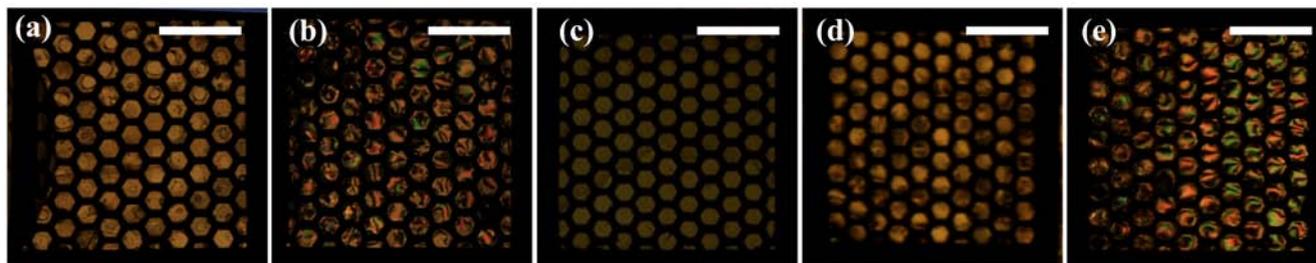
All schematics [Fig. 1(a) (c), Fig. 2, Fig. 8(d) (e), Fig. 9(b), Fig. 10(d)] are created by Microsoft Visio[®]. All simulation result plots [Fig. 3, Fig. 4, Fig. 5] are exported from ANSYS Fluent[®] 12.1. Fig. 1(b) is taken by a digital optical camera: Konica Minolta model 13655. Other figures [Fig. 6, Fig. 7, Fig. 8(a) (b) (c), Fig. 9(a), Fig. 10(a) (b) (c), Fig. 11] are taken by a polarized optical microscopy (POM), which consists of a stereoscope (Nikon, SMZ1500) and two crossed polarizers. The Gamma values of these images are increased by 22% uniformly for better visualization, by an image processing tool (ACDSee Pro2[®]).

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Repeatability test

Fig. S1 depicts the process of the repeatability test. Fig. S1(a) shows the initial state that the channel is pre-filled with the LC. Next, the LC is “cut” by the DTAB solutions [Fig. S1(b)]. Then the sensing channel was purged using ethanol so that DTAB solution as well as the LC thin film was removed from the channel [Fig. S2(c)]. N₂ stream was then applied to dry the channel for 30 min. LC was again introduced into the channel and filled the hexagonal Ni container at the bottom [Fig. S1(d)]. DTAB solution was introduced again into the channel and for the second time formed the LC thin film [Fig. S1(e)]. Comparing Fig. S1(b) and Fig. S1(e), the quality of the LC thin film is almost uniform.

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35 Figure. S1. Repeatability test of the LC thin film device.